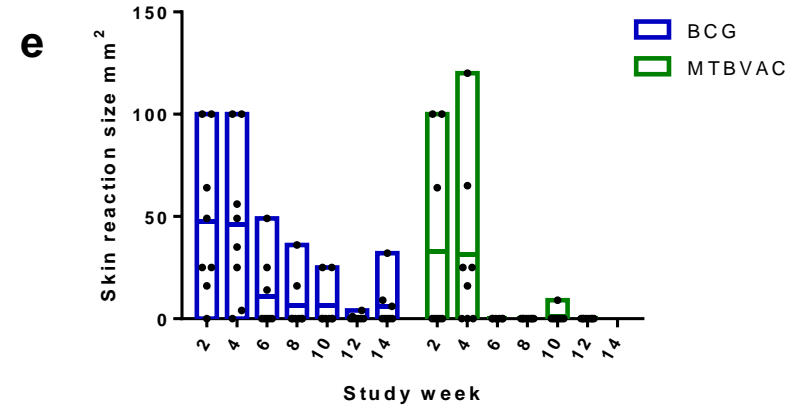
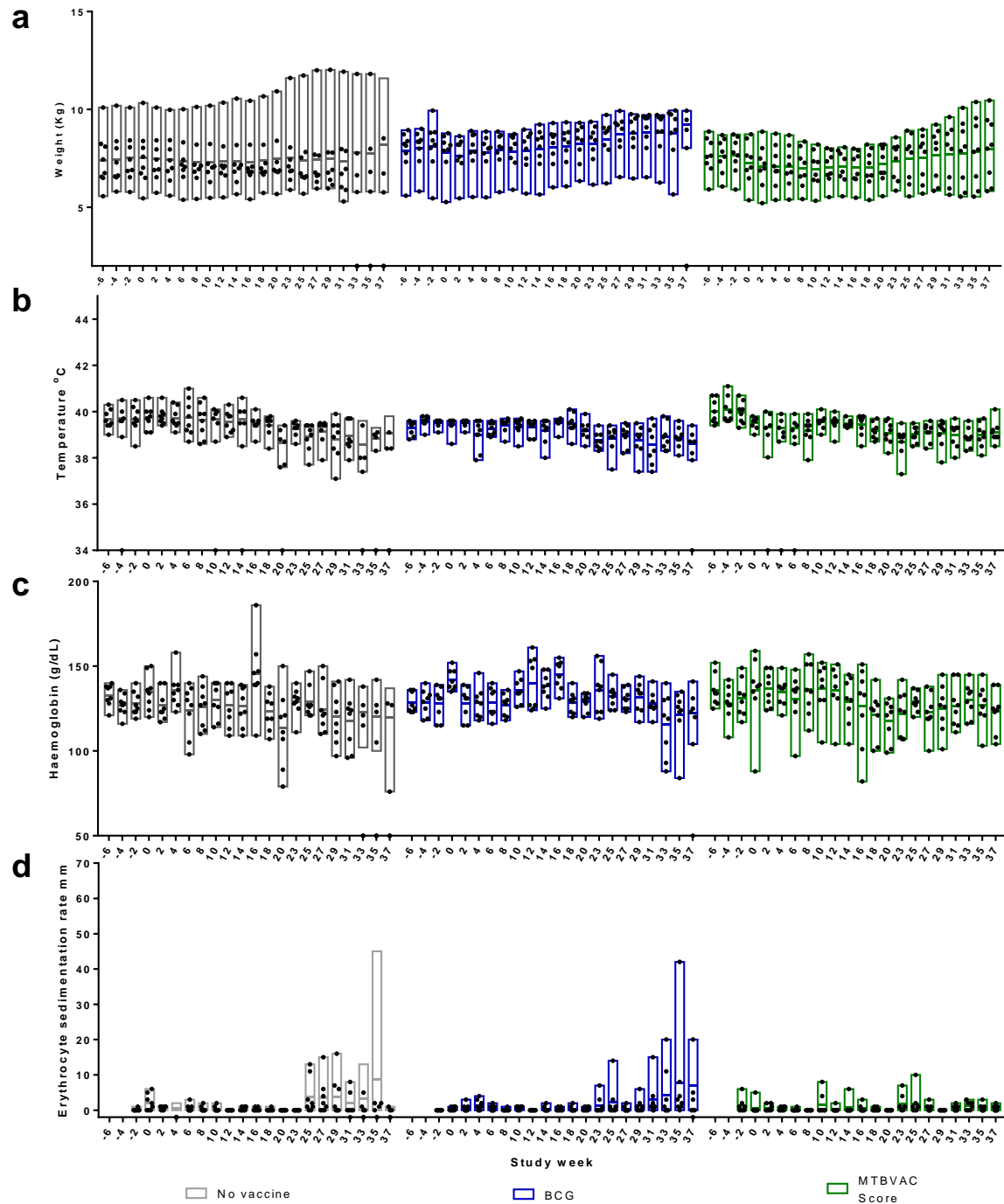


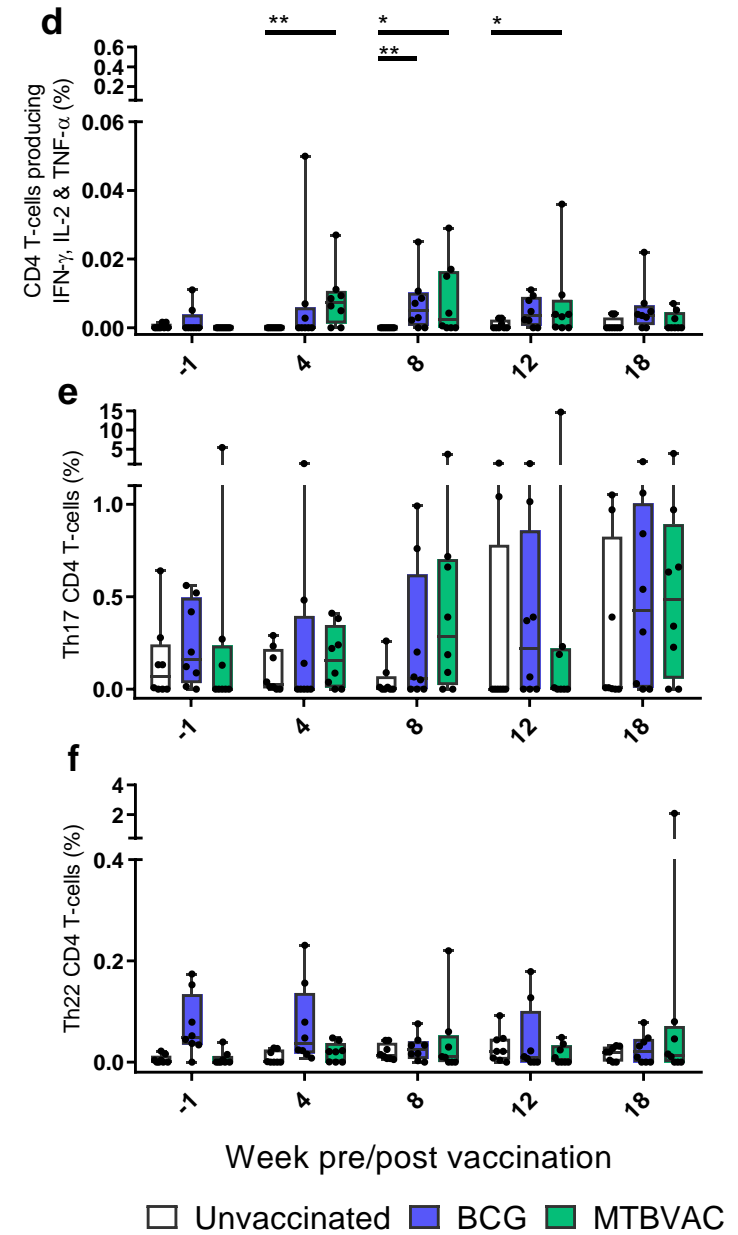
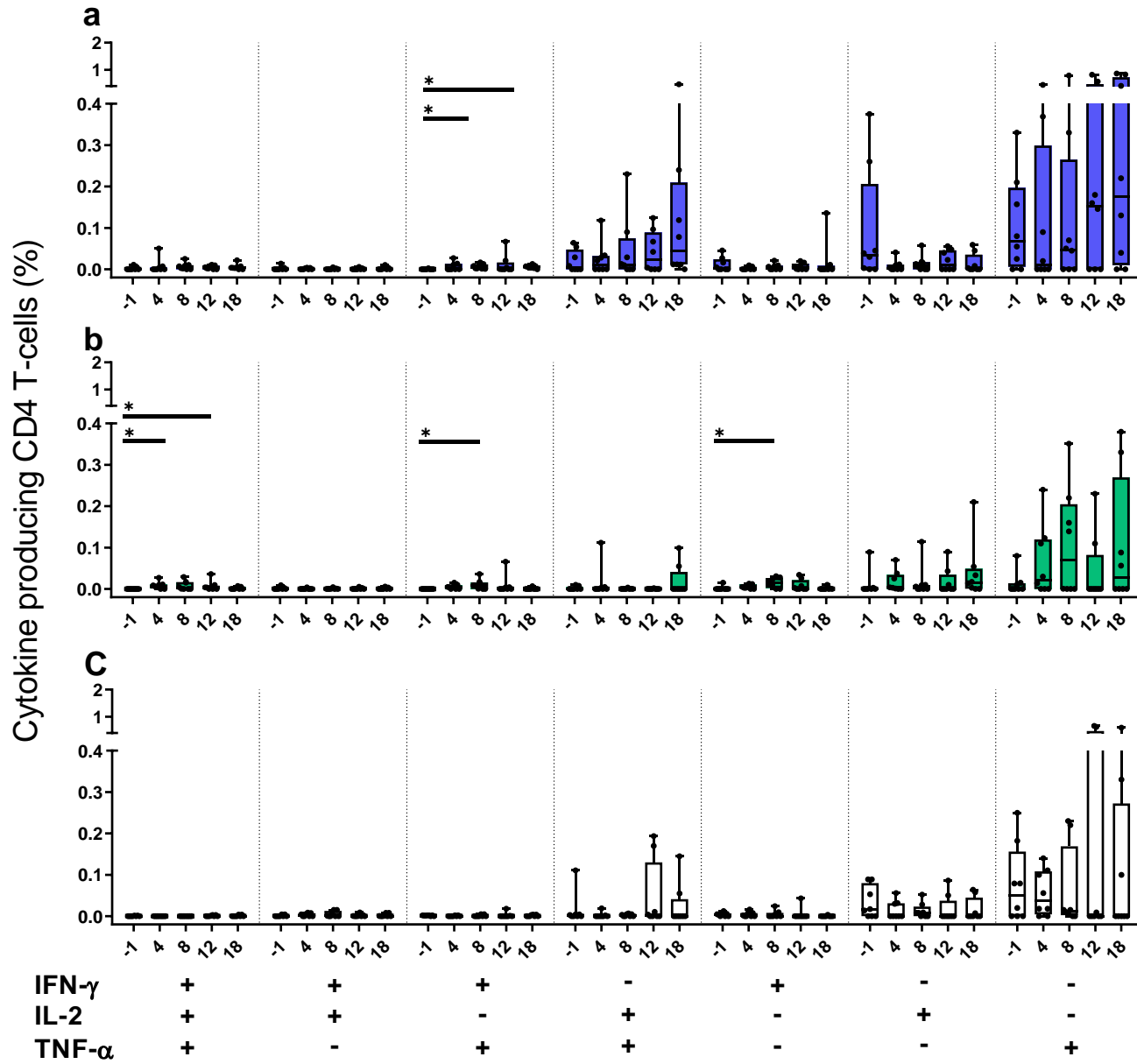
Supplementary Table 1

Average number of acid fast bacilli per granuloma of each developmental stage in lung and hilar lymph node counted in Ziehl-Neelsen stained tissue sections.

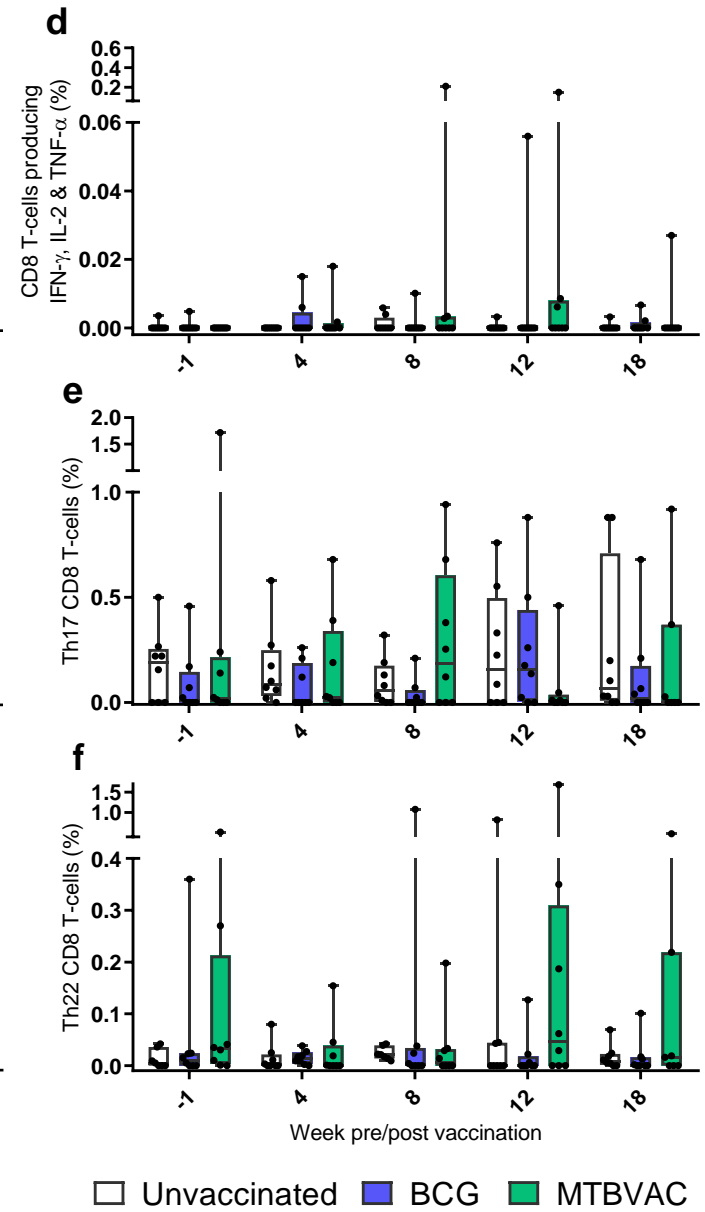
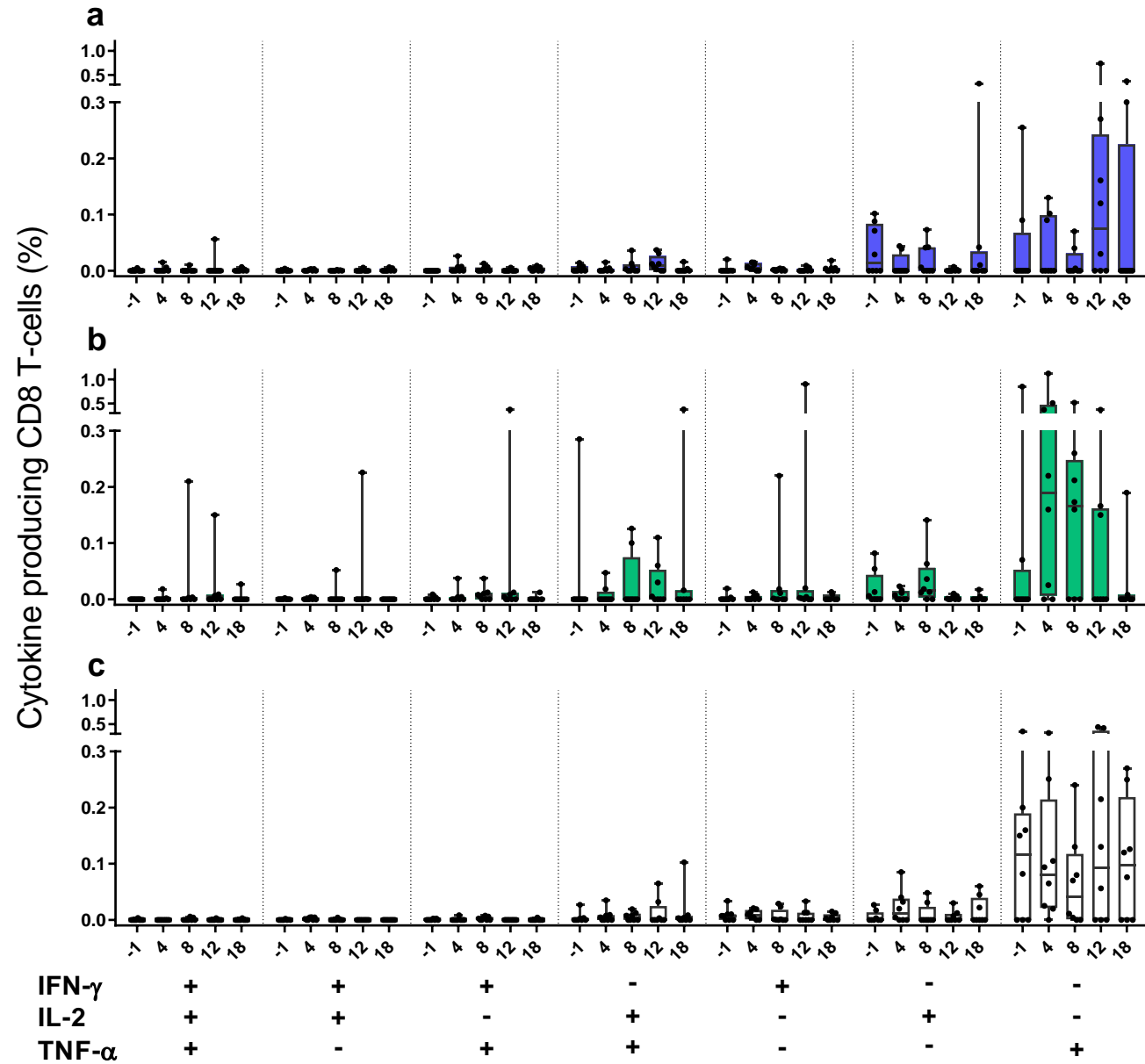
Group	Mean number of acid fast bacilli per granuloma					
	Granuloma stage					
	Type I	Type II	Type III	Type IV	Type V	Type VI
BCG	0	0	0	0	0.106	8.167
MTBVAC	0	0	0	0	0	2.214
No vaccine	0	0	0	0.009	0.400	16.577



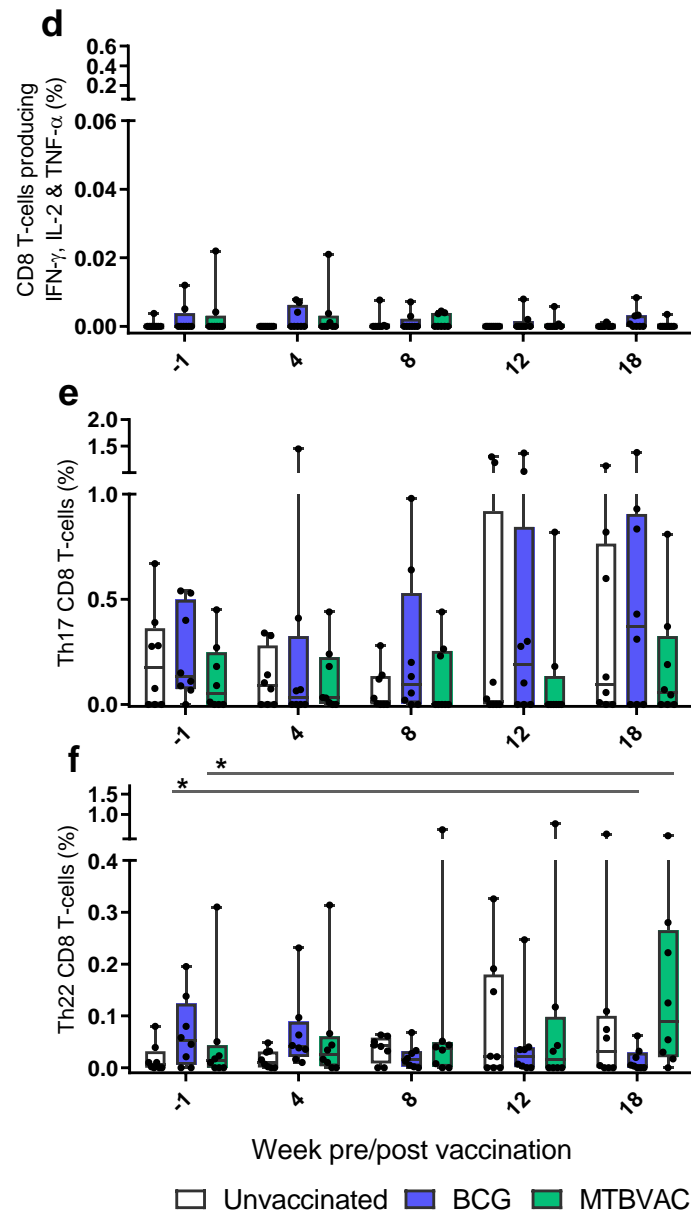
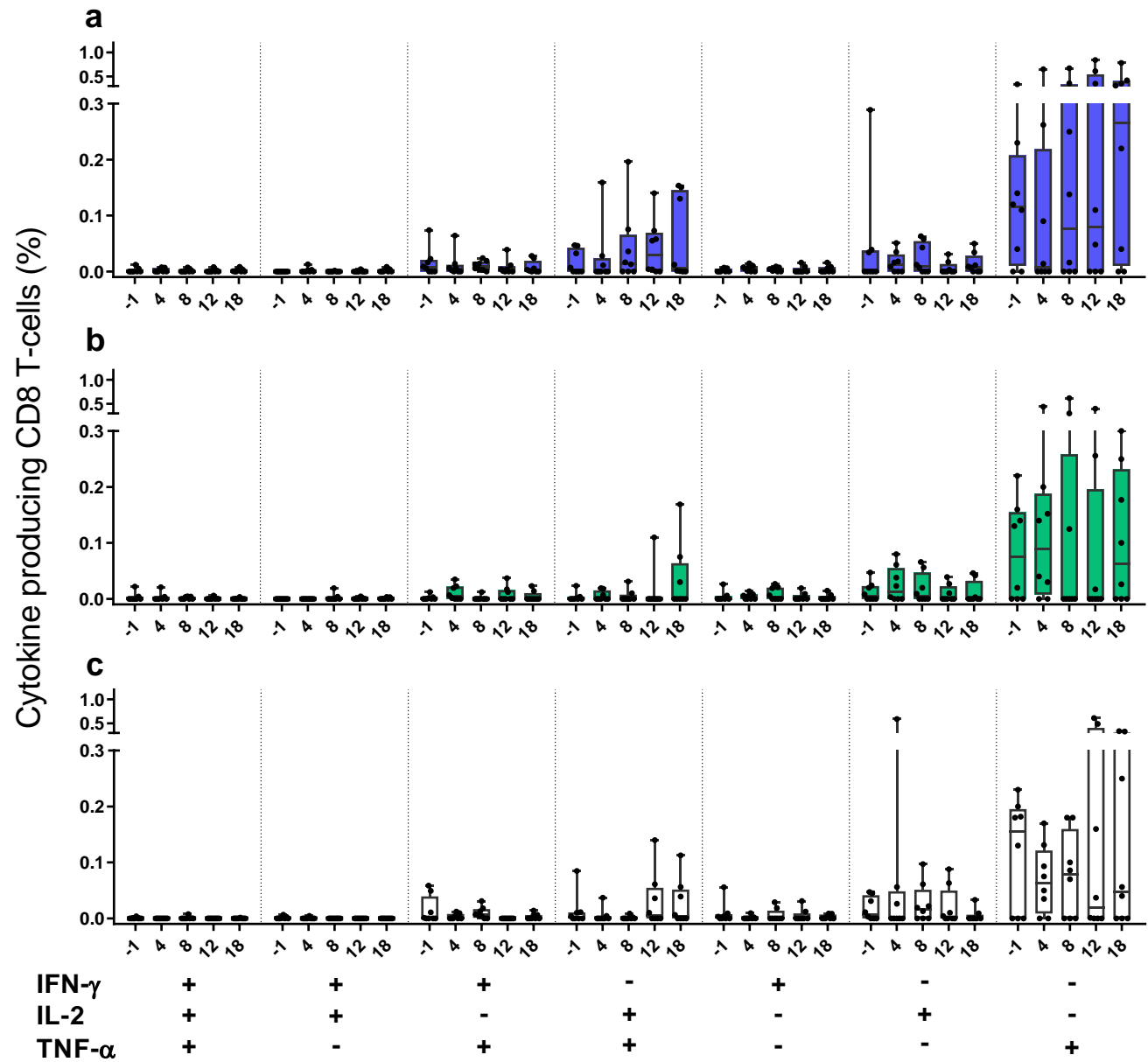
Supplementary Figure 1: Clinical parameters measured throughout the vaccination and challenge phases of the study. A: body weight; B body temperature; C: red blood cell haemoglobin concentration; D: erythrocyte sedimentation rate; E: skin reaction size at vaccination site. Box plots show the mean result for the vaccination group with minimum and maximum values, symbols represent data measured from individual subjects.



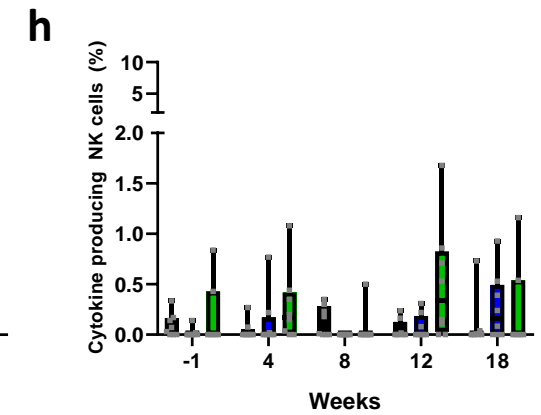
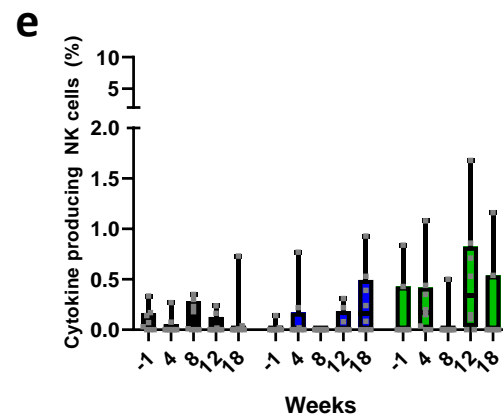
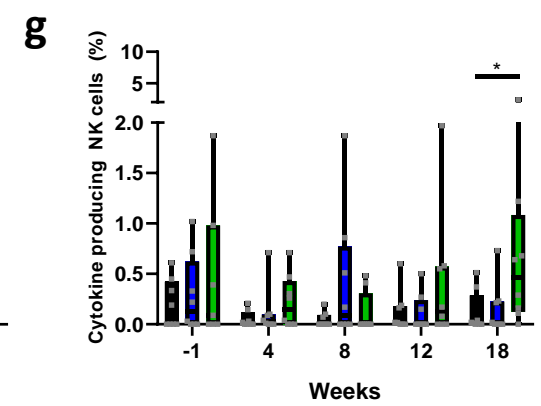
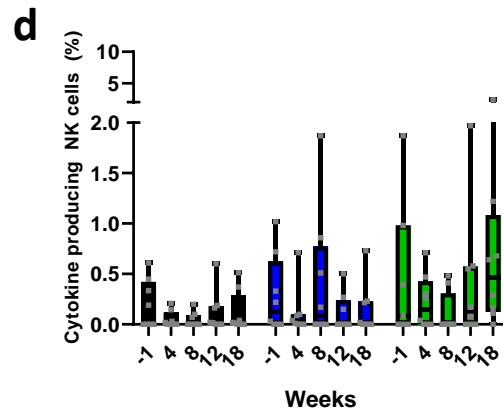
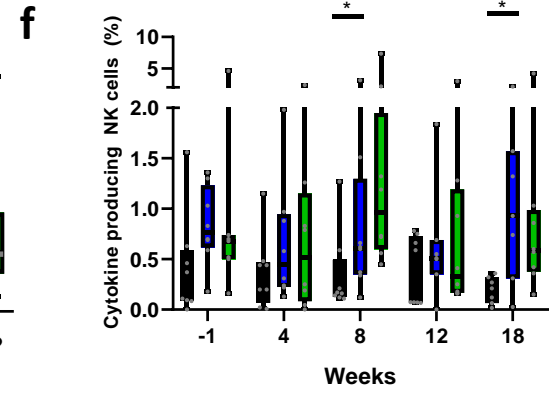
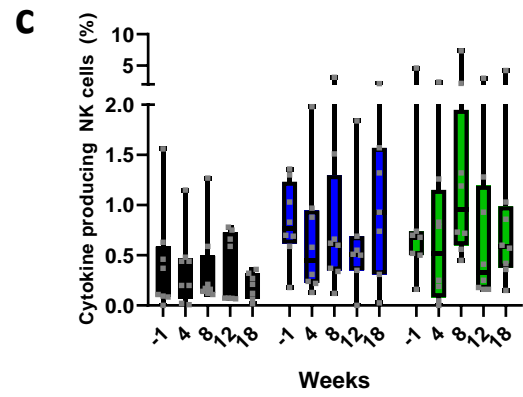
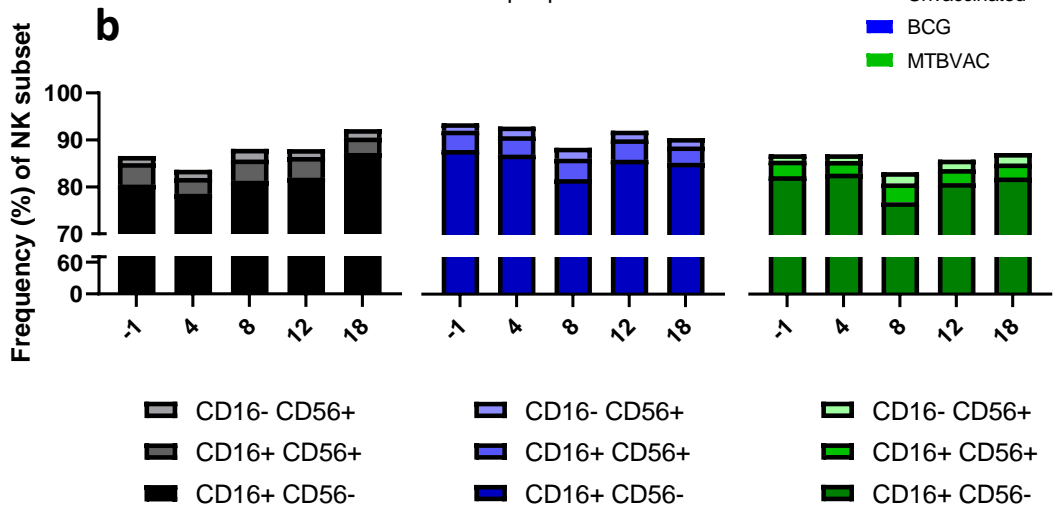
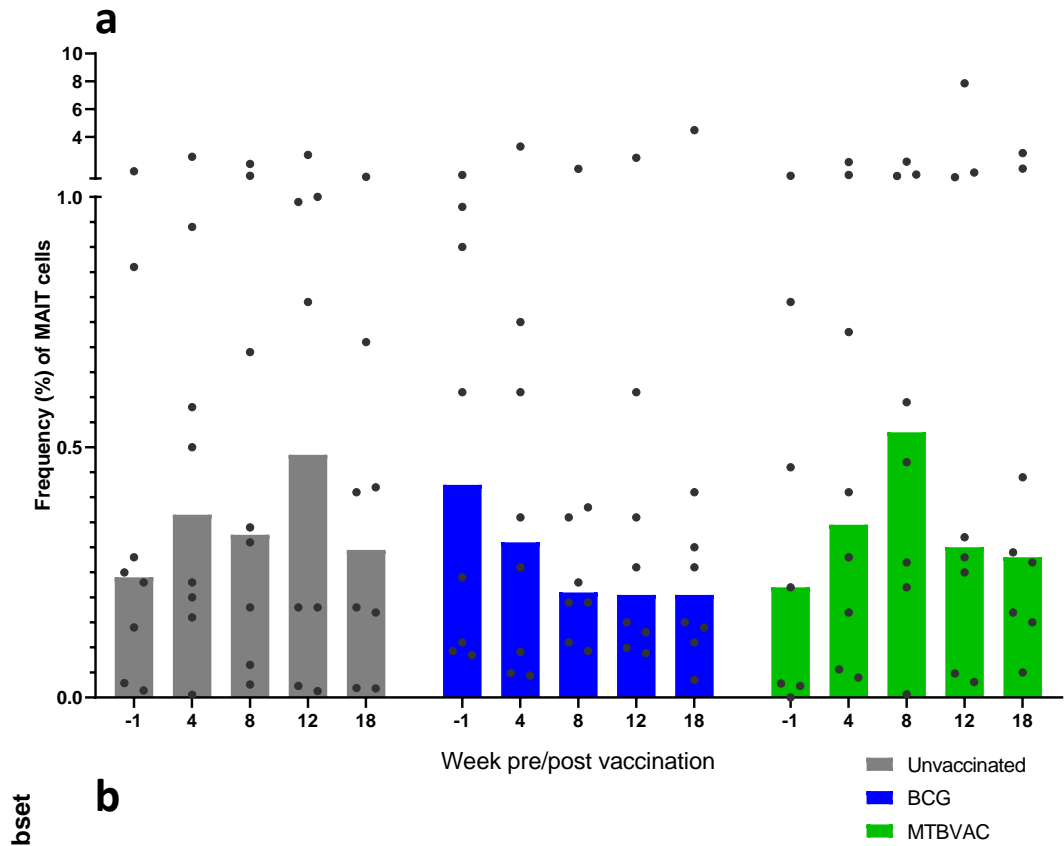
Supplementary Figure 2. MTBVAC-specific CD4 T-cell cytokine secretion profiles measured by whole blood intracellular cytokine staining. Box plots show the vaccination group median frequency of CD4 T-cells producing combinations of the cytokines IFN- γ , IL-2 and TNF- α (A, B, C, D); IL-17 (E) or IL-22 (F) +/- IQR with minimum and maximum values indicated by box whiskers. Cytokine production was measured at specific weeks prior to (-1) and following (4, 8, 12 and 18) BCG or MTBVAC vaccination. Significant differences between pre- and post-vaccination values (Wilcoxon signed-rank) and between groups (Mann-Whitney U-test) are indicated by bars and asterisks: * $p \leq 0.05$, ** $p \leq 0.01$, *** $p \leq 0.001$ (all p-values are unadjusted for multiple comparisons). Frequencies measured in individual animals are represented by dots.



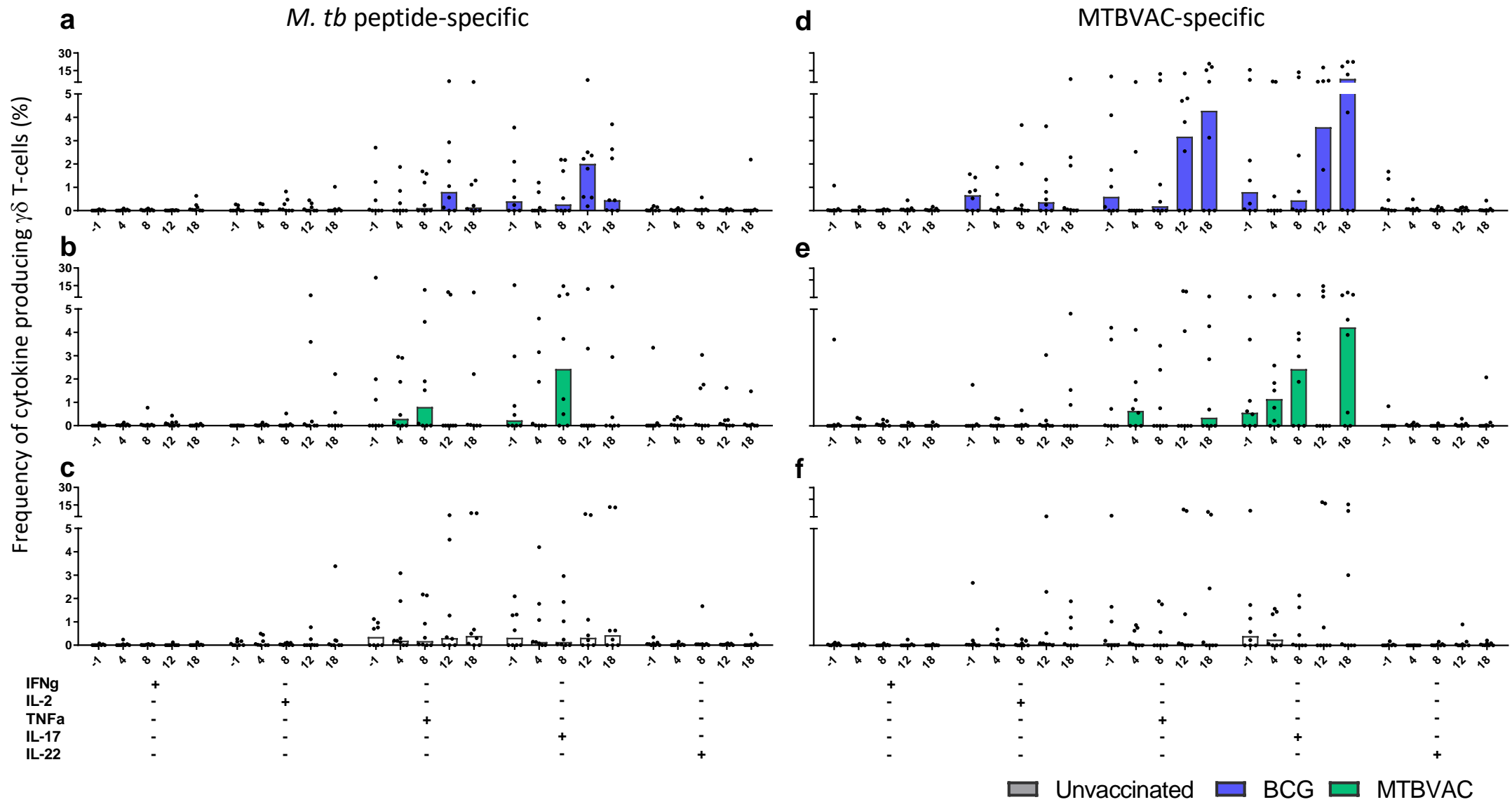
Supplementary Figure 3. *M. tuberculosis* peptide-specific CD8 T-cell cytokine secretion profiles measured by whole blood intracellular cytokine staining. Box plots show the vaccination group median frequency of CD4 T-cells producing combinations of the cytokines IFN- γ , IL-2 and TNF- α (A, B, C, D); IL-17 (E) or IL-22 (F) +/- IQR with minimum and maximum values indicated by box whiskers. Cytokine production was measured at specific weeks prior to (-1) and following (4, 8, 12 and 18) BCG or MTBVAC vaccination. Frequencies measured in individual animals are represented by dots.



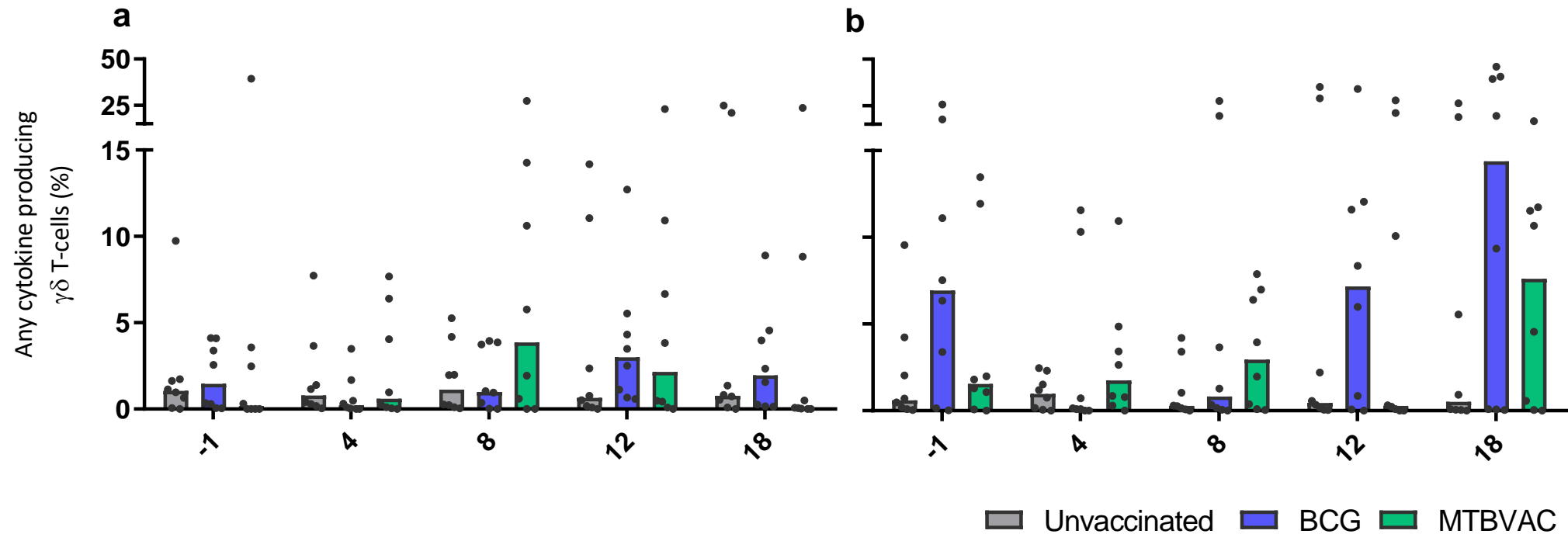
Supplementary Figure 4. MTBVAC-specific CD8 T-cell cytokine secretion profiles measured by whole blood intracellular cytokine staining. Box plots show the vaccination group median frequency of CD4 T-cells producing combinations of the cytokines IFN- γ , IL-2 and TNF- α (A, B, C, D); IL-17 (E) or IL-22 (F) +/- IQR with minimum and maximum values indicated by box whiskers. Cytokine production was measured at specific weeks prior to (-1) and following (4, 8, 12 and 18) BCG or MTBVAC vaccination. Significant differences between pre- and post-vaccination values (Wilcoxon signed-rank) are indicated by bars and asterisks: * $p \leq 0.05$. Frequencies measured in individual animals are represented by dots.



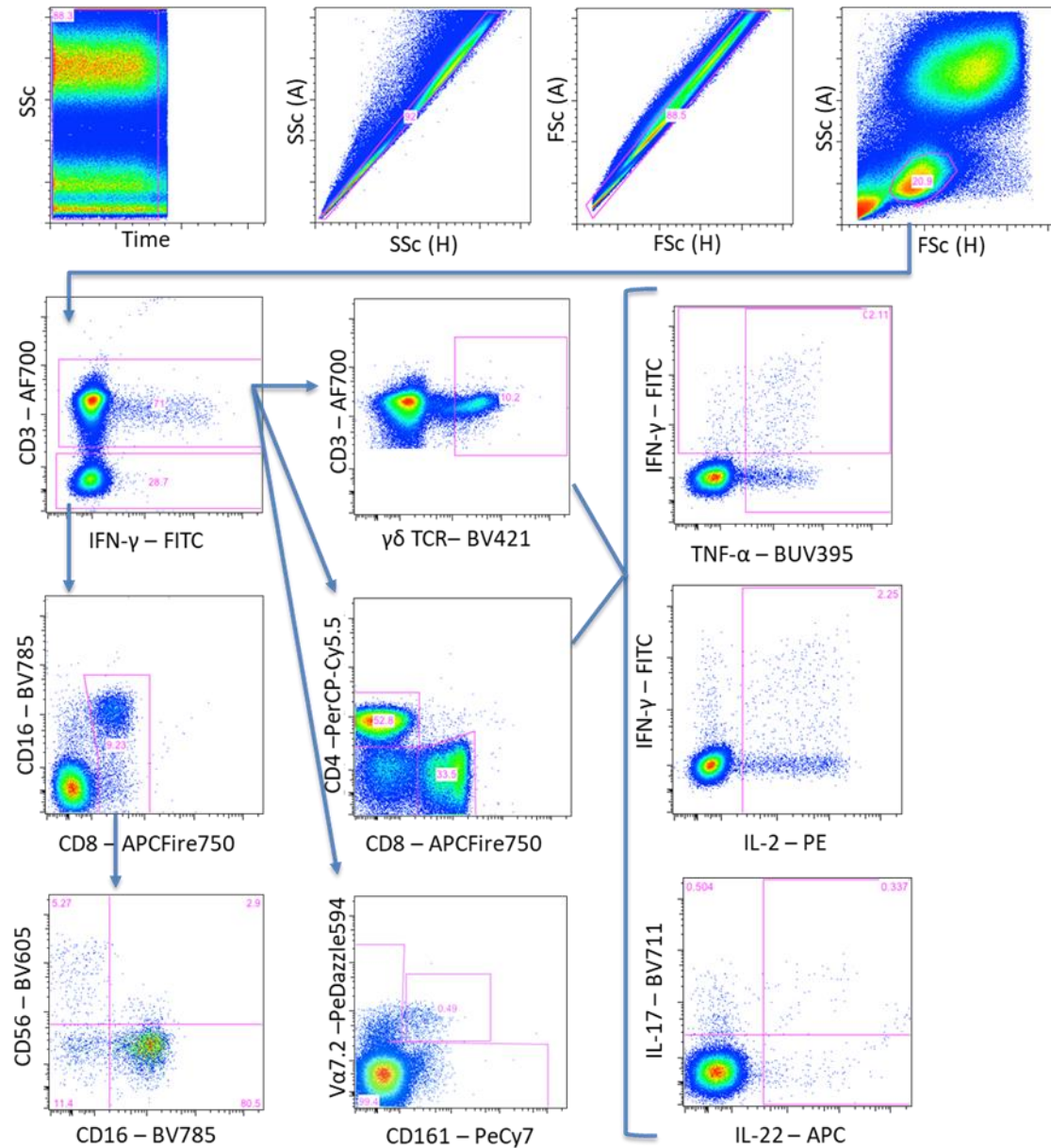
Supplementary figure 5. Frequency of innate lymphoid cell populations. The frequency of mucosal associated invariant T-cells (MAIT) (A) and Natural Killer cell (B) populations were measured by WB ICS assay prior to (-1) and at weeks four, eight, 12, and 18 after BCG or MTBVAC vaccination. Figure a) bar charts show the vaccination group median percentage of MAIT cells as a proportion of the CD3+ T-cell population, frequencies measured in individual animals are shown as dots. Figure B) stacked bars show the group median percentages of each NK subset as a proportion of the CD3-, CD8+ lymphocyte population. Figures C – H) frequency of NK cells producing IFN- γ , IL-2, TNF- α , IL-17 or IL-22 in response to stimulation with *M. tuberculosis* peptides (E, H) or MTBVAC (D, G) or without antigen or peptide stimulation (C, F). Note that plots C-E show the kinetics of NK cell cytokine production in each vaccination group, whereas plots F-H show the same data aligned by time for comparative purposes. Cytokine production was measured at specific weeks prior to (-1) and following (4, 8, 12 and 18) BCG or MTBVAC vaccination. Box plots show the vaccination group median +/- IQR with minimum and maximum values indicated by box whiskers. Frequencies measured in individual animals are represented by dots. Significant differences between vaccination groups are indicated by bars with asterisks * $p \leq 0.05$, ** $p \leq 0.01$ (all p-values are unadjusted for multiple comparisons).



Supplementary Figure 6. Cytokine production from $\gamma\delta$ T-cells. Bars show the vaccination group median frequency of $\gamma\delta$ T-cells producing IFN- γ , IL-2, TNF- α , IL-17 or IL-22 in response to stimulation with *M. tuberculosis* peptides (A, B, C) or MTBVAC (D, E, F). Cytokine production was measured at specific weeks prior to (-1) and following (4, 8, 12 and 18) BCG or MTBVAC vaccination. Frequencies measured in individual animals are represented by dots. Y-axis scales are identical in all plots.



Supplementary Figure 7. Cytokine production from $\gamma\delta$ T-cells (summed). Bars show the vaccination group median summed frequency of $\gamma\delta$ T-cells producing IFN- γ , IL-2, TNF- α , IL-17 or IL-22 in response to stimulation with *M. tuberculosis* peptides (A) or MTBVAC (B). Cytokine production was measured at specific weeks prior to (-1) and following (4, 8, 12 and 18) BCG or MTBVAC vaccination. Frequencies measured in individual animals are represented by dots. Y-axis scales are identical in all plots.



Supplementary Figure 8. Flow cytometric gating strategy. Sequential gating used for cell population analysis described in manuscript figure 4 and supplementary figures 3 – 8.