

1 **Supplementary Information for**

2 Genomic Consequences of Long-term Population Decline in Brown Eared Pheasant

3
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29 **Supplementary Information Text**

30

31 **DNA sample collection**

32 Tissue for *de novo* whole genome sequencing was obtained from a female Brown eared
33 pheasant (*Crossoptilon mantchuricum*) from the central population in the Pangquangou
34 National Natural Reserve, Jiaocheng, Shanxi Province, China. For genome re-
35 sequencing, tissue samples were obtained from 11, 18, and 11 individuals from Shanxi
36 (Central population, Brown-C), Shaanxi (Western population, Brown-W), Hebei, and
37 Beijing (Eastern population, Brown-E), respectively. The blue eared pheasant (*C.*
38 *auritum*) was used as the control group in this study, of which 11 individuals were
39 sampled for re-sequencing (the geographic distribution of the studied populations can
40 be seen in Fig. 1a and S12. Sampling information can be seen in Dataset S9). All
41 samples were stored at -80 °C at Beijing Normal University. For genome annotation,
42 we also sequenced the transcriptome of the *C. mantchuricum* individual used for the *de*
43 *nov*o genome assembly using blood samples, developing tail feathers, and primaries.
44 The tissues used to isolate the RNA were mixed with an RNA preservation solution
45 (RNA-In-Safe; Sangon Biotech, Beijing, China). All tissues used for transcriptome
46 sequencing were stored in liquid nitrogen. Sample collection was performed by an
47 accredited veterinarian, and all sampling procedures were approved by the Forestry
48 Department of China and the Ethics and Animal Welfare Committee at the College of
49 Life Science, Beijing Normal University, China (Approval Number CLS-EAW-2015-
50 012).

51

52 **DNA, RNA isolation, sequencing, and data filtering**

53 DNA and RNA isolation and library construction were performed at the Novogene
54 Sequence Center (Beijing, China). DNA and RNA quality and quantity were evaluated
55 using a Nanodrop spectrophotometer, a Qubit Fluorometer, an Agilent 2100, and gel
56 electrophoresis. Fragment libraries (insert length 250 bp) and jumping libraries (insert
57 lengths 2000 bp and 5000 bp) were constructed for *de novo* genome sequencing. For *de*

58 *nov*o genome sequencing, the targeted total amount of sequences from the fragment
59 libraries and jumping libraries was ~150 Gb (~150×) and ~100 Gb (~100×),
60 respectively. A total of 51 fragment libraries (insert length 350 bp) were constructed for
61 genome re-sequencing, targeting more than 15× coverage per individual. The targeted
62 total amount of transcriptome sequences was ~190 Gb from the RNA libraries (insert
63 length 250 bp). All libraries were sequenced on the Illumina HiSeq platform at the
64 paired-end 150 bp. Adapter content was trimmed and five base pairs were removed
65 from both the 5' and 3' ends of each read in Trim Galore v0.4.2, with other default
66 parameters (http://www.bioinformatics.babraham.ac.uk/projects/trim_galore/). FastQC
67 v0.11.5 (<http://www.bioinformatics.babraham.ac.uk/projects/fastqc/>) was used to
68 assess the quality of the raw sequencing data and clean data.

69

70 **Genome assembly and evaluation**

71 After removing the low-quality data, 220.6 Gb of DNA data (1.37 billion reads, 195.6-
72 fold sequence coverage) were used for the *de novo* genome assembly of *C.*
73 *mantchuricum* in ALLPATHS-LG v52488 (Butler, et al. 2008). Basic metrics (e.g.,
74 scaffold and contig N50) were calculated from the genome assembly with a minimum
75 contig length of 100 bp. We used BUSCO v3 (Simão, et al. 2015) to evaluate the
76 completeness of the draft genome by searching for 4915 Benchmarking Universal
77 Single-Copy Orthologs from 40 avian lineages (aves_odb9 dataset) in the *C.*
78 *mantchuricum* assembly.

79

80 **Genome annotation**

81 The *de novo* repetitive elements annotation was performed in RepeatModeler-open
82 v4.0.6 (<http://www.repeatmasker.org/>) with repeat masker libraries v20160829 (Bao,
83 et al. 2015) and repeat finding programs (RECON v1.08
84 (<http://selab.janelia.org/recon.html>) and RepeatScout v1.0.5
85 (<http://repeatscout.bioprotects.org/>)). Repetitive elements were identified with
86 RepeatMasker-open v4.0.6, in which repetitive elements were searched against *de novo*

87 repetitive elements and homologous sequences (Dfam 2.0 database (Hubley, et al. 2015)
88 and RepBase database v20160829 (Bao, et al. 2015)). Tandem repeats were identified
89 across the genome using Tandem Repeats Finder v404 (Benson 1999). The repetitive
90 elements were masked in the whole genome for annotation purposes.

91

92 We employed an integrative approach to identify a protein-coding gene set of the
93 repeated-masked genome in MAKER v2.31.8 ([http://www.yandell-](http://www.yandell-lab.org/software/maker.html)
94 [lab.org/software/maker.html](http://www.yandell-lab.org/software/maker.html)), which used homologous genes, *ab initio* predictions,
95 expressed sequence tags (ESTs), and RNA sequence evidence. The homologous gene
96 sets were from human, zebra finch, chicken, and turkey protein sequences downloaded
97 from Ensembl genome browser 87 (<http://useast.ensembl.org/info/data/ftp/index.html>).

98 The *ab initio* prediction was performed in both SNAP (Korf 2004) and AUGUSTUS
99 v3.0.3 (Stanke and Waack 2003). The 80.0-Gb clean RNA sequences were used to
100 produce splice junction information of the repeat-masked draft genome in TopHat
101 v2.0.13 (<http://ccb.jhu.edu/software/tophat/index.shtml>) and to *de novo* assemble
102 mRNA-sequences in Trinity v2.3.2 (Grabherr, et al. 2011). To improve the accuracy of
103 the annotation procedure, we ran MAKER twice. For the initial MAKER run, both the
104 AUGUSTUS chicken models and SNAP chicken models that we trained were used to
105 predict the coding genes. The assembled chromosomes and annotation file of
106 *Gallus_gallus*-5.0 (Consortium 2004) were used to train the SNAP chicken models.
107 After the initial MAKER run, we used the gene models produced during the initial
108 MAKER run to retrain AUGUSTUS and SNAP. For the final MAKER run, we used the
109 retrained gene predictors to obtain a consensus set of protein-coding genes. In both
110 MAKER runs, the minimum length required for single exon ESTs was set to 250 bp.
111 The gene set preserved genes that encoded 20 or more amino acids (Eilbeck, et al. 2009).
112 To identify protein function, Interproscan-5.27-66.0 (Zdobnov and Apweiler 2001) was
113 employed to analyze proteins from MAKER with known functional domains and GO
114 terms from the Pfam database. Then, the genes were filtered to remove those without
115 domain content support or those with a quality metric (Annotation Edit Distance, AED)

116 greater than or equal to 1 (Eilbeck, et al. 2009).

117

118 To assess annotation quality, two summary statistics were considered: the percentage
119 of the genes with an AED value less than 0.5, and the percentage of the genes with
120 recognizable domain content. The gene set from annotation was also assessed using
121 BUSCO v3 (Simão, et al. 2015), which was performed on the protein sets from
122 MAKER with 4915 Benchmarking Universal Single-Copy Orthologs from 40 avian
123 lineages (aves_odb9 dataset).

124

125 **Read mapping**

126 Cleaned reads in all the fragment libraries for each individual were mapped to the *C.*
127 *mantchuricum* reference genome produced in this project and a complete mitochondrial
128 genome that was downloaded from the GenBank database (accession number:
129 KP259807) with a Burrows–Wheeler alignment-maximal exact matches (BWA-MEM,
130 v0.7.9a) algorithm (Li and Durbin 2009). Samtools v1.3.1 was used to calculate the per-
131 base sequence depth, and was then used to calculate the mean coverage for each
132 scaffold (Li, et al. 2009). Sorting and marking of duplicate sequences and generation of
133 alignment metrics were performed with Picard Tools v2.8.0
134 (<https://broadinstitute.github.io/picard/>).

135

136 Local realignment around indels and recalibration of base quality scores was performed
137 using the Genome Analysis Toolkit v3.7 (GATK) pipeline according to the GATK best
138 practice recommendations (McKenna, et al. 2010). To obtain a database of known
139 polymorphic sites for realignment and recalibration, the raw variants were called in
140 three programs: FreeBayes v0.9.20 (Garrison and Marth 2012), which required at least
141 five supporting observations to consider a variant, the GATK UnifiedGenotyper tool
142 that set the minimum phred-scaled quality score threshold to 50, and samtools mpileup
143 v1.3.1 with default parameters (Li, et al. 2009). The genotypes identified by all three
144 approaches were extracted with bcftools v1.2 (Li, et al. 2009) and used as a database of

145 known genotypes for realignment and recalibration. To obtain convergence between the
146 average reported quality scores and empirical quality scores, this process was repeated
147 four times.

148

149 **Identification of sex chromosomes and mitochondrial genome-linked scaffolds**

150 We employed two criteria to identify scaffolds belonging to either the sex chromosomes
151 or the mitochondrial genome. For one, we used the proportion of the chicken (*Gallus*
152 *gallus*) sex chromosome and mitochondrial orthologous genes on each scaffold. We
153 employed the tblastn and blastp functions with an e-value of 1×10^{-5} in BLAST+ v2.2.28
154 (Camacho, et al. 2009) to search for the one-to-one orthologous genes of *G. gallus* and
155 *C. mantchuricum*. The sex chromosomes and mitochondrial protein sequences of *G.*
156 *gallus* were downloaded from Ensembl Release 87 ([ftp://ftp.ensembl.org/pub/release-](ftp://ftp.ensembl.org/pub/release-87/)
157 [87/](ftp://ftp.ensembl.org/pub/release-87/)). We used in-house scripts to calculate the proportion of orthologous genes on each
158 scaffold. For the second criterion, we used the ratio of scaffold coverage between males
159 and females. We calculated the per-scaffold ratio of sequence coverage between males
160 and females, which is expected to be equal to 1 on autosomes and 2 on Z-linked
161 scaffolds. Considering the coverage bias, we assumed that scaffolds with a ratio
162 between 1.6 and 2.4 belong to the Z chromosome. Based on this coverage ratio criteria,
163 we assumed that the scaffolds that have equal to or more than 25% orthologous sex and
164 mitochondrial genes are sex chromosomes and mitochondrial genome-linked scaffolds.
165 Because we mapped the re-sequencing reads to a complete sequence of *Crossoptilon*
166 *crossoptilon* mitochondrion DNA, we excluded all mitochondrial genome-linked
167 scaffolds and SNPs in the downstream analyses.

168

169 For visualization and downstream analysis purposes, all autosome-linked scaffolds
170 were ordered based on their alignment to autosomes in the *G. gallus* genome
171 (ftp://ftp.ensembl.org/pub/release-87/fasta/gallus_gallus/dna/). All autosome-linked
172 scaffolds were aligned to each *G. gallus* autosome using the PROmer function in the
173 MUMmer V 3.23 package (Kurtz, et al. 2004). After alignment, we merged all Delta

174 files together and filtered the results with the following criteria: retain the longest
175 consistent alignment, minimum alignment identity is 50, and minimum alignment
176 length is 500. After filtering, we determined the location of each scaffold based on the
177 position of the longest alignment.

178

179 **SNP calling**

180 After realignment and recalibration, sites with a phred-scaled confidence ≥ 30 were
181 used to perform joint genotyping with the GATK HaplotypeCaller and
182 GenotypeGVCFs tool. Raw variant sets on autosomes, the Z-chromosome, and the
183 mitochondrial genome were generated separately based on our designation scaffolds.
184 The quality metrics of each raw variant were extracted with vcftools v0.1.14 (Danecek,
185 et al. 2011) and plotted on the density curve of each parameter in RStudio. The
186 distribution of each parameter was used to set the filtering criteria based on the
187 suggestion of the GATK best practices. Based on the distribution of each parameter,
188 quality filtering of the raw variants on autosomes was performed with the following
189 criteria to maintain high-quality SNPs: minor allele frequency > 0.05 , strand odds ratio
190 < 3 , quality by depth > 2 , Fisher Strand < 60 , RMS mapping quality > 50 , mapping
191 quality rank sum > -4 , read position rank sum > -2 , minimum depth (summing all 51
192 samples) > 870 , and maximum depth (summing all 51 samples) < 1304 ; explanation of
193 these parameters can be found in the GATK user manual (McKenna, et al. 2010). These
194 high-quality variants were used in the analyses performed in this study. Variants on the
195 Z chromosome were filtered using the same quality thresholds as above, except that the
196 minimum and maximum depths were 545 and 1207. Considering the huge amount of
197 mitochondrial DNA in cells, we set the minimum depth to 966 for filtering the raw
198 variants. The percentage of missing SNPs in each individual was calculated using
199 vcftools v0.1.14 (Danecek, et al. 2011).

200

201 **Inference of kinship**

202 In population genetic analyses, it is often necessary to exclude closely related

203 individuals. The three populations of *C. mantchuricum* are all small isolated wild
204 populations where individuals may have familial relationships. We estimated the
205 relatedness of all samples in each population using KING v 2.1.3 (Manichaikul, et al.
206 2010) to estimate the kinship coefficient using our library of autosomal SNPs with no
207 missing sites. Pairs with a kinship coefficient > 0.177 (twins and first-degree
208 relationship) were not included in subsequent analyses. There were 14 samples with
209 close (duplicate/twins, first degree) relationships to other individuals in the data. These
210 samples were removed, leading to a total of 37 samples that were retained in
211 downstream analyses (Fig. S13; Dataset S10).

212

213 **Phylogenetic inference and the mitochondrial haplotype network**

214 Phylogenetic relationships were inferred separately using a maximum-likelihood (ML)
215 approach with autosomes, the Z chromosome, and mitochondrial SNP data. Aligned
216 autosome sequences were generated from the SNP library using the SNPhylo pipeline
217 with the filtering criteria of a linkage disequilibrium coefficient ($R^2 < 0.2$) and no
218 missing sites (Lee, et al. 2014). After filtering with SNPhylo, 2,274,011 autosome SNPs
219 were used to infer phylogenetic relationships. Sequences for the Z chromosome and
220 mitochondria were generated with in-house scripts and aligned in the Mafft v 7.271
221 program (Yamada, et al. 2016). The phylogenetic trees were constructed in PhyML v
222 3.0 (Guindon, et al. 2010) with 1000 bootstrap replicates for autosomes, the Z
223 chromosome, and mitochondrial sequences. Nucleotide frequencies, the
224 transition/transversion ratio, and the value of the gamma shape parameter were
225 estimated by ML. Tree topology, branch length, and substitution rate parameters were
226 optimized in this program. The median joining haplotype network of mitochondria was
227 constructed using Popart (Leigh and Bryant 2015).

228

229 **Population structure and principal component analysis**

230 Principal component analysis (PCA) of the genotypes from 37 individuals was
231 performed using a stringently filtered set of 49,701 autosomal SNPs (no missing sites,

232 Hardy-Weinberg equilibrium exact test p -value > 0.00001 , linkage disequilibrium
233 coefficient, $r^2 < 0.2$) in the Genome-wide Complex Trait Analysis (GCTA) v 1.91.3
234 (Yang, et al. 2011). Using the same SNP set, population structure was inferred using
235 ADMIXTURE v 1.3.0 (Alexander, et al. 2009) with cross-validation (CV) (Alexander
236 and Lange 2011) and 100 bootstraps. This approach estimates the ancestry proportion
237 of each sample based on SNP data, given the *a priori* specification of K ancestral
238 populations. To test the stability of the results, we performed the analysis using all 37
239 samples, and subsets of individuals belonging to the putative populations *C.*
240 *mantchuricum*: Blue, Brown-W, Brown-C, Brown-E. We predefined the K value from
241 1 to 10 for all individuals (including *C. auritum* and *C. mantchuricum*), and from 1 to
242 6 within *C. mantchuricum*. For the isolated populations (Blue, Brown-W, Brown-C, and
243 Brown-E), we predefined the K value from 1 to 3. For each K value, the analysis was
244 performed ten times with different random seeds, which aimed to obtain the most stable
245 results.

246

247 **Estimation of genetic diversity**

248 To assess the genetic diversity within and among *C. mantchuricum* populations, we
249 calculated the heterozygosity of each sample, defined as the proportion of heterozygous
250 sites in all callable sites across the autosomes. For each population or species, we
251 defined the genetic diversity as the mean heterozygosity per individual within each
252 population or species. We compared the diversity estimates of the two species to 21
253 other avian species for which comparable estimates were available, some of which are
254 also highly inbred, such as White tailed eagle (*Haliaeetus albicilla*), Bald eagle
255 (*Haliaeetus leucocephalus*), Crested ibis (*Nipponia nippon*), and Dalmatian pelican
256 (*Pelecanus crispus*).

257

258 We also computed population genetic statistics including pairwise nucleotide diversity
259 (θ_π) and Watterson's expected nucleotide diversity (θ_w) on both synonymous (four-fold
260 degenerate sites) and non-synonymous sites (zero-fold degenerate sites). These

261 summary statistics were calculated based on the site frequency spectrum (SFS) with in-
262 house scripts. In both the synonymous and non-synonymous sites, low-quality and
263 missing genotypes were inferred separately for each population using the program
264 BEAGLE v 4.1 (Browning and Browning 2016). To test whether *C. mantchuricum* has
265 accumulated more missense mutations than *C. auritum*, we computed Θ_{π} (zero-fold
266 degenerate sites) / Θ_{π} (four-fold degenerate sites) for each population. If the populations
267 of *C. mantchuricum* have accumulated more missense mutations, then this value is
268 expected to be higher in *C. mantchuricum* than that in *C. auritum*. To compare these
269 summary statistics between populations, these statistics were calculated using 1,000
270 bootstrap replicates (resampling with replacement). Because the variance of any
271 summary statistics between two randomly selected populations was not homoscedastic,
272 non-paired Welch two-sample t-tests were used to test whether the summary statistics
273 of the two populations were significantly different from each other.

274

275 **Estimating inbreeding patterns**

276 Estimation of inbreeding using whole genome resequencing data has been shown to
277 outperform estimates using extensive pedigree data (Kardos, et al. 2018). To understand
278 the genomic extent of inbreeding in *C. mantchuricum*, we identified genome-wide
279 patterns of runs of homozygosity (ROH), estimated linkage disequilibrium (LD), and
280 the inbreeding coefficient (F_{is}) in each of the three populations of *C. mantchuricum*.
281 For comparison, we also performed the same analysis in the sampled population of *C.*
282 *auritum*.

283

284 ROH arises through the process of inbreeding that gives offspring the opportunity to
285 inherit identical haplotypes from each parent (Ceballos, et al. 2018). The location and
286 number of ROHs were identified in each sample using plink v1.90 with default
287 parameters (Howrigan, et al. 2011). Because meiosis can break down the segments that
288 are identical-by-descent, we assume that longer ROH (≥ 0.5 Mb) arose from recent
289 inbreeding, and shorter ROH (< 0.5 Mb) arose from more ancient inbreeding (Kirin, et

290 al. 2010). To compare the relative extents of recent and ancient inbreeding, the average
291 fraction of the longer and shorter ROH on the autosomes was summarized for each
292 population.

293

294 To avoid biases caused by the different sample sizes of the sampled populations in the
295 LD analysis, we randomly selected six unrelated samples from each population and
296 repeated this analysis 10 times. We employed Haploview (Barrett, et al. 2004) to
297 calculate the correlation coefficient (R^2) between any pair of SNPs in each population.
298 The parameters were set as follows: -maxdistance 500 -dprime -minGeno 0.75 -
299 minMAF 0.05 -hwcutoff 0.001. To plot the LD decay curves, we merged the SNPs that
300 had a similar physical distance (1 kb) into one group and calculated the mean R^2 for
301 each group. We estimated the inbreeding coefficient per individual in plink v1.90
302 (Howrigan, et al. 2011) and then calculated the average inbreeding coefficient for each
303 population.

304

305 **Evaluating the genetic load**

306 Genetic load is a factor that can drive the extinction of small populations. Evaluating
307 the genetic load of endangered species is an important component of conservation
308 genetics. Thus, we sought to gauge the genetic load of *C. mantchuricum* using
309 population genomic data. We estimated the relative excess of derived loss-of-function
310 (LOF) and missense variants in three populations of *C. mantchuricum* compared to
311 those of *C. auritum* to test whether these isolated populations experience inbreeding
312 depression.

313

314 To measure the genetic load, we employed methods previously used for gorillas (Xue,
315 et al. 2015). We used SnpEff to identify LOF, missense, and synonymous variations in
316 the coding region (Cingolani, et al. 2012). We inferred the ancestral and derived allele
317 at each location based on comparison to the genome of their close relatives, the White
318 eared pheasant (*C. crossoptilon*) (Wang, et al. 2018). In order to estimate the relative

319 number of derived LOF and missense alleles found in each population of *C.*
 320 *mantchuricum* compared to those in *C. auritum*, we used the following formula to
 321 calculate the number of derived LOF or missense alleles found in population *A* rather
 322 than in population *B*:

$$323 \quad L_{A,B}(C) = \frac{\sum_{i \in C} f^{A_i}(1 - f^{B_i})}{\sum_{j \in I} f^{A_j}(1 - f^{B_j})}$$

324 We used $f^{A_i} = d^{A_i} / n^{A_i}$ to calculate the observed derived allele frequency in population
 325 *A*, where d^{A_i} is the number of derived alleles in population *A*, and n^{A_i} is the total number
 326 of alleles in population *A*. In population *B*, f^{B_i} was similarly defined. We defined *C* as
 327 a set of protein-coding sites, and *I* as a set of intergenic sites. For each comparison, the
 328 relative number of derived LOF or missense variations was measured by $R_{A/B}(C) =$
 329 $L_{A,B}(C)/L_{B,A}(C)$. If the $R_{A/B}(C)$ is larger than one, it means that the *A* population has
 330 more LOF or missense mutations than the *B* population; otherwise, the *B* population
 331 has excess corresponding mutations. To estimate the variance of $R_{A/B}(C)$, we performed
 332 a 100-block jackknife at the sites of the *C* set.

333

334 To evaluate inbreeding depression, we used three strategies to analyze the outcome of
 335 inbreeding. We first calculated whether the ROH regions accumulated much more
 336 homozygous missense mutations than those outside the ROH regions. If the
 337 homozygous lethal variations did not come from inbreeding, we expected the ratio of
 338 the number of homozygous missense genotypes to the number of homozygous
 339 synonymous genotypes to be equal to one; otherwise, the ratio should be less than one.
 340 Next, we tested whether there were more heterozygous missense mutations compared
 341 to the number of homozygous mutations. If one population accumulates homozygous
 342 lethal mutations, the ratio of the number of missense alleles to the number of
 343 synonymous alleles in the homozygous sites would be significantly less than the ratio
 344 in the heterozygous sites. To test whether there was a significant difference between the
 345 homozygous and heterozygous sites, we employed the Kolmogorov-Smirnov test to
 346 compare the distribution. For the third strategy, we compared the genetic diversity of

347 the functional region (MHC, conserved non-exonic elements, conserved intron,
348 conserved intergenic region, and coding region) and non-functional region (putative
349 non-functional region, non-conserved intron, and non-conserved intergenic region)
350 between each population of *C. mantchuricum* and *C. auritum*. If *C. mantchuricum*
351 experienced inbreeding depression, both the functional and non-functional regions
352 would have a lower genetic diversity than that of *C. auritum*.

353

354 To identify the conserved non-exonic elements (CNEE), putative non-function region,
355 conserved and non-conserved introns, and conserved and non-conserved intergenic
356 regions, we followed the method described in a comparative genomics project on
357 paleognaths (Sackton, et al. 2019) to identify the *G. gallus* (galGal5) CNEE coordinates.
358 After getting the information regarding the *G. gallus* CNEE coordinates, we aligned the
359 scaffolds of *C. mantchuricum* to the *G. gallus* genome in LAST software (Kiełbasa, et
360 al. 2011). To obtain the CNEE coordinates of *C. mantchuricum*, we lifted the *G. gallus*
361 CNEE coordinates to *C. mantchuricum*. We defined the putative non-functional region
362 as the region far away from the genes, with CNEE regions at least 50 kb and at least 1
363 kb in length. The conserved intron and conserved intergenic region are defined as the
364 regions with overlap between CNEE and the intron and intergenic regions, respectively.
365 Otherwise, the regions that do not have any overlap are considered non-conserved
366 introns and non-conserved intergenic regions.

367

368 As an additional assessment of the fitness of *C. mantchuricum*, we calculated the
369 genetic diversity in the MHC region. We identified the MHC region with the most
370 effective reciprocal hit BLAST method. By comparing the protein sequences of *C.*
371 *mantchuricum* and *G. gallus*, the ortholog genes were identified. Using the annotation
372 information of *G. gallus*, the ortholog MHC region was identified. With the in-house
373 scripts, we calculated the genetic diversity of the MHC.

374

375 In order to more intuitively understand whether purifying selection could remove

376 deleterious mutations in *C. mantchuricum*, we investigated whether or not derived
377 missense mutations in potential adaptive genes are deleterious. Because the genomic
378 islands identified by the statistic F_{ST} may occur due to reduced diversity (Cruickshank
379 and Hahn 2014), and *C. mantchuricum* typically has low genome-wide diversity, we
380 employed d_{XY} , a statistic that is independent of the genetic diversity within the
381 populations, to identify the candidate adaptive genes (Cruickshank and Hahn 2014). We
382 also used π , a measure of genetic variation, to infer the candidate adaptive genes. We
383 first employed “popgenWindows.py” written by Simon Martin at The University of
384 Edinburgh, UK, to calculate the d_{XY} between *C. auritum* and each population of *C.*
385 *mantchuricum*. The measure of genetic variation, π , was also calculated using this
386 script. We set the window size to 100 kb, and the minimum number of sites within a
387 window was 10. The size of the step for the sliding window was set to 25 kb. The
388 window size was determined by the results of the linkage disequilibrium test. Then, we
389 Z-transformed the d_{XY} and π to Zd_{XY} and $Z\pi$. Because there was almost no genetic
390 diversity in *C. mantchuricum*, we only employed the $Z\pi$ of *C. auritum* and Zd_{XY} to
391 infer the candidate adaptive genes. The windows with high (top 5%) Zd_{XY} and
392 extremum (both top 5% and bottom 5%) of $Z\pi$ were determined as the candidate
393 regions containing the genes under selection. Genes with more than 50% of their length
394 contained within the candidate regions were selected as the genes putatively under
395 positive selection. PROVEAN v 1.1.5 (Choi, et al. 2012) was used to predict whether
396 the derived missense mutations in the genes putatively under positive selection were
397 deleterious mutations. The derived missense mutations with PROVEAN scores lower
398 than -2.5 were identified as deleterious mutations. The genetic diversity of each
399 potential adaptive gene and the frequency of the derived missense mutations were also
400 calculated. KOBAS v 3.0 (Wu, et al. 2006) was used to perform KEGG and GO
401 enrichment analyses for these genes putatively under positive selection.

402

403 **Demographic History Reconstruction**

404 To explore the potential historical factors that are linked to low present-day genetic

405 diversity, we used the Multiple Sequential Markovian Coalescence (MSMC2) model
406 (Schiffels and Durbin 2014) to reconstruct demographic histories for each of our study
407 populations. Because MSMC2 uses phased haplotypes as input data, we used SHAPEIT
408 v2.904.3.10.0 (Delaneau, et al. 2012) to phase 33 scaffolds separately, with a minimum
409 length of 132 kb. Based on the results from MUMmer, we selected at least one scaffold
410 from each chromosome. The total length of these 33 scaffolds covered 18.48% of the
411 draft genome. To prepare the input file, we followed the procedure of Heng Li's
412 SNPable program (<http://lh3lh3.users.sourceforge.net/snpable.shtml>) to generate
413 Mappability mask files for each scaffold. Under the general guidelines of MSMC2
414 (<https://github.com/stschiff/msmc/blob/master/guide.md>), we ran MSMC2 and
415 calculated the effective population size (N_e) of *C. mantchuricum*. To test the
416 convergence of the result, we performed both analyses with 100 bootstrap replicates.
417 Based on the comparison of the four-fold degenerate sites among the lineages of
418 Galliformes, we set the mutation rate of *C. mantchuricum* as 4.02×10^{-9} per site per
419 generation (Zhang, et al. 2014), and the generation time as 2 years per generation
420 (Zheng 2015).

421

422 We used Migrate-n v 3.6.11 (Beerli and Felsenstein 2001) to investigate whether gene
423 flow occurs between adjacent populations of *C. mantchuricum*. We evaluated four
424 possible gene flow patterns among the three populations of *C. mantchuricum*: migration
425 between adjacent populations, migration between Brown-C and Brown-W, migration
426 between Brown-C and Brown-E, or no gene flow between adjacent populations. The
427 best-fit model was inferred by the maximum likelihood in the Migrate-n. To prepare the
428 input data for Migrate-n, we first selected the putative non-functional regions whose
429 lengths were longer than 1 kb and then randomly selected 1 kb from each of these
430 putative non-functional regions. Among the selected 1 kb putative non-functional
431 regions, we randomly selected 30 regions to create the bed file. In the vcftools software,
432 we extracted the phased vcf file for each bed and then employed the bcftools software
433 to produce sequences for each region of each sample. Using the 60 sequences of each

434 sample, we performed Migrate-n to estimate the gene flow patterns.
435
436 In order to distinguish whether the populations experienced continuous population
437 decline or consistent small population size in *C. mantchuricum*, we employed
438 fastsimcoal26 (Excoffier, et al. 2013), a simulation-based framework, to infer the
439 demographic history of the three populations of *C. mantchuricum*. Fastsimcoal26 uses
440 SFS information to infer the most suitable evolutionary model given a range of
441 scenarios. Using this method, we also tested whether *C. mantchuricum* experienced
442 inbreeding and whether there was gene flow among the adjacent populations of *C.*
443 *mantchuricum*. The three-dimensional SFS of the three populations of *C. mantchuricum*
444 was estimated using the -doSaf and -realSFS methods in ANGSD (Korneliussen, et al.
445 2014). To minimize misidentification of the derived allele in *C. mantchuricum*, we only
446 selected the sites whose alleles were fixed in the population of *C. auritum*, and excluded
447 the sites that were not located in the putative non-functional region and were not present
448 in all individuals within each population. We first estimated the unfolded one-
449 dimensional SFS for each of the *C. mantchuricum* populations. We then estimated the
450 three-dimensional SFS among the three populations. The three-dimensional SFS was
451 used to estimate the composite likelihoods of the different evolutionary models.
452 Because we wanted to test 1) whether *C. mantchuricum* experienced long-term
453 population decline (D) or consistent small population size (S), 2) whether there was
454 gene flow between the adjacent populations of *C. mantchuricum* (no gene flow between
455 adjacent populations: I; gene flow occurred between adjacent populations: WCE; gene
456 flow occurred between the Western and Central populations: CW; gene flow occurred
457 between the Central and Eastern populations: CE), and 3) whether there had been
458 inbreeding (IN) in the populations of *C. mantchuricum*, we devised 16 historical models
459 for the three populations. The 16 models were as follows: a) S+I; b) S+WCE; c) S+CW;
460 d) S+CE; e) D+I; f) D+WCE; g) D+CW; h) D+CE; i) S+I+IN; j) S+WCE+IN; k)
461 S+CW+IN; l) S+CE+IN; m) D+I+IN; n) D+WCE+IN; o) D+CW+IN; and p) D+CE+IN
462 (these models are illustrated in Fig. S11). For each model, we performed 1,000,000

463 simulations to estimate the expected derived SFS, and 100 expectation conditional
464 maximization cycles were performed. We also required the minimum console output
465 and set the minimum observed SFS entry count as 20 accounting for parameter
466 estimation. The multiSFS and infinite site options were also used for estimating the
467 model parameters. To obtain the global maximum likelihood for each model, the above
468 procedure was replicated 100 times. To infer the best-fit model, the replicate with the
469 highest estimated likelihood was used to calculate the AIC values and Akaike weights
470 (w).

471

472 **Results**

473 **Mapping and SNP quality**

474 The sequencing coverage of the re-sequence individuals varied from 13.4 to 43.2 (the
475 average coverage was 19.0), and the mapping rate varied from 83.6% to 98.9% (the
476 average mapping rate was 97.3%) (Dataset S9).

477

478 The average proportion of high-quality bases (the call quality of the bases was Q20 or
479 higher) in the BAM files was 93.30% (varied from 90.20% to 95.00%), 93.28% (varied
480 from 89.07% to 95.27%), and 95.46% (varied from 95.24% to 95.75%) of the
481 autosomes, Z chromosome, and mitochondria, respectively (Fig. S14a, Fig. S15a, Fig.
482 S16a, Dataset S11). This proportion was defined as the number of high-quality bases
483 divided by the number of aligned bases in the BAM file. In the BAM files of the
484 autosomes, Z chromosome, and mitochondrial genome, the average percentage of sites
485 covered by at least five reads was 97.58% (varied from 83.37% to 98.56%), 88.83%
486 (varied from 45.00% to 98.44%) and 100% (no variation), respectively (Fig. S14b, Fig.
487 S15b, Fig. S16b, Dataset S11).

488

489 After variant calling, we obtained 7,556,341 and 65 single-nucleotide variants for the
490 genome (including autosomes and sex chromosomes) and mitochondria, respectively.
491 After hard-filtering, the final SNP library contained 2,327,376 and 134,081 variable

492 sites on the autosome and Z chromosome, respectively. In the mtDNA dataset, we
493 retained 24 substitution sites. In the autosome, Z chromosome, and mitochondrial SNP
494 library, the mean proportion of missing SNPs was 0.047% (varied from 0.006% to
495 0.372%), 0.392% (varied from 0.003% to 4.144%), and 0 (no variation), respectively
496 (Fig. S14c, Fig. S15c, Fig. S16c, Dataset S11).

497

498 The average number of heterozygous sites of the autosomes and Z chromosome was
499 230,681 (varied from 38,525 to 902,604) and 10,764 (varied from 808 to 28,595),
500 respectively (Fig. S17, Fig. S18). In the autosome SNP library, *C. auritum* had the
501 highest heterozygosity (average= 0.083%, SD = 0.0055%). Among the three
502 populations of *C. mantchuricum*, Brown-C had the highest heterozygosity (average=
503 0.014%, SD= 0.001%), while both Brown-W and Brown-E had similar heterozygosity
504 (Fig. S14d). On the Z chromosomes, *C. auritum* had the highest genetic diversity
505 (average= 0.029%, SD= 0.007%), but the three populations of *C. mantchuricum* had
506 similar genetic diversity (Fig. S15d). For the mitochondria, the Brown-C and Brown-E
507 populations had a low average number of nucleotide differences (the value was 0) (Fig.
508 S16d).

509

510 **Scaffolds linked to the Z chromosome and mitogenome**

511 We identified 49 Z chromosome-linked scaffolds (Dataset S12), which had 71.05 Mb
512 bases with a scaffold N50 of 2432.56 kb. The length of the Z chromosome of *C.*
513 *mantchuricum* was similar to that of *G. gallus* (82.53 Mb) (Consortium 2004). The
514 distribution of single-base sequencing depths across all individuals showed two peaks
515 for all scaffolds, but there was one single peak for the scaffolds designated as autosome-
516 linked, indicating that the Z-linked scaffolds had effectively been removed from this
517 set (Fig. S19). Female individuals had lower sequencing coverage on the Z
518 chromosome compared to that of the male individuals, while the female and male
519 individuals had similar sequencing coverage of the autosomes (Fig. S20). All the
520 evidence indicated that the process of identifying and removing Z-linked scaffolds was

521 effective.

522

523 Interestingly, the W-chromosome did not appear in this assembly, even though the
524 individual used to generate the *de novo* assembly was a female. This may be due to the
525 typical characteristics of the avian W chromosomes, such as they tend to be highly
526 heterochromatic and degenerated (Zhou, et al. 2014; Peichel 2017). We identified 3
527 scaffolds that are on the mitochondrial genome (Dataset S12), which have 5,875 bp.

528

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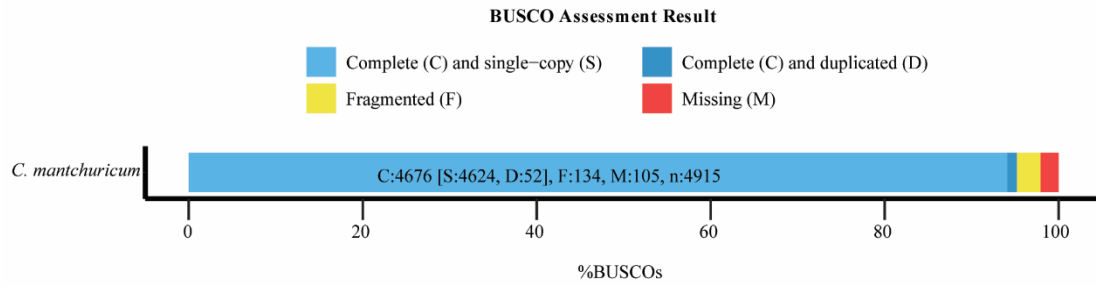
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639 **Figures**

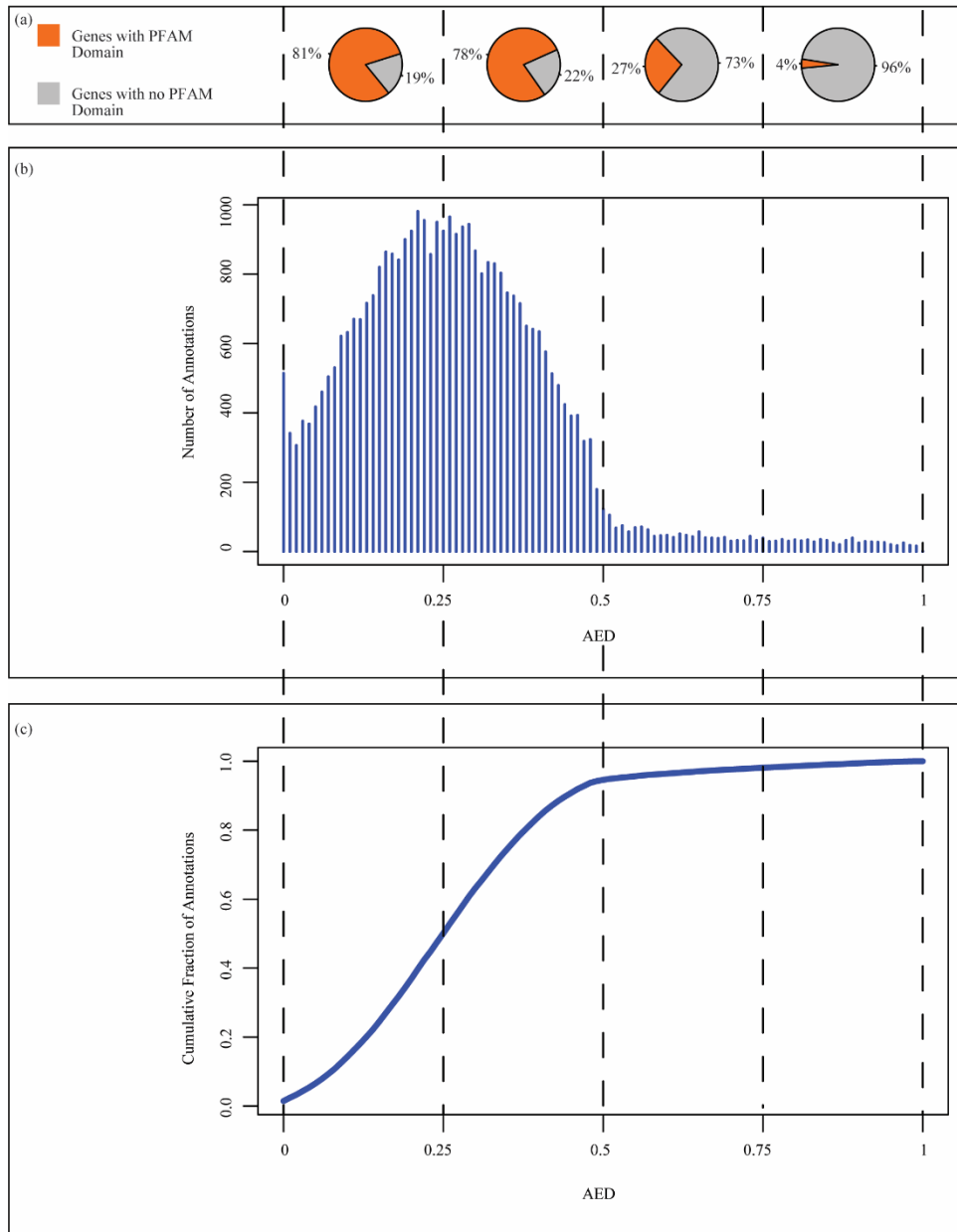


640

641 Fig. S1. Benchmarking Universal Single-Copy Orthologs (BUSCO) assessment Result.

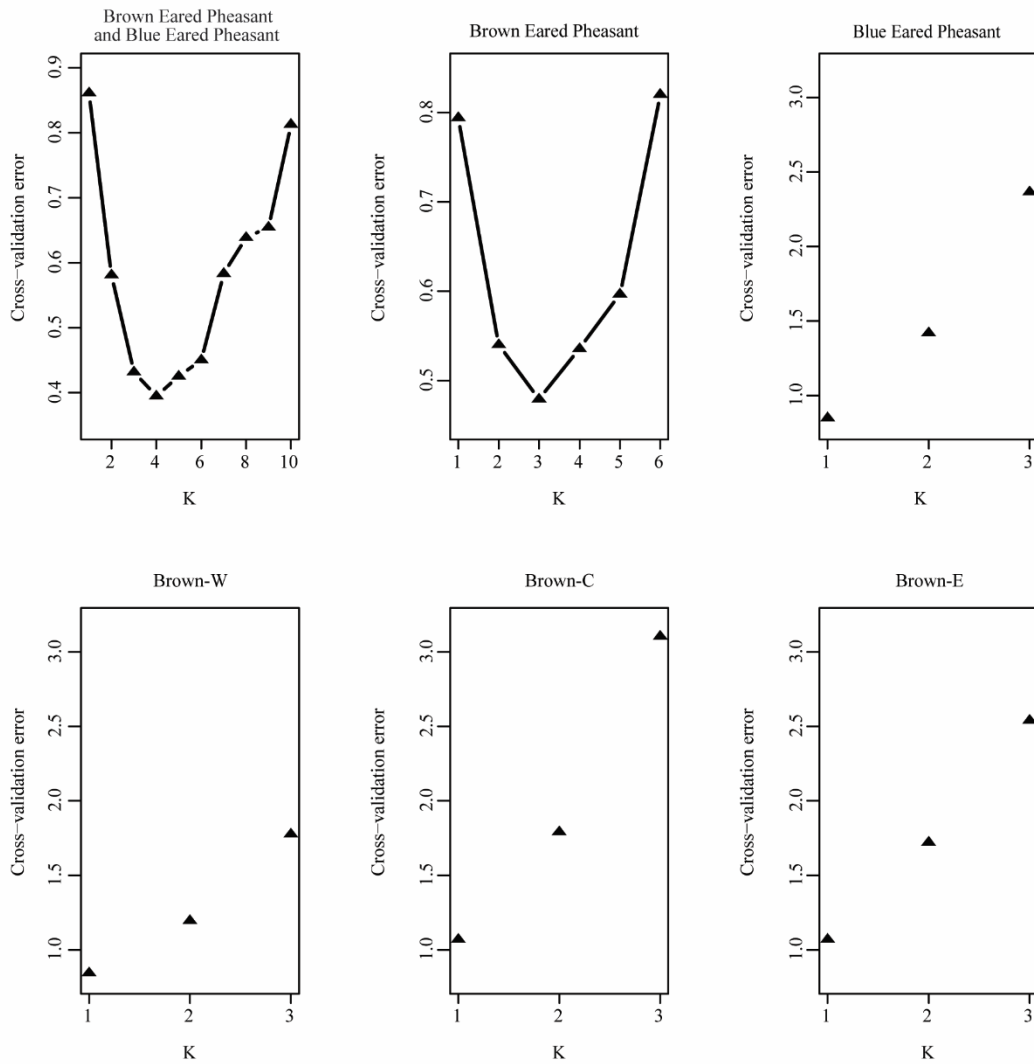
642 The numbers after the letters indicate the amount of BUSCO groups of the

643 corresponding groups. n indicates the number of BUSCO groups in the avian dataset.



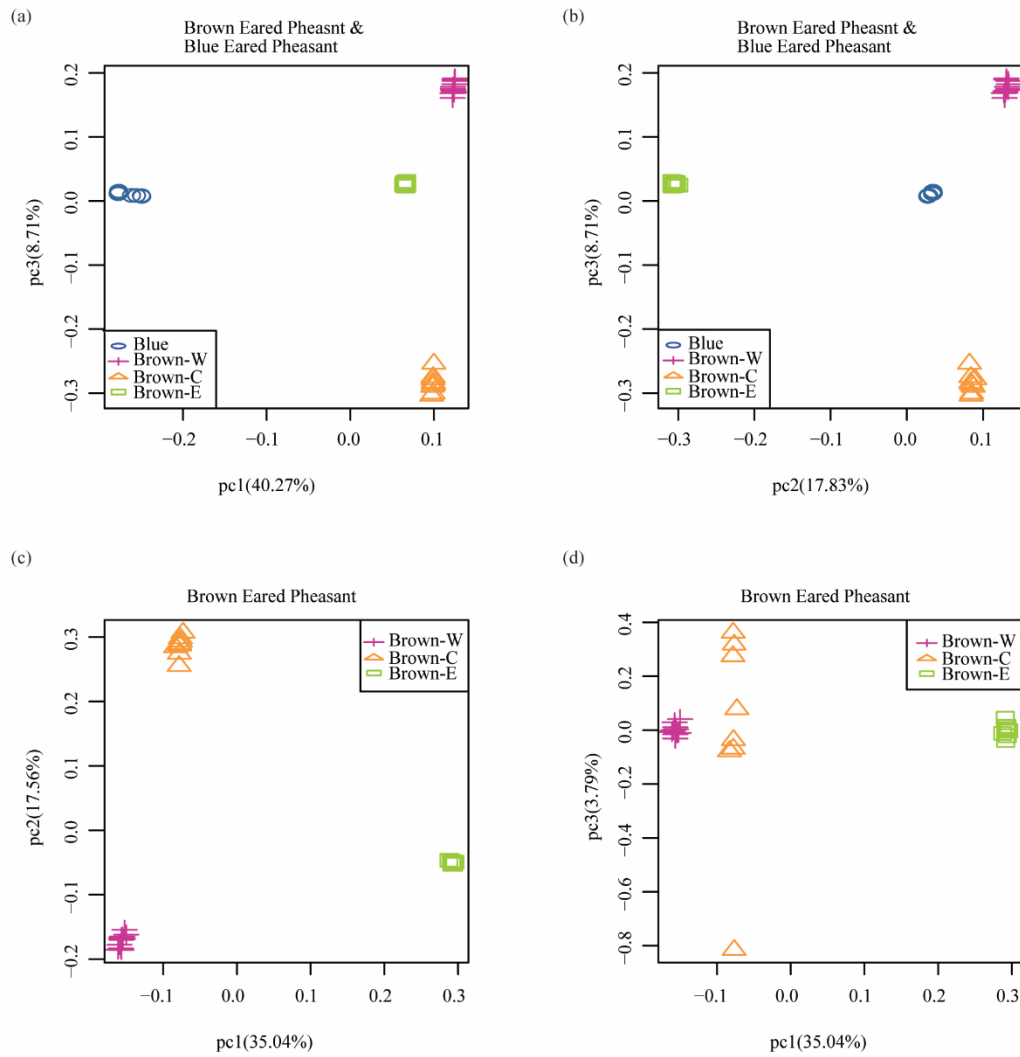
644

645 Fig. S2. Annotated gene set assessment result. AED means Annotation Edit Distance,
 646 which is used to measure the suitability between the annotated gene model and the
 647 supporting evidence of that model. AED is represented by a number between 0 and 1.
 648 An AED value of 0 indicates the annotated gene model is perfectly supported by
 649 available evidence, and an AED of 1 denotes that the annotated gene model is
 650 completely lacks supportive evidence. (a) The pie charts represent the annotated gene
 651 models with different AED values (0~0.25, 0.25~0.5, 0.5~0.75, and 0.75~1). (b) The
 652 number of annotated gene models with different AED values from 0 to 1. (c) The curve
 653 of the cumulative fraction of the annotated gene models.



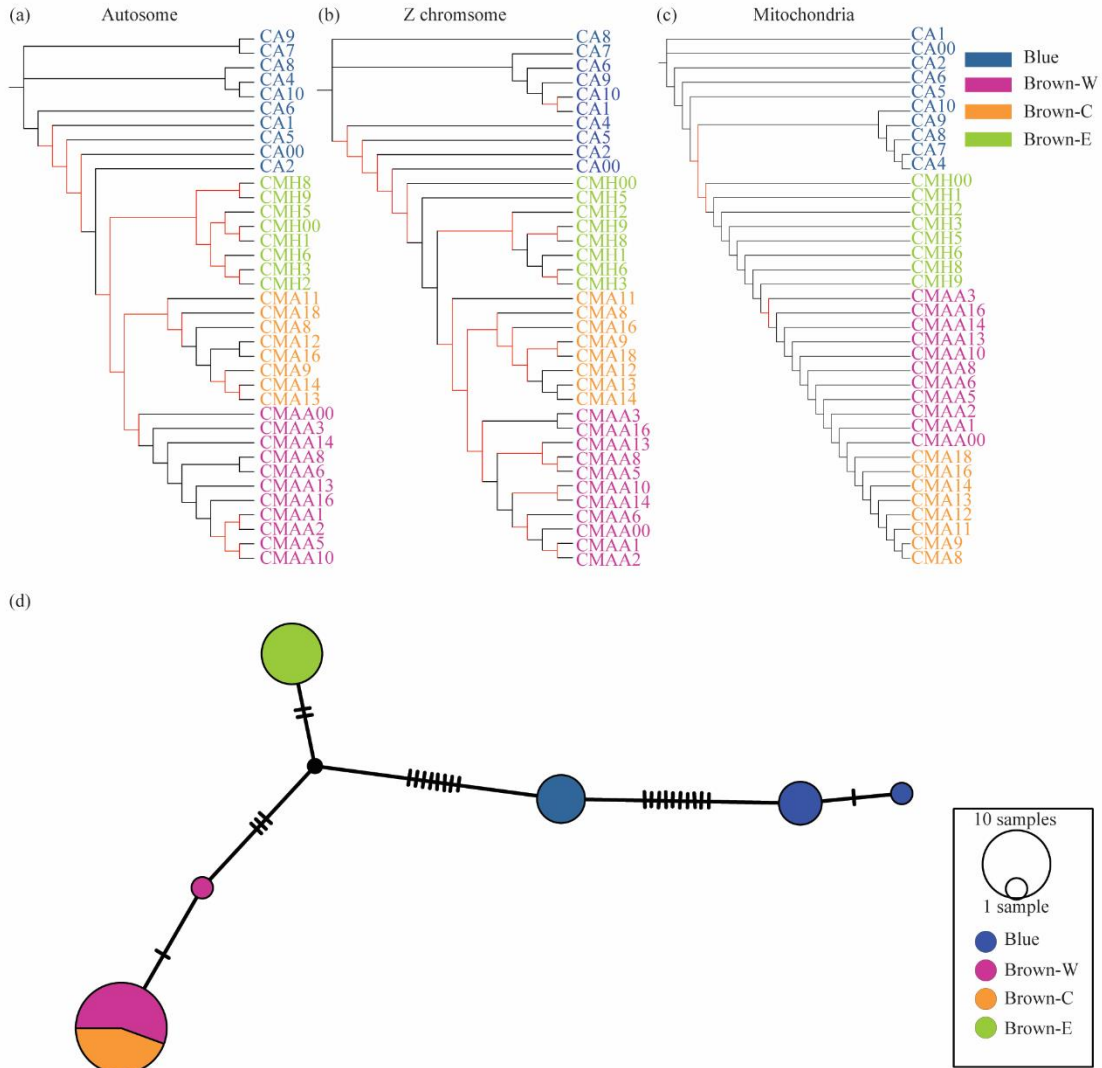
654

655 Fig. S3. Cross-validation (CV) values of different ancestral number (K) values. The CV
 656 values from the ADMIXTURE analyses performed on the different population scales
 657 with 10 independent runs with different random seeds.

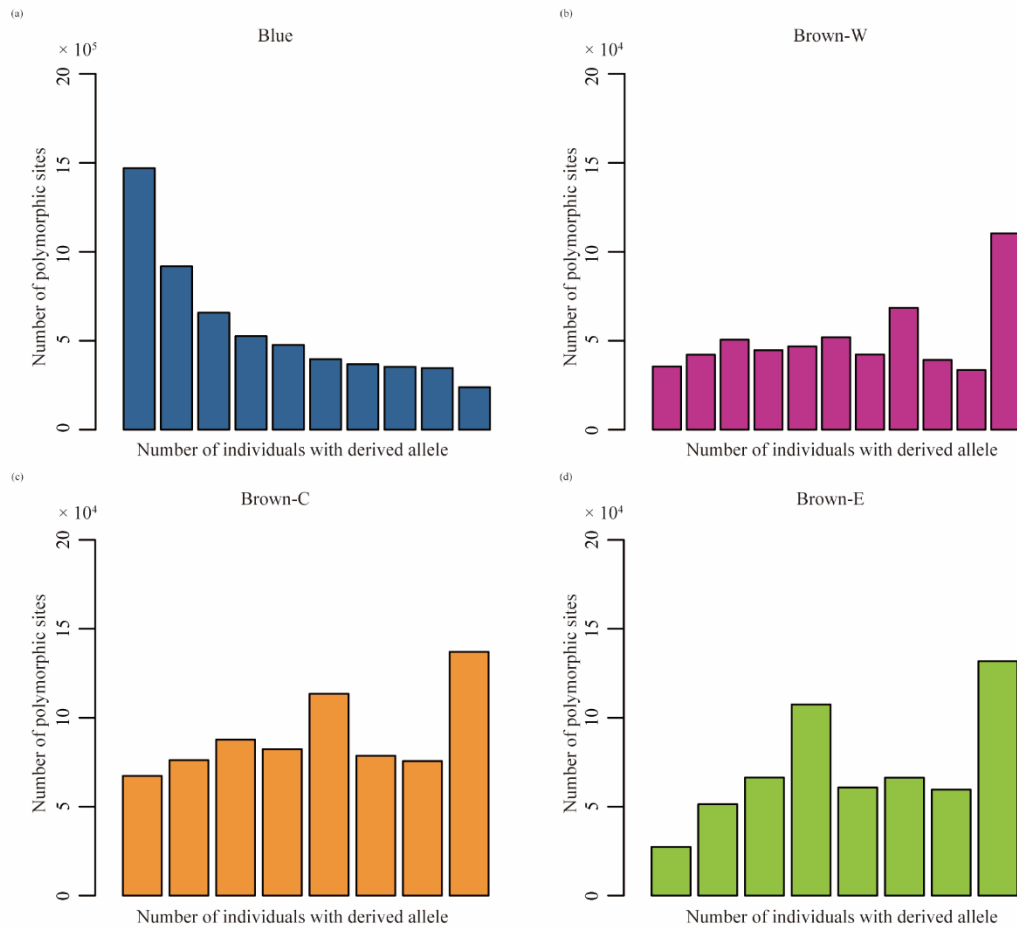


658

659 Fig. S4. Principal component analysis (PCA) of *Crossoptilon mantchuricum* and
 660 *Crossoptilon auritum*. The numbers in brackets are the contribution values of the
 661 corresponding eigenvectors.

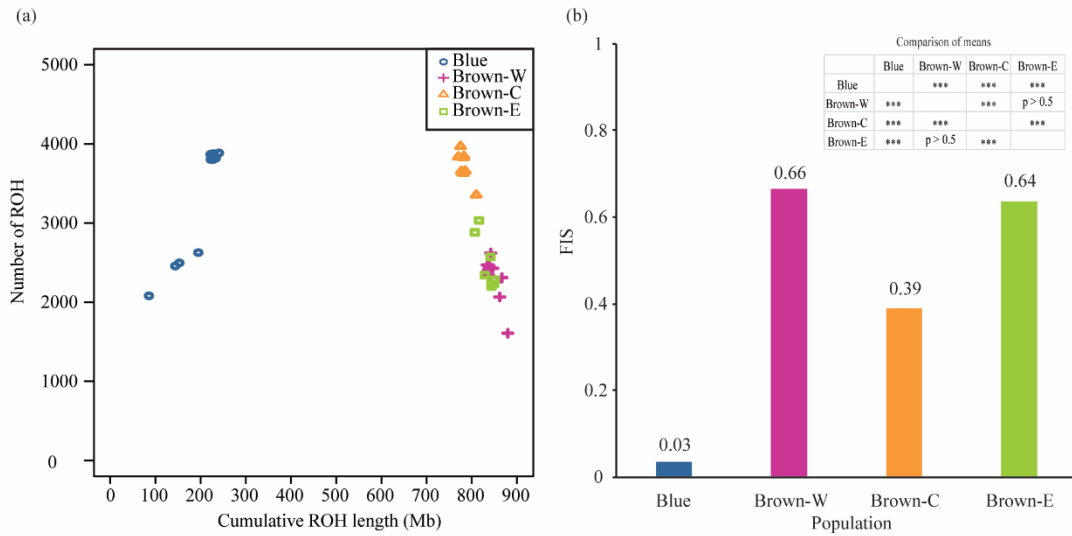


662
 663 Fig. S5. Phylogenetic relationships and haplotype networks within *Crossoptilon*
 664 *mantchuricum*. The colors represent sampling populations; blue represents
 665 *Crossoptilon auritum* (Blue) as outgroups, while orange, green, and red represent the
 666 Western (Brown-W), Central (Brown-C), and Eastern (Brown-E) populations of *C.*
 667 *mantchuricum*, respectively. (a), (b), and (c) The unrooted maximum-likelihood (ML)
 668 trees reconstructed using autosomes, the Z chromosome, and mitochondria,
 669 respectively. The bootstrap values larger than 80% of the bootstrap number were
 670 associated with the corresponding branches. (d) Median joining mitochondrial
 671 haplotype network. Lines on the linking branches indicate single base variations, and
 672 circle size represents the number of samples that share a single haplotype.



673

674 Fig. S6. Folded site frequency spectrum. (a) *C. auritum*. (b) Brown-W, the western
 675 population of *C. mantchuricum*. (c) Brown-C, the central population of *C.*
 676 *mantchuricum*. (d) Brown-E, the eastern population of *C. mantchuricum*.



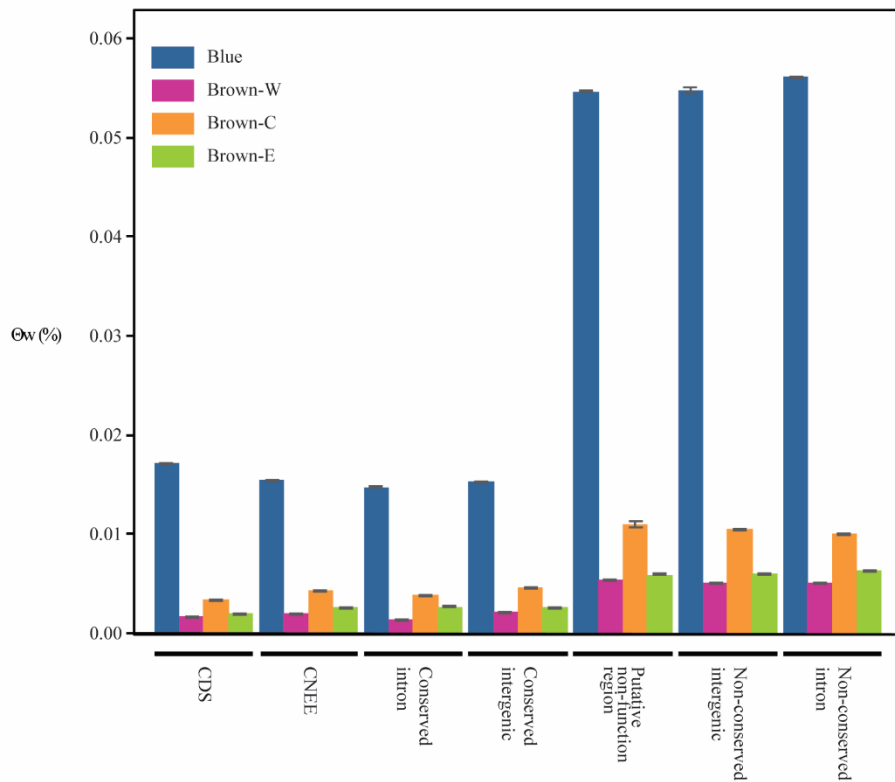
677

678 Fig. S7. Census of the Runs of Homozygosity (ROHs) and inbreeding coefficients (F_{IS}).

679 (a) The cumulative ROH length against the total number of observed ROH segments

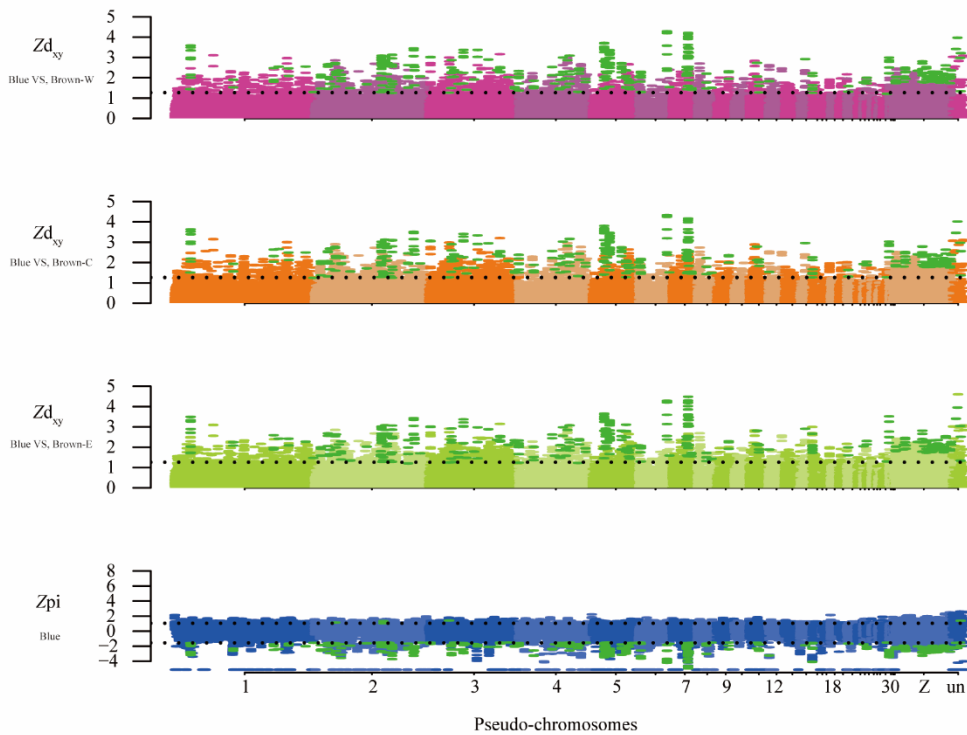
680 for each sample. (b) The mean inbreeding coefficient of each population. Three

681 asterisks indicate that the non-adjusted P values were less than 10^{-3} .



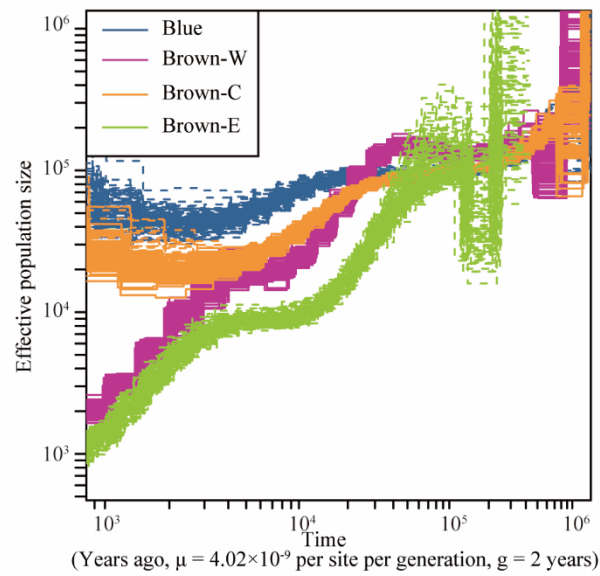
682

683 Fig. S8. Genetic diversity (θ_w) of each population in different regions (CDS: coding
 684 region; CNEE: conserved non-exonic elements; conserved intron; conserved intergenic;
 685 putative non-function region; non-conserved intergenic; and non-conserved intron).
 686 Error bars represent 95% confidence intervals estimated from 1,000 bootstrap
 687 resampling.



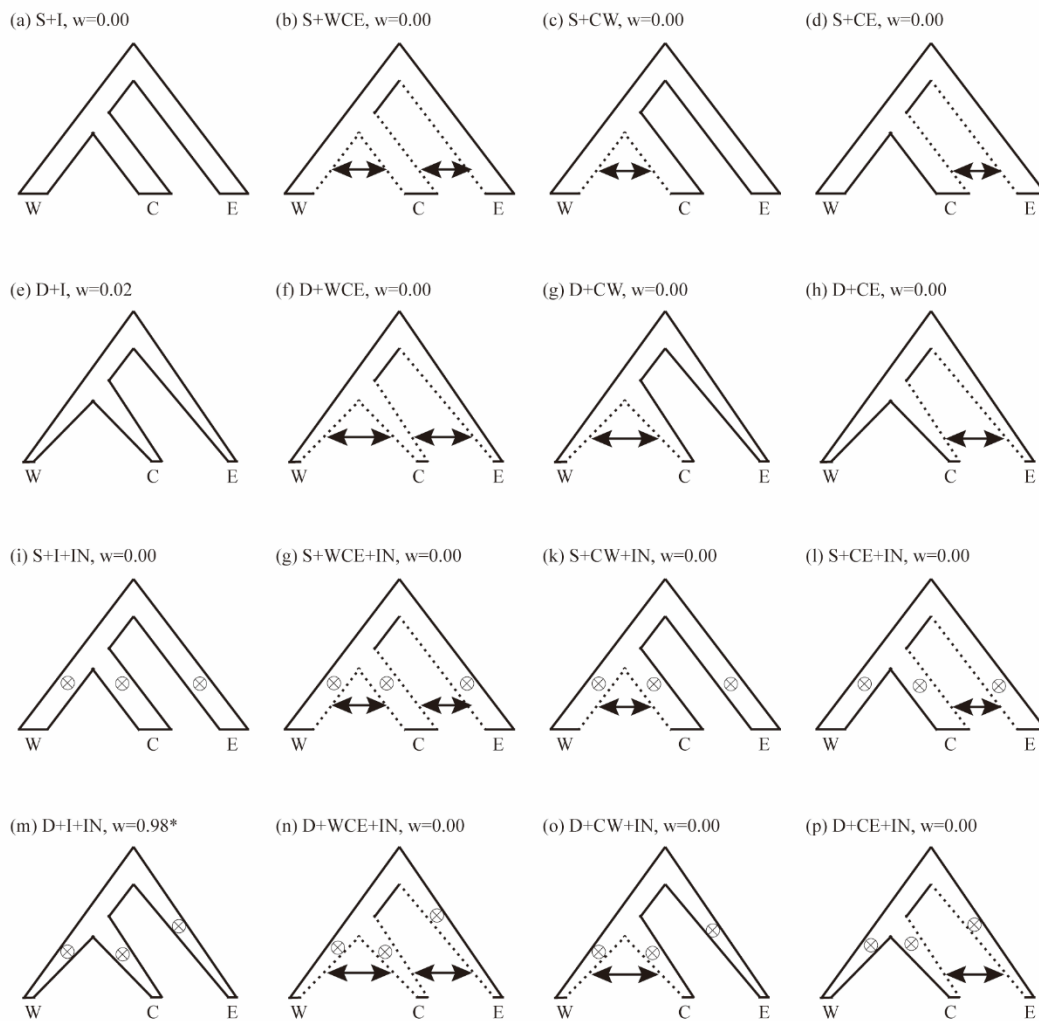
688

689 Fig. S9. Manhattan plots of the genome-wide Zd_{xy} and Zpi . The points above the dotted
 690 line in the Zd_{xy} Manhattan plots are the windows with the top 5% Zd_{xy} . The points
 691 above the upper dotted line and the points under the lower dotted line in the Zpi
 692 Manhattan plot are the windows with top 5% and bottom 5% Zpi . The green points are
 693 the windows with the genes putatively under positive selection.



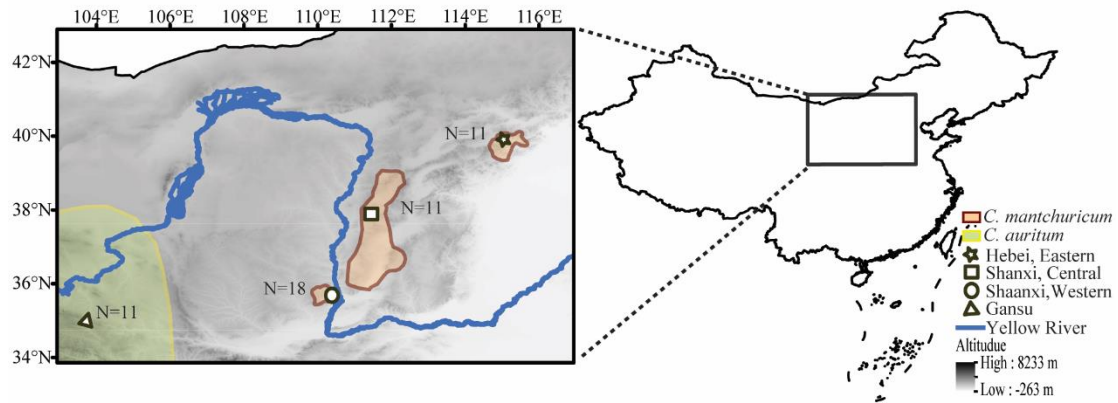
694

695 Fig. S10. Demographic history of *Crossoptilon mantchuricum* and *Crossoptilon*
696 *auritum* estimated from the 100 times MSMC bootstrap. The MSMC results show the
697 demographic history of the four populations. The blue line represents *C. auritum* (Blue);
698 the purple, brown, and green lines represent the Western (Brown-W), Central (Brown-
699 C), and Eastern (Brown-E) populations of *C. mantchuricum*, respectively.



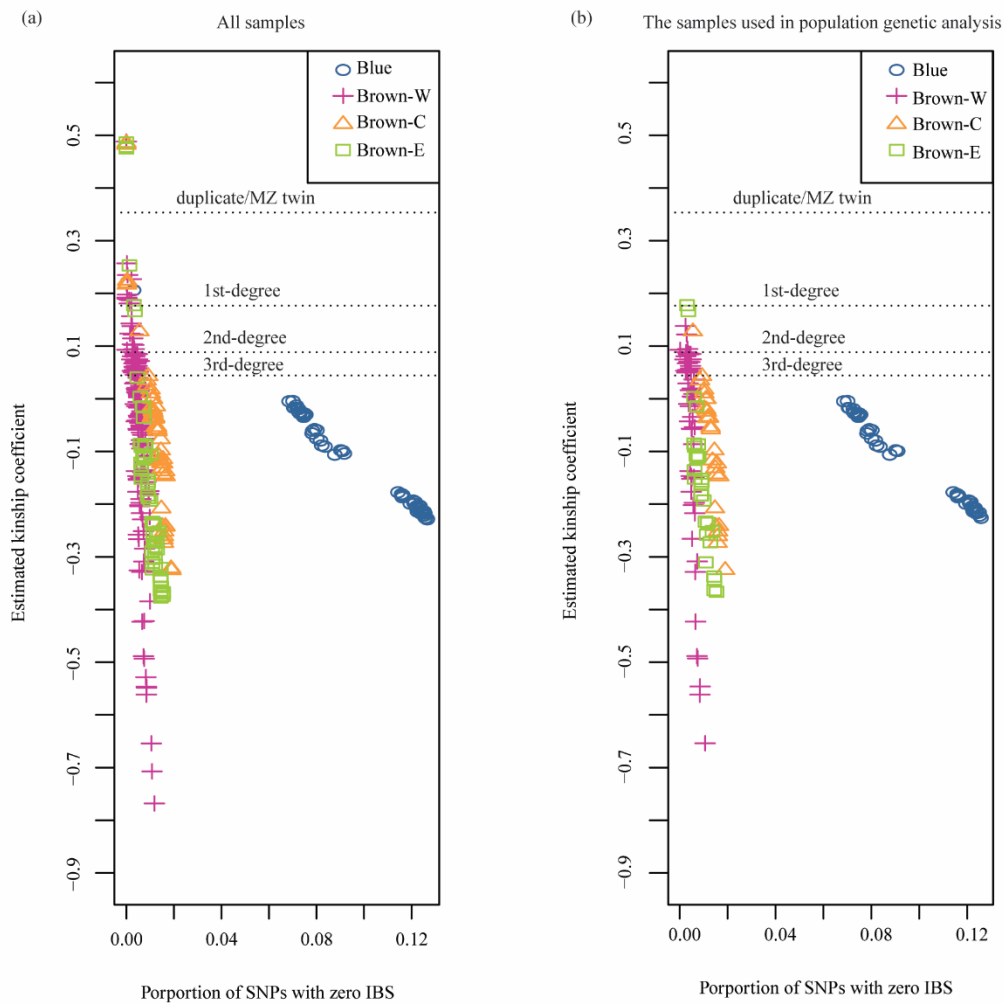
700

701 Fig. S11. The demographic models inferred by fastsimcoal26. S represents consistent
 702 small population size, D represents population decline, I indicates that there was no
 703 gene flow between adjacent populations, WCE indicates that there was gene flow
 704 between adjacent populations, CW indicates that the gene flow only existed between
 705 the Western and Central populations, CE indicates that the migration only occurred
 706 between the Central and Eastern populations, and IN indicates that inbreeding occurred.
 707 w represents the Akaike weights. The D+I+IN model obtained the ideal amount of
 708 support, which was marked by the asterisk.



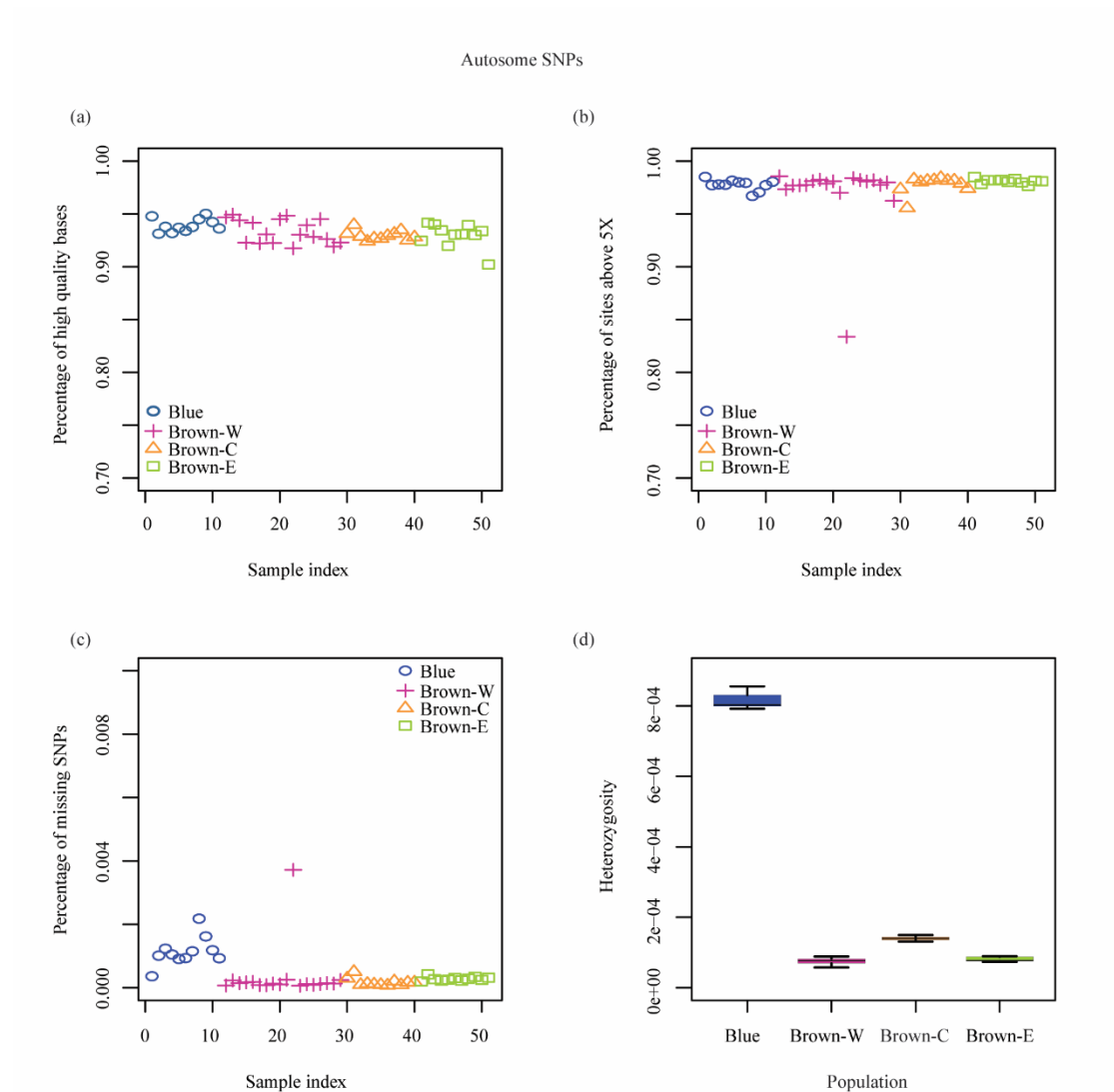
709

710 Fig. S12. Sample locations. N indicates the number of samples for each population. The
 711 brown regions indicate the distribution of *Crossoptilon mantchuricum*, while the light-
 712 yellow region indicates the distribution of *Crossoptilon auritum*. The star represents the
 713 Eastern population of *C. mantchuricum* (Brown-E), the square represents the Central
 714 population of *C. mantchuricum* (Brown-C), the circle represents the Western population
 715 of *C. mantchuricum* (Brown-W), and the triangle represents *C. auritum* (Blue).



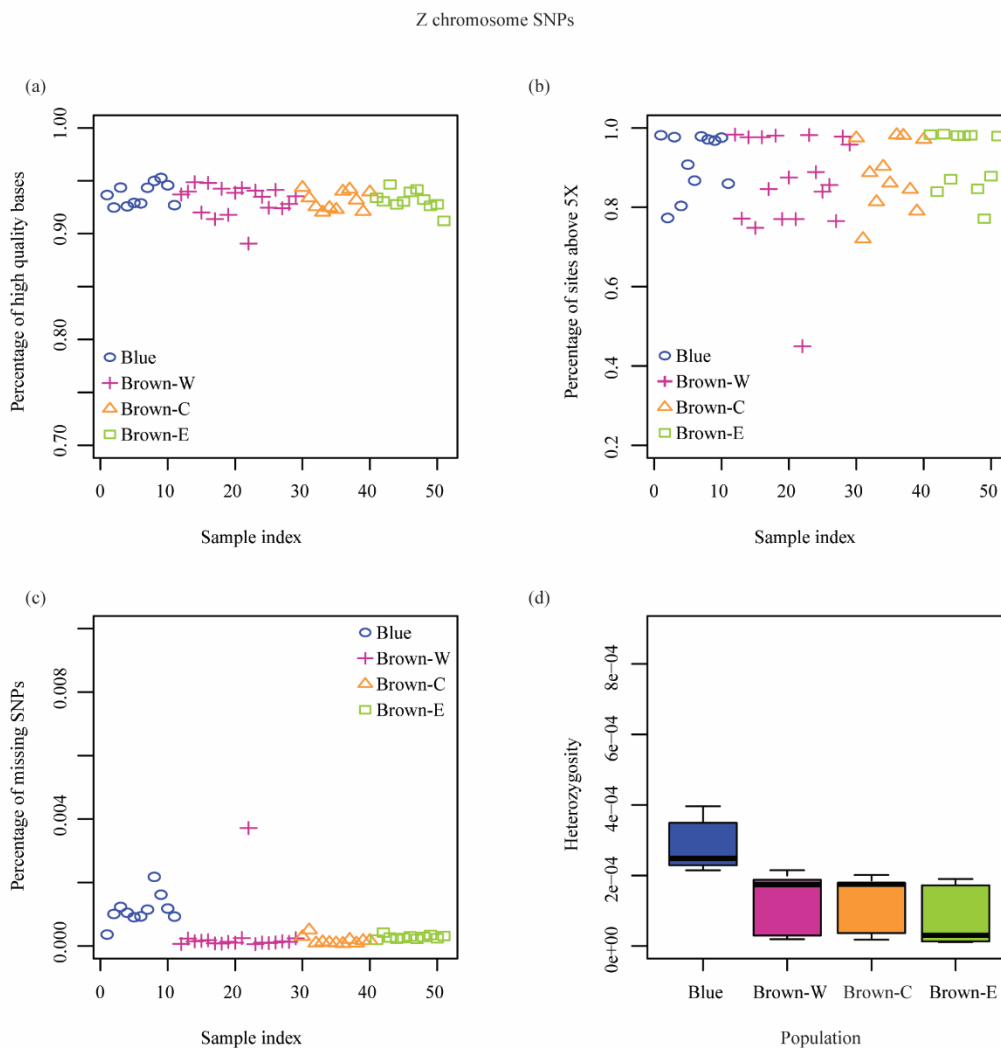
716

717 Fig. S13. Individuals' kinship within populations. Zero IBS indicates non-Identical-By-
 718 State. The dashed lines show the kinship thresholds (duplicate/MZ twin, 1st-degree, 2nd-
 719 degree, 3rd-degree). (a) and (b) show the relatedness of all samples within populations
 720 and the individuals used in the population genetic analysis in each population,
 721 respectively.



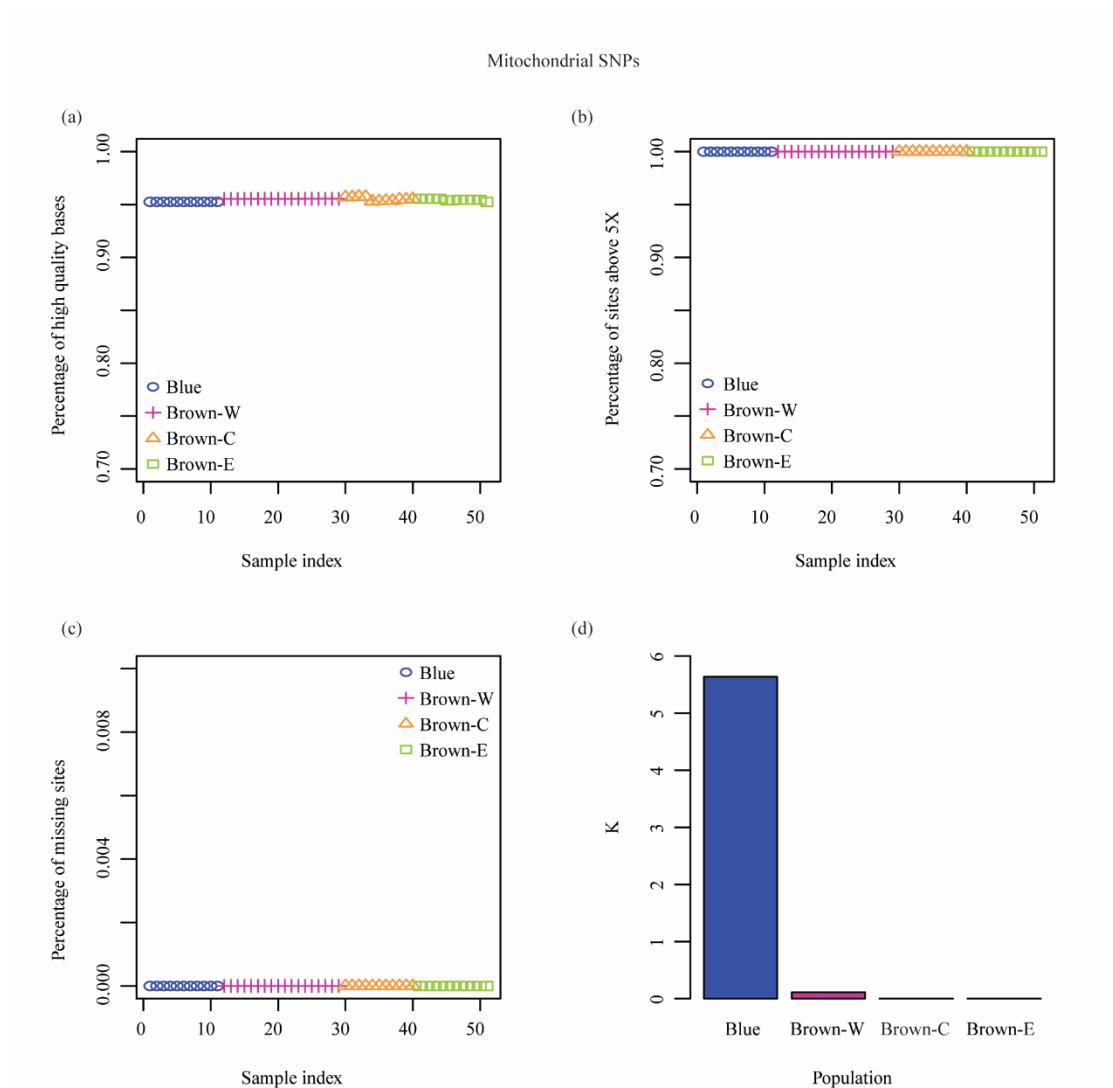
722

723 Fig. S14. Variant quality metrics of autosomes. The corresponding sample of each
 724 sample index can be seen in Supplementary Dataset S11. (a) Individual percentage of
 725 high-quality sites in the BAM file (the call quality of the bases was Q20 or higher). (b)
 726 Individual percentage of sites in the BAM file with coverage greater than 5 X. (c)
 727 Individual percentage of missing SNPs. (d) Heterozygosity of each sample in the four
 728 populations. In the box plots, the bold black line represents the median, and the boxes
 729 limit the 25th and 75th percentiles of the distributions.



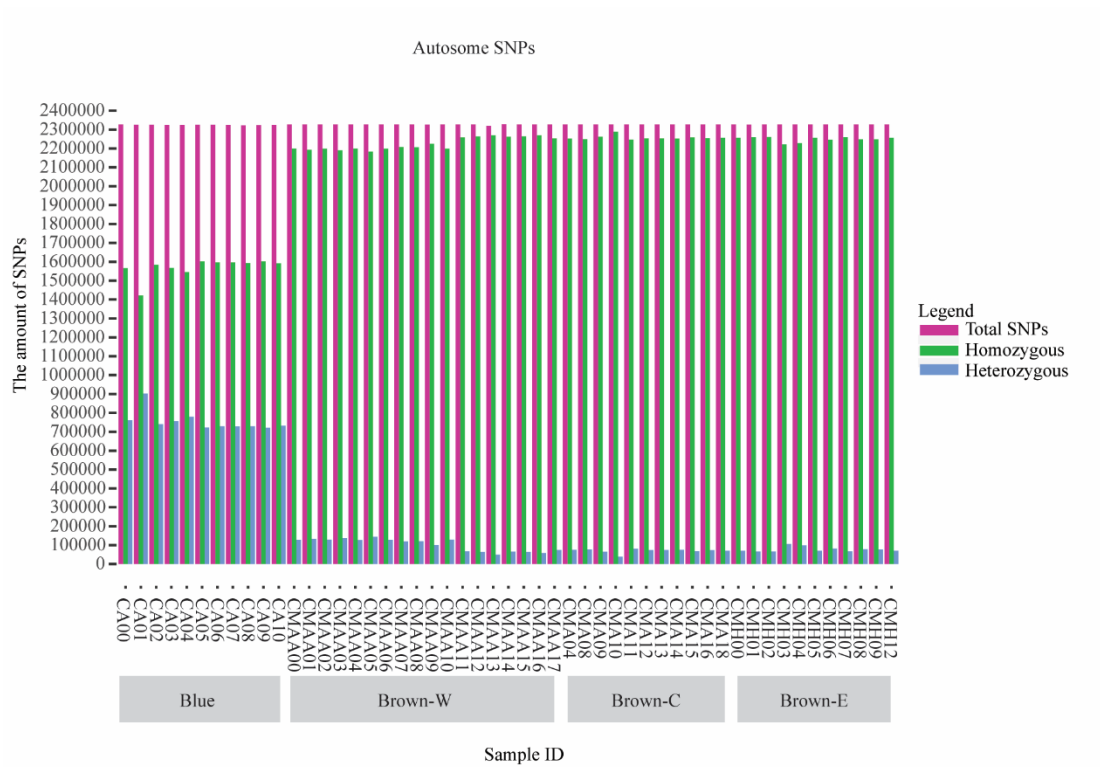
730

731 Fig. S15. Variant quality metrics of the Z chromosome. The corresponding sample of
 732 each sample index can be seen in Supplementary Dataset S11. (a) Individual percentage
 733 of high-quality sites in the BAM file (the call quality of the bases was Q20 or higher).
 734 (b) Individual percentage of sites in the BAM file with coverage greater than 5 X. (c)
 735 Individual percentage of missing SNPs. (d) Heterozygosity of each sample in the four
 736 populations. In the box plots, the bold black line represents the median, and the boxes
 737 limit the 25th and 75th percentiles of the distributions.



738

739 Fig. S16. Variant quality metrics of the mitogenome. The corresponding sample of each
 740 sample index can be seen in Supplementary Dataset S11. (a) Individual percentage of
 741 high-quality sites in the BAM file (the call quality of the bases was Q20 or higher). (b)
 742 Individual percentage of sites in the BAM file with coverage greater than 5 X. (c)
 743 Individual percentage of missing sites. (d) Average number of nucleotide differences
 744 for each population (K).



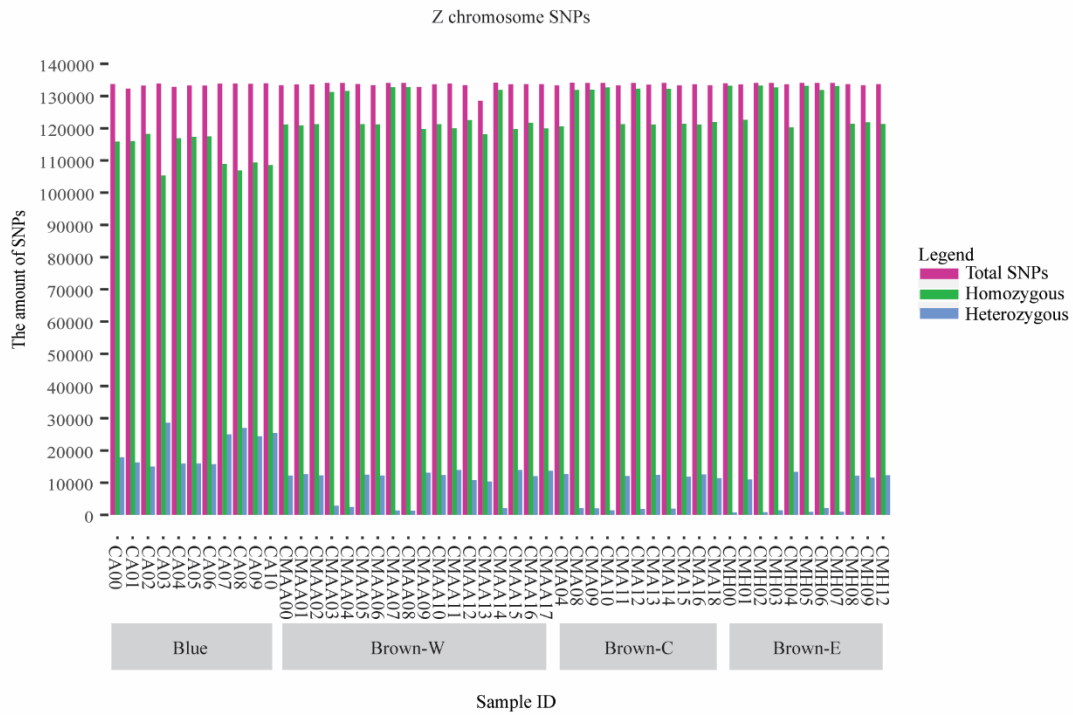
745

746 Fig. S17. Variant number of autosomes for the 51 individuals on a population-scale.

747 Sample IDs with the abbreviations Blue, Brown-W, Brown-C, and Brown-E denoted

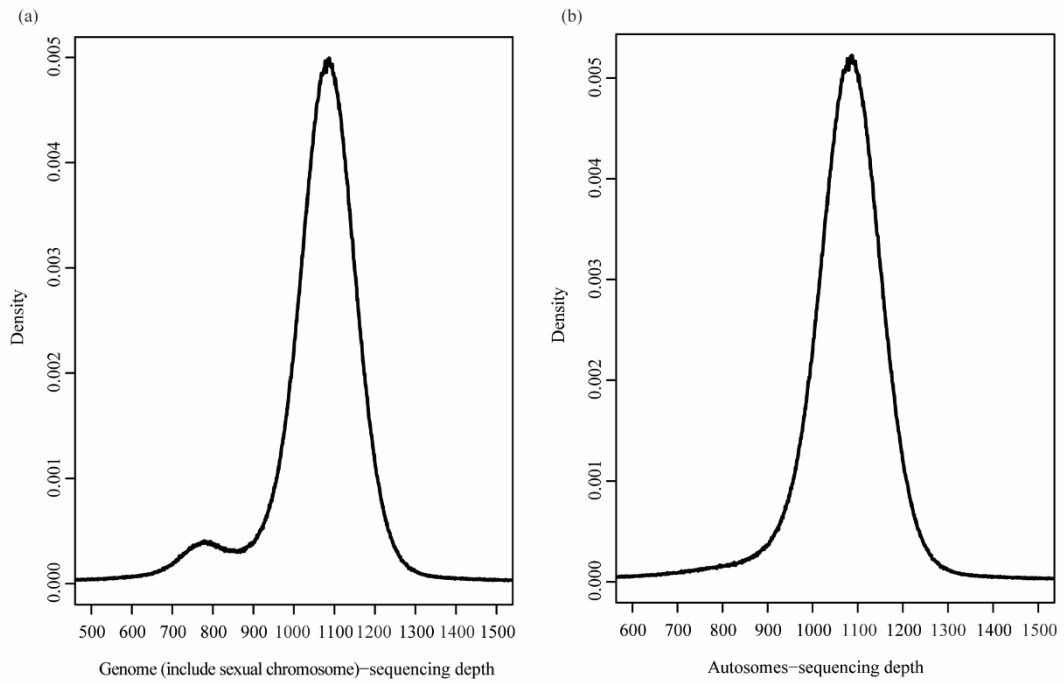
748 individuals from *Crossoptilon auritum* and the Western, Central, and Eastern

749 populations of *Crossoptilon mantchuricum*, respectively.



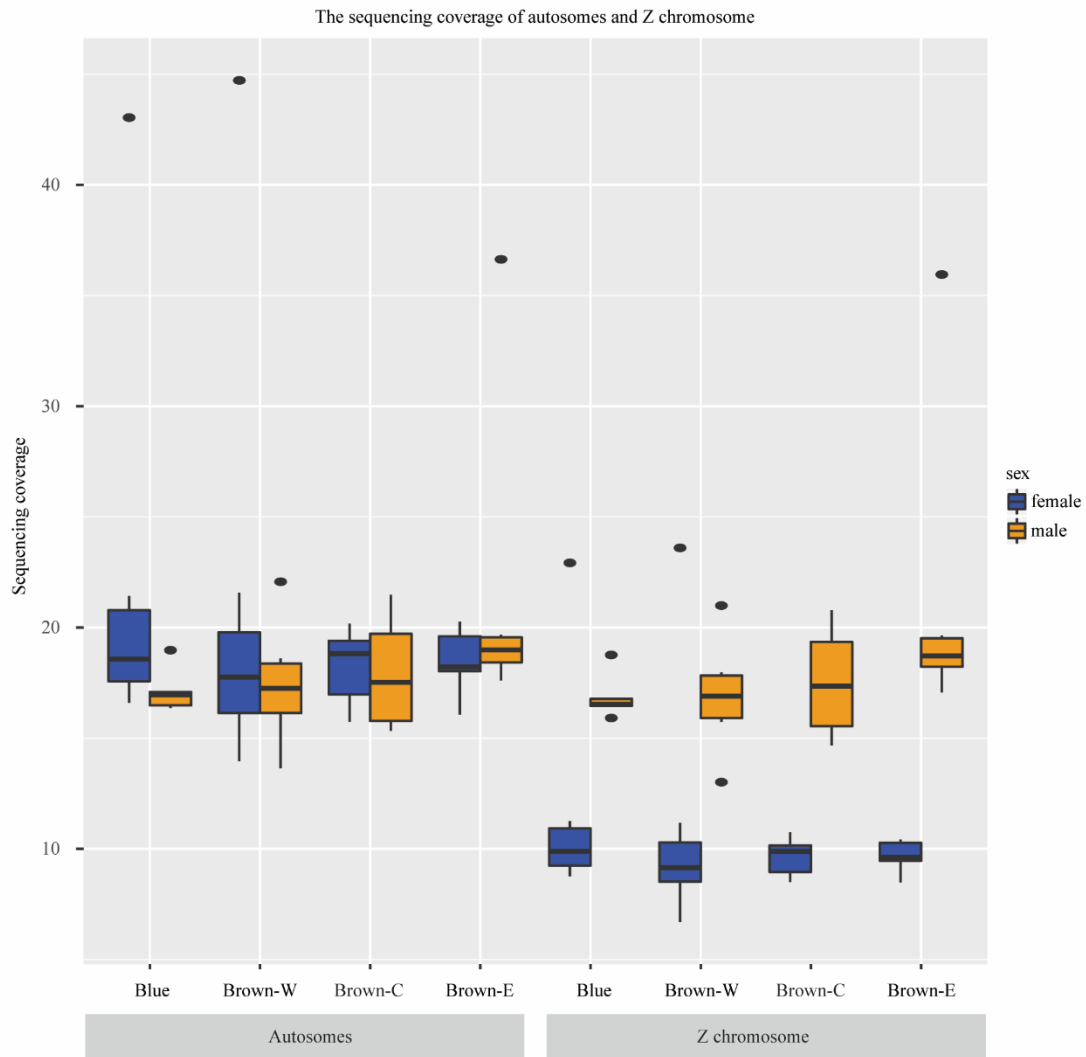
750

751 Fig. S18. Variant number of the Z chromosome for the 51 individuals on a population-
 752 scale. Sample IDs with the abbreviations Blue, Brown-W, Brown-C, and Brown-E
 753 indicate individuals from *Crossoptilon auritum* and the Western, Central, and Eastern
 754 populations of *Crossoptilon mantchuricum*, respectively.



755

756 Fig. S19. Distribution of sequencing depth across 51 individuals for (a) the whole
757 genome (including sex chromosomes) and (b) the autosomes.



758

759 Fig. S20. Sequencing coverage of males and females. Blue boxes indicate the summary
 760 of the female individuals, orange boxes indicate the summary of the male individuals.
 761 In the box plots, the bold black line represents the median, and the boxes limit the 25th
 762 and 75th percentiles of the distributions, while the points are the outlier values. The label
 763 under the plots indicates the population symbol and genomic part.

Dataset S1: Completeness evaluation results.

	Draft genome
Complete and single-copy BUSCOs (S)	4624 (94.08%)
Complete and duplicated BUSCOs (D)	52 (1.06%)
Fragmented BUSCOs (F)	134 (2.73%)
Missing BUSCOs (M)	105 (2.14%)
Total BUSCO groups searched	4915

Dataset S2: Summary of the genetic diversity (heterozygous SNP rate) of inbreeding and outbreeding birds on an autosome scale. The genetic diversity of *Crossoptilon mantchuricum* and *Crossoptilon auritum* is the average value within each isolated population. The genetic diversity of other organisms is the lowest value in the corresponding reported population.

Species	Common_Name	Class	IUCN status	Heterozygous SNP rate	Seq_Type	Source DOI
<i>Crossoptilon mantchuricum</i>	Brown eared pheasant (Western)	Aves	Vulnerable	7.33E-05	genome-wide	Curent study
<i>Crossoptilon mantchuricum</i>	Brown eared pheasant (Eastern)	Aves	Vulnerable	8.73E-05	genome-wide	Curent study
<i>Crossoptilon mantchuricum</i>	Brown eared pheasant (Center)	Aves	Vulnerable	1.37E-04	genome-wide	Curent study
<i>Haliaeetus albicilla</i>	White tailed eagle	Aves	Least Concern	4.00E-04	genome-wide	10.1186/s13059-014-0557-1
<i>Haliaeetus leucocephalus</i>	Bald eagle	Aves	Least Concern	4.30E-04	genome-wide	10.1186/s13059-014-0557-1
<i>Nipponia nippon</i>	Crested ibis	Aves	Endangered	4.30E-04	genome-wide	10.1186/s13059-014-0557-1
<i>Pelecanus crispus</i>	Dalmatian pelican	Aves	Near Threatened	6.00E-04	genome-wide	10.1186/s13059-014-0557-1
<i>Falco peregrinus</i>	Peregrine falcon	Aves	Least Concern	7.00E-04	genome-wide	10.1038/ng.2588
<i>Aptenodytes patagonicus</i>	King penguin	Aves	Least Concern	7.20E-04	RNAseq	10.1038/nature13685
<i>Eudyptes moseleyi</i>	Northern rockhopper penguin	Aves	Endangered	7.97E-04	RNAseq	10.1038/nature13685
<i>Falco cherrug</i>	Saker falcon	Aves	Endangered	8.00E-04	genome-wide	10.1038/ng.2588
<i>Crossoptilon auritum</i>	Blue eared pheasant	Aves	Least Concern	8.25E-04	genome-wide	Curent study
<i>Nestor notabilis</i>	Kea	Aves	Endangered	9.10E-04	genome-wide	10.1186/s13059-014-0557-1
<i>Eudyptes chrysocome filholi</i>	Eastern rockhopper penguin	Aves	Vulnerable	1.14E-03	RNAseq	10.1038/nature13685
<i>Ectopistes migratorius</i>	Passenger pigeon	Aves	Extinct	1.20E-03	genome-wide	10.1073/pnas.1401526111
<i>Cyanistes caeruleus</i>	Eurasian blue tit	Aves	Least Concern	1.27E-03	RNAseq	10.1038/nature13685
<i>Phalacrocorax carbo</i>	Great black cormorant	Aves	Least Concern	1.39E-03	genome-wide	10.1186/s13059-014-0557-1
<i>Taeniopygia guttata</i>	Zebra finch	Aves	Least Concern	1.40E-03	genome-wide	10.1038/nature08819

<i>Apteryx mantelli</i>	Kiwi	Aves	Vulnerable	1.50E-03	genome-wide	10.1186/s13059-015-0711-4
<i>Meleagris gallopavo</i>	Wild turkey	Aves	Least Concern	2.37E-03	genome-wide	10.1371/journal.pbio.1002112
<i>Egretta garzetta</i>	Little egret	Aves	Least Concern	2.51E-03	genome-wide	10.1186/s13059-014-0557-1
<i>Anas platyrhynchos</i>	Mallard	Aves	Least Concern	2.61E-03	genome-wide	10.1038/ng.2657
<i>Ficedula albicollis</i>	Collared flycatcher	Aves	Least Concern	3.80E-03	genome-wide	10.1371/journal.pbio.1002112
<i>Melopsittacus undulatus</i>	Budgerigar	Aves	Least Concern	4.31E-03	genome-wide	10.1186/s13059-014-0557-1
<i>Gallus gallus</i>	Chicken	Aves	Least Concern	5.88E-03	genome-wide	10.1371/journal.pbio.1002112

Dataset S3: Genetic diversity of the zero-fold degenerate sites and four-fold degenerate sites. The numbers in brackets are 95% confidence intervals, which were obtained from bootstrapping 1,000 times.

Site Class	Population	θ_{π} (%), 95% CI	θ_w (%), 95% CI	θ_{π} (zero-fold) / θ_{π} (fold-fold)
zero-fold	Blue	0.030145% [0.030136%, 0.030154%]	0.020448% [0.020442%, 0.020453%]	0.273340 [0.273220, 0.273459]
	Brown-W	0.002670% [0.002666%, 0.002675%]	0.002237% [0.002234%, 0.002240%]	0.308766 [0.307968, 0.309564]
	Brown-C	0.005474% [0.005467%, 0.005481%]	0.004528% [0.004523%, 0.004533%]	0.318080 [0.317500, 0.318661]
	Brown-E	0.003683% [0.003678%, 0.003689%]	0.002817% [0.002813%, 0.002821%]	0.369310 [0.368405, 0.370215]
four-fold	Blue	0.110287% [0.110255%, 0.110320%]	0.074654% [0.074636%, 0.074672%]	
	Brown-W	0.008657% [0.008640%, 0.008675%]	0.006903% [0.006891%, 0.006916%]	
	Brown-C	0.017218% [0.017194%, 0.017242%]	0.014164% [0.014145%, 0.014182%]	
	Brown-E	0.009983% [0.009964%, 0.010001%]	0.007687% [0.007673%, 0.007701%]	

Dataset S4: The derived loss-of-function mutations in the potential adaptive genes.

Gene ID	The frequency of derived loss-of-function (LOF) mutations				Protein ID	Protein name	The site of amino acid substitution	Function
	Blue	Brown-W	Brown-C	Brown-E				
EARP_00000097	0.8	0	0	0	EARP_00000097-RA	helix-turn-helix domain-containing protein	Q552*	regulate gene expression
EARP_00005078	0.1	1	1	1	EARP_00005078-RB	ring finger protein 146	K361*	facilitate DNA repair and protect against cell death

Dataset S5: The derived missense mutations in the potential adaptive genes. A PROVEAN score less than -2.5 indicates that the corresponding derived missense mutations were deleterious. Blue represents *Crossoptilon auritum*, while Brown-W, Brown-C, and Brown-E represent the Western, Central, and Eastern populations of *Crossoptilon mantchuricum*, respectively.

Protein ID	The site of amino acid substitution	Frequency			Proven scores	Unique fixed to <i>C. mantchuricum</i> ?	Unique fixed to <i>C. auritum</i> ?	
		Blue	Brown -W	Brown -C				Brown -E
EARP_00000044-RA	D411N	0.40	0.00	0.00	0.00	0.160	no	no
EARP_00000045-RA	R101G	0.00	1.00	1.00	1.00	0.867	yes	no
EARP_00000097-RA	A979T	0.70	0.00	0.00	0.00	-4.000	no	no
EARP_00000097-RA	C1895Y	1.00	1.00	1.00	1.00	-11.000	no	no
EARP_00000097-RA	D2118G	0.00	1.00	1.00	1.00	-7.000	yes	no
EARP_00000097-RA	D2650G	0.80	1.00	1.00	0.00	-7.000	no	no
EARP_00000097-RA	D28V	0.60	0.00	0.00	0.00	-9.000	no	no
EARP_00000097-RA	E631K	0.60	0.00	0.00	0.00	-4.000	no	no
EARP_00000097-RA	E63V	0.50	1.00	1.00	1.00	-7.000	no	no
EARP_00000097-RA	F19L	0.20	1.00	1.00	1.00	-6.000	no	no
EARP_00000097-RA	F2851S	0.40	1.00	1.00	1.00	-8.000	no	no
EARP_00000097-RA	F303Y	0.70	1.00	1.00	1.00	-3.000	no	no
EARP_00000097-RA	G1018D	0.00	1.00	1.00	0.00	-7.000	yes	no
EARP_00000097-RA	G39R	0.10	1.00	1.00	1.00	-8.000	no	no
EARP_00000097-RA	L2621F	0.00	1.00	1.00	0.00	-4.000	yes	no
EARP_00000097-RA	N777S	1.00	1.00	1.00	1.00	-5.000	no	no
EARP_00000097-RA	N785S	1.00	1.00	1.00	1.00	-5.000	no	no
EARP_00000097-RA	P1235L	0.00	1.00	1.00	1.00	-10.000	yes	no
EARP_00000097-RA	Q851K	0.00	1.00	1.00	1.00	-4.000	yes	no
EARP_00000097-RA	R29G	0.50	0.00	0.00	0.00	-7.000	no	no
EARP_00000097-RA	R29S	0.50	0.00	0.00	0.00	-6.000	no	no
EARP_00000097-RA	R364H	0.80	0.00	0.00	0.00	-5.000	no	no
EARP_00000097-RA	R374C	0.80	0.00	0.00	0.00	-8.000	no	no
EARP_00000097-RA	S1875P	0.90	1.00	1.00	1.00	-5.000	no	no
EARP_00000097-RA	S1899F	0.40	0.00	0.00	0.00	-6.000	no	no
EARP_00000097-RA	T1451I	0.90	1.00	1.00	0.00	-6.000	no	no
EARP_00000097-RA	T2466A	0.00	1.00	1.00	0.00	-5.000	yes	no
EARP_00000097-RA	T388S	0.40	1.00	1.00	1.00	-4.000	no	no
EARP_00000097-RA	V295I	0.00	1.00	1.00	1.00	-1.000	yes	no
EARP_00000097-RA	W1433C	0.70	0.00	0.00	1.00	-13.000	no	no
EARP_00000193-RA	V109A	0.30	1.00	1.00	0.00	-4.000	no	no
EARP_00000985-RA	P8L	1.00	0.00	0.00	0.00	-0.662	no	yes
EARP_00000986-RA	N62S	1.00	0.00	0.00	0.00	-0.005	no	yes

EARP_00000987-RA	R94Q	0.00	1.00	1.00	1.00	2.941	yes	no
EARP_00000987-RA	S28G	1.00	0.00	0.00	0.00	3.198	no	yes
EARP_00000989-RA	H1599N	0.00	0.00	0.00	0.50	-5.304	no	no
EARP_00000989-RA	M2378I	1.00	0.00	0.00	0.00	-0.534	no	yes
EARP_00000989-RA	S2400I	1.00	0.00	0.00	0.00	-1.674	no	yes
EARP_00002145-RB	R43G	0.70	1.00	1.00	1.00	-7.000	no	no
EARP_00002148-RA	K24E	0.70	0.00	0.00	0.00	-4.000	no	no
EARP_00002148-RA	R38H	0.60	0.00	0.00	0.00	-5.000	no	no
EARP_00002233-RA	S134N	0.90	0.00	0.00	0.00	-1.428	no	no
EARP_00002234-RB	A1163T	1.00	0.00	0.00	0.00	-2.246	no	yes
EARP_00002234-RB	L1777P	0.00	1.00	1.00	1.00	0.517	yes	no
EARP_00002634-RB	G162S	0.70	0.00	0.00	0.00	0.906	no	no
EARP_00002634-RB	T32I	1.00	0.00	0.00	0.00	0.037	no	yes
EARP_00003544-RA	A1335T	0.10	1.00	1.00	1.00	3.314	no	no
EARP_00003544-RA	L1630P	0.10	1.00	1.00	1.00	0.083	no	no
EARP_00003544-RA	S932P	0.20	1.00	1.00	1.00	2.139	no	no
EARP_00003544-RA	V988I	0.70	0.00	0.00	0.00	0.255	no	no
EARP_00003545-RC	T48A	0.90	1.00	1.00	1.00	0.433	no	no
EARP_00003547-RA	H651Q	0.70	0.00	0.00	0.00	0.401	no	no
EARP_00003548-RB	K103E	0.00	1.00	1.00	1.00	0.389	yes	no
EARP_00003548-RB	Y122D	0.00	1.00	1.00	1.00	2.078	yes	no
EARP_00005078-RB	G339R	0.20	1.00	1.00	1.00	-8.000	no	no
EARP_00005078-RB	H401R	0.10	1.00	1.00	1.00	-8.000	no	no
EARP_00005078-RB	H401Y	1.00	0.00	0.00	0.00	-6.000	no	yes
EARP_00005078-RB	N337S	0.20	1.00	1.00	1.00	-5.000	no	no
EARP_00005078-RB	Q247R	0.00	1.00	1.00	1.00	-4.000	yes	no
EARP_00005078-RB	R141W	1.00	0.00	0.00	0.00	-2.000	no	yes
EARP_00005078-RB	R444G	1.00	0.00	0.00	0.00	-1.000	no	yes
EARP_00005078-RB	S142F	0.00	0.00	0.25	0.88	-6.000	no	no
EARP_00005078-RB	T362I	0.10	1.00	1.00	1.00	-6.000	no	no
EARP_00005078-RB	V132M	1.00	0.00	0.00	0.00	-3.000	no	yes
EARP_00006312-RA	A162P	0.80	0.00	0.00	0.00	-5.000	no	no
EARP_00006312-RA	W409L	0.40	0.00	0.00	0.00	-13.000	no	no
EARP_00006312-RA	Y714H	0.20	1.00	1.00	1.00	-5.000	no	no
EARP_00006313-RA	V812A	0.40	1.00	1.00	1.00	-4.000	no	no
EARP_00006344-RA	G7D	0.70	0.00	0.00	0.00	-7.000	no	no
EARP_00006349-RB	I201V	0.60	0.00	0.00	0.00	0.427	no	no
EARP_00006349-RB	V326A	0.80	0.00	0.00	0.00	-0.455	no	no
EARP_00006356-RA	A95T	0.00	1.00	1.00	0.75	-0.252	yes	no
EARP_00006357-RC	V86L	0.00	1.00	1.00	1.00	0.494	yes	no
EARP_00006614-RD	E383K	1.00	0.00	0.00	0.00	-1.865	no	yes

EARP_00007488-RA	I62V	1.00	1.00	1.00	1.00	0.054	no	no
EARP_00007490-RA	E1110K	0.00	0.00	1.00	0.00	-0.487	yes	no
EARP_00007490-RA	V5M	0.40	0.00	0.00	0.00	-0.106	no	no
EARP_00007491-RA	R77S	0.00	1.00	1.00	1.00	-6.000	yes	no
EARP_00007492-RB	G579R	0.70	0.00	0.00	0.00	-7.828	no	no
EARP_00007492-RB	H1120N	1.00	0.00	0.00	0.00	-1.144	no	yes
EARP_00007492-RB	L479P	0.00	1.00	1.00	1.00	3.457	yes	no
EARP_00007492-RB	P353H	0.00	1.00	1.00	1.00	1.750	yes	no
EARP_00007492-RB	R559G	0.00	1.00	1.00	1.00	3.443	yes	no
EARP_00007492-RB	V418E	1.00	0.00	0.00	0.00	-5.531	no	yes
EARP_00007494-RB	L12V	1.00	1.00	1.00	1.00	-0.313	no	no
EARP_00007509-RD	L543P	0.00	1.00	1.00	1.00	1.766	yes	no
EARP_00007509-RD	M56L	0.00	1.00	1.00	1.00	1.702	yes	no
EARP_00007510-RC	A6V	1.00	0.00	0.00	0.00	-0.138	no	yes
EARP_00007511-RA	E41K	1.00	0.00	0.00	0.00	-4.000	no	yes
EARP_00007511-RC	G1815D	0.00	0.36	0.13	1.00	0.107	yes	no
EARP_00008168-RA	L1075F	0.00	1.00	1.00	1.00	1.431	yes	no
EARP_00008168-RA	L1144S	0.00	1.00	1.00	1.00	0.786	yes	no
EARP_00008598-RA	D338E	0.90	0.00	0.00	0.00	0.255	no	no
EARP_00008600-RA	V4M	0.60	1.00	1.00	1.00	1.026	no	no
EARP_00008602-RC	A34V	0.70	0.00	0.00	0.00	-0.068	no	no
EARP_00008602-RA	G51R	0.50	0.00	0.00	0.00	0.355	no	no
EARP_00008602-RC	L22W	0.10	1.00	1.00	1.00	-0.259	no	no
EARP_00008602-RC	T26A	0.50	0.00	0.00	0.00	-0.357	no	no
EARP_00008602-RA	Y78C	0.10	1.00	1.00	1.00	-0.378	no	no
EARP_00008602-RA	Y78H	0.10	1.00	1.00	1.00	0.000	no	no
EARP_00008603-RB	I842V	0.70	0.00	0.00	0.00	0.452	no	no
EARP_00008603-RB	T311I	0.00	1.00	0.25	0.00	3.975	yes	no
EARP_00008603-RB	V382M	0.20	0.00	0.00	1.00	-1.846	no	no
EARP_00009935-RA	F57S	0.30	1.00	1.00	1.00	-8.000	no	no
EARP_00009935-RA	V65M	0.90	0.00	0.00	0.00	-3.000	no	no
EARP_00010173-RA	A428V	0.00	1.00	0.50	0.00	-2.152	yes	no
EARP_00010175-RB	G285E	1.00	0.00	0.00	0.00	0.133	no	yes
EARP_00010175-RB	T949I	1.00	0.00	0.00	0.00	-0.011	no	yes
EARP_00011418-RA	L1448M	0.00	0.00	0.88	0.00	1.408	no	no
EARP_00011419-RB	T208A	0.70	0.00	0.00	0.00	0.000	no	no
EARP_00011419-RB	T446A	0.70	0.00	0.00	0.00	-0.356	no	no
EARP_00011586-RC	G208A	0.00	1.00	1.00	0.00	0.214	yes	no
EARP_00011586-RC	V33I	0.10	0.00	0.00	1.00	-0.297	no	no
EARP_00012749-RA	M467T	1.00	0.00	0.00	0.00	0.574	no	yes
EARP_00012749-RA	S3Y	0.80	0.00	0.00	0.00	-1.320	no	no

EARP_00012844-RA	T1112P	0.00	1.00	1.00	1.00	-0.129	yes	no
EARP_00012844-RA	T1198A	0.00	1.00	1.00	1.00	0.431	yes	no
EARP_00012846-RA	S376N	0.60	0.00	0.00	0.00	0.303	no	no
EARP_00012846-RA	V1344E	0.00	1.00	1.00	1.00	1.988	yes	no
EARP_00012848-RA	P104S	0.20	0.00	1.00	0.00	-4.667	no	no
EARP_00012944-RA	L33F	0.00	1.00	1.00	1.00	-0.391	yes	no
EARP_00013131-RA	R511Q	0.70	0.00	0.00	0.00	0.000	no	no
EARP_00013132-RA	I120V	1.00	0.00	0.00	0.00	1.000	no	yes
EARP_00013132-RA	L474S	0.00	1.00	1.00	1.00	6.000	yes	no
EARP_00013132-RA	M332V	0.00	1.00	1.00	1.00	3.000	yes	no
EARP_00013133-RA	P1884S	0.80	0.00	0.00	0.00	0.337	no	no
EARP_00013133-RA	S1767L	0.80	0.00	0.00	0.00	-1.419	no	no
EARP_00013133-RA	T2351A	1.00	0.00	0.00	0.00	-0.568	no	yes
EARP_00013133-RA	V2114A	0.80	0.00	0.13	0.00	0.655	no	no
EARP_00013134-RA	T711I	0.00	1.00	1.00	1.00	-0.500	yes	no
EARP_00013135-RA	T488K	0.40	0.00	0.00	0.00	-6.000	no	no
EARP_00013137-RA	I205M	0.00	1.00	1.00	1.00	1.217	yes	no
EARP_00013138-RA	T664A	0.60	1.00	1.00	1.00	-2.663	no	no
EARP_00013139-RA	E1834K	0.40	0.00	0.00	0.00	-0.870	no	no
EARP_00013139-RA	H1168Q	0.40	0.00	0.00	0.00	1.333	no	no
EARP_00013139-RA	I742V	0.80	1.00	1.00	1.00	0.467	no	no
EARP_00013139-RA	Q44R	0.00	1.00	1.00	1.00	-0.391	yes	no
EARP_00013139-RA	T1428A	0.50	0.00	0.00	0.00	-0.216	no	no
EARP_00013142-RA	T43S	1.00	0.00	0.00	0.00	-4.000	no	yes
EARP_00013786-RA	F42S	0.00	1.00	1.00	1.00	1.747	yes	no
EARP_00014053-RA	R263C	0.00	0.00	0.13	0.50	-3.196	no	no
EARP_00014072-RA	Y80F	0.70	1.00	0.88	1.00	-0.948	no	no
EARP_00014073-RD	R56H	1.00	0.00	0.00	0.00	-5.000	no	yes
EARP_00014240-RA	V119M	0.10	1.00	1.00	1.00	-0.100	no	no
EARP_00014598-RB	E7K	1.00	0.00	0.00	0.00	-0.036	no	yes
EARP_00014598-RB	V1549I	0.00	1.00	1.00	1.00	-0.021	yes	no
EARP_00015100-RA	A116E	0.00	1.00	1.00	1.00	0.886	yes	no
EARP_00015100-RA	E97D	0.90	0.00	0.00	0.00	-0.202	no	no
EARP_00015100-RA	G177D	1.00	0.00	0.00	0.00	1.921	no	yes
EARP_00015100-RA	L9P	0.00	1.00	1.00	1.00	-0.022	yes	no
EARP_00015807-RA	L58V	0.00	1.00	1.00	1.00	-3.000	yes	no
EARP_00015807-RA	L78P	0.10	1.00	1.00	1.00	-7.000	no	no
EARP_00015808-RA	C10R	1.00	0.00	0.00	0.00	0.349	no	yes
EARP_00015808-RA	H23Y	0.00	1.00	1.00	1.00	-0.193	yes	no
EARP_00015808-RA	L484P	0.00	1.00	1.00	1.00	3.432	yes	no
EARP_00016036-RA	P908S	0.00	1.00	1.00	0.50	1.395	yes	no

EARP_00016471-RA	E1545K	0.90	1.00	1.00	1.00	0.792	no	no
EARP_00016471-RA	Q1996H	0.50	1.00	1.00	1.00	-0.655	no	no
EARP_00016471-RA	V4I	0.00	1.00	1.00	1.00	-0.355	yes	no
EARP_00017025-RF	A628G	0.00	1.00	1.00	1.00	-0.100	yes	no
EARP_00017025-RF	R140H	0.70	0.00	0.00	0.00	-0.158	no	no
EARP_00017027-RB	S286P	0.70	0.00	0.00	0.00	-0.730	no	no
EARP_00017032-RA	V374I	1.00	0.00	0.00	0.00	0.314	no	yes
EARP_00017036-RA	E450D	0.10	1.00	1.00	1.00	-1.894	no	no
EARP_00017036-RA	R488C	0.90	0.00	0.00	0.00	0.244	no	no
EARP_00017036-RA	V85I	0.20	1.00	1.00	1.00	0.964	no	no
EARP_00017037-RD	T735A	0.30	1.00	1.00	1.00	0.873	no	no
EARP_00017072-RA	Q136R	0.00	1.00	1.00	1.00	-0.347	yes	no
EARP_00017811-RA	L136I	0.40	1.00	1.00	1.00	0.386	no	no
EARP_00017811-RA	M336I	1.00	1.00	1.00	0.00	-1.846	no	no
EARP_00017811-RA	V1139A	1.00	1.00	1.00	1.00	0.466	no	no

Dataset S6: Genetic diversity of the potential adaptive genes. The genetic diversity is the average value of sites-pi within each gene. Blue represents *Crossoptilon auritum*. Brown-W, Brown-C, and Brown-E represent the Western, Central, and Eastern populations of *Crossoptilon mantchuricum*, respectively.

Gene ID	Genetic diversity (II)			
	Blue	Brown-W	Brown-C	Brown-E
EARP_00000007	0.00	0.00	0.00	0.00
EARP_00000044	0.49	0.00	0.00	0.00
EARP_00000045	0.18	0.02	0.02	0.04
EARP_00000046	0.17	0.03	0.02	0.11
EARP_00000047	0.24	0.02	0.02	0.09
EARP_00000048	0.00	0.00	0.00	0.00
EARP_00000097	0.32	0.00	0.01	0.00
EARP_00000191	0.05	0.52	0.46	0.12
EARP_00000192	0.29	0.00	0.00	0.06
EARP_00000193	0.20	0.07	0.07	0.04
EARP_00000194	0.29	0.00	0.00	0.00
EARP_00009935	0.31	0.00	0.00	0.00
EARP_00010171	0.00	0.00	0.00	0.00
EARP_00010172	0.48	0.00	0.00	0.00
EARP_00010173	0.21	0.03	0.03	0.00
EARP_00010174	0.10	0.00	0.00	0.00
EARP_00010175	0.23	0.02	0.02	0.00
EARP_00002145	0.34	0.00	0.01	0.01
EARP_00002146	0.18	0.00	0.00	0.00
EARP_00002147	0.02	0.00	0.15	0.00
EARP_00002148	0.51	0.00	0.00	0.00
EARP_00002231	0.15	0.00	0.05	0.21
EARP_00002232	0.25	0.00	0.06	0.05
EARP_00002233	0.32	0.00	0.05	0.05
EARP_00002234	0.15	0.00	0.06	0.05
EARP_00002248	0.17	0.03	0.04	0.01
EARP_00002249	0.31	0.00	0.00	0.01
EARP_00002250	0.30	0.00	0.00	0.00
EARP_00011416	0.30	0.00	0.00	0.01
EARP_00011418	0.14	0.00	0.00	0.00
EARP_00011419	0.40	0.00	0.00	0.00
EARP_00011420	0.26	0.00	0.00	0.00
EARP_00011586	0.19	0.00	0.00	0.00
EARP_00011725	0.36	0.00	0.06	0.00
EARP_00002633	0.28	0.00	0.00	0.00
EARP_00002634	0.30	0.00	0.00	0.00
EARP_00002780	0.25	0.00	0.00	0.03

EARP_00002781	0.00	0.00	0.00	0.00
EARP_00002782	0.23	0.00	0.00	0.00
EARP_00002783	0.10	0.00	0.00	0.44
EARP_00012228	0.00	0.00	0.00	0.00
EARP_00012748	0.35	0.00	0.00	0.01
EARP_00012749	0.26	0.00	0.03	0.02
EARP_00012844	0.32	0.01	0.00	0.00
EARP_00012845	0.00	0.00	0.00	0.00
EARP_00012846	0.19	0.00	0.00	0.00
EARP_00012847	0.28	0.00	0.00	0.00
EARP_00012848	0.22	0.01	0.10	0.00
EARP_00012849	0.00	0.00	0.00	0.00
EARP_00012850	0.23	0.00	0.08	0.00
EARP_00012816	0.19	0.00	0.00	0.00
EARP_00012944	0.29	0.00	0.00	0.00
EARP_00012945	0.18	0.00	0.00	0.00
EARP_00012946	0.00	0.00	0.00	0.00
EARP_00013130	0.28	0.00	0.03	0.00
EARP_00013131	0.26	0.00	0.02	0.00
EARP_00013132	0.00	0.00	0.00	0.00
EARP_00013133	0.20	0.00	0.02	0.00
EARP_00013134	0.18	0.00	0.01	0.00
EARP_00013135	0.23	0.00	0.02	0.00
EARP_00013136	0.00	0.00	0.00	0.00
EARP_00013137	0.07	0.00	0.05	0.00
EARP_00013138	0.31	0.00	0.02	0.00
EARP_00013139	0.41	0.00	0.01	0.00
EARP_00013142	0.28	0.01	0.01	0.00
EARP_00003307	0.38	0.00	0.00	0.01
EARP_00013446	0.31	0.00	0.00	0.00
EARP_00013447	0.17	0.00	0.00	0.00
EARP_00013448	0.13	0.00	0.00	0.00
EARP_00013449	0.30	0.00	0.00	0.00
EARP_00013450	0.28	0.00	0.00	0.00
EARP_00003347	0.41	0.00	0.00	0.00
EARP_00003349	0.09	0.00	0.00	0.04
EARP_00013785	0.21	0.00	0.05	0.00
EARP_00013786	0.15	0.00	0.00	0.00
EARP_00013787	0.36	0.00	0.05	0.00
EARP_00000455	0.14	0.00	0.00	0.00
EARP_00003474	0.00	0.00	0.00	0.00
EARP_00013794	0.22	0.00	0.00	0.02
EARP_00014053	0.21	0.00	0.01	0.05
EARP_00014054	0.44	0.00	0.00	0.00

EARP_00014069	0.27	0.00	0.02	0.00
EARP_00014070	0.00	0.00	0.00	0.00
EARP_00014071	0.22	0.00	0.17	0.00
EARP_00014072	0.32	0.00	0.41	0.00
EARP_00014073	0.14	0.05	0.05	0.04
EARP_00014074	0.11	0.03	0.34	0.17
EARP_00014075	0.00	0.00	0.50	0.23
EARP_00003544	0.40	0.00	0.00	0.00
EARP_00003545	0.33	0.00	0.00	0.00
EARP_00003546	0.25	0.00	0.00	0.00
EARP_00003547	0.32	0.01	0.02	0.00
EARP_00003548	0.18	0.04	0.10	0.00
EARP_00003549	0.20	0.00	0.00	0.00
EARP_00003550	0.17	0.29	0.07	0.00
EARP_00014238	0.20	0.00	0.03	0.01
EARP_00014239	0.25	0.02	0.03	0.01
EARP_00014240	0.14	0.00	0.01	0.00
EARP_00014241	0.07	0.00	0.00	0.00
EARP_00014242	0.19	0.00	0.00	0.00
EARP_00014598	0.18	0.00	0.06	0.00
EARP_00014599	0.36	0.00	0.10	0.00
EARP_00014600	0.32	0.01	0.00	0.00
EARP_00014601	0.25	0.00	0.00	0.00
EARP_00015100	0.10	0.01	0.02	0.00
EARP_00015101	0.25	0.01	0.02	0.02
EARP_00015102	0.36	0.00	0.00	0.00
EARP_00015103	0.36	0.00	0.00	0.00
EARP_00003923	0.35	0.00	0.03	0.00
EARP_00015598	0.00	0.00	0.00	0.00
EARP_00015599	0.00	0.00	0.00	0.00
EARP_00015806	0.28	0.22	0.00	0.18
EARP_00015807	0.24	0.03	0.00	0.01
EARP_00015808	0.23	0.00	0.00	0.00
EARP_00016029	0.06	0.12	0.18	0.13
EARP_00016030	0.00	0.00	0.00	0.00
EARP_00016031	0.19	0.00	0.00	0.00
EARP_00016032	0.13	0.10	0.13	0.13
EARP_00016033	0.14	0.09	0.08	0.13
EARP_00016034	0.13	0.12	0.18	0.00
EARP_00016035	0.19	0.00	0.00	0.00
EARP_00016036	0.09	0.04	0.06	0.20
EARP_00016102	0.12	0.00	0.00	0.00
EARP_00000574	0.10	0.00	0.00	0.00
EARP_00000596	0.16	0.00	0.05	0.03

EARP_00000597	0.25	0.04	0.04	0.04
EARP_00000598	0.00	0.00	0.00	0.00
EARP_00004745	0.26	0.00	0.01	0.03
EARP_00004746	0.31	0.00	0.00	0.53
EARP_00016471	0.33	0.00	0.01	0.00
EARP_00016472	0.31	0.00	0.00	0.00
EARP_00016473	0.27	0.00	0.00	0.00
EARP_00016474	0.22	0.00	0.00	0.00
EARP_00016475	0.27	0.00	0.02	0.00
EARP_00017025	0.32	0.00	0.00	0.00
EARP_00017026	0.41	0.00	0.00	0.00
EARP_00017027	0.27	0.00	0.00	0.02
EARP_00017028	0.34	0.00	0.00	0.00
EARP_00017029	0.16	0.00	0.00	0.02
EARP_00017030	0.02	0.00	0.00	0.00
EARP_00017031	0.30	0.00	0.00	0.00
EARP_00017032	0.37	0.00	0.00	0.00
EARP_00017033	0.38	0.00	0.00	0.05
EARP_00017034	0.13	0.00	0.20	0.00
EARP_00017035	0.00	0.00	0.00	0.00
EARP_00017036	0.25	0.00	0.00	0.00
EARP_00017037	0.27	0.00	0.03	0.02
EARP_00017038	0.04	0.00	0.05	0.00
EARP_00017066	0.20	0.00	0.01	0.00
EARP_00017067	0.26	0.00	0.06	0.00
EARP_00017068	0.18	0.00	0.00	0.00
EARP_00017069	0.39	0.00	0.01	0.00
EARP_00017070	0.34	0.00	0.01	0.00
EARP_00017071	0.29	0.00	0.00	0.00
EARP_00017072	0.37	0.01	0.01	0.00
EARP_00005078	0.17	0.00	0.08	0.16
EARP_00005079	0.13	0.00	0.00	0.00
EARP_00005080	0.30	0.00	0.00	0.00
EARP_00017302	0.43	0.00	0.00	0.00
EARP_00017303	0.24	0.00	0.00	0.07
EARP_00017606	0.00	0.00	0.00	0.00
EARP_00017607	0.09	0.00	0.00	0.00
EARP_00017698	0.00	0.00	0.00	0.00
EARP_00017699	0.25	0.00	0.00	0.00
EARP_00017707	0.44	0.00	0.00	0.00
EARP_00017708	0.17	0.05	0.14	0.00
EARP_00017709	0.24	0.02	0.05	0.00
EARP_00017809	0.10	0.00	0.00	0.01
EARP_00017810	0.06	0.00	0.00	0.00

EARP_00017811	0.08	0.00	0.00	0.00
EARP_00017812	0.10	0.00	0.00	0.00
EARP_00006312	0.33	0.00	0.03	0.00
EARP_00006313	0.31	0.00	0.03	0.00
EARP_00006329	0.36	0.00	0.00	0.00
EARP_00006344	0.34	0.00	0.11	0.00
EARP_00006349	0.23	0.00	0.01	0.27
EARP_00006350	0.31	0.00	0.03	0.06
EARP_00006351	0.26	0.00	0.00	0.02
EARP_00006352	0.00	0.00	0.00	0.00
EARP_00006353	0.04	0.00	0.02	0.43
EARP_00006354	0.17	0.00	0.00	0.33
EARP_00006355	0.14	0.00	0.01	0.15
EARP_00006356	0.28	0.00	0.03	0.17
EARP_00006357	0.19	0.00	0.13	0.00
EARP_00000889	0.23	0.00	0.00	0.26
EARP_00006613	0.25	0.00	0.02	0.00
EARP_00006614	0.24	0.00	0.02	0.00
EARP_00006615	0.00	0.00	0.00	0.00
EARP_00006616	0.38	0.00	0.00	0.00
EARP_00018561	0.11	0.00	0.00	0.00
EARP_00018562	0.26	0.00	0.00	0.00
EARP_00018563	0.00	0.00	0.00	0.00
EARP_00018564	0.00	0.00	0.00	0.00
EARP_00006824	0.34	0.06	0.06	0.00
EARP_00006825	0.19	0.07	0.08	0.00
EARP_00000981	0.28	0.28	0.13	0.11
EARP_00000982	0.26	0.00	0.00	0.04
EARP_00000983	0.13	0.07	0.03	0.00
EARP_00000984	0.17	0.03	0.01	0.01
EARP_00000985	0.00	0.00	0.00	0.00
EARP_00000986	0.12	0.08	0.04	0.07
EARP_00000987	0.14	0.04	0.02	0.03
EARP_00000988	0.13	0.00	0.00	0.04
EARP_00000989	0.14	0.03	0.02	0.02
EARP_00000990	0.20	0.03	0.01	0.02
EARP_00007488	0.38	0.00	0.04	0.00
EARP_00007489	0.35	0.00	0.09	0.00
EARP_00007490	0.29	0.01	0.04	0.00
EARP_00007491	0.16	0.00	0.03	0.00
EARP_00007492	0.13	0.00	0.03	0.00
EARP_00007493	0.24	0.00	0.02	0.00
EARP_00007494	0.27	0.00	0.02	0.00
EARP_00007499	0.28	0.03	0.08	0.00

EARP_00007507	0.14	0.14	0.06	0.00
EARP_00007508	0.00	0.00	0.00	0.00
EARP_00007509	0.07	0.01	0.03	0.00
EARP_00007510	0.14	0.01	0.00	0.00
EARP_00007511	0.04	0.01	0.02	0.00
EARP_00007512	0.06	0.01	0.02	0.00
EARP_00007520	0.00	0.00	0.00	0.00
EARP_00007521	0.20	0.00	0.01	0.00
EARP_00007522	0.20	0.00	0.00	0.00
EARP_00007523	0.15	0.09	0.11	0.00
EARP_00018845	0.52	0.00	0.00	0.00
EARP_00018891	0.48	0.00	0.00	0.00
EARP_00018892	0.00	0.00	0.00	0.00
EARP_00018893	0.46	0.00	0.00	0.00
EARP_00018894	0.37	0.00	0.00	0.00
EARP_00018895	0.00	0.00	0.00	0.00
EARP_00018896	0.47	0.00	0.00	0.00
EARP_00018902	0.05	0.00	0.00	0.00
EARP_00008168	0.23	0.00	0.16	0.00
EARP_00008169	0.09	0.00	0.02	0.00
EARP_00008170	0.00	0.00	0.00	0.00
EARP_00008578	0.19	0.08	0.06	0.00
EARP_00008598	0.33	0.00	0.01	0.00
EARP_00008599	0.31	0.00	0.02	0.00
EARP_00008600	0.24	0.00	0.07	0.00
EARP_00008601	0.37	0.00	0.02	0.00
EARP_00008602	0.21	0.00	0.09	0.00
EARP_00008603	0.29	0.00	0.06	0.00
EARP_00008888	0.23	0.09	0.02	0.10
EARP_00011127	0.00	0.00	0.00	0.00
EARP_00011128	0.00	0.00	0.00	0.00
EARP_00011129	0.15	0.03	0.01	0.00
EARP_00011680	0.05	0.00	0.00	0.00
EARP_00011865	0.12	0.00	0.01	0.03
EARP_00011866	0.05	0.00	0.00	0.00
EARP_00011867	0.15	0.00	0.00	0.00
EARP_00011868	0.02	0.00	0.00	0.05
EARP_00011869	0.07	0.00	0.00	0.00
EARP_00012502	0.00	0.00	0.00	0.05
EARP_00012505	0.16	0.00	0.00	0.00
EARP_00012506	0.00	0.00	0.00	0.00
EARP_00012507	0.00	0.00	0.00	0.00
EARP_00012508	0.07	0.00	0.00	0.00
EARP_00012509	0.22	0.00	0.00	0.00

EARP_00012510	0.10	0.00	0.00	0.01
EARP_00012511	0.18	0.04	0.07	0.01
EARP_00012512	0.20	0.00	0.00	0.00
EARP_00012513	0.13	0.00	0.00	0.01
EARP_00012514	0.31	0.00	0.00	0.00
EARP_00012515	0.07	0.00	0.00	0.00
EARP_00012516	0.01	0.00	0.00	0.00
EARP_00012517	0.05	0.00	0.00	0.01
EARP_00012518	0.07	0.00	0.00	0.00
EARP_00012519	0.12	0.00	0.00	0.00
EARP_00012520	0.13	0.00	0.00	0.00
EARP_00012521	0.09	0.00	0.00	0.01
EARP_00013700	0.20	0.00	0.00	0.00
EARP_00013701	0.22	0.00	0.00	0.00
EARP_00013702	0.00	0.00	0.00	0.00
EARP_00013703	0.02	0.00	0.00	0.00
EARP_00013704	0.12	0.07	0.06	0.03
EARP_00014256	0.10	0.00	0.00	0.00
EARP_00014257	0.12	0.00	0.01	0.01
EARP_00014258	0.10	0.00	0.00	0.02
EARP_00014259	0.12	0.00	0.01	0.02
EARP_00014269	0.37	0.00	0.00	0.00
EARP_00014270	0.32	0.17	0.18	0.18
EARP_00015298	0.22	0.00	0.00	0.00
EARP_00015299	0.10	0.00	0.00	0.00
EARP_00015303	0.21	0.00	0.01	0.00
EARP_00015304	0.05	0.00	0.00	0.03
EARP_00015305	0.14	0.00	0.00	0.01
EARP_00015306	0.11	0.00	0.00	0.00
EARP_00015426	0.27	0.00	0.00	0.01
EARP_00005020	0.34	0.00	0.00	0.00
EARP_00005021	0.00	0.00	0.00	0.00
EARP_00005022	0.05	0.00	0.04	0.00
EARP_00005028	0.06	0.01	0.02	0.00
EARP_00017436	0.00	0.00	0.00	0.00
EARP_00017437	0.05	0.00	0.00	0.02
EARP_00017591	0.12	0.01	0.01	0.01
EARP_00017592	0.08	0.00	0.00	0.00
EARP_00017593	0.28	0.31	0.30	0.24
EARP_00006966	0.17	0.00	0.00	0.00
EARP_00006967	0.10	0.00	0.00	0.00
EARP_00008636	0.20	0.00	0.00	0.01
EARP_00008638	0.00	0.00	0.00	0.00
EARP_00008639	0.07	0.00	0.00	0.00

EARP_00008640	0.00	0.00	0.00	0.00
EARP_00008641	0.00	0.00	0.00	0.00
EARP_00008643	0.26	0.00	0.00	0.02
EARP_00008644	0.10	0.18	0.20	0.00
EARP_00008645	0.08	0.00	0.00	0.00
EARP_00008646	0.08	0.00	0.00	0.00
EARP_00009516	0.31	0.00	0.02	0.00
Average	0.19	0.01	0.03	0.02

Dataset S7: Likelihood analysis of different evolutionary scenarios. In the models, S represents consistent small population size, D represents population decline, I indicates that there was no gene flow between adjacent populations, WCE indicates that there was gene flow between adjacent populations, CW indicates that the gene flow only existed between the Western and Central populations, CE indicates that migration only occurred between the Central and Eastern populations, and IN indicates that inbreeding occurred. The likelihood (ln) was estimated using fastsimcoal2. d represents the number of parameters in the different models. AIC represents the Akaike information criterion. w represents Akaike weight.

Models	ln(Likelihood)	d	AIC _i	Δ_i	w _i
S+I	-47915.76	9.00	95849.52	6566.44	0.00
S+WCE	-45039.69	23.00	90125.37	842.30	0.00
S+CW	-45231.52	16.00	90495.04	1211.96	0.00
S+CE	-44783.52	16.00	89599.04	315.97	0.00
D+I	-44617.25	28.00	89290.49	7.42	0.02
D+WCE	-54029.39	34.00	108126.79	18843.71	0.00
D+CW	-44689.21	31.00	89440.41	157.34	0.00
D+CE	-44653.58	31.00	89369.16	86.09	0.00
S+I+IN	-47926.01	12.00	95876.01	6592.94	0.00
S+WCE+IN	-45031.20	26.00	90114.40	831.32	0.00
S+CW+IN	-45232.04	19.00	90502.07	1219.00	0.00
S+CE+IN	-44771.16	19.00	89580.32	297.25	0.00
D+I+IN	-44610.54	31.00	89283.07	0.00	0.98
D+WCE+IN	-47953.29	37.00	95980.58	6697.50	0.00
D+CW+IN	-44692.62	34.00	89453.24	170.16	0.00
D+CE+IN	-44657.87	34.00	89383.75	100.67	0.00

Dataset S8: Model probability estimated in Migrate-N. Brown-W, Brown-C, and Brown-E indicate the Western, Central, and Eastern populations of *Crossoptilon mantchuricum*, respectively. ml is the marginal likelihood. bf is the Bayes factor.

Model	Log(ml)	Log(bf)	Model-probability	Custom migration model
Migration between adjacent populations	-1403.43	-7.87	0.0004	Brown-W, Brown-C, Brown-E={**0 *** 0**}
Migration between Brown-W and Brown-C	-1402.01	-6.45	0.0015	Brown-W, Brown-C, Brown-E={**0 **0 00*}
Migration between Brown-C and Brown-E	-1398.59	-3.03	0.046	Brown-W, Brown-C, Brown-E={*00 0** 0**}
Isolation	-1395.56	0	0.9521	Brown-W, Brown-C, Brown-E={*00 0*0 00*}

Dataset S9: Sample information, coverage, and mapping rate. The asterisks mark the individuals that were excluded from the population genetic analysis because they had a close relationship (duplicate/MZ twin, 1st-degree) with at least one individual in the corresponding population. The sequence data of all individuals were deposited in the National Genomics Data Center (<https://bigd.big.ac.cn/?lang=en>). Readers can use the accession number to find the data in the National Genomics Data Center Databases.

Species	Location	Tissue type	Gender	Sample index	Sample ID	Sequencing type	Sequencing coverage (×)	Mapping rate	Accession number
<i>C. auritum</i>	Gansu	Blood	Female	1	CA00	Re-sequencing	41.57	98.53%	SAMC226688
<i>C. auritum</i>	Gansu	Blood	Female	2	CA01	Re-sequencing	16.03	97.86%	SAMC226689
<i>C. auritum</i>	Gansu	Blood	Male	3	CA02	Re-sequencing	17.06	98.48%	SAMC226690
<i>C. auritum</i>	Gansu	Blood	Female	4	CA03*	Re-sequencing	16.73	98.24%	SAMC226691
<i>C. auritum</i>	Gansu	Blood	Female	5	CA04	Re-sequencing	20.68	98.49%	SAMC226692
<i>C. auritum</i>	Gansu	Blood	Female	6	CA05	Re-sequencing	17.67	98.05%	SAMC226693
<i>C. auritum</i>	Gansu	Blood	Male	7	CA06	Re-sequencing	18.96	98.58%	SAMC226694
<i>C. auritum</i>	Gansu	Blood	Male	8	CA07	Re-sequencing	16.48	98.74%	SAMC226695
<i>C. auritum</i>	Gansu	Blood	Male	9	CA08	Re-sequencing	16.33	98.68%	SAMC226696
<i>C. auritum</i>	Gansu	Blood	Male	10	CA09	Re-sequencing	16.93	98.54%	SAMC226697
<i>C. auritum</i>	Gansu	Blood	Female	11	CA10	Re-sequencing	18.20	98.09%	SAMC226698
<i>C. mantchuricum</i>	Shaanxi(Western population)	Blood	Female	12	CMAA00	Re-sequencing	43.17	98.59%	SAMC226699
<i>C. mantchuricum</i>	Shaanxi(Western population)	Blood	Female	13	CMAA01	Re-sequencing	15.60	98.54%	SAMC226700

<i>C. mantchuricum</i>	Shaanxi(Western population)	Blood	Male	14	CMAA02	Re-sequencing	15.89	98.65%	SAMC226701
<i>C. mantchuricum</i>	Shaanxi(Western population)	Muscle	Female	15	CMAA03	Re-sequencing	15.31	97.56%	SAMC226702
<i>C. mantchuricum</i>	Shaanxi(Western population)	Blood	Male	16	CMAA04*	Re-sequencing	17.64	98.85%	SAMC226703
<i>C. mantchuricum</i>	Shaanxi(Western population)	Blood	Female	17	CMAA05	Re-sequencing	18.86	97.42%	SAMC226704
<i>C. mantchuricum</i>	Shaanxi(Western population)	Blood	Male	18	CMAA06	Re-sequencing	18.56	93.89%	SAMC226705
<i>C. mantchuricum</i>	Shaanxi(Western population)	Blood	Female	19	CMAA07*	Re-sequencing	16.07	95.51%	SAMC226706
<i>C. mantchuricum</i>	Shaanxi(Western population)	Blood	Female	20	CMAA08	Re-sequencing	20.81	98.60%	SAMC226707
<i>C. mantchuricum</i>	Shaanxi(Western population)	Blood	Female	21	CMAA09*	Re-sequencing	15.53	98.75%	SAMC226708
<i>C. mantchuricum</i>	Shaanxi(Western population)	Blood	Female	22	CMAA10	Re-sequencing	13.42	86.63%	SAMC226709
<i>C. mantchuricum</i>	Shaanxi(Western population)	Blood	Male	23	CMAA11*	Re-sequencing	21.99	96.29%	SAMC226710
<i>C. mantchuricum</i>	Shaanxi(Western population)	Blood	Female	24	CMAA12*	Re-sequencing	19.77	96.96%	SAMC226711
<i>C. mantchuricum</i>	Shaanxi(Western population)	Blood	Female	25	CMAA13	Re-sequencing	18.19	97.43%	SAMC226712
<i>C. mantchuricum</i>	Shaanxi(Western population)	Blood	Female	26	CMAA14	Re-sequencing	18.36	96.47%	SAMC226713
<i>C. mantchuricum</i>	Shaanxi(Western population)	Blood	Female	27	CMAA15*	Re-sequencing	15.59	93.59%	SAMC226714
<i>C. mantchuricum</i>	Shaanxi(Western population)	Blood	Male	28	CMAA16	Re-sequencing	16.82	97.81%	SAMC226715
<i>C. mantchuricum</i>	Shaanxi(Western population)	Blood	Male	29	CMAA17*	Re-sequencing	13.60	83.55%	SAMC226716
<i>C. mantchuricum</i>	Shanxi(Central population)	Muscle	Male	30	CMA04*	Re-sequencing	15.94	98.39%	SAMC226717

<i>C. mantchuricum</i>	Shanxi(Central population)	Blood	Female	31	CMA08	Re-sequencing	15.21	98.24%	SAMC226718
<i>C. mantchuricum</i>	Shanxi(Central population)	Muscle	Female	32	CMA09	Re-sequencing	19.23	97.51%	SAMC226719
<i>C. mantchuricum</i>	Shanxi(Central population)	Liver	Female	33	CMA10*	Re-sequencing	16.77	97.81%	SAMC226720
<i>C. mantchuricum</i>	Shanxi(Central population)	Muscle	Female	34	CMA11	Re-sequencing	19.53	96.86%	SAMC226721
<i>C. mantchuricum</i>	Shanxi(Central population)	Liver	Female	35	CMA12	Re-sequencing	18.26	97.80%	SAMC226722
<i>C. mantchuricum</i>	Shanxi(Central population)	Liver	Male	36	CMA13	Re-sequencing	21.46	97.98%	SAMC226723
<i>C. mantchuricum</i>	Shanxi(Central population)	Liver	Male	37	CMA14	Re-sequencing	19.13	98.39%	SAMC226724
<i>C. mantchuricum</i>	Shanxi(Central population)	Liver	Female	38	CMA15*	Re-sequencing	18.19	97.54%	SAMC226725
<i>C. mantchuricum</i>	Shanxi(Central population)	Muscle	Female	39	CMA16	Re-sequencing	16.03	97.79%	SAMC226726
<i>C. mantchuricum</i>	Shanxi(Central population)	Muscle	Male	40	CMA18	Re-sequencing	15.32	97.62%	SAMC226727
<i>C. mantchuricum</i>	Hebei(Eastern population)	Blood	Male	41	CMH00	Re-sequencing	36.59	98.26%	SAMC226728
<i>C. mantchuricum</i>	Hebei(Eastern population)	Blood	Female	42	CMH01	Re-sequencing	17.40	98.53%	SAMC226729
<i>C. mantchuricum</i>	Hebei(Eastern population)	Blood	Male	43	CMH02	Re-sequencing	19.22	98.65%	SAMC226730
<i>C. mantchuricum</i>	Hebei(Eastern population)	Blood	Female	44	CMH03	Re-sequencing	19.54	98.12%	SAMC226731
<i>C. mantchuricum</i>	Hebei(Eastern population)	Muscle	Male	45	CMH04*	Re-sequencing	18.79	97.85%	SAMC226732
<i>C. mantchuricum</i>	Hebei(Eastern population)	Blood	Male	46	CMH05	Re-sequencing	18.30	98.63%	SAMC226733
<i>C. mantchuricum</i>	Hebei(Eastern population)	Muscle	Male	47	CMH06	Re-sequencing	19.69	97.85%	SAMC226734

<i>C. mantchuricum</i>	Hebei(Eastern population)	Blood	Female	48	CMH07*	Re-sequencing	17.60	98.34%	SAMC226735
<i>C. mantchuricum</i>	Hebei(Eastern population)	Blood	Female	49	CMH08	Re-sequencing	15.51	97.95%	SAMC226736
<i>C. mantchuricum</i>	Hebei(Eastern population)	Blood	Female	50	CMH09	Re-sequencing	18.92	98.19%	SAMC226737
<i>C. mantchuricum</i>	Hebei(Eastern population)	Muscle	Male	51	CMH12*	Re-sequencing	17.60	97.77%	SAMC226738
<i>C. mantchuricum</i>	Shanxi(Central population)	Blood	Female	52	CMA00*	De novo sequencing	201.75	N	SAMC226739

Dataset S10: The relationship of each sample in each population inferred by the King program. Blue indicates *Crossoptilon auritum*. Brown-W, Brown-C, and Brown-E indicate the Western, Central, and Eastern populations of *Crossoptilon manchuricum*, respectively. ID1: The first individual of the pair; ID2: The second individual of the pair; N_SNP: The number of SNPS that do not have missing SNPS in either of the individuals; HetHet: Percentage of SNPs with double heterozygotes; IBS0: Proportion of SNPs with 0-IBS (identical-by-state); Kinship: Kinship coefficient estimated by the program. The asterisks mark the individuals that were excluded in the downstream analysis.

Population	ID1	ID2	N_SNP	HetHet	IBS0	Kinship
Blue	CA00	CA1	2275444	15.29%	8.02%	-0.06
Blue	CA00	CA10	2275444	12.22%	12.15%	-0.20
Blue	CA00	CA2	2275444	13.75%	7.01%	0.00
Blue	CA00	CA3*	2275444	13.94%	7.15%	-0.01
Blue	CA00	CA4	2275444	11.88%	12.58%	-0.23
Blue	CA00	CA5	2275444	12.65%	9.10%	-0.10
Blue	CA00	CA6	2275444	12.14%	12.50%	-0.22
Blue	CA00	CA7	2275444	11.95%	12.38%	-0.21
Blue	CA00	CA8	2275444	12.02%	12.52%	-0.22
Blue	CA00	CA9	2275444	12.15%	12.33%	-0.21
Blue	CA1	CA10	2275444	15.29%	7.84%	-0.06
Blue	CA1	CA2	2275444	15.24%	7.92%	-0.06
Blue	CA1	CA3*	2275444	17.07%	0.30%	0.21
Blue	CA1	CA4	2275444	14.83%	8.22%	-0.09
Blue	CA1	CA5	2275444	14.68%	8.77%	-0.11
Blue	CA1	CA6	2275444	15.06%	8.01%	-0.08
Blue	CA1	CA7	2275444	15.06%	8.17%	-0.08
Blue	CA1	CA8	2275444	14.94%	8.36%	-0.09
Blue	CA1	CA9	2275444	15.15%	7.82%	-0.07
Blue	CA10	CA2	2275444	12.19%	12.07%	-0.19
Blue	CA10	CA3*	2275444	12.59%	12.35%	-0.20
Blue	CA10	CA4	2275444	13.34%	7.26%	-0.03
Blue	CA10	CA5	2275444	12.04%	11.43%	-0.18
Blue	CA10	CA6	2275444	13.54%	7.25%	-0.02
Blue	CA10	CA7	2275444	13.39%	7.44%	-0.03
Blue	CA10	CA8	2275444	13.30%	7.30%	-0.03
Blue	CA10	CA9	2275444	13.39%	7.49%	-0.03
Blue	CA2	CA3*	2275444	13.73%	7.19%	-0.02
Blue	CA2	CA4	2275444	12.00%	12.38%	-0.22
Blue	CA2	CA5	2275444	12.55%	9.02%	-0.10
Blue	CA2	CA6	2275444	12.05%	12.22%	-0.21
Blue	CA2	CA7	2275444	12.16%	12.21%	-0.20
Blue	CA2	CA8	2275444	11.95%	12.25%	-0.21
Blue	CA2	CA9	2275444	12.20%	12.12%	-0.20
Blue	CA3*	CA4	2275444	12.33%	12.66%	-0.23
Blue	CA3*	CA5	2275444	12.95%	9.17%	-0.10
Blue	CA3*	CA6	2275444	12.36%	12.56%	-0.22
Blue	CA3*	CA7	2275444	12.51%	12.56%	-0.22
Blue	CA3*	CA8	2275444	12.39%	12.66%	-0.23
Blue	CA3*	CA9	2275444	12.57%	12.53%	-0.21
Blue	CA4	CA5	2275444	11.66%	11.95%	-0.20

Blue	CA4	CA6	2275444	13.02%	7.41%	-0.03
Blue	CA4	CA7	2275444	13.20%	7.05%	-0.02
Blue	CA4	CA8	2275444	13.06%	7.44%	-0.03
Blue	CA4	CA9	2275444	13.01%	7.46%	-0.03
Blue	CA5	CA6	2275444	11.78%	11.62%	-0.18
Blue	CA5	CA7	2275444	11.91%	11.58%	-0.18
Blue	CA5	CA8	2275444	11.73%	11.61%	-0.19
Blue	CA5	CA9	2275444	11.96%	11.63%	-0.18
Blue	CA6	CA7	2275444	13.28%	7.55%	-0.03
Blue	CA6	CA8	2275444	13.00%	7.30%	-0.03
Blue	CA6	CA9	2275444	13.17%	7.09%	-0.02
Blue	CA7	CA8	2275444	13.15%	7.40%	-0.03
Blue	CA7	CA9	2275444	13.46%	6.85%	0.00
Blue	CA8	CA9	2275444	13.21%	7.50%	-0.03
Brown-W	CMAA00	CMAA1	2275444	0.98%	0.53%	-0.03
Brown-W	CMAA00	CMAA10	2275444	0.79%	0.61%	-0.20
Brown-W	CMAA00	CMAA11*	2275444	1.24%	0.20%	0.14
Brown-W	CMAA00	CMAA12*	2275444	1.02%	0.69%	-0.09
Brown-W	CMAA00	CMAA13	2275444	1.16%	0.34%	0.05
Brown-W	CMAA00	CMAA14	2275444	1.39%	0.43%	0.07
Brown-W	CMAA00	CMAA15*	2275444	1.33%	0.40%	0.07
Brown-W	CMAA00	CMAA16	2275444	1.26%	0.27%	0.09
Brown-W	CMAA00	CMAA17*	2275444	1.10%	0.60%	-0.03
Brown-W	CMAA1	CMAA10	2275444	1.05%	0.22%	0.07
Brown-W	CMAA1	CMAA11*	2275444	1.06%	0.45%	0.02
Brown-W	CMAA1	CMAA12*	2275444	0.90%	0.93%	-0.18
Brown-W	CMAA1	CMAA13	2275444	0.96%	0.35%	0.03
Brown-W	CMAA1	CMAA14	2275444	1.19%	0.44%	0.02
Brown-W	CMAA1	CMAA15*	2275444	1.30%	0.25%	0.10
Brown-W	CMAA1	CMAA16	2275444	1.46%	0.33%	0.09
Brown-W	CMAA1	CMAA17*	2275444	1.07%	0.49%	0.01
Brown-W	CMAA10	CMAA11*	2275444	0.70%	0.50%	-0.15
Brown-W	CMAA10	CMAA12*	2275444	0.65%	0.77%	-0.28
Brown-W	CMAA10	CMAA13	2275444	0.76%	0.60%	-0.15
Brown-W	CMAA10	CMAA14	2275444	0.89%	0.50%	-0.15
Brown-W	CMAA10	CMAA15*	2275444	0.98%	0.61%	-0.19
Brown-W	CMAA10	CMAA16	2275444	0.96%	0.48%	-0.15
Brown-W	CMAA10	CMAA17*	2275444	0.73%	0.68%	-0.23
Brown-W	CMAA11*	CMAA12*	2275444	0.98%	0.63%	-0.06
Brown-W	CMAA11*	CMAA13	2275444	1.40%	0.05%	0.23
Brown-W	CMAA11*	CMAA14	2275444	1.12%	0.43%	0.01
Brown-W	CMAA11*	CMAA15*	2275444	1.45%	0.12%	0.18
Brown-W	CMAA11*	CMAA16	2275444	1.57%	0.22%	0.16
Brown-W	CMAA11*	CMAA17*	2275444	1.18%	0.27%	0.11
Brown-W	CMAA12*	CMAA13	2275444	0.84%	0.70%	-0.14
Brown-W	CMAA12*	CMAA14	2275444	1.03%	0.65%	-0.09
Brown-W	CMAA12*	CMAA15*	2275444	1.10%	0.63%	-0.08
Brown-W	CMAA12*	CMAA16	2275444	1.13%	0.82%	-0.16
Brown-W	CMAA12*	CMAA17*	2275444	0.82%	0.98%	-0.22
Brown-W	CMAA13	CMAA14	2275444	1.16%	0.44%	-0.01
Brown-W	CMAA13	CMAA15*	2275444	1.28%	0.15%	0.12
Brown-W	CMAA13	CMAA16	2275444	1.33%	0.26%	0.08
Brown-W	CMAA13	CMAA17*	2275444	1.10%	0.35%	0.05

Brown-W	CMAA14	CMAA15*	2275444	1.27%	0.43%	0.06
Brown-W	CMAA14	CMAA16	2275444	1.42%	0.24%	0.14
Brown-W	CMAA14	CMAA17*	2275444	1.11%	0.50%	-0.02
Brown-W	CMAA15*	CMAA16	2275444	1.73%	0.02%	0.26
Brown-W	CMAA15*	CMAA17*	2275444	1.37%	0.04%	0.20
Brown-W	CMAA16	CMAA17*	2275444	1.94%	0.21%	0.23
Brown-W	CMAA00	CMAA2	2275444	0.62%	1.05%	-0.65
Brown-W	CMAA00	CMAA3	2275444	1.48%	0.59%	0.00
Brown-W	CMAA00	CMAA4*	2275444	1.20%	0.52%	0.00
Brown-W	CMAA00	CMAA5	2275444	1.33%	0.63%	-0.01
Brown-W	CMAA00	CMAA6	2275444	1.21%	0.52%	0.00
Brown-W	CMAA00	CMAA7*	2275444	0.90%	0.69%	-0.09
Brown-W	CMAA00	CMAA8	2275444	1.52%	0.60%	0.04
Brown-W	CMAA00	CMAA9*	2275444	1.02%	0.68%	-0.07
Brown-W	CMAA1	CMAA2	2275444	0.86%	0.02%	0.09
Brown-W	CMAA1	CMAA3	2275444	1.08%	0.58%	-0.09
Brown-W	CMAA1	CMAA4*	2275444	1.19%	0.62%	-0.05
Brown-W	CMAA1	CMAA5	2275444	0.83%	0.41%	-0.04
Brown-W	CMAA1	CMAA6	2275444	1.20%	0.62%	-0.05
Brown-W	CMAA1	CMAA7*	2275444	1.07%	0.95%	-0.18
Brown-W	CMAA1	CMAA8	2275444	1.08%	0.30%	0.05
Brown-W	CMAA1	CMAA9*	2275444	1.10%	0.61%	-0.05
Brown-W	CMAA10	CMAA2	2275444	0.69%	0.73%	-0.31
Brown-W	CMAA10	CMAA3	2275444	1.03%	0.63%	-0.22
Brown-W	CMAA10	CMAA4*	2275444	0.82%	0.57%	-0.20
Brown-W	CMAA10	CMAA5	2275444	0.81%	0.42%	-0.14
Brown-W	CMAA10	CMAA6	2275444	0.83%	0.56%	-0.20
Brown-W	CMAA10	CMAA7*	2275444	0.79%	0.99%	-0.38
Brown-W	CMAA10	CMAA8	2275444	0.73%	0.48%	-0.18
Brown-W	CMAA10	CMAA9*	2275444	0.85%	0.72%	-0.25
Brown-W	CMAA11*	CMAA2	2275444	0.71%	0.75%	-0.42
Brown-W	CMAA11*	CMAA3	2275444	1.30%	0.40%	0.03
Brown-W	CMAA11*	CMAA4*	2275444	1.23%	0.36%	0.06
Brown-W	CMAA11*	CMAA5	2275444	1.20%	0.38%	0.05
Brown-W	CMAA11*	CMAA6	2275444	1.24%	0.35%	0.06
Brown-W	CMAA11*	CMAA7*	2275444	1.02%	0.49%	-0.01
Brown-W	CMAA11*	CMAA8	2275444	1.37%	0.47%	0.05
Brown-W	CMAA11*	CMAA9*	2275444	1.18%	0.52%	0.01
Brown-W	CMAA12*	CMAA2	2275444	0.40%	1.08%	-0.71
Brown-W	CMAA12*	CMAA3	2275444	1.00%	0.67%	-0.14
Brown-W	CMAA12*	CMAA4*	2275444	1.20%	0.52%	-0.02
Brown-W	CMAA12*	CMAA5	2275444	1.48%	0.12%	0.19
Brown-W	CMAA12*	CMAA6	2275444	1.21%	0.52%	-0.02
Brown-W	CMAA12*	CMAA7*	2275444	1.00%	0.76%	-0.12
Brown-W	CMAA12*	CMAA8	2275444	1.40%	0.79%	-0.08
Brown-W	CMAA12*	CMAA9*	2275444	1.26%	0.69%	-0.05
Brown-W	CMAA13	CMAA2	2275444	0.64%	0.65%	-0.33
Brown-W	CMAA13	CMAA3	2275444	1.19%	0.49%	-0.06
Brown-W	CMAA13	CMAA4*	2275444	1.08%	0.51%	-0.06
Brown-W	CMAA13	CMAA5	2275444	1.04%	0.37%	-0.01
Brown-W	CMAA13	CMAA6	2275444	1.09%	0.50%	-0.06
Brown-W	CMAA13	CMAA7*	2275444	0.80%	0.62%	-0.14
Brown-W	CMAA13	CMAA8	2275444	1.09%	0.25%	0.05

Brown-W	CMAA13	CMAA9*	2275444	0.98%	0.46%	-0.04
Brown-W	CMAA14	CMAA2	2275444	0.66%	0.75%	-0.49
Brown-W	CMAA14	CMAA3	2275444	1.44%	0.45%	0.06
Brown-W	CMAA14	CMAA4*	2275444	1.25%	0.60%	0.01
Brown-W	CMAA14	CMAA5	2275444	1.19%	0.37%	0.07
Brown-W	CMAA14	CMAA6	2275444	1.26%	0.59%	0.01
Brown-W	CMAA14	CMAA7*	2275444	1.26%	0.88%	-0.11
Brown-W	CMAA14	CMAA8	2275444	1.36%	0.29%	0.12
Brown-W	CMAA14	CMAA9*	2275444	1.24%	0.65%	-0.02
Brown-W	CMAA15*	CMAA2	2275444	0.81%	0.54%	-0.33
Brown-W	CMAA15*	CMAA3	2275444	1.37%	0.40%	0.07
Brown-W	CMAA15*	CMAA4*	2275444	1.34%	0.50%	0.05
Brown-W	CMAA15*	CMAA5	2275444	1.57%	0.55%	0.07
Brown-W	CMAA15*	CMAA6	2275444	1.35%	0.49%	0.06
Brown-W	CMAA15*	CMAA7*	2275444	1.16%	0.66%	-0.05
Brown-W	CMAA15*	CMAA8	2275444	1.28%	0.31%	0.10
Brown-W	CMAA15*	CMAA9*	2275444	1.43%	0.60%	0.02
Brown-W	CMAA16	CMAA2	2275444	1.01%	0.52%	-0.27
Brown-W	CMAA16	CMAA3	2275444	1.50%	0.44%	0.08
Brown-W	CMAA16	CMAA4*	2275444	1.30%	0.36%	0.08
Brown-W	CMAA16	CMAA5	2275444	1.33%	0.47%	0.05
Brown-W	CMAA16	CMAA6	2275444	1.31%	0.36%	0.09
Brown-W	CMAA16	CMAA7*	2275444	1.24%	0.56%	-0.01
Brown-W	CMAA16	CMAA8	2275444	1.35%	0.37%	0.08
Brown-W	CMAA16	CMAA9*	2275444	1.28%	0.64%	-0.03
Brown-W	CMAA17*	CMAA2	2275444	0.73%	0.50%	-0.26
Brown-W	CMAA17*	CMAA3	2275444	1.30%	0.42%	0.02
Brown-W	CMAA17*	CMAA4*	2275444	1.13%	0.70%	-0.09
Brown-W	CMAA17*	CMAA5	2275444	1.05%	0.51%	-0.03
Brown-W	CMAA17*	CMAA6	2275444	1.14%	0.69%	-0.08
Brown-W	CMAA17*	CMAA7*	2275444	0.79%	0.79%	-0.16
Brown-W	CMAA17*	CMAA8	2275444	1.05%	0.56%	-0.05
Brown-W	CMAA17*	CMAA9*	2275444	0.93%	0.77%	-0.14
Brown-W	CMAA2	CMAA3	2275444	0.78%	0.84%	-0.56
Brown-W	CMAA2	CMAA4*	2275444	0.69%	0.85%	-0.55
Brown-W	CMAA2	CMAA5	2275444	0.71%	0.66%	-0.42
Brown-W	CMAA2	CMAA6	2275444	0.70%	0.84%	-0.55
Brown-W	CMAA2	CMAA7*	2275444	0.52%	1.18%	-0.77
Brown-W	CMAA2	CMAA8	2275444	0.60%	0.72%	-0.49
Brown-W	CMAA2	CMAA9*	2275444	0.61%	0.82%	-0.53
Brown-W	CMAA3	CMAA4*	2275444	1.19%	0.33%	0.06
Brown-W	CMAA3	CMAA5	2275444	1.40%	0.45%	0.06
Brown-W	CMAA3	CMAA6	2275444	1.20%	0.32%	0.07
Brown-W	CMAA3	CMAA7*	2275444	1.19%	0.61%	-0.05
Brown-W	CMAA3	CMAA8	2275444	1.60%	0.46%	0.08
Brown-W	CMAA3	CMAA9*	2275444	1.24%	0.35%	0.05
Brown-W	CMAA4*	CMAA5	2275444	1.40%	0.47%	0.07
Brown-W	CMAA4*	CMAA6	2275444	3.07%	0.00%	0.49
Brown-W	CMAA4*	CMAA7*	2275444	1.27%	0.03%	0.19
Brown-W	CMAA4*	CMAA8	2275444	1.24%	0.39%	0.07
Brown-W	CMAA4*	CMAA9*	2275444	1.34%	0.42%	0.07
Brown-W	CMAA5	CMAA6	2275444	1.42%	0.47%	0.08
Brown-W	CMAA5	CMAA7*	2275444	1.00%	0.54%	-0.04

Brown-W	CMAA5	CMAA8	2275444	1.18%	0.52%	0.02
Brown-W	CMAA5	CMAA9*	2275444	1.29%	0.49%	0.04
Brown-W	CMAA6	CMAA7*	2275444	1.28%	0.02%	0.19
Brown-W	CMAA6	CMAA8	2275444	1.25%	0.38%	0.07
Brown-W	CMAA6	CMAA9*	2275444	1.35%	0.41%	0.07
Brown-W	CMAA7*	CMAA8	2275444	1.18%	0.65%	-0.04
Brown-W	CMAA7*	CMAA9*	2275444	1.22%	0.54%	0.01
Brown-W	CMAA8	CMAA9*	2275444	1.58%	0.20%	0.19
Brown-C	CMA10*	CMA11	2275444	2.08%	1.31%	-0.06
Brown-C	CMA10*	CMA12	2275444	5.28%	0.00%	0.49
Brown-C	CMA10*	CMA13	2275444	2.18%	1.19%	-0.04
Brown-C	CMA10*	CMA14	2275444	1.86%	1.03%	-0.02
Brown-C	CMA11	CMA12	2275444	2.10%	1.30%	-0.06
Brown-C	CMA11	CMA13	2275444	2.40%	0.92%	0.04
Brown-C	CMA11	CMA14	2275444	2.37%	1.12%	0.00
Brown-C	CMA12	CMA13	2275444	2.19%	1.19%	-0.03
Brown-C	CMA12	CMA14	2275444	1.86%	1.02%	-0.02
Brown-C	CMA13	CMA14	2275444	2.69%	0.55%	0.13
Brown-C	CMA10*	CMA15*	2275444	2.10%	1.08%	-0.04
Brown-C	CMA10*	CMA16	2275444	1.98%	1.14%	-0.03
Brown-C	CMA10*	CMA18	2275444	1.71%	1.37%	-0.12
Brown-C	CMA10*	CMA4*	2275444	1.73%	1.35%	-0.11
Brown-C	CMA10*	CMA8	2275444	1.76%	1.64%	-0.24
Brown-C	CMA10*	CMA9	2275444	1.98%	1.11%	-0.02
Brown-C	CMA11	CMA15*	2275444	2.71%	0.02%	0.22
Brown-C	CMA11	CMA16	2275444	2.12%	1.29%	-0.05
Brown-C	CMA11	CMA18	2275444	1.90%	1.53%	-0.14
Brown-C	CMA11	CMA4*	2275444	1.92%	1.50%	-0.13
Brown-C	CMA11	CMA8	2275444	1.62%	1.58%	-0.26
Brown-C	CMA11	CMA9	2275444	1.95%	1.45%	-0.10
Brown-C	CMA12	CMA15*	2275444	2.11%	1.07%	-0.03
Brown-C	CMA12	CMA16	2275444	1.99%	1.13%	-0.02
Brown-C	CMA12	CMA18	2275444	1.72%	1.36%	-0.12
Brown-C	CMA12	CMA4*	2275444	1.74%	1.35%	-0.11
Brown-C	CMA12	CMA8	2275444	1.76%	1.64%	-0.24
Brown-C	CMA12	CMA9	2275444	1.99%	1.11%	-0.02
Brown-C	CMA13	CMA15*	2275444	2.46%	1.24%	-0.01
Brown-C	CMA13	CMA16	2275444	2.25%	1.22%	-0.04
Brown-C	CMA13	CMA18	2275444	2.17%	1.65%	-0.15
Brown-C	CMA13	CMA4*	2275444	2.20%	1.63%	-0.14
Brown-C	CMA13	CMA8	2275444	1.57%	1.58%	-0.27
Brown-C	CMA13	CMA9	2275444	2.26%	0.85%	0.03
Brown-C	CMA14	CMA15*	2275444	2.39%	0.88%	0.03
Brown-C	CMA14	CMA16	2275444	2.18%	1.09%	0.00
Brown-C	CMA14	CMA18	2275444	2.06%	1.13%	-0.04
Brown-C	CMA14	CMA4*	2275444	2.08%	1.12%	-0.03
Brown-C	CMA14	CMA8	2275444	1.58%	1.59%	-0.25
Brown-C	CMA14	CMA9	2275444	2.25%	1.02%	0.02
Brown-C	CMA15*	CMA16	2275444	2.18%	1.22%	-0.05
Brown-C	CMA15*	CMA18	2275444	2.82%	0.04%	0.22
Brown-C	CMA15*	CMA4*	2275444	2.86%	0.03%	0.22
Brown-C	CMA15*	CMA8	2275444	1.73%	1.56%	-0.27
Brown-C	CMA15*	CMA9	2275444	2.37%	1.44%	-0.08

Brown-C	CMA16	CMA18	2275444	1.82%	1.49%	-0.13
Brown-C	CMA16	CMA4*	2275444	1.84%	1.47%	-0.12
Brown-C	CMA16	CMA8	2275444	1.52%	1.51%	-0.24
Brown-C	CMA16	CMA9	2275444	2.16%	1.03%	0.01
Brown-C	CMA18	CMA4*	2275444	4.93%	0.00%	0.48
Brown-C	CMA18	CMA8	2275444	1.39%	1.90%	-0.33
Brown-C	CMA18	CMA9	2275444	2.03%	1.55%	-0.12
Brown-C	CMA4*	CMA8	2275444	1.40%	1.88%	-0.32
Brown-C	CMA4*	CMA9	2275444	2.05%	1.54%	-0.11
Brown-C	CMA8	CMA9	2275444	1.72%	1.47%	-0.21
Brown-E	CMH00	CMH1	2275444	1.69%	0.31%	0.18
Brown-E	CMH00	CMH12*	2275444	1.02%	1.29%	-0.29
Brown-E	CMH00	CMH2	2275444	1.47%	0.63%	-0.09
Brown-E	CMH00	CMH3	2275444	1.32%	0.60%	-0.09
Brown-E	CMH00	CMH4*	2275444	1.03%	1.27%	-0.26
Brown-E	CMH00	CMH5	2275444	1.28%	0.77%	-0.09
Brown-E	CMH00	CMH6	2275444	1.03%	1.28%	-0.27
Brown-E	CMH1	CMH12*	2275444	1.00%	1.55%	-0.37
Brown-E	CMH1	CMH2	2275444	1.22%	0.63%	-0.15
Brown-E	CMH1	CMH3	2275444	1.43%	0.89%	-0.18
Brown-E	CMH1	CMH4*	2275444	1.02%	1.53%	-0.37
Brown-E	CMH1	CMH5	2275444	1.25%	0.90%	-0.15
Brown-E	CMH1	CMH6	2275444	1.01%	1.54%	-0.37
Brown-E	CMH12*	CMH2	2275444	1.25%	0.63%	-0.15
Brown-E	CMH12*	CMH3	2275444	1.04%	1.09%	-0.32
Brown-E	CMH12*	CMH4*	2275444	2.76%	0.00%	0.48
Brown-E	CMH12*	CMH5	2275444	1.06%	1.14%	-0.27
Brown-E	CMH12*	CMH6	2275444	2.77%	0.00%	0.49
Brown-E	CMH2	CMH3	2275444	2.26%	0.36%	0.17
Brown-E	CMH2	CMH4*	2275444	1.27%	0.62%	-0.12
Brown-E	CMH2	CMH5	2275444	1.72%	0.59%	0.00
Brown-E	CMH2	CMH6	2275444	1.27%	0.62%	-0.14
Brown-E	CMH3	CMH4*	2275444	1.07%	1.08%	-0.29
Brown-E	CMH3	CMH5	2275444	1.69%	0.71%	-0.01
Brown-E	CMH3	CMH6	2275444	1.06%	1.09%	-0.31
Brown-E	CMH4*	CMH5	2275444	1.08%	1.12%	-0.24
Brown-E	CMH4*	CMH6	2275444	2.80%	0.00%	0.48
Brown-E	CMH5	CMH6	2275444	1.07%	1.13%	-0.26
Brown-E	CMH00	CMH7*	2275444	1.28%	0.81%	-0.09
Brown-E	CMH00	CMH8	2275444	1.04%	1.01%	-0.19
Brown-E	CMH00	CMH9	2275444	1.25%	1.37%	-0.25
Brown-E	CMH1	CMH7*	2275444	1.03%	0.93%	-0.19
Brown-E	CMH1	CMH8	2275444	1.07%	1.08%	-0.23
Brown-E	CMH1	CMH9	2275444	1.01%	1.43%	-0.34
Brown-E	CMH12*	CMH7*	2275444	0.88%	1.21%	-0.32
Brown-E	CMH12*	CMH8	2275444	1.13%	1.46%	-0.36
Brown-E	CMH12*	CMH9	2275444	0.83%	1.45%	-0.38
Brown-E	CMH2	CMH7*	2275444	1.85%	0.68%	-0.01
Brown-E	CMH2	CMH8	2275444	1.45%	0.79%	-0.11
Brown-E	CMH2	CMH9	2275444	1.52%	0.70%	-0.11
Brown-E	CMH3	CMH7*	2275444	1.67%	0.73%	-0.04
Brown-E	CMH3	CMH8	2275444	1.23%	1.17%	-0.24
Brown-E	CMH3	CMH9	2275444	1.34%	0.90%	-0.18

Brown-E	CMH4*	CMH7*	2275444	0.89%	1.20%	-0.28
Brown-E	CMH4*	CMH8	2275444	1.16%	1.45%	-0.32
Brown-E	CMH4*	CMH9	2275444	0.85%	1.44%	-0.35
Brown-E	CMH5	CMH7*	2275444	1.41%	1.02%	-0.11
Brown-E	CMH5	CMH8	2275444	0.90%	0.92%	-0.16
Brown-E	CMH5	CMH9	2275444	1.10%	0.90%	-0.16
Brown-E	CMH6	CMH7*	2275444	0.89%	1.20%	-0.30
Brown-E	CMH6	CMH8	2275444	1.15%	1.45%	-0.34
Brown-E	CMH6	CMH9	2275444	0.84%	1.44%	-0.36
Brown-E	CMH7*	CMH8	2275444	1.20%	0.46%	0.04
Brown-E	CMH7*	CMH9	2275444	1.95%	0.13%	0.25
Brown-E	CMH8	CMH9	2275444	0.96%	0.73%	-0.11

Dataset S11: Summary metrics of the SNP of each sample. The high-quality SNPs indicate the bases with a call quality of Q20 or higher. The high-coverage sites are at least covered by five reads. Heterozygosity was defined as the number of heterozygous SNPs divided by the total callable sites. Blue indicates *Crossoptilon auritum*, and Brown indicates *Crossoptilon mantchuricum*. A represents autosomes, Z represents the Z chromosome, and M represents the mitogenome.

Species	Sample ID	Sample index	The percent of high quality SNPs			The percent of high coverage sites			The percent of missing SNPs			The number of heteozygous / substitutions		Heterozygosity	
			A	Z	M	A	Z	M	A	Z	M	A	Z	A	Z
Blue	CA00	1	94.78%	93.66%	95.25%	98.50%	98.16%	100.00%	0.04%	0.28%	0.00%	760421	17881	0.00083	0.00025
Blue	CA01	2	93.13%	92.48%	95.25%	97.72%	77.31%	100.00%	0.10%	1.34%	0.00%	902604	16246	0.00099	0.00024
Blue	CA02	3	93.78%	94.36%	95.26%	97.79%	97.68%	100.00%	0.12%	0.12%	0.00%	756677	28595	0.00083	0.00040
Blue	CA03	4	93.19%	92.59%	95.26%	97.76%	80.35%	100.00%	0.10%	0.93%	0.00%	779739	15933	0.00086	0.00023
Blue	CA04	5	93.69%	92.91%	95.26%	98.15%	90.75%	100.00%	0.09%	0.62%	0.00%	722721	15973	0.00079	0.00023
Blue	CA05	6	93.39%	92.87%	95.26%	97.99%	86.72%	100.00%	0.09%	0.63%	0.00%	728717	15723	0.00080	0.00022
Blue	CA06	7	93.77%	94.35%	95.26%	97.93%	97.88%	100.00%	0.11%	0.14%	0.00%	728200	24951	0.00080	0.00035
Blue	CA07	8	94.50%	94.99%	95.26%	96.70%	97.11%	100.00%	0.22%	0.13%	0.00%	728940	26981	0.00080	0.00037
Blue	CA08	9	95.00%	95.27%	95.26%	97.04%	96.82%	100.00%	0.16%	0.22%	0.00%	721363	24411	0.00079	0.00034
Blue	CA09	10	94.22%	94.58%	95.26%	97.72%	97.57%	100.00%	0.12%	0.08%	0.00%	732472	25416	0.00080	0.00035
Blue	CA10	11	93.63%	92.70%	95.26%	98.05%	85.95%	100.00%	0.09%	0.63%	0.00%	740524	14987	0.00081	0.00021
Brown	CMAA00	12	94.68%	93.71%	95.54%	98.56%	98.30%	100.00%	0.01%	0.12%	0.00%	68230	13903	0.00007	0.00019
Brown	CMAA01	13	94.92%	93.99%	95.54%	97.34%	77.13%	100.00%	0.02%	0.53%	0.00%	63205	10840	0.00007	0.00016
Brown	CMAA02	14	94.39%	94.87%	95.54%	97.68%	97.67%	100.00%	0.01%	0.02%	0.00%	38525	1403	0.00004	0.00002
Brown	CMAA03	15	92.27%	92.00%	95.54%	97.69%	74.81%	100.00%	0.02%	0.59%	0.00%	80829	12038	0.00009	0.00018
Brown	CMAA04	16	94.16%	94.80%	95.54%	97.73%	97.68%	100.00%	0.02%	0.02%	0.00%	73818	1840	0.00008	0.00003
Brown	CMAA05	17	92.22%	91.39%	95.54%	98.10%	84.59%	100.00%	0.01%	0.41%	0.00%	74476	12417	0.00008	0.00018
Brown	CMAA06	18	93.07%	94.25%	95.54%	98.24%	98.08%	100.00%	0.01%	0.02%	0.00%	75001	1898	0.00008	0.00003

Brown	CMAA07	19	92.24%	91.79%	95.54%	97.86%	77.04%	100.00%	0.01%	0.57%	0.00%	68547	11875	0.00008	0.00018
Brown	CMAA08	20	94.48%	93.87%	95.54%	98.09%	87.52%	100.00%	0.01%	0.34%	0.00%	73400	12521	0.00008	0.00018
Brown	CMAA09	21	94.84%	94.33%	95.54%	97.00%	77.02%	100.00%	0.03%	0.53%	0.00%	70336	11406	0.00008	0.00017
Brown	CMAA10	22	91.75%	89.07%	95.54%	83.37%	45.00%	100.00%	0.37%	4.14%	0.00%	49204	10382	0.00006	0.00021
Brown	CMAA11	23	93.03%	94.07%	95.55%	98.39%	98.21%	100.00%	0.01%	0.00%	0.00%	65444	2150	0.00007	0.00003
Brown	CMAA12	24	93.93%	93.48%	95.55%	98.21%	88.87%	100.00%	0.01%	0.36%	0.00%	63348	13922	0.00007	0.00020
Brown	CMAA13	25	92.83%	92.46%	95.55%	98.08%	83.94%	100.00%	0.01%	0.32%	0.00%	58156	12005	0.00006	0.00017
Brown	CMAA14	26	94.50%	94.15%	95.55%	98.19%	85.63%	100.00%	0.01%	0.30%	0.00%	74402	13709	0.00008	0.00020
Brown	CMAA15	27	92.62%	92.41%	95.55%	97.76%	76.50%	100.00%	0.01%	0.59%	0.00%	75143	12700	0.00008	0.00019
Brown	CMAA16	28	91.92%	92.82%	95.55%	97.98%	97.82%	100.00%	0.01%	0.01%	0.00%	77779	2145	0.00009	0.00003
Brown	CMAA17	29	92.29%	93.54%	95.55%	96.23%	95.82%	100.00%	0.02%	0.02%	0.00%	64647	2054	0.00007	0.00003
Brown	CMA04	30	93.13%	94.36%	95.69%	97.34%	97.45%	100.00%	0.03%	0.04%	0.00%	120600	1294	0.00013	0.00002
Brown	CMA08	31	93.96%	93.37%	95.71%	95.55%	72.05%	100.00%	0.05%	0.97%	0.00%	101425	13094	0.00011	0.00020
Brown	CMA09	32	92.85%	92.53%	95.75%	98.28%	88.71%	100.00%	0.01%	0.34%	0.00%	128238	12343	0.00014	0.00017
Brown	CMA10	33	92.40%	92.02%	95.73%	98.00%	81.37%	100.00%	0.01%	0.53%	0.00%	127557	12243	0.00014	0.00018
Brown	CMA11	34	92.69%	92.46%	95.30%	98.15%	90.29%	100.00%	0.01%	0.39%	0.00%	133880	12653	0.00015	0.00018
Brown	CMA12	35	92.66%	92.29%	95.32%	98.21%	86.08%	100.00%	0.01%	0.39%	0.00%	128242	12283	0.00014	0.00018
Brown	CMA13	36	92.93%	94.01%	95.36%	98.41%	98.24%	100.00%	0.01%	0.02%	0.00%	136818	2837	0.00015	0.00004
Brown	CMA14	37	93.14%	94.20%	95.36%	98.16%	98.08%	100.00%	0.02%	0.02%	0.00%	126997	2454	0.00014	0.00003
Brown	CMA15	38	93.47%	93.18%	95.48%	98.18%	84.49%	100.00%	0.01%	0.27%	0.00%	144027	12458	0.00016	0.00018
Brown	CMA16	39	92.50%	92.09%	95.49%	97.86%	79.01%	100.00%	0.02%	0.52%	0.00%	127677	12222	0.00014	0.00018
Brown	CMA18	40	92.79%	93.93%	95.53%	97.38%	97.07%	100.00%	0.02%	0.03%	0.00%	119486	1346	0.00013	0.00002
Brown	CMH00	41	92.46%	93.40%	95.55%	98.49%	98.31%	100.00%	0.02%	0.08%	0.00%	70061	808	0.00008	0.00001
Brown	CMH01	42	94.17%	93.06%	95.55%	97.84%	83.94%	100.00%	0.04%	0.37%	0.00%	66657	10978	0.00007	0.00016
Brown	CMH02	43	94.00%	94.66%	95.55%	98.18%	98.44%	100.00%	0.03%	0.03%	0.00%	105279	1392	0.00012	0.00002
Brown	CMH03	44	93.44%	92.77%	95.55%	98.19%	87.08%	100.00%	0.02%	0.35%	0.00%	99277	13337	0.00011	0.00019
Brown	CMH04	45	91.98%	93.04%	95.41%	98.23%	98.09%	100.00%	0.03%	0.02%	0.00%	70004	912	0.00008	0.00001
Brown	CMH05	46	93.03%	93.95%	95.41%	98.03%	98.01%	100.00%	0.03%	0.03%	0.00%	81476	2178	0.00009	0.00003

Brown	CMH06	47	93.08%	94.18%	95.44%	98.30%	98.16%	100.00%	0.02%	0.03%	0.00%	67604	953	0.00007	0.00001
Brown	CMH07	48	93.94%	93.23%	95.44%	97.97%	84.65%	100.00%	0.03%	0.31%	0.00%	78353	12207	0.00009	0.00018
Brown	CMH08	49	92.99%	92.63%	95.44%	97.64%	77.13%	100.00%	0.03%	0.53%	0.00%	77740	11520	0.00009	0.00017
Brown	CMH09	50	93.38%	92.78%	95.45%	98.13%	87.84%	100.00%	0.02%	0.32%	0.00%	69882	12318	0.00008	0.00018
Brown	CMH12	51	90.21%	91.22%	95.24%	98.09%	97.95%	100.00%	0.03%	0.04%	0.00%	66594	812	0.00007	0.00001

Dataset S12: Scaffolds on the sex chromosomes and mitochondrial genome.

Z chromosome linked scaffolds		Mitochondria linked scaffolds	
Scaffolds	The percent of homologous gene (%)	Scaffolds	The percent of homologous gene (%)
scaffold_74	25	scaffold_1343	100
scaffold_142	41.38	scaffold_1701	100
scaffold_34	46.24	scaffold_1847	100
scaffold_394	50		
scaffold_397	50		
scaffold_399	57.14		
scaffold_215	68.75		
scaffold_247	70.59		
scaffold_157	73.33		
scaffold_79	73.68		
scaffold_241	75		
scaffold_380	75		
scaffold_401	75		
scaffold_254	76.47		
scaffold_187	77.08		
scaffold_62	77.5		
scaffold_212	80		
scaffold_333	80		
scaffold_403	80		
scaffold_122	80.36		
scaffold_196	81.48		
scaffold_291	81.48		
scaffold_192	82.93		
scaffold_336	85.71		
scaffold_349	85.71		
scaffold_160	86.36		
scaffold_236	86.36		
scaffold_113	86.67		
scaffold_156	86.67		
scaffold_57	87.04		
scaffold_93	87.23		
scaffold_37	88.46		
scaffold_259	88.89		
scaffold_136	89.66		

scaffold_252	90
scaffold_1249	100
scaffold_1272	100
scaffold_194	100
scaffold_285	100
scaffold_351	100
scaffold_357	100
scaffold_367	100
scaffold_393	100
scaffold_427	100
scaffold_438	100
scaffold_453	100
scaffold_582	100
scaffold_777	100
scaffold_913	100
