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## Supplementary Materials for

### **Substrate-specific recognition of IKKs mediated by USP16 facilitates autoimmune inflammation**

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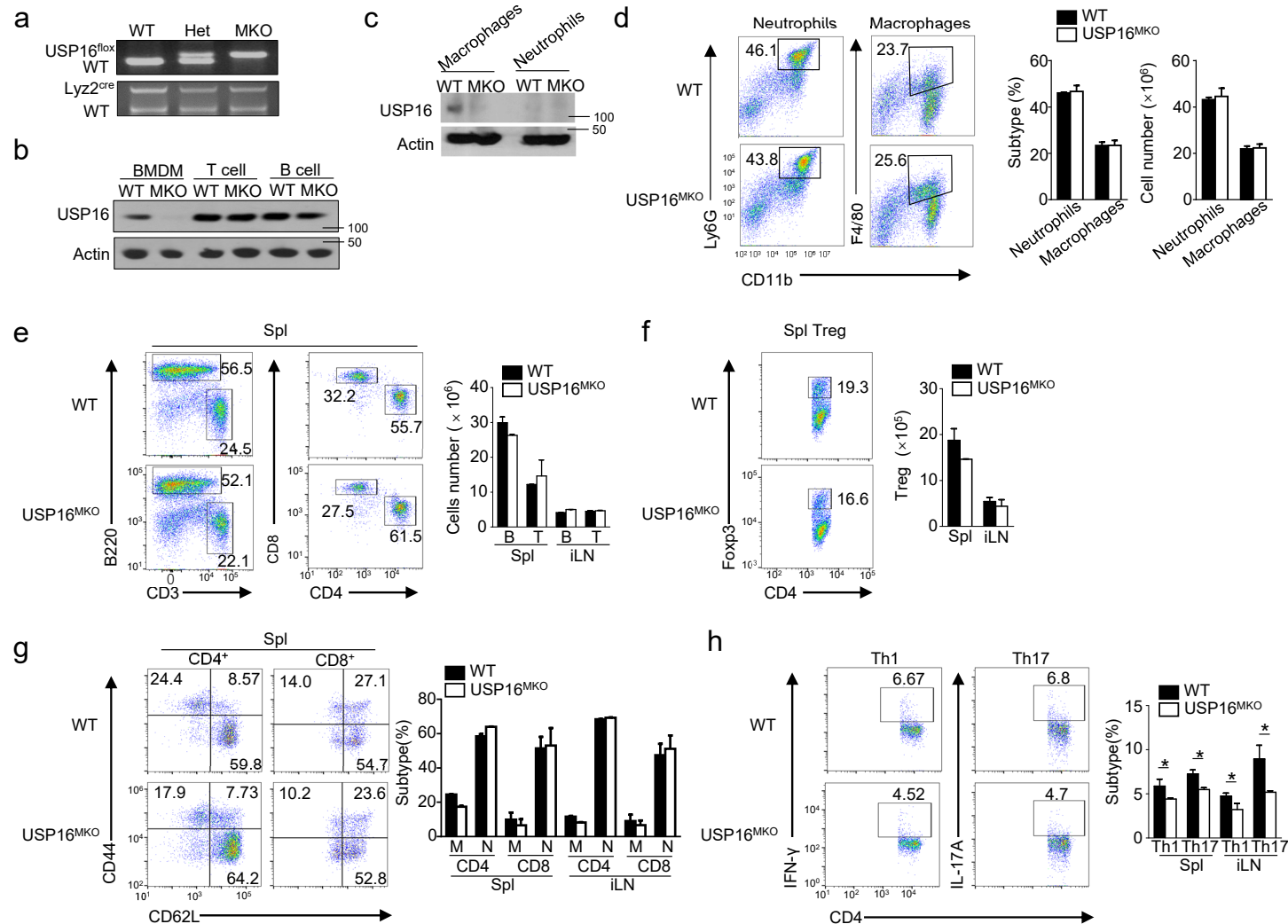
#### **The PDF file includes:**

Figs. S1 to S6  
Legend for table S1

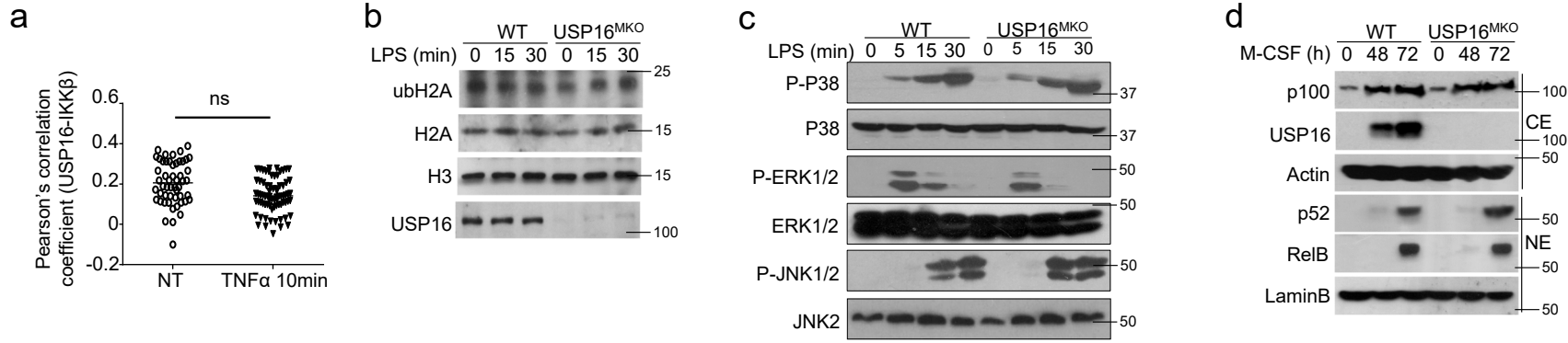
#### **Other Supplementary Material for this manuscript includes the following:**

(available at [advances.sciencemag.org/cgi/content/full/7/3/eabc4009/DC1](https://advances.sciencemag.org/cgi/content/full/7/3/eabc4009/DC1))

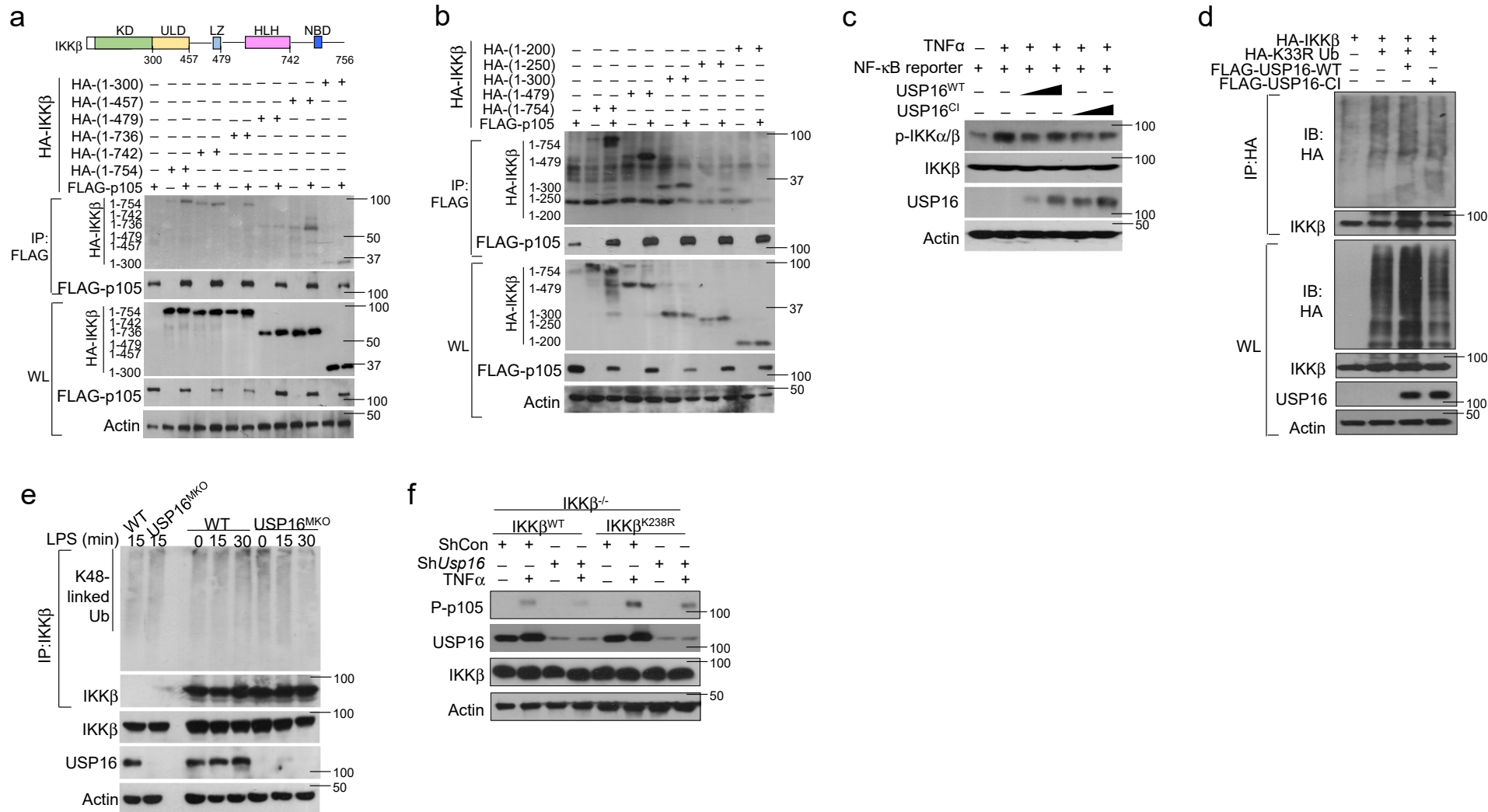
Table S1



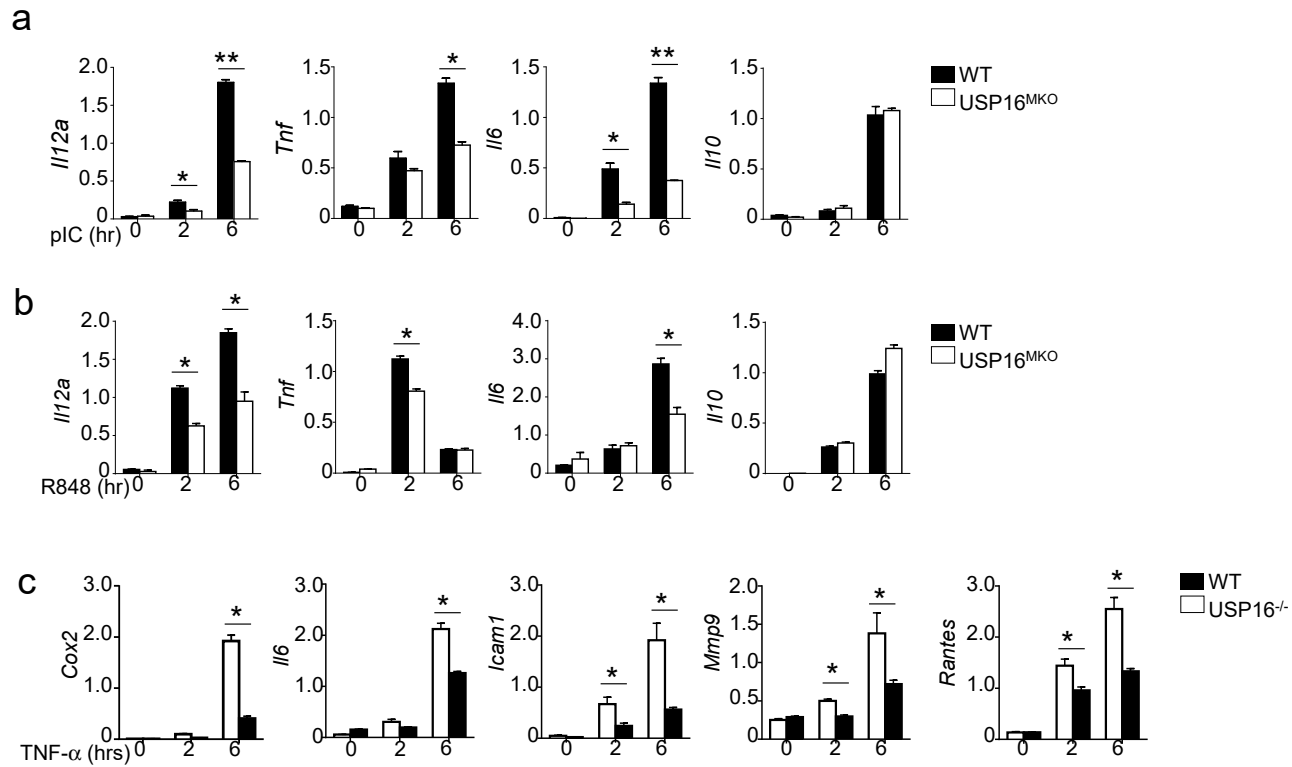
**Supplementary Figure 1 USP16 did not affect the development and homeostasis of the immune system.** (a) Genotyping PCR of WT ( $USP16^{fllox/+}$ /Lyz2-Cre) and myeloid-conditional USP16-knockout ( $USP16^{MKO}$ ,  $USP16^{fllox/0}$ /Lyz2-Cre) mice. (b) IB assays showing specific ablation of USP16 in the M-CSF-induced BMDMs of  $USP16^{MKO}$  mice. (c) IB assays showing the levels of USP16 in the macrophages and neutrophils isolated from the bone marrow (BM) of WT and  $USP16^{MKO}$  mice. (d) Flow cytometry analysis of macrophages ( $CD11b^+F4/80^+$ ) and neutrophils ( $CD11b^+Ly6G^+$ ) in BM from 6- to 8-week-old WT and  $USP16^{MKO}$  mice (n=3). (e-h) Flow cytometry analysis of the absolute numbers of different immune cells in the spleen (Spl) (e) and of Treg cells (f) and naïve and memory T cells (g) from WT or  $USP16^{MKO}$  mice (n=3). (h) Flow cytometric analysis of the percentages of IFN- $\gamma$  and IL-17-producing  $CD4^+$  T cells in the Spl of WT and  $USP16^{MKO}$  mice. All FACS data are presented in a representative plot and summary graph of the subpopulation percentages. All data are representative of three independent experiments. The bars and error bars show the means  $\pm$  SEMs. Significance was determined by the two-tailed Student's t test. \*P < 0.05.



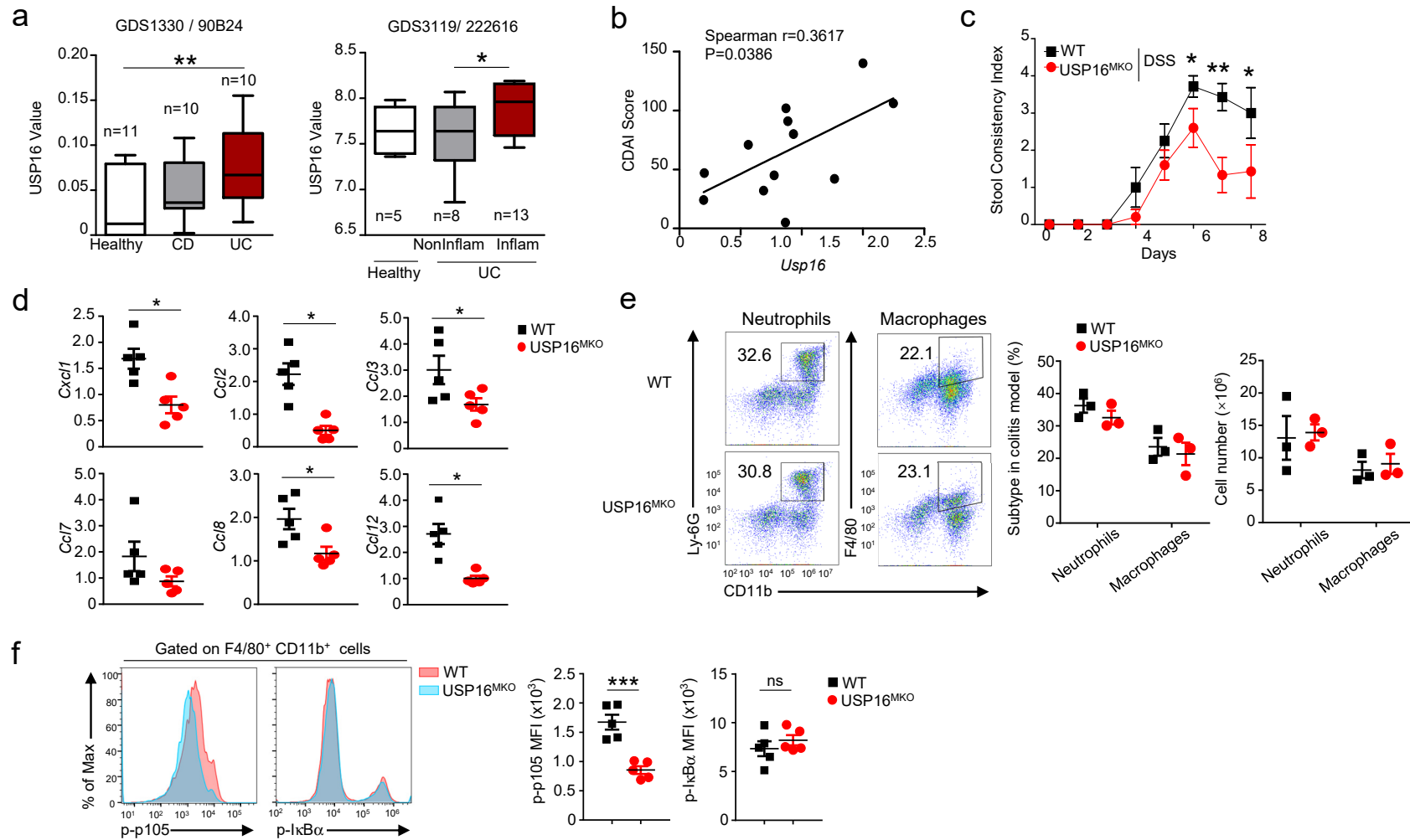
**Supplementary Figure 2 USP16 has no effect on the chromatin level and noncanonical NF- $\kappa$ B activation.** (a) Confocal microscopy analysis of IKK $\beta$  and USP16 in the WT BMDMs from **Figure 2c**. Quantification of IKK $\beta$ /USP16 colocalization was performed as described in the Materials and Methods using Pearson's correlation coefficient (n=50). (b) Immunoblot (IB) analysis of lys119-monoubiquitinated H2A (UbH2A), total H2A, and H3 (loading control) in lysates of WT and USP16-deficient macrophages stimulated with LPS as indicated. (c) The phosphorylation of MAPKs in whole-cell lysates of WT and USP16-deficient BMDMs was measured by IB analysis. (d) Immunoblot analysis of the indicated proteins in the cytoplasmic (CE) and nuclear (NE) extracts of WT or USP16<sup>MKO</sup> bone marrow cells stimulated with M-CSF (50 ng/ml) at the indicated time points. The data are representative of at least three independent experiments.



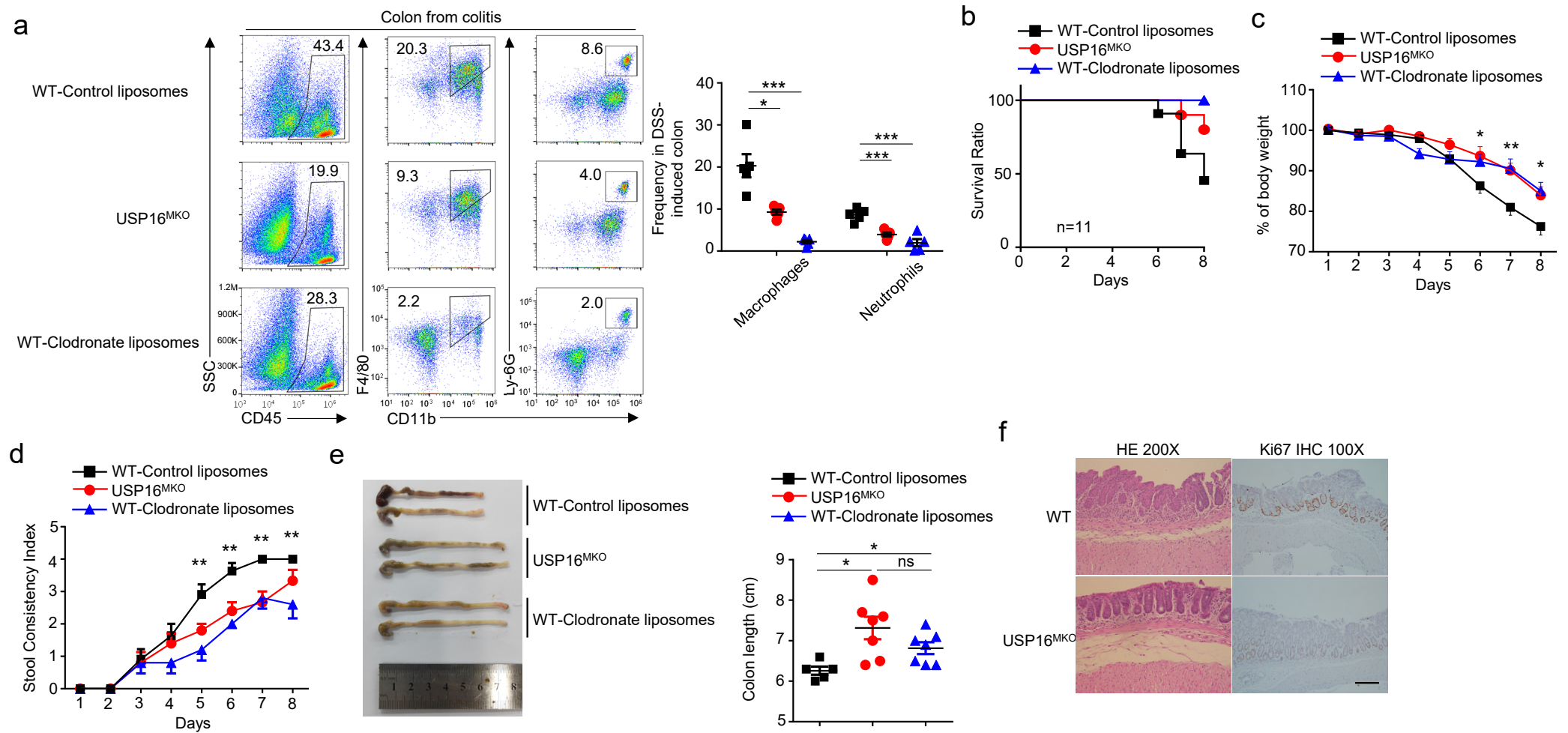
**Supplementary Figure 3 USP16 specifically regulates the ubiquitination of IKKβ.** (a-b) HEK293T cells were transfected with the indicated plasmids. The associations between p105 and various truncation mutants of IKKβ (a) or its kinase domain (b) were detected through the indicated IP and IB analyses. (c) HEK293T cells were transfected with an NF-κB-luciferase reporter plasmid in the presence (+) or absence (-) of the indicated empty vector or expression plasmids. Protein lysates were also subjected to direct IB. (d) HEK293T cells were transfected with K33R along with the indicated expression plasmids. FLAG-tagged IKKβ was isolated by IP, and the ubiquitination level was then detected by IB. (e) IKKβ was isolated by immunoprecipitation (IP) (under denaturing conditions) from whole-cell lysates of WT and USP16-deficient BMDMs treated with MG132, and then subjected to IB assays using anti-K48 linked ubiquitin (top panels). Protein lysates were also subjected to direct IB (bottom panels). (f) IKKβ-deficient MEFs were reconstituted with IKKβ<sup>WT</sup> or IKKβ<sup>K238R</sup>, and then Usp16 was knocked down by shRNA. IB analysis of the phosphorylation of p105 under TNF-α (50 ng/ml) stimulation was performed as indicated. The data are representative of at least three independent experiments.



**Supplementary Figure 4 USP16 deficiency inhibits proinflammatory cytokines.** (a-b) qRT-PCR analysis of mRNA (vertical axes) in WT or USP16-deficient BMDMs unstimulated (0 hr) or stimulated for 2 or 6 hr with pIC (200  $\mu$ g/ml; a) or R848 (1  $\mu$ g/ml; b). (c) qRT-PCR analysis of the indicated genes in WT or USP16-deficient MEFs stimulated for 2 or 6 hr with TNF- $\alpha$  (50 ng/ml). All qRT-PCR data are presented as the fold induction relative to the *Actb* mRNA level. The data are presented as the means  $\pm$  SEMs and are representative of at least three independent experiments. The statistical analysis results show the variations among experimental replicates. Two-tailed unpaired t-tests were performed. \*P < 0.05, \*\*P < 0.01.



**Supplementary Figure 5 USP16 is associated with the onset of colitis.** (a) Box plot of USP16 expression in the patients with CD or UC (using public data set GDS1330; n = 10, left). Box plot of USP16 mRNA level in the different area from the patients with UC (using public data set GDS3119, Right). (b) Spearman correlation analysis of USP16 mRNA levels in colon macrophages and CD activity index (CDAI) scores. (c) WT and USP16<sup>MKO</sup> mice were treated with 3% DSS described as in **main Figure 6d**. Stool Consistency Index of DSS-treated WT and USP16<sup>MKO</sup> mice. (d) qRT-PCR assay of chemokine levels in colon tissue of DSS-treated WT and USP16<sup>MKO</sup> mice. All qRT-PCR data are presented as the fold induction relative to the *Actb* mRNA level. (e) Flow cytometry analysis of macrophages (CD11b<sup>+</sup>F4/80<sup>+</sup>) and neutrophils (CD11b<sup>+</sup>Ly6G<sup>+</sup>) in bone marrow (BM) from WT and USP16<sup>MKO</sup> mice in DSS-induced colitis model (n=3). FACS data are presented in a representative plot and summary graph of the subpopulation percentages. (f) Phosphorylation (p-) of p105 and IκBα in macrophages from the colons of DSS-induced WT and USP16<sup>MKO</sup> mice (n=5), assessed by flow cytometry (gated on CD11b<sup>+</sup>F4/80<sup>+</sup> cells). The data are presented as the mean ± SEM values and representative of at least three independent experiments. Statistical analyses represent variations in experimental replicates. Two-tailed unpaired t-tests were performed. \*P < 0.05; \*\*, P < 0.01; \*\*\*, P < 0.005.



**Supplementary Figure 6 USP16 in macrophages is required for the onset of colitis. (a-e)** Six- to eight-week-old WT mice were i.v. injected with clodronate liposomes for three times every other day. WT and USP16<sup>MKO</sup> mice were treated with 3.5% (b) or 3% (a and c-e) DSS (in drinking water) for 5 continuous days and then supplied with normal drinking water. (a) FACS analysis results for the total immune cells (CD45<sup>+</sup>), macrophages (CD11b<sup>+</sup>F4/80<sup>+</sup>) and neutrophils (CD11b<sup>+</sup>Ly6G<sup>+</sup>), presented in a representative plot for multiple mice treated with clodronate liposomes (n=5). Survival rates (b), body weight loss (c), stool consistency index (d) and colon length (e) of DSS-treated WT and USP16<sup>MKO</sup> mice. (f) Representative images of Ki-67 staining of colon tissues of DSS-treated WT and USP16<sup>MKO</sup> mice (Scale bar: 20 $\mu$ m). Photo credits: Jian-shuai Yu and Yu Zhang (Zhejiang University). The data are presented as the means  $\pm$  SEMs and are representative of at least three independent experiments. The statistical analysis results show the variations among experimental replicates. Two-tailed unpaired t-tests were performed. \*P < 0.05, \*\*P < 0.01; \*\*\*P < 0.005.

**Supplementary Table 1 The gene-specific PCR primers.** The forward and reverse sequences of the primers used for the qPCR assay. All of primers were designed by Oligo 7 v7.56 based on the mouse genes.