
Supplemental Information

**Cytoplasmic SHMT2 drives the progression and metastasis of colorectal cancer
by inhibiting β -catenin degradation**

Supporting information includes:

8 Supplementary Figures

3 Supplementary Tables

Figure S1

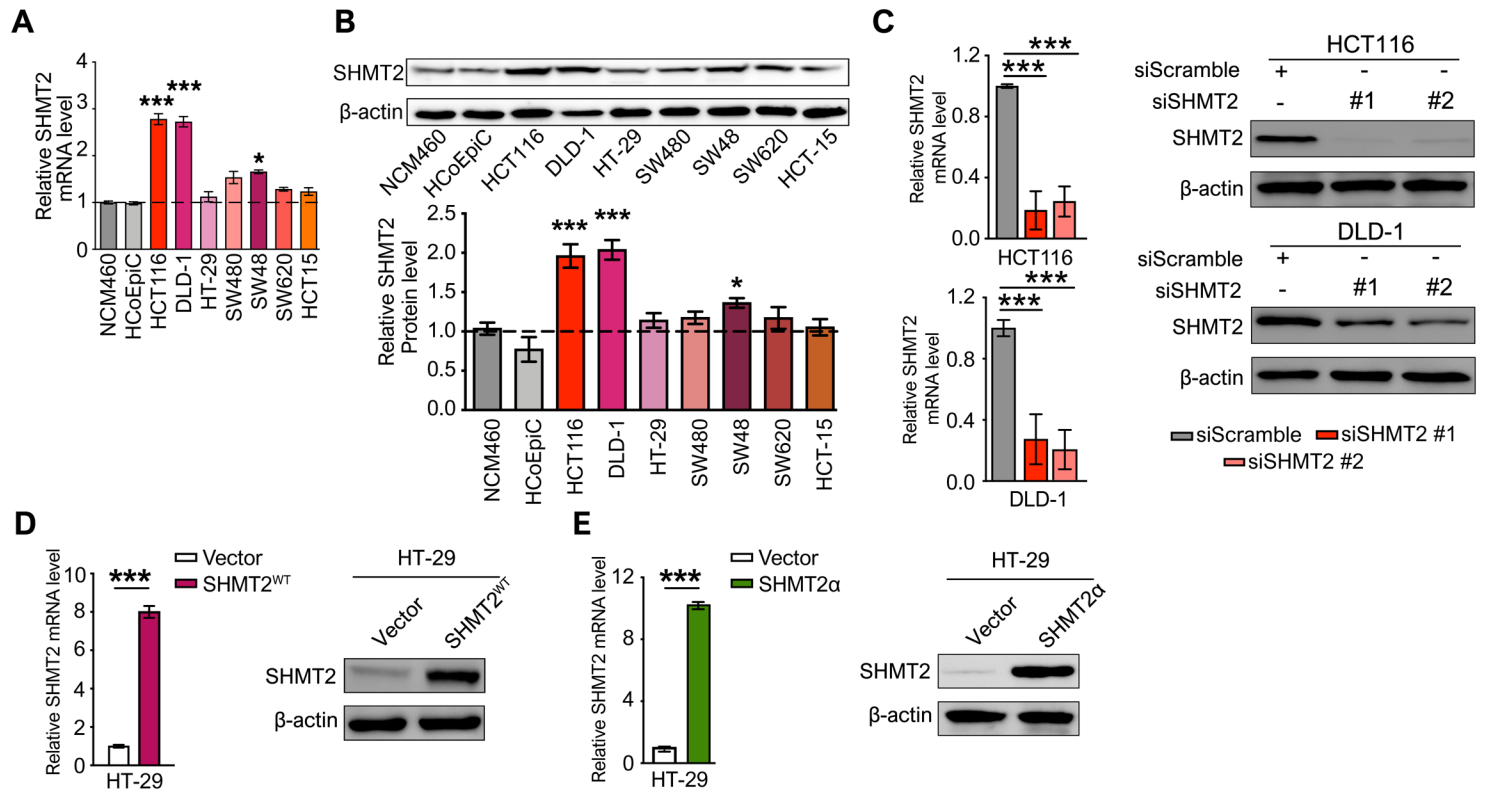
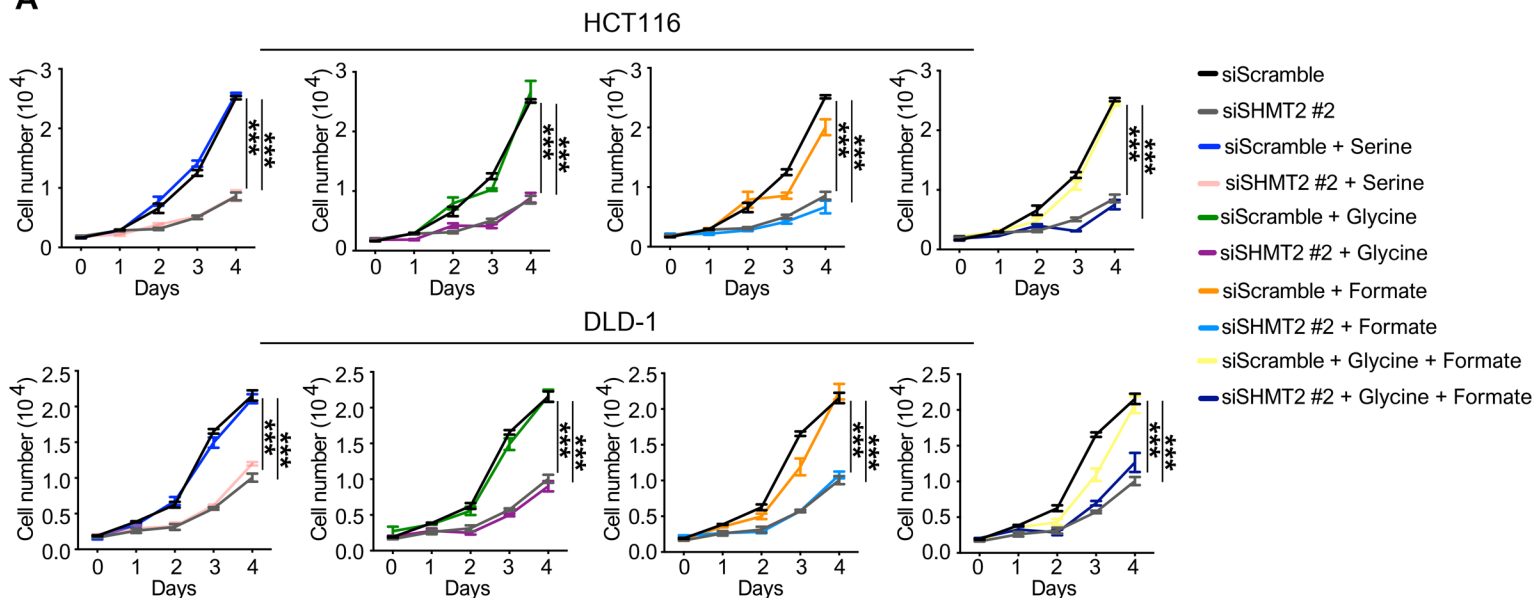
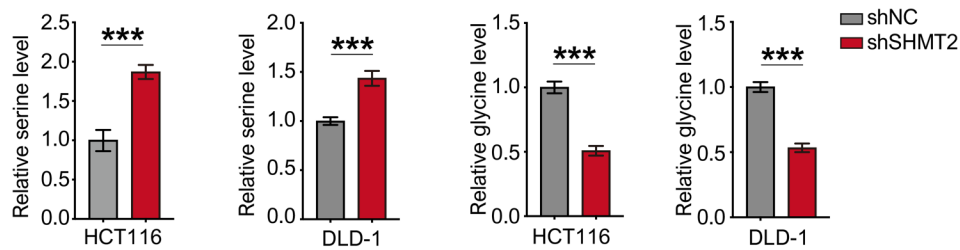


Figure S2

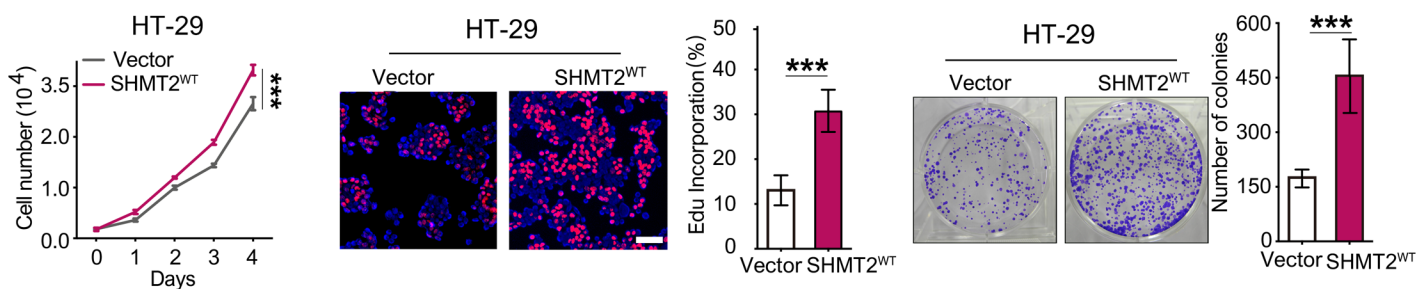
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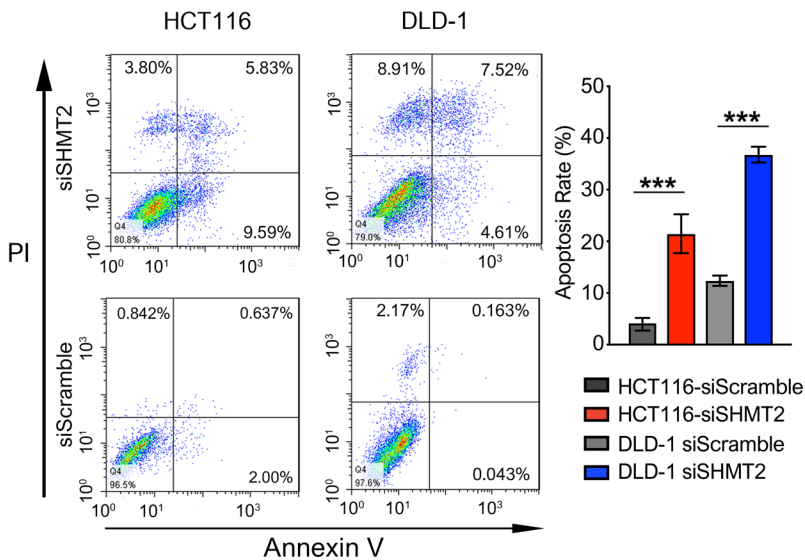
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C



D



E

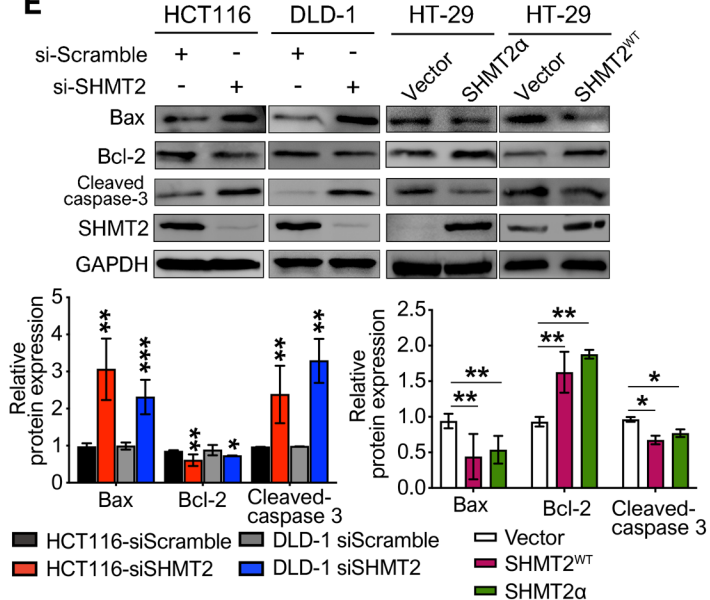


Figure S3

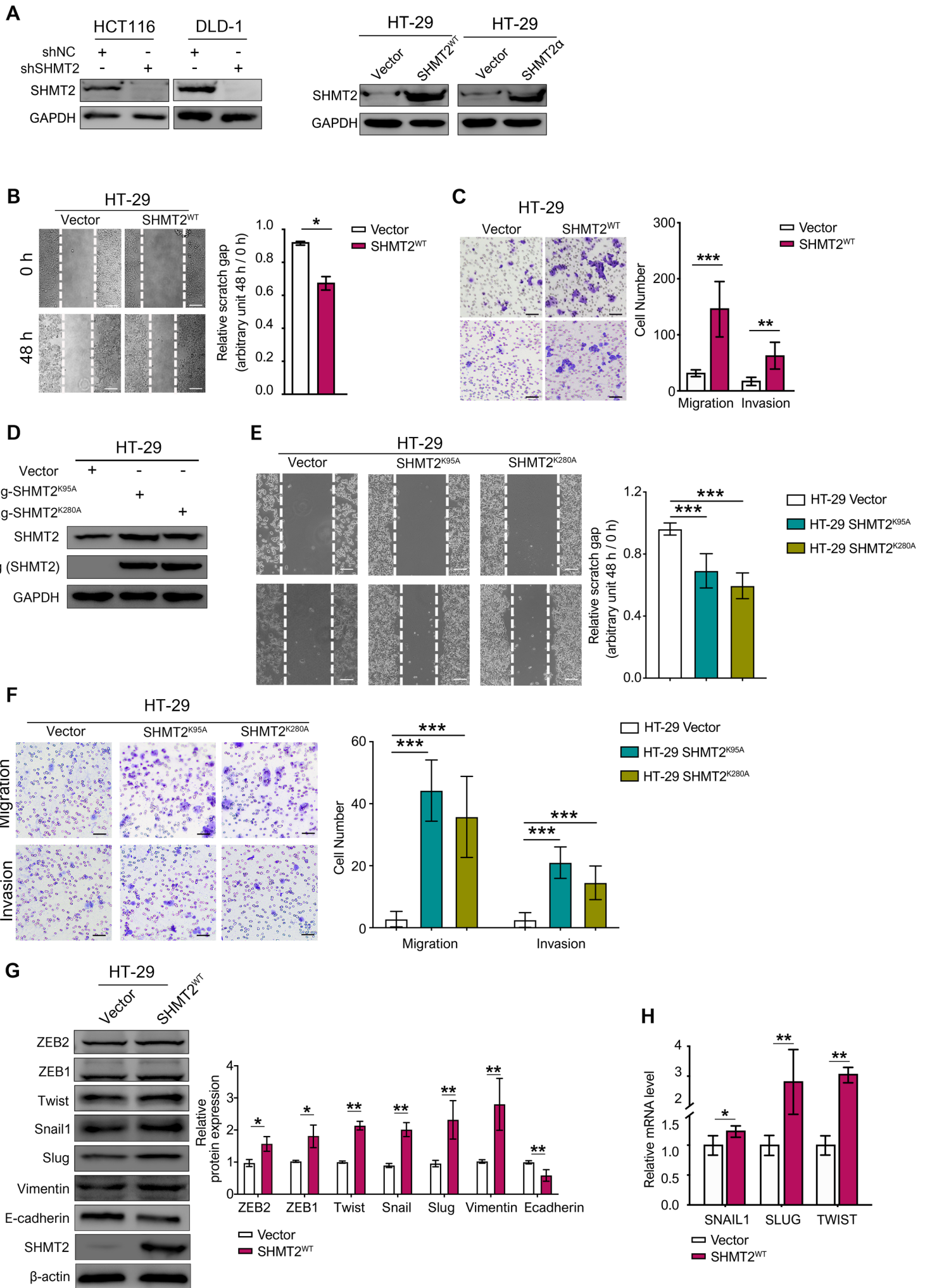


Figure S4

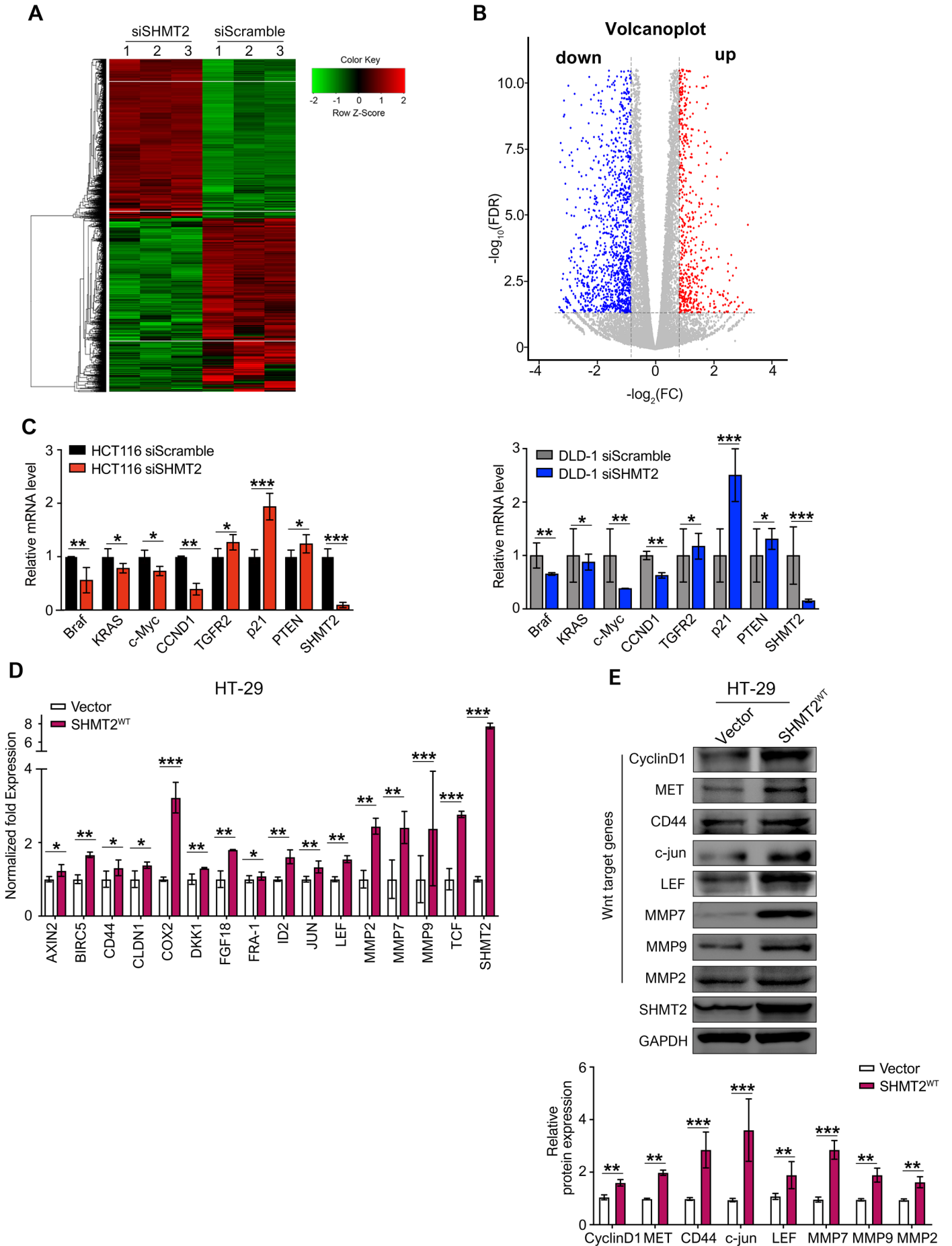


Figure S5

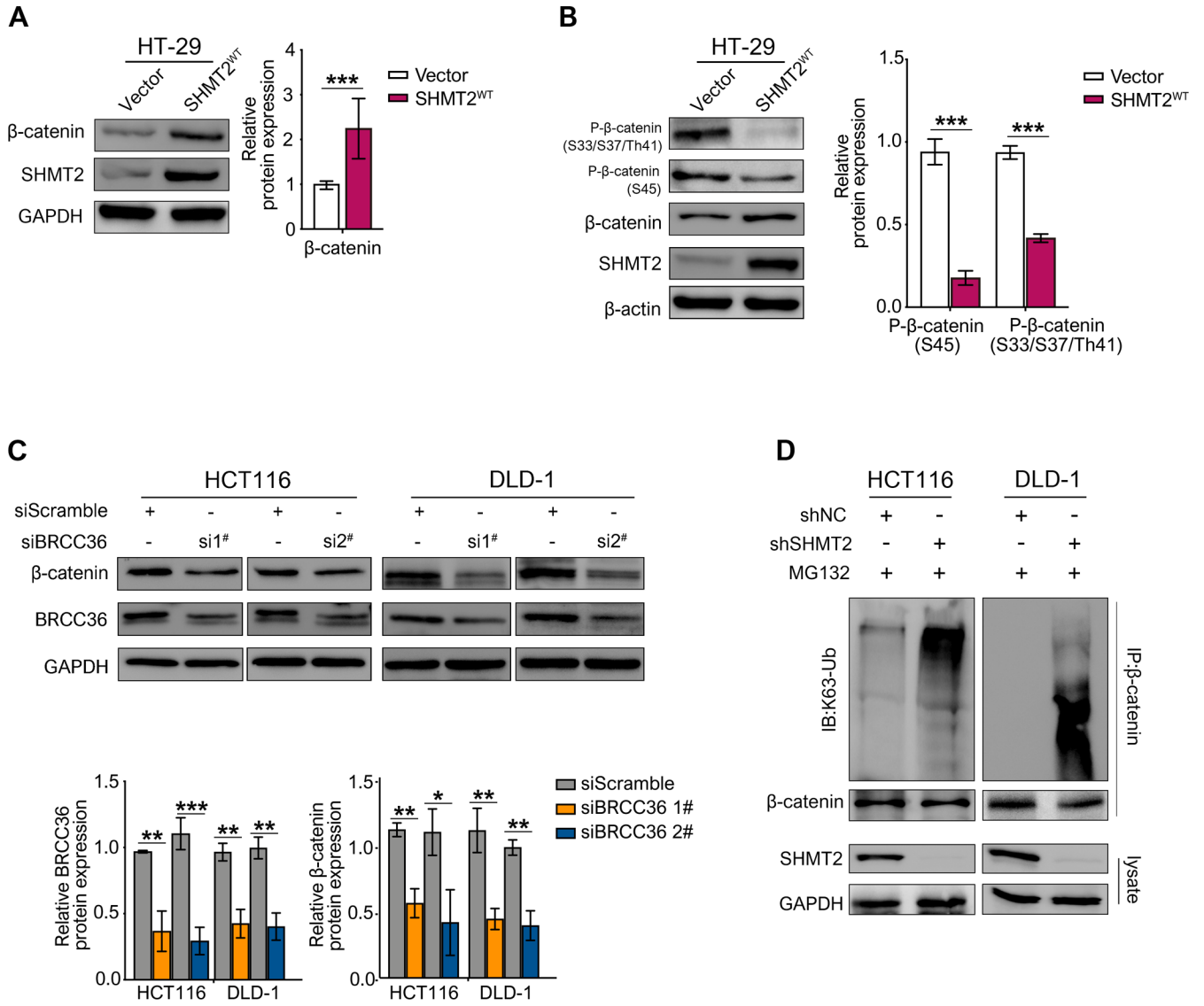


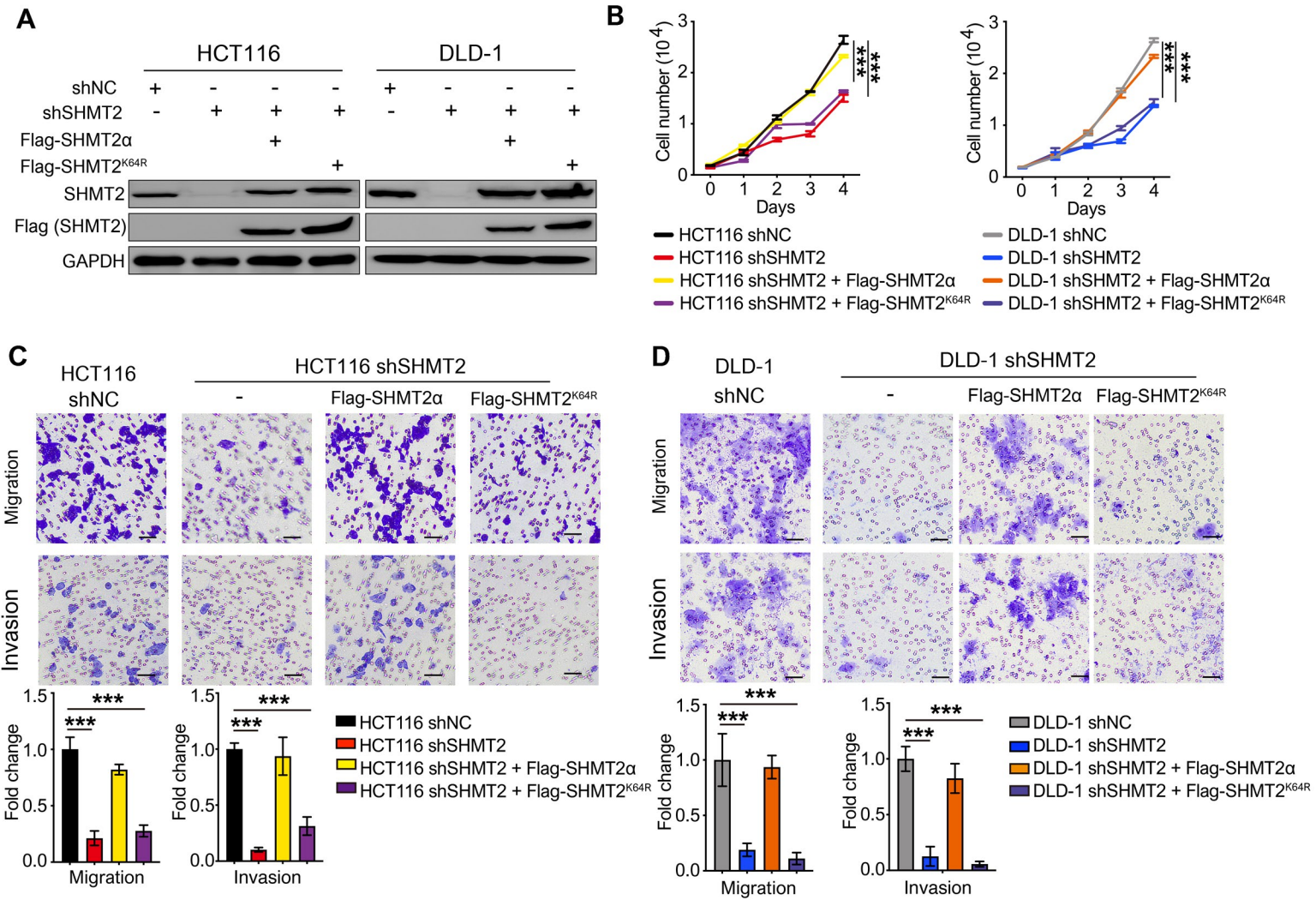
Figure S6

Figure S7

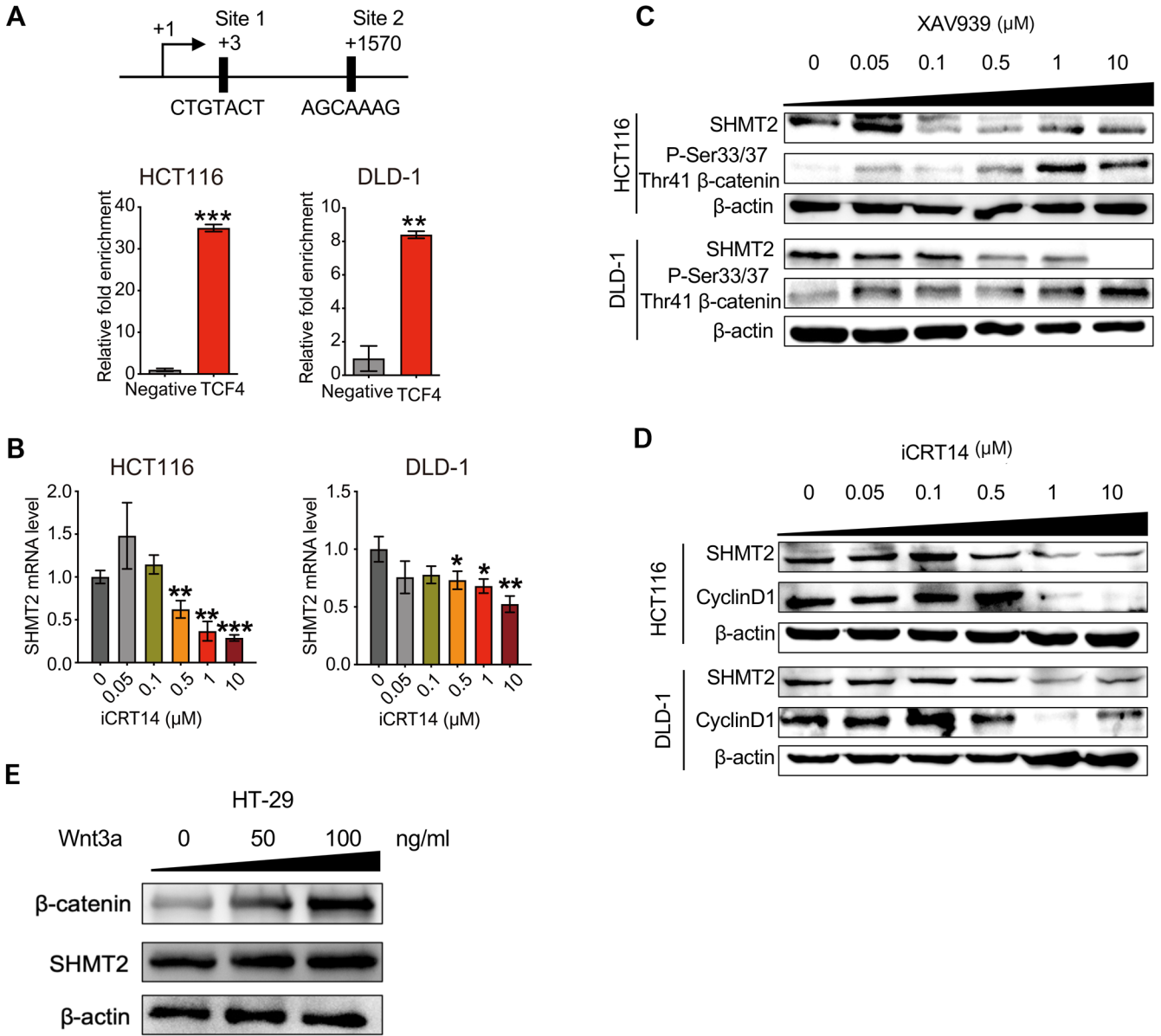


Figure S8

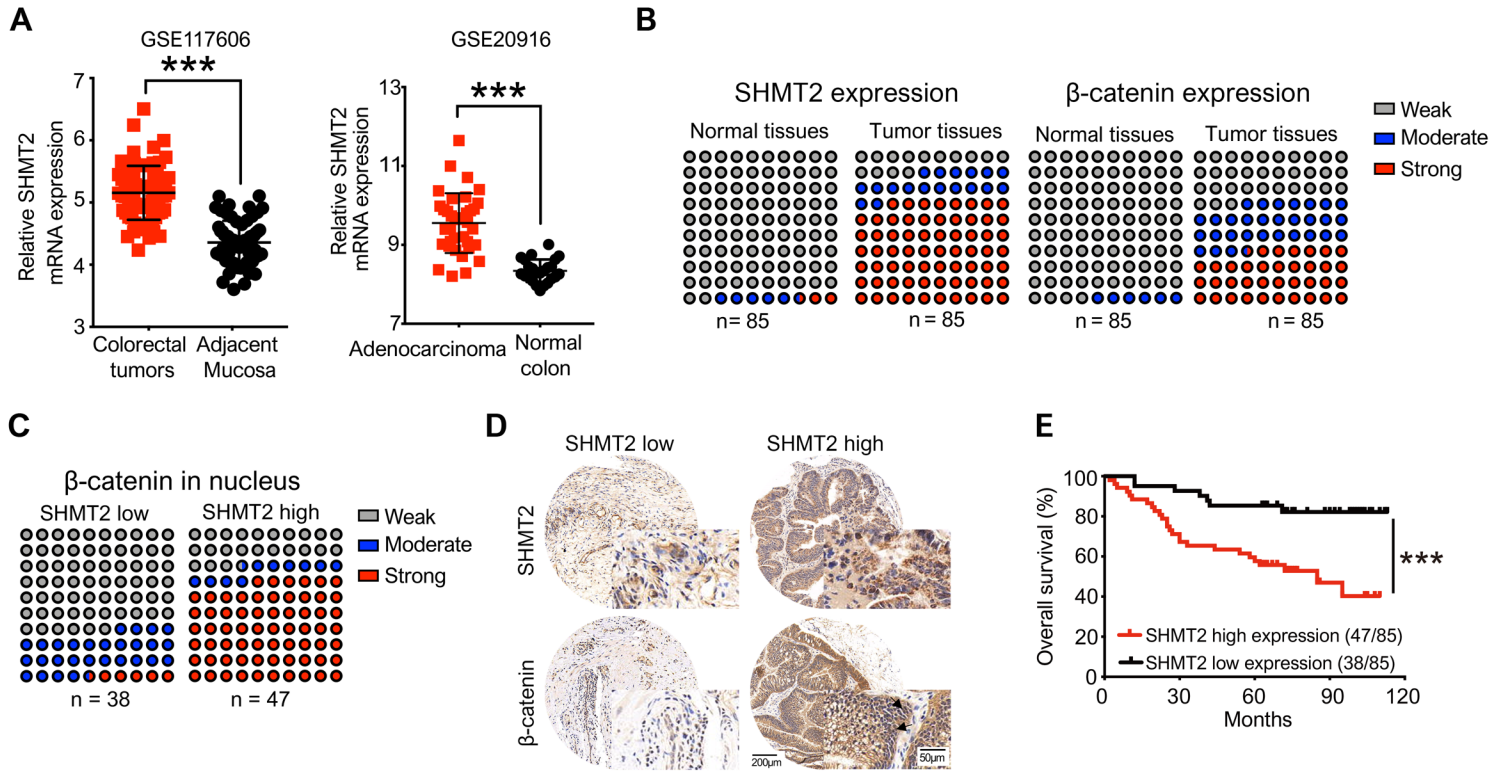


Figure S1. SHMT2 is overexpressed in human CRC cells. Related to Figure 1.

(A) The relative mRNA level of SHMT2 in human CRC cells and normal colonic epithelial cell NCM460 and HCoEpiC. * $p < 0.05$, *** $p < 0.001$; Student's t-test.

(B) Western blot analysis (up) and quantification (bottom) of SHMT2 in human CRC cells and normal colonic epithelial cell NCM460 and HCoEpiC. * $p < 0.05$, *** $p < 0.001$; Ordinary one-way ANOVA multiple comparisons test.

(C) The knockdown of SHMT2 by siRNA in HCT116 and DLD-1 cells were confirmed by RT-qPCR and western blot analysis. *** $p < 0.001$; Ordinary one-way ANOVA multiple comparisons test.

(D) (E) The overexpression of SHMT2^{WT} and SHMT2 α in HT-29 cells was confirmed by RT-qPCR and western blot analysis. *** $p < 0.001$; Student's t-test.

Data are presented as the mean \pm SD from at least three independent experiments. The error bars show the SD.

Figure S2. SHMT2 promotes CRC cell proliferation and regulates cell apoptosis. Related to Figure 1.

(A) Cells proliferation was detected by CCK8 assay by siSHMT2 #2, with or without 400 μ M serine, 400 μ M glycine or 1mM formate supplementation (n = 3). *** $p < 0.001$; Ordinary one-way ANOVA multiple comparisons test.

(B) CRC cells with shSHMT2 were harvested in 80% confluence. Serine and glycine levels in cells were analyzed by LC-MS (n = 3). The relative quantities of metabolites were shown. *** $p < 0.001$; Student's t-test.

(C) Representative images and quantification of cell proliferation (left) (n = 3), EdU incorporation assay (middle) (n = 5) and colony formation assay (right) (n = 3) using HT29 cells with SHMT2^{WT} overexpression. Scale bar, 50 μ m. ** $p < 0.01$; Student's t-test.

(D) HCT116 and DLD-1 cells with siSHMT2 were stained by annexin V/propidium iodide (PI), followed by flow cytometry (n = 3). The relative quantities of the apoptosis rate (%) were shown. *** $p < 0.001$; Student's t-test.

(E) Western blot analysis (up) and quantification (bottom) of SHMT2, BAX, Bcl-2 and Cleaved-caspase-3 in CRC cells with SHMT2 knockdown, SHMT2 α and SHMT2^{WT} overexpression. * $p < 0.1$, ** $p < 0.05$, *** $p < 0.001$; Student's t-test and Ordinary one-way ANOVA multiple comparisons test.

Data are presented as the mean \pm SD from at least three independent experiments. The error bars show the SD.

Figure S3. The catalytic function of SHMT2 is not indispensable for the migration and invasion ability of HT-29 cells. Related to Figure 2.

(A) The knockdown of SHMT2 by shRNA in HCT116 and DLD-1 cells were confirmed by western blot analysis. The overexpression of SHMT2^{WT} and SHMT2 α in HT-29 cells was confirmed by western blot analysis.

(B) Representative images (left) and quantification (right) of the wound healing assay using HT-29 cells with SHMT2^{WT} overexpression (n = 3). Scale bar, 200 μ m. * $p < 0.05$; Student's t-test.

(C) Representative images (left) and quantification (right) of the migration and invasion assay using HT-29 cells with SHMT2^{WT} overexpression (n = 3). Scale bar, 200 μ m. ** $p < 0.01$; Student's t-test.

(D) The overexpression of Flag-SHMT2^{K95A} and Flag-SHMT2^{K280A} in HT-29 cells were confirmed by western blot analysis.

(E) Representative images (left) and quantification (right) of the wound healing assay using HT-29 cells with Flag-SHMT2^{K95A} and Flag-SHMT2^{K280A} overexpression (n = 3). Scale bar, 200 μ m. *** $p < 0.001$; Ordinary one-way ANOVA multiple comparisons test.

(F) Representative images (left) and quantification (right) of migration and invasion assay using HT-29 cells with Flag-SHMT2^{K95A} and Flag-SHMT2^{K280A} overexpression (n = 3). Scale bar, 100 μm. *** $p < 0.001$; Ordinary one-way ANOVA multiple comparisons test.

(G) Western blot analysis (left) and quantification (right) of vimentin, E-cadherin and EMT-associated transcription factors in HT-29 cells after SHMT2^{WT} overexpression. * $p < 0.05$, ** $p < 0.01$; Student's t-test.

(H) RT-qPCR analysis of EMT-associated transcription factors in HT-29 cells after SHMT2^{WT} overexpression (n = 3). * $p < 0.05$, ** $p < 0.01$; Student's t-test.

Data are presented as the mean ± SD from at least three independent experiments. The error bars show the SD.

Figure S4. SHMT2 knockdown affects the expression of multiple genes in CRC cells. Related to Figure 3.

(A) Heatmap and sample clustering analysis of the mRNA-seq analysis of HCT116 cells after SHMT2 knockdown. Each column represents a sample, and each row represents a gene. Red represents upregulated genes, and the green represents downregulated genes (n = 3).

(B) Volcano plots of the mRNA-seq analysis of HCT116 cells after SHMT2 knockdown. Blue dots represent downregulated genes, and red dots represent upregulated genes (FDR < 0.05) (n = 3).

(C) RT-qPCR was performed to verify the representative genes obtained from mRNA-seq results in HCT116 and DLD-1 cells (n = 3). * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$; Student's t-test.

(D) RT-qPCR analysis of Wnt target genes with SHMT2^{WT} overexpression (n = 3). * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$; Student's t-test.

(E) Western Blot analysis (top) and quantification (bottom) of Wnt target genes in HT-29 cells with SHMT2^{WT} overexpression. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$;

Student's t-test.

Data are presented as the mean \pm SD from at least three independent experiments. The error bars show the SD.

Figure S5. BRCC36 knockdown inhibits the β -catenin expression, and SHMT2 knockdown increases K63-Ub of β -catenin. Related to Figure 4.

(A) Western blot analysis (left) and quantification (right) of β -catenin in HT-29 cells after SHMT2^{WT} overexpression. *** $p < 0.001$; Student's t-test.

(B) Western blot analysis (left) and quantification (right) of phosphorylated β -catenin after SHMT2^{WT} overexpression in HT-29 cells. *** $p < 0.001$; Student's t-test.

(C) Western blot analysis (up) and quantification (bottom) of β -catenin after BRCC36 knockdown in CRC cells. * $p < 0.1$, ** $p < 0.05$, *** $p < 0.001$; Student's t-test.

(D) CRC cells stably expressing shNC or shSHMT2 were treated with MG132 for 16 h before harvesting. β -catenin was immunoprecipitated and immunoblotted with the indicated antibodies.

Data are presented as the mean \pm SD from at least three independent experiments. The error bars show the SD.

Figure S6. K64 of SHMT2 is critical for CRC cell proliferation, migration, and invasion. Related to Figure 5.

(A) The overexpression of Flag-SHMT2 α and Flag-SHMT2^{K64R} in HCT116 and DLD-1 was confirmed by western blot analysis.

(B) Cell proliferation was detected by a CCK8 assay after stable SHMT2 knockdown with the re-expression of shRNA-resistant SHMT2 α or SHMT^{K64R} (n = 3). *** $p < 0.001$; Ordinary one-way ANOVA multiple comparisons test.

(C-D) Representative images (top) and statistically analyzed (bottom) of the transwell migration and invasion assay using CRC cells after stable SHMT2 knockdown with the

re-expression of shRNA-resistant SHMT2 α or SHMT^{K64R} (n = 3). Scale bar, 100 μ m.
*** $p < 0.001$; Ordinary one-way ANOVA multiple comparisons test.

Data are presented as the mean \pm SD from at least three independent experiments. The error bars show the SD.

Figure S7. SHMT2 is a target gene of the Wnt/ β -catenin pathway

(A) The software PROMO was used to identify a putative binding site of TCF4 (TCF4 BS) in the SHMT2 gene (top). ChIP-qPCR analysis of the enrichment of TCF-4 on the SHMT2 promoter in HCT116 and DLD-1 cells (bottom) (n = 3). ** $p < 0.01$, *** $p < 0.001$; Student's t-test.

(B) RT-qPCR analysis of SHMT2 expression in HCT116 and DLD-1 cells treated with Wnt signal inhibitors iCRT14 (n = 3). * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$; Ordinary one-way ANOVA multiple comparisons test.

(C) Western blot analysis of SHMT2 and phosphorylated β -catenin expression in HCT116 and DLD-1 cells treated with the Wnt signal inhibitor XAV939.

(D) Western blot analysis of SHMT2 and Cyclin D1 expression in HCT116 and DLD-1 cell treated with the Wnt signal inhibitor iCRT14.

(E) Western blot analysis of SHMT2 and β -catenin in HT-29 cells with Wnt3a treatment. Data are presented as the mean \pm SD from at least three independent experiments. The error bars show the SD.

Figure S8. SHMT2 and β -catenin expression in CRC patients. Related to Figure 8.

(A) The relative mRNA expression of SHMT2 in two GEO datasets (GSE117606 and GSE20916). Data are presented as the mean \pm SD. The error bars show the SD. *** $P < 0.001$; Student's t-test.

(B) Quantitation of SHMT2 and β -catenin expression in CRC tissues and NATs in 85 human CRC specimens.

(C) The expression of nucleus β -catenin in CRC tissues with low or high expression of

SHMT2.

(D) Representative IHC staining images of SHMT2 and β -catenin in human CRC tissues. The arrow points to the position where β -catenin enters the nucleus: scale bar, 200 μm , or 50 μm .

(E) Survival analysis of the correction of SHMT2 expression in CRC tissues with patients' survival rate. *** $P < 0.001$; Log-rank test.

Table S1. Go pathway analysis. Related to Figure 3.

Style	Pathway Term	DifGene	P-Value	Enrichment
down	Wnt signaling pathway	15	0.0233	1.3669
down	TGF- β signaling pathway	12	0.0190	1.9454
down	Arginine and proline metabolism	8	0.0281	1.8905
down	Alanine, aspartate and glutamate metabolism	6	0.0308	2.3237
down	Glycine, serine, and threonine metabolism	6	0.0272	2.1449
down	Notch signaling pathway	6	0.0326	1.7428
down	beta-Alanine metabolism	5	0.0298	2.3237
down	Histidine metabolism	5	0.0373	2.2487

Table S2. Clinicopathologic characters of CRC patients. Related to Figure 8.

Characteristic	Cohort 1, n
Patients	85
Age, yr (median, range)	63 (33-86)
Gender	
Male	49 (57%)
Female	36 (43%)
T stages	
T1-T2	3 (3%)
T3-T4	82 (97%)
Lymph node involvement	
No	51 (60%)
Yes	34 (40%)
Distance metastasis	
No	85 (100%)
Yes	N
AJCC stages	
I-II	49 (57%)
III-IV	36 (43%)

Table S3. Primers used in this study.

Primer set	Forward	Reverse
<i>β-actin</i>	TGACGTGGACATCCGCAAAG	CTGGAAGGTGGACAGCGAGG
<i>SHMT2</i>	CCTGCCCTGAGTTTCCATTA	GTCTGGTGTCTGGCTCTGAT
<i>TGFR2</i>	CGGTAA TAACGACATGATAGTCAC	TCATGGCAAACGTCTCTAGTGTTA
<i>p21</i>	GGACAGCAGAGGAAGACCATGT	TGGAGTGGTA GAAATCTGTCTATGC
<i>c-Myc</i>	GCCACGTCTCCACACATCAG	TGGTGCATTTTCGGTTGTTG
<i>PTEN</i>	CAATGTTTCACTGGCGGAA	TCGTGTGGGTCCTGAATT
<i>KRAS</i>	ACAGAGAGTGGAGGATGCTTT	TTTCACACAGCCAGGAGTCTT
<i>BRAF</i>	CAAACCTATAGATATTGCACA	TCTGGTGCCATCCACAAAATG
<i>CCND1</i>	CCCTCGGTGTCCTACTTCA	CTCCTCGCACTTCTGTTTCT
<i>TCF</i>	CTCCGATGTCCACTTTCCAT	CGCTTCCTCTATTTGCCATT
<i>CTNFB1</i>	TGGTGACAGGGAAGACATCA	CCATAGTGAAGGCGAAGTGC
<i>CD44</i>	TCAGAGGAGTCGGAGAGAGGAAAC	GAAAAGTCAAAGTAACAATAACGTGG
<i>COX2</i>	CGGCGGACTAATCTTCAACT	CGGTGAAAGTGGTTTGGTTT
<i>FGF18</i>	AGTGGGAAACACATCCAGGT	CGGACTTGACTACCGAAGG
<i>FRA-1</i>	TGACCACACCCTCCCTAACT	CTGCTGCTACTCTTGCGATG
<i>c-JUN</i>	AAAGGAAGCTGGAGAGAATCG	TGTTCCCTGAGCATGTTGG
<i>MET</i>	TGCACAGTTGGTCTGCCATGA	CAGCCATAGGACCGTATTTTCGG
<i>LEF</i>	ACCTCTTGGCTGGTTTTCCGTCA	GTCAGTGTGGGGATGTTTCT
<i>MMP2</i>	CCATCGAGACCATGCGGAAGC	CATCGCTCCAGACTTGGAAGGC
<i>MMP7</i>	ACCGTGCTGTGTGCTGTGT	GTCCTGAGCCTGTTCCCACT
<i>MMP9</i>	TTCTGCCAGGACCGCTTCTAC	AGGATGTCATAGGTCACGTAGCC
<i>Twist</i>	CGGACAAGCTGAGCAAGATT	ACGGAAGGCGTAGCTGA
<i>Slug</i>	CAGCTCCACTCCACTCTCCT	TGAACCACTGTGATCCTTGG
<i>Snail</i>	CACACTGGTGAGAAGCCATT	GCCTGGCACTGGTATCTCTT