# D,L-methadone causes leukemic cell apoptosis via an OPRM1-triggered increase in IP3R-mediated ER Ca<sup>2+</sup> release and decrease in Ca<sup>2+</sup> efflux, elevating [Ca<sup>2+</sup>]<sub>i</sub>

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#### **Supplementary Materials and Methods**

**mPTP opening assay** Calcein fluorescence was measured using the Image-IT live mitochondria permeability transition pore assay kit (Thermo Fisher Sci.) as per the manufacturer's protocol. Briefly, cells ( $1 \times 10^4$ ) pre-incubated with cyclosporin A (CsA; 1 µM) for 1 hr then treated with D,L-methadone for 16 hrs were incubated with or without ionomycin (1 µM). Cells were then stained with calcein AM (1 µM) for 15 min then with CoCl<sub>2</sub> (1 mM) in Hank's balanced salt solution (HBSS) for 15 min. Cells were washed with HBSS, resuspended in ice-cold PBS and analyzed by flow cytometry using a fluorescein isothiocyanate filter (530 nm) for measuring calcein fluorescence.

#### Supplementary Figure Legend

**Supplementary Figure 1**. \*+pRS-sh*OPRM1* cells exhibit resistance to L-asparaginase (ASNase) and D,Lmethadone (Met). Cells were treated with 50 mIU of ASNase (left panel) or 0.5  $\mu$ g/ml Met (right panel) for 24, 48, and 72 hrs. Surviving cell fractions were quantified using Alamar blue assay. Values are means  $\pm$  SEM from three independent experiments. \*\*p<0.05.

**Supplementary Figure 2.** OPRM1 loss inhibits L-asparaginase- and D,L-methadone-induced apoptosis. Flow cytometry analysis was performed in cells treated with 50 mIU ASNase or 0.5  $\mu$ g/ml Met for 12 hrs. Data are presented in pseudocolor plots, with green and brown dots correspond to live and apoptotic cells, respectively. Dataset represent one of three experiments presented in Figure 1D.

**Supplementary Figure 3.** The pattern of  $Ca^{2+}$  extrusion following D,L-methadone treatment in the presence of 500  $\mu$ M external  $Ca^{2+}$  is similar to that of 2 mM (as shown in Figure 4A) except that amplitudes of  $Ca^{2+}$  entry in both OPRM1-depleted and control cells are reduced.

**Supplementary Figure 4.** Activation of calpain-1 is involved in D,L-methadone-induced leukemic cell apoptosis. Cells pretreated (or untreated) with 10  $\mu$ M calpeptin then treated with D,L-methadone for 16 hrs were analyzed by flow cytometry following Annexin V-FITC and propidium iodide staining. Values are means ± SEM from three (n = 3) independent experiments. \*\*p<0.05.

**Supplementary Figure 5.** mPTP opening is not involved in D,L-methadone-induced leukemic cell apoptosis. Cells ( $1 \times 10^4$ ) treated with D,L-methadone were stained with 1 µM calcein-AM then CoCl<sub>2</sub> and analyzed by flow cytometry (left panel). 1 µM ionomycin and 1 µM cyclosporin A (CsA) were used as positive and negative controls, respectively. Quantitative analysis of the relative calcein fluorescence is shown on the right panel. Values for cells loaded with calcein-AM alone were normalized to 1.0. The relative calcein fluorescence intensity in cells treated with CoCl<sub>2</sub> were then calculated. Values are means ± SEM from three (n = 3) independent experiments. \*\*p<0.05 by unpaired Student's t test.

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Supplementary Fig. 1



Supplementary Fig. 2



Supplementary Figure 3



Supplementary Fig. 4



Supplementary Figure 5

Note: some of the blots were cut to allow immunoblotting with different antibodies without reblotting the whole membrane.







-Figure-6B.

### Figure 6B

## Fig. 1A additional data







#2

#3





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Fig. 6B additional data



For right panel

