

## Supporting Information for:

# Ring-Expansion Leads to A More Potent Analogue of Ipomoeassin F

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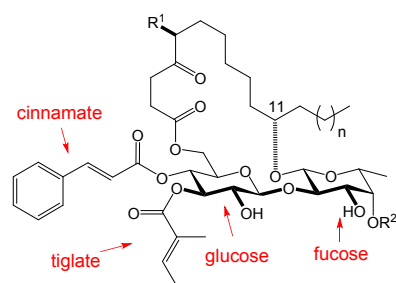
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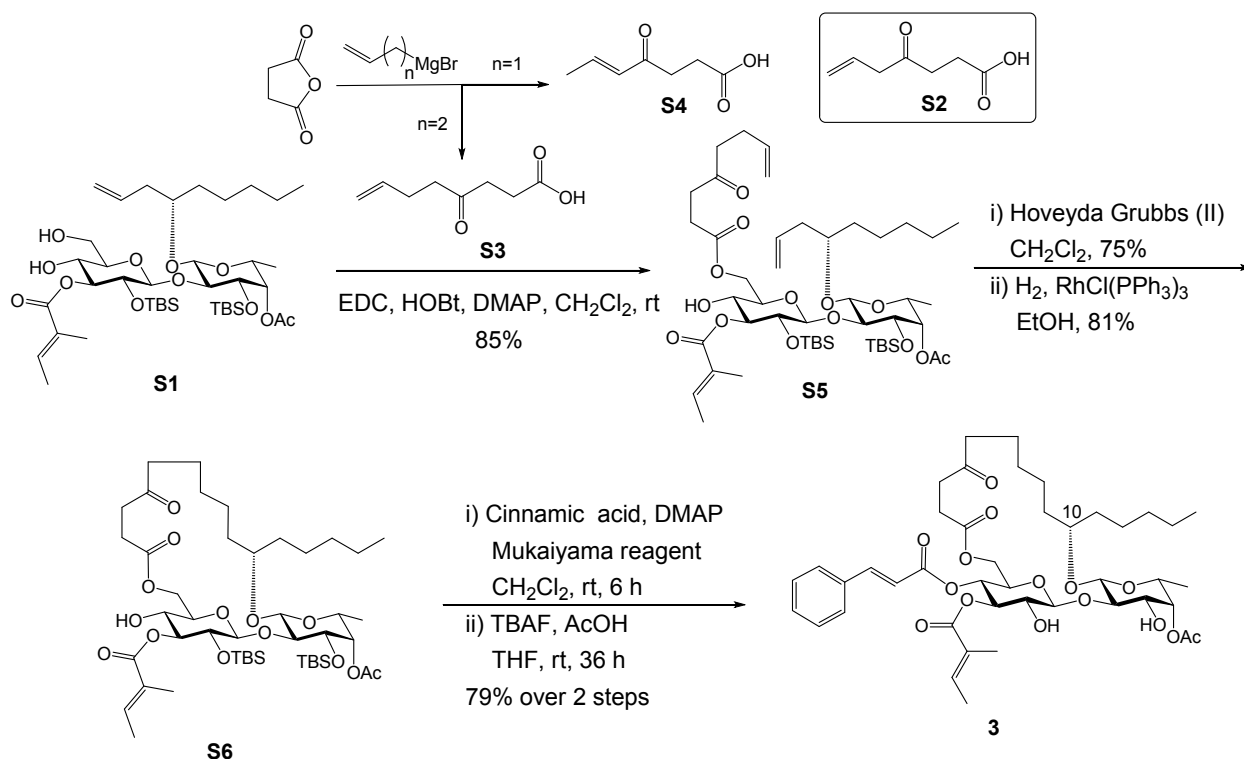
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Ipomoeassin	Structure		
	R <sup>1</sup>	R <sup>2</sup>	n
A	H	Ac	1
B	H	H	1
C	OH	Ac	1
D	OAc	Ac	1
E	OAc	H	1
F	H	Ac	3

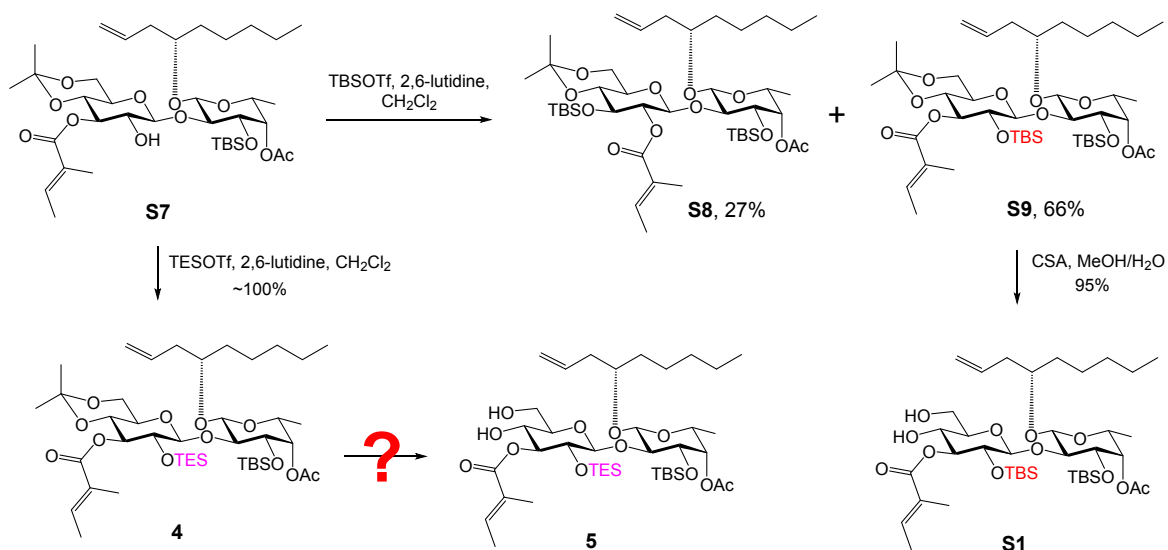
**Figure S1.** Structures of ipomoeassins A–F.

### Scheme S1. Synthesis of 19-Membered Ring Analogue 3.



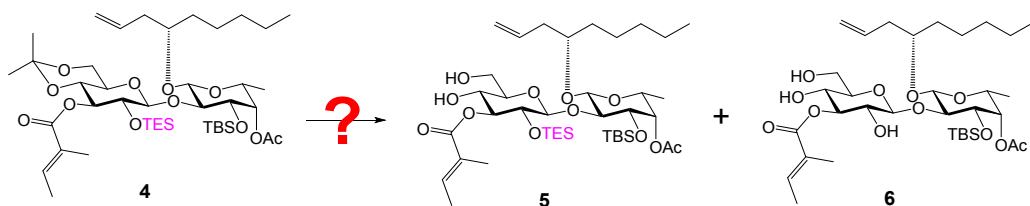
Initially, we thought that the 18-membered ring analogue **1** could be easily obtained by coupling **S1** with 4-oxo-6-heptenoic acid **S2**. After a literature search, however, we realized that the reaction of succinic anhydride with allylmagnesium bromide would cause alkene migration, leading to (*E*)-4-oxo-5-heptenoic acid **S4** (see the inset, Scheme S1) due to the acidic  $\alpha$ -Hs of the carbonyl group and higher stability of internal vs. terminal alkenes. This was confirmed by our initial attempts. Therefore, we revised our plan to synthesize the 19-membered ring analogue **3** (Figure 1) instead. As expected, 4-oxo-7-octenoic acid **S3** was successfully prepared from succinic anhydride and 3-butenylmagnesium bromide. EDC-mediated esterification installed the acid **S3** onto the primary alcohol in **S1** to generate the diene **S5** in good yield. Subsequent ring-closing metathesis (RCM), followed by chemo-selective hydrogenation catalyzed by Wilkinson's catalyst, successfully produced the 19-membered ring alcohol precursor **S6**. After cinnamic acid was introduced to 4-OH-Glc<sub>p</sub> (glucopyranose), both TBS protecting groups were removed by tetra-*n*-butylammonium fluoride (TBAF) buffered with acetic acid to deliver the final 19-membered ring analogue **3**.

## Scheme S2. Syntheses of Diol Intermediates S1 and 5.



Although the desired TBS-protected compound **S9** was the major product, a significant amount of by-product **S8**, with tiglate migration from 3-*O*-Glcp (glucopyranose) to 2-*O*-Glcp, was also generated (~27%). More importantly, given the very small polarity difference between **S8** and **S9**, purification of **S9** by column chromatography was time-consuming and labor-intensive. To overcome these two shortcomings, we first explored switching the 2-OH-Glcp protecting group of **4** from TBS to TES, envisioning that the comparatively reduced steric bulk of the TES group would encounter less steric hindrance from the existing bulky 3-*O*-Fucp (fucosepyranose) TBS group and, hence, be more efficiently incorporated than TBS. Encouragingly, we saw an almost quantitative transformation of **S7** to **4** when TESOTf, in place of TBSOTf, was employed as the silylation reagent.

**Table S1. Optimization of Chemoselective Removal of Isopropylidene.<sup>a</sup>**

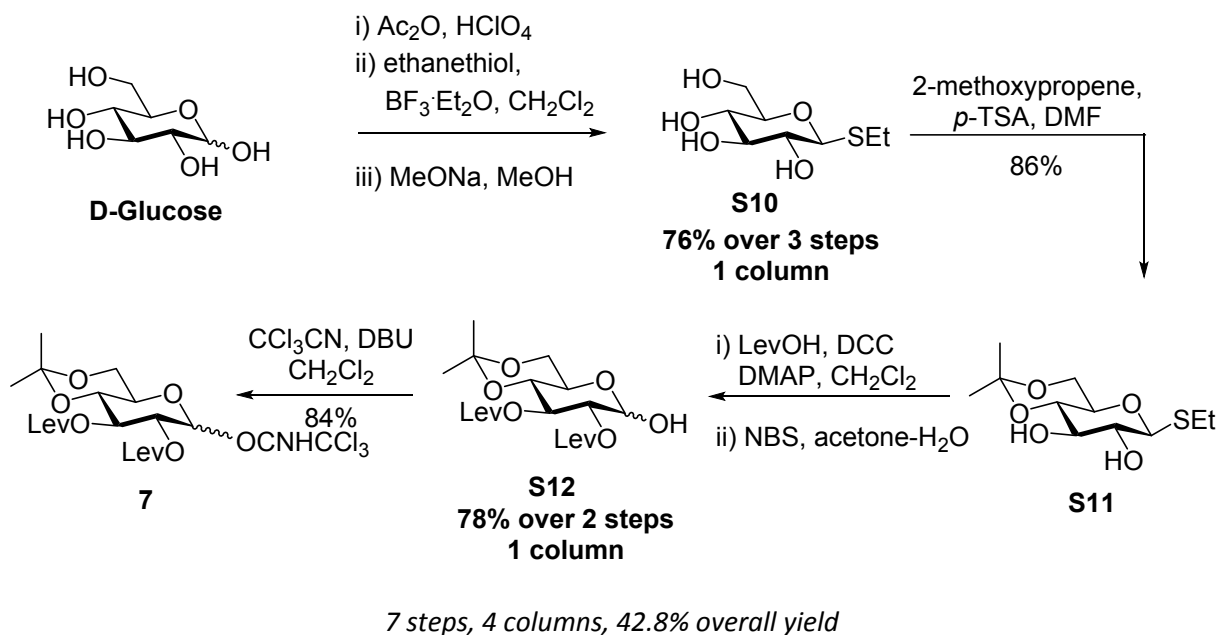


Entry	Acid		Solvent	Time	Products <sup>b</sup>		
	Name	Equivalents			4	5	6
1	CSA	0.2	MeOH/CH <sub>2</sub> Cl <sub>2</sub> v/v, 1/1	6 h	~20%	~70%	~10%
2	AcOH	3/1, v/v	H <sub>2</sub> O	1.5 h	none	70-80%	20-30%
3	AcOH	50	MeOH	84 h	~10%	60-70%	20-30%
4	AcOH	50	CH <sub>2</sub> Cl <sub>2</sub>	72 h	100%	none	none

<sup>a</sup> Reaction conditions: 50 mg of **4** in 1 mL solvent. The reactions were all carried out at room temperature.

<sup>b</sup> The ratios were determined by <sup>1</sup>H NMR analyses of the crude reaction mixture.

**Scheme S3. Synthesis of glucosyl donor 7.**



**Scheme S4. Synthesis of fucosyl acceptor 8.**

