#### A Mathematical Model of Tumor Regression and Recurrence After Therapeutic Oncogene Inactivation

#### SUPPLEMENTARY INFORMATION

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**Supplementary Fig. 1.** Submodels of the 3-compartment model. (a) The simplified 3-compartment model with net-growth and net-death rates. (b) A 2-compartment submodel without net-growth of the MYC-On compartment. (c) A 2-compartment submodel including net-growth of the MYC-On compartment.



**Supplementary Fig. 2.** Mice implanted with subcutaneous xenografts of  $2 \times 10^5$  4188-PGK-RLuc/Tet-FLuc cells based on a single clonal population (a) B11 or (b) E12, with doxycycline (Dox) administration (dotted black vertical line) at Day 20, 23 or 33. Top rows show the fitting of the simplified 3-compartment model (dashed black line) to RLuc signal (white circles), superimposed with the predicted population of MYC-On cells (blue line), MYC-Off cells (red line), and Escaped cells (green line). Shaded grey region (below  $1.60 \times 10^5$  photon/s) indicates imaging signal is below the limit of detection. Middle rows show the model-predicted MYC-expressing tumor cell population (orange line) superimposed

with Tet-FLuc signal (yellow squares). Bottom rows show the unweighted (white triangles) or weighted (black triangles) residual sum of squares indicating the deviation between experimental data and predicted value at each time point.



**Supplementary Fig. 3.** Western blots of **(a)** MYC and **(b)** Firefly luciferase (FLuc) expression in the tumors collected immediately before (day 0), or days 2 or 7 after doxycycline (Dox) was administered to inactivate MYC. Cultured 4188-PGK-RLuc/Tet-FLuc cells (Clone B11) were used as a positive control. MYC, FLuc and actin bands are indicated by arrows. Bands for days 0, 2, and 7 are shown in **Fig. 2g**.



**Supplementary Figure 4**. Mice implanted with subcutaneous xenografts of  $2 \times 10^5$  4188-PGK-RLuc-Tet-FLuc cells based on a single clonal population (a) B11 or (b) E12, with doxycycline (Dox) administration (dotted black vertical line) at Day 0. Top rows show the fitting of the

simplified 3-compartment model (dashed black line) to RLuc signal (white circles), superimposed with the predicted population of MYC-On cells (blue line), MYC-Off cells (red line), and Escaped cells (green line). Shaded grey region (below 1.60×10<sup>5</sup> photon/s) indicates imaging signal is below the limit of detection. Middle rows show the model-predicted MYC-expressing tumor cell population (orange line) superimposed with Tet-FLuc signal (yellow squares). Bottom rows show the unweighted (white triangles) or weighted (black triangles) residual sum of squares indicating the deviation between experimental data and predicted value at each time point.



**Supplementary Figure 5**. Mice implanted with subcutaneous xenografts of  $2 \times 10^5$  4188-PGK-RLuc-Tet-FLuc cells based on a single clonal population (a) B11 or (b) E12, with doxycycline (Dox) administration (dotted black vertical line) at indicated times (Day 7, 9, or 11). Top rows show the fitting of the simplified 3-compartment model (dashed black line) to RLuc signal (white circles), superimposed with the predicted population of MYC-On cells (blue line), MYC-Off cells (red line), and Escaped cells (green line). Shaded grey region (below  $1.60 \times 10^5$  photon/s) indicates imaging signal is below the limit of detection. Middle rows show the model-predicted MYC-expressing tumor cell population (orange line) superimposed with Tet-FLuc signal (yellow squares). Bottom rows show the unweighted (white triangles) or weighted (black triangles) residual sum of squares indicating the deviation between experimental data and predicted value at each time point.



**Supplementary Figure 6**. Natural killer (NK) cells delay tumor growth. NSG mice received either (a) a saline injection or (b)  $1 \times 10^6$  NK cells via tail vein three days before intravenous transplantation of  $3 \times 10^6$  4188-PGK-FLuc cells. Tumor growth was then monitored by firefly luciferase (FLuc) imaging.



**Supplementary Figure 7**. Natural killer (NK) cells delay tumor recurrence. NSG mice (n=16) were intravenously injected with  $3 \times 10^6$  4188-PGK-FLuc cells and monitored by firefly luciferase (FLuc) imaging. Doxycycline (Dox) was administered 7 days later to inactivate MYC oncogene. On the day of Dox administration (black dotted vertical line) and weekly thereafter (red dotted vertical lines), mice received either (a) a saline injection or (b) adoptive transfer of  $1 \times 10^6$  NK cells. Top rows show the fitting of the simplified 3-compartment model (dashed black line) to FLuc signal (white circles), superimposed with the predicted population of MYC-On cells (blue line), MYC-Off cells (red line), and Escaped cells (green line). Shaded grey region (below  $1.60 \times 10^5$  photon/s) indicates imaging signal is below the limit of detection. Bottom rows show the unweighted (white triangles) or weighted (black triangles) residual sum of squares indicating the deviation between experimental data and predicted value at each time point.

#### Supplementary Tables

The percent coefficient of variation (%CV), also called the relative standard deviation, is defined to be the standard deviation of the parameter estimate divided by the mean of the parameter estimate. The %CV provides a measure of the uncertainty of a parameter estimate, with a higher %CV corresponding to a greater uncertainty.

#### Supplementary Table 1. Clone B11

Dox administered on Day 0 post-implantation of 2×10<sup>5</sup> cells

Doromotor	Linito	NSG1	NSG2	NSG3	NSG4	NSG5
Farameter	Units	Value (%CV)	Value (%CV)	Value (%CV)	Value (%CV)	Value (%CV)
<b>N</b> On,0	photon/s	2.95×10 <sup>5</sup> (26)	3.84×10 <sup>5</sup> (23)	1.62×10 <sup>6</sup> (20)	4.98×10 <sup>5</sup> (22)	5.72×10 <sup>5</sup> (16)
kesc	day⁻¹	0 (fixed)	0 (fixed)	0 (fixed)	0 (fixed)	0 (fixed)
<b>k</b> netdeath,Off	day⁻¹	1.71×10 <sup>-1</sup> (108)	2.89×10 <sup>-1</sup> (22)	3.18×10 <sup>-1</sup> (15)	4.88×10 <sup>-1</sup> (33)	2.45×10 <sup>-1</sup> (26)
<b>k</b> netgr,On	day⁻¹	5.73 (111)	1.15×10 <sup>1</sup> (13)	0 (fixed)	1.17×10 <sup>1</sup> (17)	0 (fixed)
<i>k</i> netgr,Esc	day⁻¹	0 (fixed)	0 (fixed)	0 (fixed)	0 (fixed)	0 (fixed)

## Supplementary Table 2. Clone B11

Dox administered on Day 7 post-implantation of 2×10<sup>5</sup> cells

Deremeter	Lipita	NSG6	NSG7	NSG8	
Parameter	Units	Value (%CV)	Value (%CV)	Value (%CV)	
N <sub>On,0</sub>	photon/s	9.51×10 <sup>3</sup> (62)	1.05×10 <sup>5</sup> (79)	2.30×10 <sup>4</sup> (162)	
kesc	day <sup>-1</sup>	0 (fixed)	0 (fixed)	0 (fixed)	
<b>k</b> netdeath,Off	day <sup>-1</sup>	5.83×10 <sup>-1</sup> (8)	6.97×10 <sup>-1</sup> (6)	6.15×10 <sup>-1</sup> (13)	
<b>k</b> netgr,On	day <sup>-1</sup>	1.08 (9)	1.17 (11)	1.16 (23)	
<b>k</b> netgr,Esc	day <sup>-1</sup>	0 (fixed)	0 (fixed)	0 (fixed)	

### Supplementary Table 3. Clone B11

Dox administered on Day 9 post-implantation of 2×10<sup>5</sup> cells

Deremeter	Linito	NSG9	NSG10	
Farameter	Units	Value (%CV)	Value (%CV)	
N <sub>On,0</sub>	photon/s	4.44×10 <sup>5</sup> (39)	2.43×10 <sup>5</sup> (23)	
<i>k</i> esc	day <sup>-1</sup>	0 (fixed)	0 (fixed)	
<b>k</b> netdeath,Off	day <sup>-1</sup>	5.14×10 <sup>-1</sup> (8)	6.45×10 <sup>-1</sup> (6)	
<b>k</b> netgr,On	day <sup>-1</sup>	5.95×10 <sup>-1</sup> (10)	5.85×10 <sup>-1</sup> (6)	
<i>k</i> netgr,Esc	day <sup>-1</sup>	0 (fixed)	0 (fixed)	

# Supplementary Table 4. Clone B11

Dox administered on Day 11 post-implantation of 2×10<sup>5</sup> cells

Deremeter	Lipita	NSG11	NSG12	NSG13
Farameter	Units	Value (%CV)	Value (%CV)	Value (%CV)
N <sub>On,0</sub>	photon/s	6.66×10 <sup>3</sup> (121)	3.58×10 <sup>5</sup> (71)	1.98×10 <sup>5</sup> (38)
k <sub>esc</sub>	day <sup>_1</sup>	1.79×10 <sup>-7</sup> (208)	2.12×10 <sup>-6</sup> (112)	0 (fixed)
knetdeath,Off	day <sup>_1</sup>	4.66×10 <sup>-1</sup> (17)	6.73×10 <sup>-1</sup> (16)	5.33×10 <sup>-1</sup> (11)
<b>k</b> netgr,On	day <sup>_1</sup>	1.04 (13)	6.45×10 <sup>-1</sup> (15)	5.44×10 <sup>-1</sup> (9)
k <sub>netgr,Esc</sub>	day <sup>_1</sup>	4.83×10 <sup>-1</sup> (17)	3.05×10 <sup>-1</sup> (11)	0 (fixed)

## Supplementary Table 5. Clone B11

Dox administered on Day 20 post-implantation of 2×10<sup>5</sup> cells

Deremeter	Linito	NSG14	NSG15	
Farameter	Units	Value (%CV)	Value (%CV)	
N <sub>On,0</sub>	photon/s	5.11×10 <sup>5</sup> (130)	2.88×10 <sup>5</sup> (105)	
kesc	day <sup>-1</sup>	3.18×10 <sup>-7</sup> (721)	8.38×10 <sup>-7</sup> (534)	
<b>k</b> netdeath,Off	day <sup>-1</sup>	3.67×10 <sup>-1</sup> (43)	4.41×10 <sup>-1</sup> (31)	
<b>k</b> netgr,On	day <sup>-1</sup>	4.33×10 <sup>-1</sup> (20)	4.52×10 <sup>-1</sup> (16)	
<b>k</b> netgr,Esc	day <sup>-1</sup>	4.12×10 <sup>-1</sup> (53)	3.83×10 <sup>-1</sup> (45)	

## Supplementary Table 6. Clone B11

Dox administered on Day 23 post-implantation of 2×10<sup>5</sup> cells

Deremeter	Lipito	NSG16	NSG17	NSG18	
Farameter	Units	Value (%CV)	Value (%CV)	Value (%CV)	
N <sub>On,0</sub>	photon/s	3.20×10 <sup>5</sup> (52)	6.82×10 <sup>4</sup> (64)	4.57×10 <sup>4</sup> (123)	
k <sub>esc</sub>	day <sup>_1</sup>	3.95×10 <sup>-9</sup> (678)	5.52×10 <sup>-6</sup> (162)	2.54×10 <sup>-9</sup> (315)	
knetdeath,Off	day <sup>_1</sup>	4.19×10 <sup>-1</sup> (19)	5.29×10 <sup>-1</sup> (16)	4.99×10 <sup>-1</sup> (15)	
<b>k</b> netgr,On	day <sup>_1</sup>	4.12×10 <sup>-1</sup> (8)	4.96×10 <sup>-1</sup> (8)	4.94×10 <sup>-1</sup> (13)	
<b>k</b> netgr,Esc	day <sup>_1</sup>	5.92×10 <sup>-1</sup> (31)	3.28×10 <sup>-1</sup> (18)	5.06×10 <sup>-1</sup> (16)	

### Supplementary Table 7. Clone B11

Dox administered on Day 33 post-implantation of 2×10<sup>5</sup> cells

Doromotor	Linito	NSG19		
Farameter	Units	Value (%CV)		
N <sub>On,0</sub>	photon/s	3.11×10 <sup>4</sup> (83)		
k <sub>esc</sub>	day <sup>_1</sup>	1.41×10 <sup>-13</sup> (432)		
$k_{ m netdeath,Off}$	day <sup>_1</sup>	5.64×10 <sup>-1</sup> (12)		
<i>k</i> netgr,On	day <sup>_1</sup>	3.09×10 <sup>-1</sup> (11)		
k <sub>netgr,Esc</sub>	day <sup>_1</sup>	6.03×10 <sup>-1</sup> (13)		

## Supplementary Table 8. Clone E12

Dox administered on Day 0 post-implantation of 2×10<sup>5</sup> cells

Deremeter	Linito	NSG20	NSG21	NSG22	NSG23	NSG24
Parameter	Units	Value (%CV)	Value (%CV)	Value (%CV)	Value (%CV)	Value (%CV)
N <sub>On,0</sub>	photon/s	4.98×10 <sup>5</sup> (16)	2.41×10 <sup>5</sup> (12)	1.95×10 <sup>5</sup> (9)	1.39×10 <sup>6</sup> (21)	2.07×10 <sup>5</sup> (24)
<i>k</i> esc	day⁻¹	0 (fixed)	0 (fixed)	0 (fixed)	0 (fixed)	0 (fixed)
<b>k</b> netdeath,Off	day⁻¹	2.66×10 <sup>-1</sup> (9)	4.91×10 <sup>-2</sup> (59)	1.83×10 <sup>-1</sup> (14)	4.01×10 <sup>-1</sup> (14)	1.62×10 <sup>-2</sup> (42)
<b>k</b> netgr,On	day⁻¹	1.38×10 <sup>1</sup> (3)	0 (fixed)	1.09×10 <sup>1</sup> (7)	1.07×10 <sup>1</sup> (15)	9.72 (25)
<b>k</b> netgr,Esc	day⁻¹	0 (fixed)	0 (fixed)	0 (fixed)	0 (fixed)	0 (fixed)

**Supplementary Table 9.** Clone E12 Dox administered on Day 7 post-implantation of 2×10<sup>5</sup> cells

Deremeter Unite	NSG25	NSG26	NSG27	NSG28	NSG29	NSG30	NSG31	
Farameter	Units	Value (%CV)	Value (%CV)	Value (%CV)	Value (%CV)	Value (%CV)	Value (%CV)	Value (%CV)
N <sub>On,0</sub>	photon/s	1.49×10 <sup>5</sup> (144)	5.07×10 <sup>5</sup> (44)	4.32×10 <sup>5</sup> (44)	3.47×10 <sup>5</sup> (44)	2.69×10 <sup>5</sup> (50)	5.32×10 <sup>3</sup> (121)	2.46×10 <sup>4</sup> (123)
<i>k</i> esc	day <sup>-1</sup>	0 (fixed)	0 (fixed)	0 (fixed)	0 (fixed)	0 (fixed)	0 (fixed)	0 (fixed)
$k_{ m netdeath,Off}$	day <sup>-1</sup>	3.40×10 <sup>-1</sup> (15)	2.91×10 <sup>-1</sup> (10)	3.86×10 <sup>-1</sup> (10)	4.72×10 <sup>-1</sup> (8)	3.30×10 <sup>-1</sup> (14)	3.20×10 <sup>-1</sup> (13)	4.29×10 <sup>-1</sup> (10)
<b>k</b> netgr,On	day <sup>-1</sup>	8.21×10 <sup>-1</sup> (28)	5.82×10 <sup>-1</sup> (13)	8.01×10 <sup>-1</sup> (10)	1.07 (7)	5.93×10 <sup>1</sup> (15)	1.28 (15)	1.22 (16)
knetgr,Esc	day <sup>-1</sup>	0 (fixed)	0 (fixed)	0 (fixed)	0 (fixed)	0 (fixed)	0 (fixed)	0 (fixed)

**Supplementary Table 10.** Clone E12 Dox administered on Day 20 post-implantation of 2×10<sup>5</sup> cells

Deremeter Unite	NSG32	NSG33	NSG34	NSG35	NSG36	NSG37	NSG38	
Farameter	Units	Value (%CV)	Value (%CV)	Value (%CV)	Value (%CV)	Value (%CV)	Value (%CV)	Value (%CV)
N <sub>On,0</sub>	photon/s	3.74×10 <sup>5</sup> (67)	9.23×10 <sup>5</sup> (66)	3.13×10 <sup>5</sup> (64)	3.47×10 <sup>5</sup> (67)	1.47×10 <sup>5</sup> (139)	1.03×10 <sup>5</sup> (103)	6.46×10 <sup>5</sup> (58)
<i>k</i> esc	day <sup>-1</sup>	5.11×10 <sup>-8</sup> (364)	9.13×10 <sup>-11</sup> (660)	2.18×10 <sup>-7</sup> (481)	1.31×10 <sup>-10</sup> (369)	2.63×10 <sup>-8</sup> (270)	8.29×10 <sup>-11</sup> (318)	3.41×10 <sup>-11</sup> (441)
<b>k</b> netdeath,Off	day <sup>-1</sup>	3.57×10 <sup>-1</sup> (17)	3.73×10 <sup>-1</sup> (16)	2.99×10 <sup>-1</sup> (23)	4.55×10 <sup>-1</sup> (11)	4.10×10 <sup>-1</sup> (16)	4.86×10 <sup>-1</sup> (13)	4.07×10 <sup>-1</sup> (12)
<b>k</b> netgr,On	day <sup>-1</sup>	4.52×10 <sup>-1</sup> (10)	3.46×10 <sup>-1</sup> (12)	4.54×10 <sup>-1</sup> (9)	4.43×10 <sup>-1</sup> (10)	5.03×10 <sup>-1</sup> (16)	5.21×10 <sup>-1</sup> (13)	4.30×10 <sup>-1</sup> (9)
<b>k</b> netgr,Esc	day <sup>-1</sup>	4.52×10 <sup>-1</sup> (23)	6.11×10 <sup>-1</sup> (28)	4.79×10 <sup>-1</sup> (31)	5.33×10 <sup>-1</sup> (17)	4.18×10 <sup>-1</sup> (18)	3.35×10 <sup>-1</sup> (17)	5.18×10 <sup>-1</sup> (20)