Supplementary Materials for

Chromosome-level genome assembly of *Ophiorrhiza pumila* reveals the evolution of camptothecin biosynthesis

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Chromosome	Short arm	Long arm	Total length	Relative length of chromosome (%)	Total length (Mb)	Relative length of sequence (%)
1	08.4 ± 1.2	13.8 ± 4.8	22.2 ± 5.1	8.4	37.1	8.4
2	10.6 ± 1.5	13.7 ± 3.8	24.3 ± 5.2	9.1	42.3	9.6
3	09.6 ± 4.2	13.9 ± 2.1	23.5 ± 6.3	8.8	30.5	6.9
4	05.7 ± 1.5	14.9 ± 0.9	20.5 ± 2.1	7.7	32.8	7.5
5	07.7 ± 1.3	12.5 ± 1.6	20.2 ± 2.3	7.6	32.3	7.3
6	15.2 ± 1.4	17.3 ± 1.9	32.5 ± 3.0	12.2	60	13.6
7	10.5 ± 0.5	13.3 ± 1.5	23.8 ± 1.5	9	40.7	9.3
8	12.2 ± 2.7	14.2 ± 2.5	26.4 ± 4.9	9.9	39	8.9
9	12.3 ± 0.8	13.0 ± 1.7	25.3 ± 2.5	9.5	51.8	11.8
10	08.5 ± 2.9	16.1 ± 4.4	24.6 ± 6.9	9.2	32.7	7.4
11	09.1 ± 2.7	13.6 ± 2.4	22.7 ± 4.9	8.5	40.6	9.2

Supplementary Table 1. Karyotype measurements for Ophiorrhiza pumila chromosomes.

Three pachytenes were used for measurements.

The relative length of chromosome (%) is in the percentage of the karyotype length.

Total length (Mb) is the length of assembled sequences for each chromosome.

The relative length of the sequence (%) is calculated using the assembled genome size.



Supplementary Fig. 1. Strictosidine derived monoterpene indole alkaloid (MIA) biosynthetic

pathways. Color codes represent specific enzymes assigned to *Ophiorrhiza pumila* genome. Metabolite names with * were chemically assigned and identified in metabolite profiling datasets of O. pumila in this study. The vellow shaded branch pathway represents camptothecin biosynthesis pathway. Abbreviations- STR: strictosidine synthase; SGD: strictosidine-B-D-glucosidase; THAS: tetrahydroalstonine synthase; HS: heteroyohimbine synthase; AS: alstonine synthase; GS: geissoschizine synthase; SBE: sarpagan bridge enzyme; GO: geissoschizine oxidase; PNAE: polyneuridine-aldehyde esterase; VS: vinorine synthase; CYP5437: vinorine hydroxylase; VR: vomilenine reductase; SAT: stemmadenine O-acetyltransferase; RG: raucaffricine-β-Dglucosidase; AAE: acetylajmaline esterase; ASO/PAS: O-acetylstemmadenine oxidase; DPAS: dihydroprecondylocarpine synthase; HL3: hydrolase 3; HL4: hydrolase 4; CS: catharanthine tabersonine synthase: vincadifformine-19-hydroxylase; synthase: *TS*: *V19H*: MAT: minovincinine-O-acetyltransferase; TEX: tabersonine-6,7-epoxidase; T16H: tabersonine-16hydroxylase; T19H: tabersonine-19-hydroxylase; T16OMT: tabersonine-16-O-methyltransferase; T3O: tabersonine-3-oxygenase; T3R: tabersonine-3-reductase; TAT: 19-hydroxytabersonine,19-Oacetyltransferase; DhtNMT: 3-hydroxy-16-methoxy-2,3-dihydrotabersonine-N-methyltransferase; D4H: deacetoxyvindoline-4-hydroxylase; DAT: deacetylvindoline-O-acetyltransferase; T19AT: tabersonine-19-hydroxy-O-acetyltransferase; Prx1:peroxidase 1



Supplementary Fig. 2. Assembly statistics for chromosome-scale plant genome assemblies deposited at the NCBI genome database. Chromosome-scale plant genome assemblies being published and deposited in NCBI genome database are included here, while *Ophiorrhiza pumila* genome was assembled in this study. All publicly available genome stats of monoterpene indole alkaloids producing plants were highlighted through red circle.



Supplementary Fig. 3. Contig-level genome assembly validation using Bionano de novo genome assembly for *Ophiorrhiza* genome. The Canu PacBio contigs are shown as green bar, Bionano de novo contigs are shown as light blue (turquoise) bar. The regions, on both of the NGS and Bionano assemblies, with consensus are shown as dark blue shades on the bar connected through grey lines, the regions with no support either for NGS assembly or Bionano assembly are shown through yellow lines, and the regions which has conflicts based on NGS assembly and Bionano alignments are shown through red box.



Supplementary Fig. 4. *Ophiorrhiza pumila* **karyotype and genome size estimation. (a)** Meiotic metaphase I chromosomes of *O. pumila*. The chromosomes form 11 bivalents. The scale bar corresponds to 10 μ m. (b) Mitotic chromosomes of *O. pumila*, representing a diploid genome with 2n = 22 chromosomes. The scale bar corresponds to 5 μ m. The karyotyping was performed twice, and for each experiment, at least 10 metaphase plates were analyzed. (c) Genome size estimation of *O. pumila* genome using KmerGenie software. Genome size at kmer-195 was estimated as 440 Mb. (d) Flow cytometer-based genome estimation, with genome size predicted as 420 Mb. Here count refers to the number of nuclei.



%similarity

Supplementary Fig. 5. Alignment of phased and reference genome assemblies of Ophiorrhiza pumila. Phased genome assemblies of O. pumila, haplotig1 (a) and haplotig2 (b), were aligned to the reference genome using PROmer from MUMmer package, and the alignments are represented by dot-plots.



Supplementary Fig. 6. Fluorescence *in situ* hybridization (FISH) analysis for chromosome 2 of *Ophiorrhiza pumila*. The four probes (red text) correspond to the four different sites on chromosome 2 of *O. pumila*. While probes on the same chromosome arms were consistent with the expected assigned scaffolds, their positions at the assembly gap provided evidence of misalignment. The scaffolds at the assembly gap of chromosome 2 were re-oriented in the final reference genome of *O. pumila*. The experiment was performed twice, and for each experiment, we validated chromosomes for at least 10 slides.



Supplementary Fig. 7. Identification of *Ophiorrhiza pumila* chromosomes and its karyotype. (a) DAPI stained *O. pumila* pachytene chromosomes and Fluorescence *in situ* hybridization (FISH) analysis-based identifications of the chromosomes. The scale bar corresponds to $5\mu m$. (b) Ideogram of the *O. pumila* chromosome, are drawn based on Supplementary Table 1 results. Red bars indicate the position of FISH signals used in the identification of respective chromosomes, as shown in (a). The experiment was repeated twice, and for each experiment, at least 10 chromosome slides were analyzed.



Supplementary Fig. 8. The *Ophiorrhiza pumila* reference genome assembly validation using Bionano de novo genome assembly as orthogonal evidence. All 11 chromosomes were supported by Bionano de novo assembly (blue color bars), with green and red color shades between these two assembly alignments representing regions of insertions or deletions, respectively.



Supplementary Fig. 9. Hi-C contact map representing chromosome-scale phased genomes of *Ophiorrhiza pumila*. Hi-C contact matrix visualization for the *O. pumila* phased genomes, haplotig1 (a) and haplotig2 (b). The pixel intensity represents the count of Hi-C links at 150 Kb size-windows on the chromosomes on a logarithmic scale. Darker red color indicates higher contact probability, while white space represents no or fewer contacts. The blue lines separate chromosomes for each phased genome.



Supplementary Fig. 10. Synteny dot-plot for *Ophiorrhiza pumila* genome within, and with selected plant species including ancestral eudicot karyotype (AEK). (a) Dot-plot based synteny visualization showed a near-perfect 1:3 synteny relationship between AEK and *O. pumila* genome. Synteny dot-plot between (b) *O. pumila - O. pumila*, (c) *O. pumila - Vitis vinifera*, (d) *Coffea canephora - V. vinifera*, (e) *O. pumila - C. canephora*, and (f) *O. pumila - Solanum lycopersicum* showed conserved gene blocks within these species. Synteny dot-plot between *O. pumila* and *C. canephora* showed a 1:1 synteny depth with an interesting interaction between chromosome 9 of the *Ophiorrhiza* genome and chromosome 2 of coffee genome, as shown in the red box.









Supplementary Fig. 12. Macrosynteny analysis for *Ophiorrhiza pumila* and *Coffea canephora* genome assemblies. Karyotypes with synteny associations, gene density distribution, and assembly gaps for *Ophiorrhiza* and coffee genome are represented. The color of synteny relationships are based on associations with *Ophiorrhiza* chromosomes.



Supplementary Fig. 13. Fluorescence *in situ* hybridization (FISH) analysis to test potential mis-assembly in the chromosome 2 of *Coffea arabica*. (a) Sequence positions for two FISH probes in *Coffea canephora* chromosome 2 representing two regions that showed syntenic relationships with chromosome 2 and 9 of *Ophiorrhiza pumila* as shown in Supplementary Fig. 12. The chromosome 2 map of *C. canephora* with assigned ancestral blocks was obtained from Coffee Genome hub (http://coffee-genome.org/coffeacanephora). The probe names are corresponding to the gene ID in the *C. canephora* genome. (b) FISH mapping using two FISH probes on *C. arabica* chromosomes (2n = 4x = 44). The Cc02_g01170 probe shows four FISH signals, and the Cc02_g28160 probe shows two signals on 44 chromosome. The experiment was repeated twice, and for each experiment, we verified at least six slides. The scale bar corresponds to 5 µm.



Supplementary Fig. 14. Characteristics of *Ophiorrhiza pumila* **predicted gene models. (a)** Gene models predicted across 11 chromosomes of *O. pumila*. (b) Exon length distribution plot. (c) Gene length distribution plot. (d) Protein length distribution plot. (e) % GC distribution plot for predicted gene models. (f) Distribution of annotation edit distance (AED).



Supplementary Fig. 15. Functional classification of *Ophiorrhiza pumila* gene models. (a) Top-hit species distribution plot. (b) Species distribution plot based on database-based annotation. (c) Sequence similarity distribution plot. (d) Gene length distribution plot with number of assigned gene ontologies (GO). (e) GO mapping distribution plot. (f) GO distribution as biological process (BP), Molecular function (MF), and Cellular component (CC) categories. Annotation based functional classification was performed using Omics box software. Source data are provided as a Source Data File.



Supplementary Fig. 16. Chromosome primary constriction and centromere localization in *Ophiorrhiza pumila* chromosomes using Fluorescence *in situ* hybridization (FISH) analysis. (a) Detection of OpCEN (red) at the heterochromatic chromocenters in the nuclei (white). (b) Centromeric detection of OpCEN (red) in somatic metaphase chromosomes (blue). (c) Detection of OpCEN (red) at centromeric regions of prometaphase chromosomes (blue). (d)-(f) Cut-off image for chromosome selected in the white color box in (c). (d) DAPI-stained chromosome. (e) Detection of OpCEN signal. (f) Merged image. Arrowheads indicated primary constriction. Repeat analysis using *O. pumila* reference genome identified pericentromeric repeat sequence, conserved across all chromosomes, and positioned sat putative centromere region, named OpCEN. Probes designed for OpCEN labeled putative centromere of *O. pumila*. The scale bar corresponds to 5μ m. The FISH experiment was performed twice, and for each experiment, at least 10 slides were analyzed.



Supplementary Figure. 17. Distribution of repeat elements across eight plant species. Percentage of genome content consisting of (a) DNA element repeats. (b) LINEs repeat; and (c) Other repeats. At: Arabidopsis thaliana, Ca: Camptotheca acuminata, Cc: Coffea canephora, Cr: Catharanthus roseus, Nb: Nicotiana benthamiana, Op: Ophiorrhiza pumila, Ps: Papaver somniferum, SI: Solanum lycopersicum. Source data are provided as a Source Data file.





Supplementary Fig. 18. Estimated divergence and whole genome duplication for monoterpene indole alkaloids producing species. The lineage divergence time as branch length is indicated at each of the branch points in Million years ago (Mya). Divergence time for *Ophiorrhiza pumila* with coffee and *Camptotheca acuminata* was estimated at 47 Mya and 120 Mya, respectively. The whole-genome duplication for *C. acuminata* was identified and the duplication time was estimated. "#" and "\$" depicts species from Rubiaceae and Apocynaceae families, respectively.



Supplementary Fig. 19. Density plot depicting Ks (synonymous substitution rate) distribution of paralogous gene pairs for selected eudicot plant genome assemblies.



Supplementary Fig. 20. Density plot depicting Ks (synonymous substitute rate) distribution for orthologous gene pairs identified between *Ophiorrhiza pumila* and 12 other eudicot plant genome assemblies.



Supplementary Fig. 21. Metabolite profiling for complete nitrogen labeled (¹⁵N) or non-labeled hairy roots to identify nitrogen-containing metabolites in *Ophiorrhiza pumila*. (a) Principle component analysis for labeled and non-labeled hairy root metabolite profiling datasets. Five biological replicates for each condition were used for metabolite profiling. (b) S-plot based multivariate analysis approach to select characteristic metabolite features of labeled and non-labeled hairy root samples. Metabolite features colored red and green represents stable isotope labeled and non-labeled metabolites, respectively.



Supplementary Fig. 22. Metabolite profiling for six tissues of *Ophiorrhiza pumila. O. pumila* metabolome database was established by assigning chemical identity to metabolite features using previously reported complete carbon labeling approach and complete nitrogen labeling approach used in this study. (a) Principle component analysis. (b) Loadings plot for metabolite profiling datasets of six tissues. Five biological replicates for each tissue were used for metabolite profiling.



Supplementary Fig. 23. Accumulation of specialized metabolites in six tissues of *Ophiorrhiza pumila*. * and ** represent chemically assigned metabolites based on MS/MS analysis using the public database and pure standards, respectively; m/z-mass by charge ratio. The color intensity refers to the scaled median intensity of metabolites across tissues of *O. pumila* using five biological replicates for each tissue.



Supplementary Fig. 24. Coexpression based hierarchical clustering for genes assigned to enzymes from the secoiridoid biosynthesis pathway in *Ophiorrhiza pumila*. Spearman's correlation coefficients were calculated using transcript expression data for seven tissues of *O. pumila*. Hierarchical clustering identified complete secoiridoid biosynthesis pathway as a gene cluster at the second order of the dendrogram (selected within a yellow-colored box). Source data are provided as a Source Data file.



Supplementary Fig. 25. Coexpression based hierarchical clustering for genes assigned to enzymes from monoterpene indole alkaloid (MIA) biosynthesis in Ophiorrhiza pumila. Spearman's correlation coefficients were calculated using transcripts expression data for seven tissues of *O. pumila*. Genes associated with MIA biosynthesis and membered within a coexpressed gene-cluster (selected within a yellow-colored box) including genes assigned to secoiridoid biosynthesis pathway were selected. Genes colored as green are genes assigned to secoiridoid biosynthesis pathways. Source data are provided as a source Data file.



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							Opuchr04_g0011350-1.1 (OG0000538)	100101
							Opuchr09 a0016460-1.1 (OG0000087)	AAE
							Opuchr05_g0005010-1.1 (OG0003863)	
							Opuchr05_g0005050-1.1 (OG0003863)	
							Opuchr09_g0088900-1.1 (OG0000009)	ASO/PAS
							Opuchr09_g0089210-1.1 (OG0000009)	
							Opuchr09_g0091220-1.1 (OG0000009)	
							Opuchr03_g0007140-1.1 (OG0000019)	
	_						Opuchr03_g0007400-1.1	D4H
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							Opuchr06_g0004570=1.1 (OG0000709) • C1592	HL3
							Opuchr06 a0093830-1.1 (OG0010376) • C1624	
							Opuchr06_g0103980-1.1 (OG0010376)	
							Opuchr06_g0104080-1.1 (OG0010376)	PNAE
							Opuchr07_g0007230-1.1 (OG0000066) • C1684	
							Opuchr02_g0057870-1.1 (OG0001044) • C1401	
							Opuchr03_g0014520-1.1 (OG0000154)	
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							Opuchr01_g0010310=1.1 (OG0007462)	
							Opuchr06_g0001350-1.1 (OG0001600)	
							Opuchr06 g0110230-1.1 (OG0000780)	Prx1
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							Opuchr09_g0012180-1.1 (OG0000966)	
							Opuchr09_g0099200-1.1 (OG0000625)	
							Opuchr11_g0083390-1.1 (OG0000222)	Redox 2
							Opuchr05_g0065570-1.1 (OG0000007)	
							Opuchr06_g0097460-1.1 (OG0000118) C0000	
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							Opuchr08 a0011270-1.1 • C1749	SBE
							Opuchr01_g0011710-1.1 (OG0000057)	
							Opuchr02_g0011010-1.1 (OG0000057) • C1381	000
							Opuchr06_g0093740-1.1 (OG0000057) • C1624	SGD
							Opuchr09_g0086020-1.1 (OG0016082)	
		_					Opuchr04_g0013560-1.1 (OG0000291) • C1501	Τ19Δ Τ
			_				Opuchr05_g0000/90-1.1 (OG0000291) C1527	
							Opuchr04_g0014990=1.1 (OG0000079) C1504	T19H/CYP71BJ1
							Opuchr01_g0014830=1.1 ($OG0000007$)	
							Opuchr01_g0014980-1.1 (OG0000007)	
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							Opuchr08_g0009600-1.1 (OG0000007) • C1746	130
							Opuchr08_g0009750-1.1 (OG0000007)	
							Opuchr08_g0009760−1.1 ⊥	
							Opuchr08_g0013260-1.1 (OG0000007)	
							Opuchr11_g0012010-1.1 (OG0000007) • C1914	· •
							Opuchr05_g0004280-1.1 (OG0000007)	TEY1
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b

Strictosidine

Supplementary Fig. 26. Expression of genes assigned to secoiridoid and monoterpene indole alkaloid (MIA) biosynthesis pathway across seven tissues of Ophiorrhiza pumila. Expression of genes assigned to secoiridoid biosynthesis pathway (a) and MIA biosynthesis (b) across seven tissues of O. pumila. The color code used for individual genes corresponds to their position in MIA biosynthesis pathway in Supplementary Fig. 1. Genes were assigned to respective biosynthesis pathways based on co-expression and sequence homology against previously characterized genes. Cell suspension culture (CSC) and hairy root tissues are known to accumulate no and high MIA, respectively. Gene expression analysis showed tissue specific expression of genes, which were correlated with accumulation of MIA in Ophiorrhiza tissues. Associated orthogene families (OGs) and gene-cluster IDs are provided together with the gene IDs. Abbreviations- GPPS: geranyl diphosphate synthase; GES: geraniol synthase; G10H: geraniol 10hydroxylase; 10-HGO: 10- hydroxygeraniol oxidoreductase; IS: iridoid synthase; IO: iridoid oxidase; 7-DLGT: 7-deoxyloganetin glucosyltransferase, 7-DLH: 7-deoxyloganic acid 7-hydroxylase, LAMT: loganic acid O-methyltransferase; SLS: secologanin synthase; TDC: tryptophan decarboxylase; STR: strictosidine synthase; SGD: strictosidine beta-D-glucosidase; *THAS*: tetrahydroalstonine synthase: PNAE: polyneuridine- aldehyde esterase; ASO/PAS: O-acetylstemmadenine oxidase; AAE: acetylajmaline TEX1: tabersonine 6,7-epoxidase 1; HL3: hydrolase 3; 10OMT: 10- hydroxycamptothecin Oesterase; tabersonine-19-hydroxy-O-acetyltransferase; methyltransferase; *T19H*: *RH11H*: rankinidine/ humantenine-11-hydroxylase 3; D4H: deacetoxyvindoline 4-hydroxylase; Prx1: peroxidase 1; PR: perakine reductase. Source data are provided as Source Data file.

Supplementary Fig. 27. Gene-to-metabolite relationship network associated with MIA biosynthesis. The Pearson's correlation matrix was calculated using the psych package in R and drawn using Cytoscape (v3.6.0). The circle and square nodes represent genes assigned to secoiridoid and MIA biosynthesis pathways (shown in the Supplementary Fig. 26), and the metabolites, respectively. Edges represent gene-metabolite pairs with paired Pearson Correlation coefficient scores over 0.7 and corrected *p*-value < 0.05. Source data are provided as a Source Data file.

Supplementary Fig. 28. Orthogene families across monoterpene indole alkaloid (MIA) producing plant species. OrthoFinder based gene family classification was performed for 13 plant species, and orthogenes assigned to MIA producing plants were used for comparison. (a) Orthogenes shared among four MIA producing plant species. (b) Orthogenes specific to MIA producing plants. (c) Gene ontology enrichment analysis for *Ophiorrhiza pumila* genes assigned to MIA specific orthogene families as test set and *O. pumila* genome as reference set. Statistical significance was estimated using Fisher's Exact test (two-sided) with the corrected *p*-value cutoff set as 0.05. GO enrichment graph for statistically significant GO terms were drawn using Omics box software.

a Orthogene family OG0000148

b Orthogene family OG0015245

Supplementary Fig. 29. Gain, loss, expansion, or contraction for gene families annotated as strictosidine synthase (STR) across 33 plant species. (a) Orthogene family OG0000148 representing genes annotated as STR and consisting of member genes across multiple plant species. (b) Orthogene family OG0015245 consisting of functionally characterized STRs and specifically gained in the strictosidine derived monoterpene indole alkaloids producing plant species. Evolutionary analysis of gene content was performed using COUNT software. Phylogenetic tree and orthogene classifications were performed as described in the material and method section. Empty rectangles at the nodes denote the absence of gene from an ancestral node, shaded node denotes presence. The green bar represents gene gain or expansion, while the red bar represents gene loss or contraction at the node.

Supplementary Fig. 30. Phylogenetic analysis of loganic acid O-methyltransferase (LAMT) assigned to the orthogene families, OG0000252 and OG0014621. Gene family classification was performed based on 13 plant genomes, and the genes are colored based on the associated plant species. Functionally characterized genes are represented by "#" with the GenBank accession IDs. Strongly supported nodes with bootstrap value above 60% are shown.

Supplementary Fig. 31. Phylogenetic analysis of secologanin synthase (SLS) assigned to the orthogene families, OG0002438 and OG0013616. Gene family classification was performed based on 13 plant genomes, and the genes are colored based on the associated plant species. Functionally characterized genes are represented by "#" with the GenBank accession IDs. Strongly supported nodes with bootstrap value above 60% are shown.

Supplementary Fig. 32. Phylogenetic analysis of tryptophan decarboxylase (TDC) assigned to the orthogene family OG0000823. Gene family classification was performed based on 13 plant genomes, and the genes are colored based on the associated plant species. Functionally characterized genes are represented by "#" with the GenBank accession IDs. Strongly supported nodes with bootstrap value above 60% are shown.

Supplementary Fig. 33. Phylogenetic analysis of strictosidine synthase (STR) assigned to the orthogene families, OG0000148 and OG0015245. Gene family classification was performed based on 13 plant genomes, and the genes are colored based on the associated plant species. Functionally characterized genes are represented by "#" with the GenBank accession IDs. Strongly supported nodes with bootstrap value above 60% are shown.

Supplementary Fig. 34. Phylogenetic analysis of strictosidine beta-D-glucosidase (SGD) assigned to the orthogene families, OG0000057 and OG0016082. Gene family classification was performed based on 13 plant genomes, and the genes are colored based on the associated plant species. Functionally characterized genes are represented by "#" with the GenBank accession IDs. Strongly supported nodes with bootstrap value above 60% are shown.

Supplementary Fig. 35. Phylogenetic analysis of tetrahydroalstonine synthase (THAS) assigned to the orthogene family, OG0013119. Gene family classification was performed based on 13 plant genomes, and the genes are colored based on the associated plant species. Functionally characterized genes are represented by "#" with the GenBank accession IDs. Strongly supported nodes with bootstrap value above 60% are shown.

Supplementary Fig. 36. Phylogenetic analysis of sarpagan bridge enzyme (SBE) assigned to the orthogene family, OG0011713. Gene family classification was performed based on 13 plant genomes, and the genes are colored based on the associated plant species. Functionally characterized genes are represented by "#" with the GenBank accession IDs. Strongly supported nodes with bootstrap value above 60% are shown.

Tabersonine-19-hydroxy-Oacetyltransferase (T19AT)

Supplementary Fig. 37. Phylogenetic analysis of tabersonine-19-hydroxy-O-acetyltransferase (T19AT) assigned to the orthogene family, OG0000291. Gene family classification was performed based on 13 plant genomes, and the genes are colored based on the associated plant species. Functionally characterized genes are represented by "#" with the GenBank accession IDs. Strongly supported nodes with bootstrap value above 60% are shown.

Supplementary Fig. 38. Phylogenetic analysis of polyneuridine-aldehyde esterase (PNAE) assigned to the orthogene families, OG0000066 and OG0010376. Gene family classification was performed based on 13 plant genomes, and the genes are colored based on the associated plant species. Functionally characterized genes are represented by "#" with the GenBank accession IDs. Strongly supported nodes with bootstrap value above 60% are shown.

Supplementary Fig. 39. Phylogenetic analysis of O-acetylstemmadenine oxidase (ASO/PAS) assigned to the orthogene families, OG000009 and OG0003863. Gene family classification was performed based on 13 plant genomes, and the genes are colored based on the associated plant species. Functionally characterized genes are represented by "#" with the GenBank accession IDs. Strongly supported nodes with bootstrap value above 60% are shown.

Supplementary Fig. 40. Phylogenetic analysis of polyneuridine-aldehyde esterase (PR) assigned to the orthogene families, OG0000154, and OG0001044. Gene family classification was performed based on 13 plant genomes, and the genes are colored based on the associated plant species. Functionally characterized genes are represented by "#" with the GenBank accession IDs. Strongly supported nodes with bootstrap value above 60% are shown.

Supplementary Fig. 41. Evolution of secoiridoid and MIA biosynthesis associated orthogene families in camptothecin producing plants and coffee genome. Median for synonymous substitutions per synonymous sites (*Ks*) were calculated using genes assigned to a given orthogene family for a plant species as described in method section. Small *Ks*-median values for several key enzymes associated with MIA biosynthesis in *O. pumila* and *C. acuminata*, while high *Ks*-median values for genes from coffee genome suggests faster and active evolution of the functional genes in the camptothecin producing plants post established STR enzymes. Results suggested evolution of STR as key event that preceded with evolution of genes associated with MIA biosynthesis, a factor that was missing for coffee genome. Source data are provided as a Source Data file.

Supplementary Fig. 42. Monoterpene indole alkaloid (MIA) gene-clusters identified in chromosome 1 of the *Ophiorrhiza* genome assembly. The position of the gene-cluster is scaled based on chromosome size and its physical position on the chromosome. The gene description, colored as orange, represents genes with functions associated with MIA biosynthesis. O.p.: *Ophiorrhiza pumila;* C.a: *Camptotheca acuminata;* C.r: *Catharanthus roseus;* G.s: *Gelsemium sempervirens.*

32811491 bps	Gene id Orthogroup Opuchr04_g0014720-1.1 Opuchr04_g0014730-1.1 Opuchr04_g0014760-1.1 OG0005946 Opuchr04_g0014700-1.1 OG0013021 Opuchr04_g0014700-1.1 OG0013021 Opuchr04_g0014800-1.1 OG0005823 Opuchr04_g0014800-1.1 OG0005823 Opuchr04_g0014800-1.1 OG0005223 Opuchr04_g0014800-1.1 OG0005822 Opuchr04_g0014800-1.1 OG0005823 Opuchr04_g0014900-1.1 OG00086840 Opuchr04_g0014900-1.1 OG0000079 Opuchr04_g0014900-1.1 OG0000079 Opuchr04_g0015000-1.1 OG0000079 Opuchr04_g0015000-1.1 OG0000079 Opuchr04_g0015000-1.1 OG0000079 Opuchr04_g0015200-1.1 <	$\begin{array}{c} \bullet \bullet$	 6 Construction Indole-3-glycerol phosphate synthase, conserved site uncharacterized protein Glycaal oxidase, N-terminal endoglucanase 11-like uncharacterized protein LOC 113761003 isoform X2 UDP-3-O-I3-hydroxymyrisov/I glucosamine N-acyltransferase Endonuclease/exonuclease/phosphatase superfamily uncharacterized protein E3 ubiguitin-protein ligase RGL G2-like NADH dehydrogenase Nucleoside phosphorylase/phosphoribosyltransferase catalytic domain superfamily Aminoacyl-IRNA synthetase, class I, conserved site tabersonine/lochnericine 19-hydroxylase cytochrome P450 71A3-like Co-acetylstemmademine-11-hydroxylase 3 Tabersonine/lochnericine 19-hydroxylase uncharacterized protein LOC 113737449 hypothetical protein LOC 113737449 hypothetical protein LOC 113737449 hypothetical protein Ging factor 1a Tabersonine/lochnericine 19-hydroxylase multiprotein-bridging factor 1a Tabersonine/lochnericine 19-hydroxylase Protein LYKS-like quinolinate synthase chloroplastic high-affinity nitrate transporter 3.1-like high-affinity nitrate t
C1504 (8812227-9246645 bps) C1501 (7954296-8085873 bps) C1497 (6751199-7952760 bps) C1493 (5697088-5998559 bps)	Opuchr04 g0013420-11 OG000082 Opuchr04 g0013430-11 OG000082 Opuchr04 g001340-11 OG000082 Opuchr04 g001340-11 OG00003283 Opuchr04 g001340-11 OG0000342 Opuchr04 g001340-11 OG0000342 Opuchr04 g0013530-11 OG000092 Opuchr04 g0013530-11 OG000092 Opuchr04 g0013530-11 OG000092 Opuchr04 g0013630-11 OG000082 Opuchr04 g0013700-11 OG0007265 Opuchr04 g0011760-11 OG0007265 Opuchr04 g0011760-11 OG0007265 Opuchr04 g0011760-11 OG0007265 Opuchr04 g0011760-11 OG0007265 Opuchr04 g001180-11 OG0007265 Opuchr04	$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	 stemmadenine O-acetyltransferase stemmadenine O-acetyltransferase stemmadenine O-acetyltransferase Zinc finger, BED-type Winorine synthase Cytochrone P450, E-class.group I hypothetical protein MIMGU mgVra0268911mg, partial tabersonine-19-hypothytoxy-O-acetyltransferase NAD/P)H-guinone oxidoreductase subunit 2 A, chloroplastic PREDICTED: NAD(P)H-guinone oxidoreductase subunit 2 B, chloroplastic, partial stemmadenine O-acetyltransferase hypothetical protein CUMW 134320 stemmadenine O-acetyltransferase uncharacterized protein product DELLA protein RCI-1ike Protein kinase domain * 7-deoxyloganetic acid synthase unnamed protein product NAC domain Hupothetic elpident protein At3g28410 uncharacterized protein product NAC domain probable tRNA N6-adenosine threonylcarbamoyltransferase probable tRNA N6-adenosine threonylcarbamoyltransferase elongation factor 1-alpha-like Mitchondrial import inner membrane translocase subunit TM14-3, partial guitative D-oysteine desulfhydrase 1, mitchondrial guitamate decarboxylase-like DNA polymerase alpha/epsilon, subunit B protein NRDE2 hornolog protein LOC113742114 isoform X5 uncharacterized protein LOC11374214 isoform X5 uncharacterized protein LOC11374214 . uncharacterized protein LOC11374214 isoform X5 uncharacterized protein LOC11374214 isoform X5 uncharacterized protein LOC112177048 isoform X2 Redox 2 # redox 2 vinchine synthase-like
0 bps	Opuchr04 g0010120-1.1 OG0003212 Opuchr04 g0010130-1.1 OG0001436 Opuchr04 g0010140-1.1 OG0001436 Opuchr04 g0010160-1.1 OG0014255 Opuchr04 g0010160-1.1 OG0002333 Opuchr04 g0010190-1.1 OG00012821 Opuchr04 g0010220-1.1 OG0012323 Opuchr04 g0010220-1.1 OG0002590 Opuchr04 g0010220-1.1 OG0002590 Opuchr04 g0010220-1.1 OG0003363 Opuchr04 g0010220-1.1 OG0003363 Opuchr04 g0010220-1.1 OG0005062 Opuchr04 g0010230-1.1 OG0005062 Opuchr04 g0010230-1.1 OG0005061 Opuchr04 g0010330-1.1 OG0005061 Opuchr04 g0010330-1.1 OG0010273 Opuchr04 g0010330-1.1 OG00010273 Opuchr04 g0010400-1.1 OG00010273 Opuchr04 g0010400-1.1 OG000102521 Opuchr04 g0010440-1.1 OG000102521 <th>$\begin{array}{cccccccccccccccccccccccccccccccccccc$</th> <th> strictosidine beta-D-glucosidase uncharacterized protein LOC113740142 oleosin 1-like protein CHUP1, chloroplastic-like translation initiation factor elF-28 subunit delta-like isoform X3 UspA (IPR006016) / Rossmann-like alpha/beta/alpha sandwich fold protein decapping 5-like single-stranded DNA-binding protein, mitochondrial Zinc finger, C3HC-like piriformospora indica-insensitive protein 2-like isoform X1 cytochrome c-type biogenesis CcmH-like mitochondrial protein mavicyanin-like DEAD-box ATP-dependent RNA helicase 510 kinesin-like protein KIN-8A isoform X1 protein MI2U-KUSE1-like G patch domain-containing protein TGH-like putative phospholipid-transporting ATPase 9 isoform X10 uncharacterized protein LOC113741806 isoform X2 ethylene-responsive transcription factor WIN1-like F-box/kelch-repeat protein Al3g23880-like calcium sensing receptor, chloroplastic isoform X1 AMP-binding domain-containing protein unnamed protein product strictosidine synthase strictosidine synthase<!--</th--></th>	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	 strictosidine beta-D-glucosidase uncharacterized protein LOC113740142 oleosin 1-like protein CHUP1, chloroplastic-like translation initiation factor elF-28 subunit delta-like isoform X3 UspA (IPR006016) / Rossmann-like alpha/beta/alpha sandwich fold protein decapping 5-like single-stranded DNA-binding protein, mitochondrial Zinc finger, C3HC-like piriformospora indica-insensitive protein 2-like isoform X1 cytochrome c-type biogenesis CcmH-like mitochondrial protein mavicyanin-like DEAD-box ATP-dependent RNA helicase 510 kinesin-like protein KIN-8A isoform X1 protein MI2U-KUSE1-like G patch domain-containing protein TGH-like putative phospholipid-transporting ATPase 9 isoform X10 uncharacterized protein LOC113741806 isoform X2 ethylene-responsive transcription factor WIN1-like F-box/kelch-repeat protein Al3g23880-like calcium sensing receptor, chloroplastic isoform X1 AMP-binding domain-containing protein unnamed protein product strictosidine synthase strictosidine synthase<!--</th-->

Number of genes assigned to an orthogroup from a specific species
 Presence or absence of genes from a syteny relationships with *O. pumila* { • Present Absent }

Supplementary Fig. 43. Monoterpene indole alkaloid (MIA) gene-clusters identified in chromosome 4 of the *Ophiorrhiza* genome assembly. The position of the gene-cluster is scaled based on chromosome size and its physical position on the chromosome. The gene description, colored as orange, represents genes with functions associated with MIA biosynthesis. O.p: *Ophiorrhiza pumila*; C.a: *Camptotheca acuminata*; C.r: *Catharanthus roseus*; G.s: *Gelsemium sempervirens*.

	Gene idOrthogroupO C C C C C C DescriptionOpuchr05_g0058170-1.1OG00120901000••Opuchr05_g0058180-1.1OG0012090111••Itertaricopeptide-like helical domain superfamilyOpuchr05_g0058180-1.1OG00012090111••Itertaricopeptide-like helical domain superfamilyOpuchr05_g0058190-1.1OG0003740111••Itertaricopeptide-like helical domain superfamilyOpuchr05_g0058200-1.1OG0000203740111••Itertaricopeptide-like helical domain superfamilyOpuchr05_g0058210-1.1OG0002037211••Itertaricopeptide-like helical domains on the helical domains
32257636 bps C1565 (26825172-26990957 bps) C1559 (24139397-24663912 bps)	Opuchr05_g0053440-1.1 OG000022_OS 25 13 10 15 * * secologanic acid 7-hydroxylase famesyl diphosphate synthase Opuchr05_g0053510-1.1 OG000022_OS 25 13 10 15 * * 7-deoxyloganic acid 7-hydroxylase Opuchr05_g005350-1.1 OG000022_OS 25 13 10 5 * AAA-TPase Opuchr05_g005380-1.1 OG000022_OS 25 13 10 5 * secologanin synthase Opuchr05_g005380-1.1 OG0000022_OS 25 13 10 15 * secologanin synthase Opuchr05_g005380-1.1 OG0000022_OS 25 13 10 15 * secologanin synthase Opuchr05_g005380-1.1 OG0000022_OS 25 13 10 15 * secologanin synthase Opuchr05_g005380-1.1 OG0000022_OS 13 10 15 * secologanin synthase Opuchr05_g005380-1.1 OG0000022_OS 13 10 5 * * unnamed pro
	Opuchr05_g0008060-1.1_OG0000421_5_7_7_7_6_**•Multi antimicrobial extrusion protein Multi antimicrobial extrusion protein Multi antimicrobial extrusion protein Opuchr05_g0008090-1.1_OG0004051_2_2_2_1_1_x*••Opuchr05_g0008100-1.1_Og0004051_2_2_2_1_1_x*******Opuchr05_g0008130-1.1_OG0000191_47_0_0_3_3** <td< th=""></td<>
C1541 (4233711-4239798 bps) C1538 (3339095-3340799 bps) C1537 (3084158-3137612 bps) C1532 (1313179-1402999 bps) C1527 (170887-361673 bps) 0 bps	Opuchr05_00005530-1:1 OG0008024 1 <t< th=""></t<>
	OpuchrolsSourcestImage: SourcestOpuchrolsSourcestSourcestImage: SourcestOpuchrolsSourcestImage: SourcestImage: SourcestOpuchrolsSourcestSourcestImage: SourcestOpuchrolsSourcestImage: SourcestImage: SourcestOpuchrolsSourcestImage: So

(2) Presence or absence of genes from a syteny relationships with O. pumila { • Present × Absent }

Supplementary Fig. 44. Monoterpene indole alkaloid (MIA) gene-clusters identified in chromosome 5 of the *Ophiorrhiza* **genome assembly.** The position of the gene-cluster is scaled based on chromosome size and its physical position on the chromosome. The gene description, colored as orange, represents genes with functions associated with MIA biosynthesis. O.p: *Ophiorrhiza pumila*; C.a: *Camptotheca acuminata*; C.r: *Catharanthus roseus*; G.s: *Gelsemium sempervirens*.

				(1)	_	()	
	Gene id	Orthogroup	0.	ς γ°ς	, , 0	۶ ۲	ຈ ບັບ	ر ، 	e Description
59986529 bps	Opuchr06_90113730-1.1 Opuchr06_90113760-1.1 Opuchr06_90113770-1.1 Opuchr06_90113780-1.1 Opuchr06_90113780-1.1 Opuchr06_90113800-1.1 Opuchr06_90113810-1.1 Opuchr06_90113820-1.1	OG0002046 OG0002046 OG0007265 OG0007545 OG0006798 OG0003811 OG0002234 OG0003796	2222112222	66122422	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	×	× ×	•••••	7-deoxyloganetic acid synthase # 7-deoxyloganetic acid synthase 7-deoxyloganetic acid synthase probable 3-deoxy-D-manno-octulosonic acid transferase leucine-rich repeat receptor-like protein kinase PXC1 uncharacterized protein LOC113776178 isoform X2 Pyruvate kinase uncharacterized protein LOC113694709 isoform X1
40761 bps) 75133 bps)	Opuchr06_g0113530-11 Opuchr06_g0113540-1 Opuchr06_g0113550-1 Opuchr06_g0113350-1 Opuchr06_g0113350-1 Opuchr06_g0113360-1 Opuchr06_g011360-1 Opuchr06_g011370-1 Opuchr06_g0113710-1 Opuchr06_g0113710-1 Opuchr06_g0113720-1	OG0008470 OG0000633 OG0000807 OG0004674 OG000598 OG000598 OG0004591 OG0004536 OG000273 OG0006603 OG0012375	13212351311	15311721422	1 1 1 1 2 1 1 3 1 2 3 1 2 3 1 1 1 1 2 3 1 1 1 1 2 3 1 1 1 1			• • • • • • • •	delta(14)-sterol reductase histone deacetylase HDT1-like isoform X1 splicing factor U2af small subunit B-like enolase peroxiredoxin-2E-2, chloroplastic-like glucomannan 4-beta-mannosyltransferase 9 uncharacterized protein Al2g27730, mitochondrial-like isoform X3 uncharacterized protein LOC113695442 uncharacterized protein LOC113697438 isoform X2 AT-hook motif nuclear-localized protein 7-like unnamed protein product
58498 bps) 74411 bps)	Opuchr06_g0097330-11 Opuchr06_g0097420-11 Opuchr06_g0097420-11 Opuchr06_g0097430-11 Opuchr06_g0097450-11 Opuchr06_g0097450-11 Opuchr06_g0097450-11 Opuchr06_g0097600-11 Opuchr06_g0097600-11 Opuchr06_g0097600-11 Opuchr06_g0097700-11 Opuchr06_g0097700-11 Opuchr06_g0097700-11 Opuchr06_g0097700-11 Opuchr06_g0097700-11 Opuchr06_g0097700-11 Opuchr06_g0097700-11 Opuchr06_g0097700-11 Opuchr06_g0097700-11 Opuchr06_g0097700-11 Opuchr06_g0097820-11 Opuchr06_g0097800-11 Opuchr06_g0097800-11 Opuchr06_g0098100-11 Opuchr06_g0098150-11 Opuchr06_g0098150-11 Opuchr06_g0098150-11 Opuchr06_g0098150-11 Opuchr06_g0098150-11 Opuchr06_g0098210-11 Opuchr06_g0098210-11 Opuchr06_g0098210-11 Opuchr06_g0098250-11 Opuchr06_g0098250-11 Opuchr06_g0098250-11 Opuchr06_g0098250-11 Opuchr06_g0098250-11	OG0000118 OG0000118 OG0004775 OG000473 OG0010047 OG000116 OG0000872 OG0000872 OG000632 OG0000632 OG000118 OG00014129 OG00014129 OG000018	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	8 0 0 2 2 2 8 0 0 0 0 0 0 0 0 8 0 0 0 0 0 8 0 0 0 0 0 8 0 0 0 0 0 8 0 0 0 0 0 8 0 0	1 3 1 3 0 02 1 1 1 3 0 2 1 1 1 3 0 0 1 1 3 0 0 0 0 0 0 0 0 0 0 0 0 0 0			* * * * * * * * * * * * * * * * * * * *	rankinidine/humantenine-11-hydroxylase 3 uncharacterized protein LOC113708247 uncharacterized protein uncharacterized protein LOC113723305 rankinidine/humantenine-11-hydroxylase 3 uncharacterized protein LOC108660713 hypothetical protein TorRG33x02_048750 ABC transporter-like putative U-box domain-containing protein 50 isoform X2 putative U-box domain-containing protein 50 isoform X2 putative U-box domain-containing protein 50 isoform X2 putative U-box domain-containing protein 50 isoform X2 probable F-box protein At4g22030 rankindine/humantenine-11-hydroxylase 3 hypothetical protein CRG98 035434, partial actin-related protein 5 isoform X2 Calcineurin-like phosphoesterase domain, ApaH type uncharacterized protein LOC108220559 Defensin-like protein 1_OC108220559 Defensin-like protein 1_OC113696826 hypothetical protein CDL12_03455 tabersonine 6,7-epox/dase T ABC transporter G family member 29-like ACT domain hypothetical protein TanjilG_11455 Ribosomal protein TANL8 Protein kinase domain rankindine/humantenine-11-hydroxylase 3 uncharacterized protein LOC113719845, partial rankindine/humantenine-11-hydroxylase 3 uncharacterized protein TANL8 Protein kinase domain rankindine/humantenine-11-hydroxylase 3 uncharacterized protein LOC113719845, partial rankindine/humantenine-11-hydroxylase 3
	Opuchr06_g0093040-1.1 Opuchr06_g0093060-1 Opuchr06_g0093060-1 Opuchr06_g0093106-1.1 Opuchr06_g0093160-1.1 Opuchr06_g0093270-1.1 Opuchr06_g0093330-11 Opuchr06_g0093330-11 Opuchr06_g0093330-11 Opuchr06_g0093340-1.1 Opuchr06_g0093470-1.1 Opuchr06_g0093610-1.1 Opuchr06_g0093620-1.1 Opuchr06_g0093620-1.1 Opuchr06_g0093620-1.1 Opuchr06_g0093630-1.1 Opuchr06_g009370-1.1 Opuchr06_g009370-1.1 Opuchr06_g009370-1.1 Opuchr06_g009370-1.1 Opuchr06_g009370-1.1 Opuchr06_g009370-1.1 Opuchr06_g009370-1.1 Opuchr06_g009370-1.1 Opuchr06_g009370-1.1 Opuchr06_g0093810-1.1 Opuchr06_g0093810-1.1 Opuchr06_g0093810-1.1 Opuchr06_g0093810-1.1 Opuchr06_g0093810-1.1 Opuchr06_g0093810-1.1 Opuchr06_g0093810-1.1 Opuchr06_g0093810-1.1	OG0000291 OG00051135 OG001285 OG0003150 OG000180 OG00011699 OG000748 OG000080 OG000080 OG000080 OG000180 OG000180 OG000180 OG000180 OG000187 OG000187 OG000187 OG000187 OG000187 OG00018229 OG0000914 OG0002829	7 332 78 13 266 248412 3 5 6363 33	2 1 1 1 2 0 0 0 3 118 1 1 1 0 0 0 0 0 0 0 0 0 0 0 0 0	3 6 3C 1 1 0 0 1			* * * * * * * * * * * * * * * * * * * *	vinorine synthase uncharacterized protein pentatricopeptide repeat-containing protein At5g57250, mitochondrial B3 domain-containing protein RLSI-like 60S ribosomal protein L37-3 uncharacterized protein LOC113739276 protein LOC113739276 protein LOC113689119 uncharacterized protein uncharacterized protein DCC109183381 dihydropyrimidine dehydrogenase (NADP(+)), chloroplastic EF-Hand 1, calcium-binding site Protein kinase domain Malectin-like carbohydrate-binding domain uncharacterized protein At2227730, mitochondrial-like isoform X3 DDT domain-containing protein DDB_G0282237-like strictosidine beta-D-glucosidase SKP1-like protein TA Nypothetical protein G2 G382530 uncharacterized protein LOC113754190 Protein misot like polyneuridine-aldehyde esterase cadmium/zinc-transporting ATPase HMA3-like Endonuclease/exonuclease/phosphatase superfamily cadmium/zinc-transporting ATPase HMA3-like

Opuchr06_g0007620-1.1 OG008895 1 1 1 • • ABC transporter-like Opuchr06_g0007630-1.1 OG0017640-1.1 OG0007630-1.1 • • * KRNA A642'O-Tibosylphosphate transferase-like isoform X1 Opuchr06_g0007640-1.1 OG0008946 1 1 • • Senescence/spartin-associated Opuchr06_g0007670-1.1 OG0008946 1 1 1 • • Intervence/spartin-associated Opuchr06_g0007670-1.1 OG0008946 1 1 1 • • Intervence/spartin-associated Opuchr06_g0007680-1.1 OG0000709 4 2 13 4 × * hydrolase 3 Opuchr06_g0007680-1.1 OG000057 2 4 1 * • probable 3-hydroxyisobutyrate dehydrogenase-like 1, mitochondrial Opuchr06_g0007720-1.1 OG00002253 2 2 * • hydrolase-2-like Probable 3-hydroxyisobutyrate dehydrogenase-like 1, mitochondrial Opuchr06_g0007740-1.1 OG0006924 1 1 1 * • receptor-like protein kinase HSL1 Opuchr06_g0007780-1.1 OG0006924
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C1592 (4975628-5129694 bps)

C1652 (55776348-55840761 bps) C1651 (55691098-55775133 bps)

C0000 (46097318-46558498 bps)

C1624 (44145373-44674411 bps)

0 bps

(2)

Presence or absence of genes from a syteny relationships with O. pumila { • Present Absent }

Supplementary Fig. 45. Monoterpene indole alkaloid (MIA) gene-clusters identified in chromosome 6 of the *Ophiorrhiza* **genome assembly.** The position of the gene-cluster is scaled based on chromosome size and its physical position on the chromosome. The gene description, colored as orange, represents genes with functions associated with MIA biosynthesis. O.p: *Ophiorrhiza pumila*; C.a: *Camptotheca acuminata*; C.r: *Catharanthus roseus*; G.s: *Gelsemium sempervirens*.

Supplementary Fig. 46. Monoterpene indole alkaloid (MIA) gene-clusters identified in chromosome 7 **of the** *Ophiorrhiza* **genome assembly.** The position of the gene-cluster is scaled based on chromosome size and its physical position on the chromosome. The gene description, colored as orange, represents genes with functions associated with MIA biosynthesis. O.p: *Ophiorrhizapumila;* C.a: *Camptotheca acuminata;* C.r: *Catharanthus roseus;* G.s: *Gelsemium sempervirens.*

Supplementary Fig. 47. Monoterpene indole alkaloid (MIA) gene-clusters identified in chromosome 8 of the *Ophiorrhiza* **genome assembly.** The position of the gene-cluster is scaled based on chromosome size and its physical position on the chromosome. The gene description, colored as orange, represents genes with functions associated with MIA biosynthesis. O.p: *Ophiorrhiza pumila*; C.a: *Camptotheca acuminata*; C.r: *Catharanthus roseus*; G.s: *Gelsemium sempervirens*.

Supplementary Fig. 48. Monoterpene indole alkaloid (MIA) gene-clusters identified in chromosome 11 of the *Ophiorrhiza* **genome assembly.** The position of the gene-cluster is scaled based on chromosome size and its physical position on the chromosome. The gene description colored as orange represents genes with functions associated with MIA biosynthesis. O.p: *Ophiorrhiza pumila*; C.a: *Camptotheca acuminata*; C.r: *Catharanthus roseus*; G.s: *Gelsemium sempervirens*.