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Supplemental Information

**Coevolution of enamel, ganoin, enameloid, and their
matrix SCPP genes in osteichthyans**

Kazuhiko Kawasaki, Joseph N. Keating, Mitsushiro Nakatomi, Monique Welten, Masato Mikami, Ichiro Sasagawa, Mark N. Puttick, Philip C.J. Donoghue, and Mikio Ishiyama

A

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exon2          exon3          exon4          exon5
AMELX_human    MGTWILFACL LGAAFAMP  LPPHPGHPG---YINFYE  NSHSQAINVDRTAL  VLTPLKWKYQS-IR---PP
AMEL_pig       MGTWIFFACL LGASLAMP  LPPHPGHPG---YINFYE  -----          VLTPLKWYNMIR---HP
AMEL_platypus  MGTWILFTCL LIGAAFAIP  LPPHPAHPG---YINFYE  -----          VLTPLQWYONMKR---QQ
AMEL_Anolis    MEGWTLVMCL LSTTFAIP  LFPQ---HPG---YINFYE  -----          VMTPLKWKYOSLIG---HQ
AMEL_caiman    MEGWMLITCL LGATFAIP  LPPHPHPHPG---YVNFYE  -----          VLTPLKWKYOSLMR---QP
AMEL_axolotl   MGPWILLTCL LLSASCAMP  LPPHPNHPG---YINFYE  -----          VMTPLKWKYOSMMR---QQ
AMEL_Xenopus   MRLWVMLTAL TGAAFSVP  LPPHPHPHPG---YVNFYE  -----          ILSPLKWKYOSMMK---HQ
AMEL_lungfish  MKTLVLVACL LSSSIIVP  LPPAGQH-GNPGVINIYE  -----          LLSPLYAWLNKMMRQOQP
AMEL_coelacanth MKSLSLITCF LGAALALP  LGPQQQPQGPASIVNFYE  -----          MMSPLHWAD-WIROOSQP
                LGPHRQNPGTANVVNYE
<- aromatic residue rich region (YY)

exon2          exon3          exon4          exon5
SCPP5_medaka   MKFAILCLCL VSTACAVE  FQYLPHFET---GSRQQQP---EVC--  GNNP-FSSGF--PQE-----GLPGAYNIEL  IYFHRFPGGPTGGANET-Q
SCPP5_tilapia  MKLAILCLCL ASTACAAP  FQYLPHYT---GSRQQMP---TLC--  MNNP-FTAGF--PHT-----GLPGGYNVEL  IYFHRFPGGA-GVSSBA-Q
SCPP5_fugu     MKLAILCLCL ASTASAAP  S---FHYLPHYGGPQQVAPA---PQQ--  VKTP-FTAAQTLTQP-----GLVGGYNVEL  LYFHRFPGTF--GANTA-Q
SCPP5_stickleback MKLVIFCLCL ASTACAAP  SI--FHYLPHYAGSRQQVE---PQQVI  VGNP-FTAGQSLPPP-----GAAGAYNVEL  IYFHRVAGGV-GGTNAG-Q
SCPP5_zebrafish MWTSLLLCLL LAGAVSAA  LSPFFNYLPHYGSPR-----Q--  GNTGGFQGMSPQHP-----AMNAPIIMEI  IFPPRFANPAGGAAGT-S
SCPP5_eel      MKTAIACLCF ASTTICAA  MTSLYDYLPQIGVPPQPSQ---PQQ--  RTTNVVAPAPPAQPP-----GMAAPIIMEI  VFPPRFPSQVTGGQQTG-S
SCPP5_gar      MKTIILLTCL FGSIFAAP  MQPLLYEFLPOVETP---QCTAPYRQQPQ--  -NNPYAPPSTPQQQ-----GPTRPAPFEI  LFYGYPRPSPFGGPEHAGA
SCPP5_bichir   MKATVLLMCL LGLTTFAVE  MQRLYEFLPOVNSP---QSAQPS-QQP--  QSNPERAPIVPEAPOTPAAPPAEPAGPAPPALEI  MFYGYPGQTLGQP---SS
                <- aromatic residue rich region (YY)

exon6
AMELX_human    YPSYGYEPMG GWLHHQIIPVLSQQHPPTHT-LQPHHHIPVVPAQQPV-IPQQPMMFVPGQHSMTPIQHHQPNLP---E--
AMEL_pig       YTSYGYEPMG GWLHHQIIPVVSQQTPQSHA-LQPHHHIPMVPAQQPG-IPQQPMMELPGQHSMTPTQHHQPNLP---L--
AMEL_platypus  YPSYGYEPMG GWLHHQIIPVLSQQQTPHTA-LQPHHHIPVMAAQQEM-IPQQPMMEMPGQPSVTPTOHHQSNLPQBAQQ--
AMEL_Anolis    YPRYGFEPMG GWMHHAAGP-TMHTTFQSH-PSVHSTLHMQQPPHR---ALNPHMQPSGHNEFGSMFGQNTLMP-----
AMEL_Paleosuchus YSSYGYEPMG GWLHQPMPL-IAQQHPPIQT-LTPHHQIPFLS-----PQHFLMQMPGPHQMMPIPOQQPSLOMPVQEPV
AMEL_axolotl   YPSYGYEPMG GWLQQMMP-GSEPMQCHM---QPIPMQPHHEMMLPQQMPLP---HGQMMPLPQHQQLLPQPHQC--
AMEL_Xenopus   YPNYGYEPMG SWLQNVVPAPEMMPQQQH---PQQHVVPKLPFHHPFLIPQQEVVVEP--HPEVILTPQHTHQLKPTYL--
AMEL_lungfish  YPAFGFEPMQ PW-----
AMEL_coelacanth YPVIEIE-----VKPRFPMSPQEPISPMQPPQPMPE-----
                YY -><- E/Q-rich core region (E/Q)

exon6          exon7          exon8          exon9
SCPP5_medaka   FSSYGFIKYS IPQQPPGRQLEI FYEYDFSQ-QR  IVQNLPEMSNPIM-----PD  VFP-IEYEPQNM-----PQQTIN
SCPP5_tilapia  FSSFLIKYSI PQQPPGRQVEV FYAYDFSQ-QR  IIPNIPMTKGPV-----PE  VLP-FDYPPQNI-----PQQNMN
SCPP5_fugu     FFSHGFIKYS IPQQPPGRQVEV YYPYDFSQ-QR  ILTNIPLTNVPHL-----PS  VLP-IEYPAHI-----PN
SCPP5_stickleback SF---GFIKYS IPQQPPGRQVEV YYPYDFSQ-QR  IMTNLPEMTNSPQM-----EN  VFP-FEYEPQNI-----PQQIPN
SCPP5_zebrafish SFPQAFIKYS LKAPGRKVEI FYPYDFGR-AQ  DQPNVPLI---POL-----PN  IFF-FDLMPQTV-----PQQPPV
SCPP5_eel      AFPSCAFIKY IKAPGRQVEI FYPYDLIQQQQ  MLPNVQMPAMPNIPQFPQI-----PN  LFP-YGFLPPTG-----PQQN-N
SCPP5_gar      MFSQAFIRQK IPQPPGRQLEI LFYYSFGQHQ  MFPGASNMPQPPQMQQLPQLPQQN  MFP-YGNMPTGG-----PQQNPN
SCPP5_bichir   MFSQGFIRQK IPQPPGRQLEI LYFYGFQQ-QQ  MFPGFSNLSFPVPMQ-----FP  MLPN  MFP-YGSMVPM---PQQSPQTPN
                (YY)-><- E/Q-rich core region (E/Q)
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B

	exon2	exon3	exon4
AMBN_human	---MKDLILILCLEMSFAVP	--FFFO-----	CSGTPGMALE TMRQLGSL-QLNLTLSQ
AMBN_platypus	---MENLVLLSLFGTSFAIP	--VFLO-----	PGGPPGMGLE TMRHLG---NLNLLFO
AMBN_caiman	---MNVWMLTLCLLGTGFALP	--MYFO-----	HTGTRGMAMLE -MROYGR--QNMNMLEFO
AMBN_Anolis	---MKQWMLVSSLLGICSAIP	--LFFO-----	HAGIPGMGLE RTROLGGL-NIPPAVFO
AMBN_Xenopus	---MEPLVLVLCLLSTAMAYP	--MHFO-----	APGTQGLAILE TMRQQAANTLTAFPSQ
AMBN_lungfish	---MKALSLLMCLLGTIHTLFP	--LMFO-----	QAGTFGVALLE RMRKYGQ--FGMSFDDQ
AMBN_coelacanth	---MKRWILGCFLGTAFSMP	--MFFO-----	IPMARVLE KMRQYGG--EYMPQSPQ
AMBN_bichir	---MKAAILLICALGTTWAMP	MQVYQOMQOQIQLOQQFO	---HGGIPALATIQE MMRDIARYKSLMCHYPE
AMBN_sturgeon	---MKAVVILMCLLGTTCVVP	LOVYHQIQOQOQIQAQEQAQOQFO	PGMLRYPILE MMQDIARYKRFKQOHPQ
AMBN_gar	---MRSATVLMCLIGTTLTSMF	LQVYQQLQROMLQIQHHQOQFO	QOQAGNPRYFLE TMRMARYKAFMQOQSM
AMBN_zebrafish	MTQMRLIVMIVCFLAGTSAVP	-----	-----
AMBN_medaka	---MIIILLLSCITLLVSAVP	-----	-----
AMBN_tilapia	---MIIITLLSCFTIVVSAVP	-----	-----
AMBN_fugu	---MIVPTLLSWFIIMASAAP	-----	-----

	exon5	
AMBN_human	YSRYGFGKSFNSLMMHGFL	PPHSSLPWMRPR--EHETQQ--
AMBN_platypus	FSRFGFGKQFNSLMMHGFL	PPHSNYPWLSPR--DQETQQ--
AMBN_caiman	YGRYDYGEPFNSVWLHGFL	PPHSSFPWLQQR-POEHETQQ--
AMBN_Anolis	ALPYSYADPFHSMWGRSFL	PSHTFLWMTGGPQRQDTQQ--
AMBN_Xenopus	ISRFGYNDPVSVLWLHGFL	PPHSSYPWLHOR-PQLSDNQQ--
AMBN_lungfish	FFVNGYNQFQSLLPWYQQL	HLFPALVPWAQOP--HQENQQ--
AMBN_coelacanth	NPFWGYNPVTSOQDMSWL	-----THLVNSMFPOLRLYPWLQPP--SQHDNQQ--
AMBN_bichir	LHFQLNFPDLQPGAPMNEAQLFQVPHGQAPSMLPPQOQPSQVPSQLPPL	PPQLPSVL--TPPLNQLPPQMPHFQLIE-AQPGWEQQ--
AMBN_sturgeon	MFPSLNLPQAQPLPPKKTGQOQPPQOQSQSPSLRPPQOQPOQPPQOQPPQOQ	YQOQPPQOQPOSSPOLPPNFQVVEYLPGLQOE-RLINRNQQ--
AMBN_gar	MFPSLNMPHSTGLQPPQIPQFTGVQLPHNTGLQPPQIQAPSQTPPTSQAP	SQLPQOQPSQLPHQMTPSQOQVEYQOQGLQOM-QLVHAEQQ--
AMBN_zebrafish	ISPNDLKESSLVSG--DMQO	FOQPSRMLSGLHOOVNEPNASQPNP
AMBN_medaka	VAPNTHARMLQGGGA--TORIHVQSTNHKPEVQTPAFLSPIEQAQSGIQOQPPDQAVLEPSFSLFOHTWYQGGSTINEMFQNVPPFLPANHLTLGQLPL	STAVRQHEAQF
AMBN_tilapia	VNSNVRRPQLLQGGETATQGAQHVAANQKPESETPASISPSVQFO	AGGLQOQDPPQPI
AMBN_fugu	TGEPVSHFESVIIQGE-ATQGTDNEDYHTPAPPSSQVQOQALSLOQDPRGGFHLI	SE-QFLWSVVGGAFTF

	exon6	
AMBN_human	YEYSLEVVHPPPLPS-QESLKPQOQGLKPEFLQS--AAATTNQAT-ALKEALQPP	IHLGHLFLOEGKLELVQQRVAPSDKPPKPE
AMBN_platypus	YEYALPVHPPPPRPS-QRTLOPQOQPKQNAFLPF--TVAFSAQGGPPAQNGD	QOPIILHLOPPLQOQEKPSVQEQVAFPKRPPKPE
AMBN_caiman	YEYAMPVHPPPLPSLOTPLQOQPRIQANPSLRSSTLPTKQGGIQLNEALLP	VOVGQPPLOQOQGLFGIQOQLIPADKQL-C
AMBN_Anolis	YEYAMPVHPPPLPSLOTPIQFAEPRLEPEQEFFQITNLIPTRVQIPRYAIP	QPSIQOQTFQOQVEAHPVVDQOQQLPAERQTHC
AMBN_Xenopus	FEYALPIHPPLPGAQSEFAQTEKPGQHAQNMPE	---QOQAAASTDQVSHQPSLPLGFEILOQADPAMMPKGGPADGQI-QTVALYM
AMBN_lungfish	YEYALPVHPPQOQPPQPHLPPQOQPEVSPQOQPT	-----DQOQPSAQPSQOQGHSPQ
AMBN_coelacanth	YEYTLPIHPEVQF-QQPPAQOQPPVQOQIPT	-----QKTEQOQGOQPEVQSPVQOQHQPQLDLOQL
AMBN_bichir	YEYDAPVLLAHGPDQQSEALPTAGNVVVPATQOAVVVEQOQVQOQCGGT	QLNQPGQTPSAQGHLEVPQOQPMFTFGFVPEVPEOT---AKQOQ
AMBN_sturgeon	YEYESPVLFHQVPPQOQSTAVPATQOASVQPAQOQOQPGRQDQPEHQIQIPNAPGQOQPSHQ	-----GQOQPIQFTFGFLEMMHYQVQOQOTQO
AMBN_gar	YEYETEFLLEQVPEQOQTEQVQATAGSAPATPTAPAAPROQOQAGQOQHEQAQAPQPGVQVLEQENIQIQGTGINGQOQ	-----GTQGHQPTLPAQLPPE
AMBN_zebrafish	-----	-----
AMBN_medaka	-----	-----
AMBN_tilapia	-----	-----
AMBN_fugu	-----	-----

B (continued)

	exon7	exon8	exon9	exon10
AMBN_human	LPRVDFADPQGPS	LPGMDFPDQGPS	LPGLDFADPQGST	IFCIAR-LISHG-----PMPQN--KOSP
AMBN_platypus	LPALDFTVQLROS	-----	-----	IFPIAOKLISOG-----PROKA--EOSP
AMBN_caiman	LPALVYSGHLGOV	-----	-----	MYFIVHQLVHOG-----PMQEQ--QCPA
AMBN_Anolis	-----	-----	-----	IYPALH-GTQOD-----PSQQL--QQS
AMBN_Xenopus	-----	-----	-----	YQTIMNKLLQQGAGETIIPD-PAGTLEVNQQH
AMBN_lungfish	-----	-----	-----	IEVEIF--OTFOVAQQP-QPFQOM--IFPA
AMBN_coelacanth	-----	-----	-----	VSPSNGNAVQTPQVTQQPQQG--QQPQ
AMBN_bichir	-----	-----	-----	-----
AMBN_sturgeon	-----	-----	-----	-----
AMBN_gar	-----	-----	-----	-----
AMBN_zebrafish	-----	-----	-----	-----
AMBN_medaka	-----	-----	-----	-----
AMBN_tilapia	-----	-----	-----	-----
AMBN_fugu	-----	-----	-----	-----

	exon11
AMBN_human	L--YPGMLYVPF-----GANQL-----
AMBN_platypus	L--YPELVYVPF-----GANQQ-----
AMBN_caiman	L--HPALFYMSY-----AANQG-----
AMBN_Anolis	I--YPSLYMPY-----VANQG-----
AMBN_Xenopus	P--YPLGFFMQY-----GGGPG-----
AMBN_lungfish	F--E--YFYQF--FVQQGQOPM MFPFENSPP-QPFGQQGQYFVMLF TSDILPFVIQQPDIQQPQLPQVPP TNSFLP SIVQQGGQQQQQQQQQQQPQQ
AMBN_coelacanth	IQFFFTYEYLQMLPQVPGQQGQQPG GYPPYSFFP--ALACQPLDF-----QQQQQQQQQQQQQQQQQQQQQQPQQQP
AMBN_bichir	-----FSGYGLLL-PASFPPQEV-----QEGLEQQPPQQQPSQS
AMBN_sturgeon	-----FPGYSFLI-PAAPFQQ-----QAQPGQEQQQQ
AMBN_gar	V--FNTF--GILPIMPPQACDEQQ FPGYGVFF-NTAFPCQEV-----QPPQAAQPGAPPQQQ
AMBN_zebrafish	F--F--YYPILGQSS---RNET FS-YGVSAFQHAASQQYFANAAPALREARE-----QQGNAHQMRQFPQQQQQQQQ
AMBN_medaka	M--FPPYGFYPLFAPPY---GNQM FSPYGFPSIRESF FLOAPTQQLNNQAPPAENVV-----PAGAARQQTQQ
AMBN_tilapia	I--FPPYGYLPLFSSPY---GNQL FSPYGYPMILESGLPQIPANQLNSPVLPAENAGVAV-----DPSIDATQGIQQPQQQLQQQ
AMBN_fugu	I-SSPR-SFTPLISEAF---INQL-----

	exon12
AMBN_human	-----N-APARLGIMSSEEVA
AMBN_platypus	-----NAAPARLGKVSSEEMF
AMBN_caiman	-----G-APARLGIVSSEEML
AMBN_Anolis	-----A-APARQGVISSEEMQ
AMBN_Xenopus	-----G-PPARLGAMSSEET
AMBN_lungfish	GFPFIQFYLAALGHQ-NLPPQ V-----QGAISSEETC
AMBN_coelacanth	PPPSFVYVPLPARQEQLPQG G-FPNTFGMSSEEMQ
AMBN_bichir	MFPOLVYVPLQV---GVMA G---AIGVSSSEEMQ
AMBN_sturgeon	MYPOLVYIQLOP---GMAG G---AGVSSSEELQ
AMBN_gar	MIPQILYLIQCPM--RSAMG G---GGFGASSEEMQ
AMBN_zebrafish	FQPIF-YMVEQMP---QRM A---GSYGGLSSEELQ
AMBN_medaka	QNPPIVYMLQ-----QPM N---PAIGGLSSEELE
AMBN_tilapia	N-PQILYMLQ-----QPM S---SELGLSSEELQ
AMBN_fugu	-----D--STLSFSEEFEE

B (continued)

exon13
 AMBN_human GGRE-DPMAYGAMFPGFGMRP-----GFEQMPHNPAMGGDFLEFDSEVAATKGPEN-----EGGAC---GSPMPEANPDNLENPAFLTELEPAPHAG-LLV
 AMBN_platypus GGGG-EELAYGSIFFGFGSMRP-----GFGAMQINPALGKDFLEDDSETVAGK-PPG-----CGEVL---GSRDGAIPAGHPNPGLLPERISRLRSGAALT
 AMBN_caiman GGRG-GVFAFGAMIF-----GFRGMQDPAIQGDFTEDDNPATAHN-PAI-----CGGANCGFSRGRSFAVNRAGHSAILLPDGTAGHEG----
 AMBN_Anolis GGGF-GAPAYRAFPPDLFAMDT-----RFGNSPLN--CPGDYTVEDDTLGLITECPDV-----KGGANA-----GANPSGKGNNAVLLDGTGGALP---
 AMBN_Xenopus GGRTGAAHAFSSLYPGLLGMP-----RLESHQDPAIQGDFTEDDSEVAGQK-PTG-----CEVSCG-----PIESPQGVGSSIMIPLGLESPGCG-GET
 AMBN_lungfish GFPIQFYLCALGHQNLQONVQGAIEEIQANRAAGAAAAAVFNSMFAFAANYFGFGSVAARFISVDATIEDDHFVVGQGEQAGCGTGTGFGVGNLNAVGT-----PSSPAVNIPIPEGNPTG
 AMBN_coelacanth AAGGGLS-PALGGIIPGYVGNIPG-FGAGVPSFGGSVPGAANIPMF-----GFNTAVNPAGCGGVTVEDDPPPAEK-PASFGN-SPGT-GANPSGQGGSAFNPENPTGQVGSIPNPQANPPPSEGTPS
 AMBN_bichir QSAGDMS-PEMAGFIPGFATPT--FGGGPNFVGSQPAVGGTVFV--AGGI--PTAQGTQPGVLEEV-----TAQGTQPGVLEEV-----KLEIPV
 AMBN_sturgeon AAGG-IS-PGFAGFIPGFAGQA--FGGSIPSFGGSVPGAEGSIF--AGGV--PTGQGTNP-VVPEA-----GLPTVEIILPA
 AMBN_gar AAGG-VA--GFGGFVPGFAGSG--YGGFGGTVPGTE--GVFPTGQGIIEVAPDAGLF-PVQGG-----SPTAFVAFPA
 AMBN_zebrafish NMGR-VNMHLPAIRGNVFVSGPQTVVPIGFVRPPNEFPATGMTYPGITELQQPSIIIAVPPSRDAIEATGGSH-----PNSAVLESRISGTDQRDRAPCVIAEIEPLTNSFALD--SGHGAFISADVE--
 AMBN_medaka MQAK-LNOMSV--YSVLTNLPAGAGFVQPEAQAGLTHPEQGGVQPAVGTSA-----AGVRSIQGACSGSLLNSNSVFAQVKTAVEANAVQTQNHAK
 AMBN_tilapia MAAT-MGQLGV--YMNVLTNQFAGAVQPEAQAGLTHPEQGGVQPAVGTSA-----AGAQPIKALPC-SGSGFNASGFSAG-LAGPDAATAQTAEFPQ
 AMBN_fugu -----VNI--YLSTALTDPSAATVQVVKQAAGL---QNLVSTVGTFS-----TGAFQNC-VFTSSGPPMDTNGVAVG-SEKFAEEMATVQHEVE

exon13 (continued)
 AMBN_human -----FLKDDIPLPRDPSGKMKGLPSVTPAAVDPLMTEPELADVYRTYDADMTTSVDFQEEATMDTTMAFNSLQTSMPGNKAC-----EPEMMHDAWHFOEPP
 AMBN_platypus -----FSDFIPIHRQGGPAGQSKLPPGVTPPPADPLMTPEGSDLEFFEGDGTPLGLQREALDTTMTPTDQSTYMEGNEAQ-----QMQVTDQDQVQFQEP
 AMBN_caiman -----FLPNINDMPGQGVNVPVGG-RGTPEVTPATAPELTQGIQDSFMTFGAEGTVPLGIQKEVTADPTMFPPEAQHTLMAGNGAE-----QEQVMQDVWHFOEPP
 AMBN_Anolis -----NANSHLFSFAGCSQGLSGLAQATAAPHATQG--EFLPLDAADES-----MPLDATMFPD-LYTSNVGNEAG-----LAQVGDQDAWHFOEPP
 AMBN_Xenopus -----VFPFNINSFNMGFNEQSKIPEPGVTE-ANAPRLTHDTGAGYVPEFALDDTMEFGIQRENVISGDI TRANNAKGIESEIMQH-----EVHLDQNHNNYFOEPP
 AMBN_lungfish QIGNPQIIP-----QSNPSSFGGQIMQNPDINPSNQLPATPADI PAASAGNPSAATASFNEGGL-----FVIYYEGLNHFAARGPWFPP
 AMBN_coelacanth VQGVPPQPAPEGNLPNLEGN-----PTGQVGNQPAPOSKGPMGGATTPASTKNLTPANVPGGPEGGIVPFAGDITASAAARKVPTIIPAFFNPSLGGTATENNNAVRSNAAGVLPPIVVNEGLN-PLGGGWIIYP
 AMBN_bichir GQG-----GSPTIPEVPIIGQGNMPEKTEGAVPAQGGQQPSFQMKLENMTDGVLLDGVVPEEGTVQSFAPGSTQDTIGKKSPTAAPDPAATVE-----KAQAPTPEEVEMPDLDYDTANFYLHP
 AMBN_sturgeon GQG-----GGPTIPEVPIPTGQSSLP-----VMSKTNQPAPOSKLFTVGLTTPAVWRMLTPEGNSFAAA.
 AMBN_gar GHGNLPPTPGS-TVFV--QGS-----NNOFAPQSRDPAFSMTTPAGVRAATVGNPPAAAAAGTACPNAAAAATAAAE-----RVHVNHAEARNIPADDEMPQADDYRAE-FIYFP
 AMBN_zebrafish -----NNOFAPQSRDPAFSMTTPAGVRAATVGNPPAAAAAGTACPNAAAAATAAAE-----RVHVNHAEARNIPADDEMPQADDYRAE-FIYFP
 AMBN_medaka SLPTNRNLVRSKAN.
 AMBN_tilapia LQPMQETLV.
 AMBN_fugu AEELLPLTQTRSV.

C

	exon3	exon4	exon5	
ENAM_human	---MK-ILLVFLGLLGNSVAMP	MHMPRMPGFSKKEE--	-----MMR-YNQFN--FMNGPH-	
ENAM_pig	---MK-ILLVFLGLLCYSAAMP	MQMPRMPGFSKKEE--	-----MMR-YGHFN--FMNAPH-	
ENAM_platypus	---MK-SFLLFLGLFGASTAMP	MHMPRMPGFGKKEE--	-----MMF-YGQYN--FMNSPH-	
ENAM_Anolis	MQRMK-QLLLYLCLIGTSWAVR	LRKPRKAGFGKKEE--	-----MMQ-YGPFQ--YLNSPQ-	
ENAM_Xenopus	---MMV-PIILLCLFLISSNSVF	MHF--SHMSNKEE--	-----GME-YGPGY--YQN-PQ-	
ENAM_coelacanth	---MK-TLLLVCLLGTSLAGL	ARL--VRKARNKEE--	-----MVQKFTSYNRHMFPQORL	
ENAM_bichir	---MK-RGLLLLCLFLTALAFV	V----GVALAAQEE--	-----ALMCO-RWNALLGAFQOO	
ENAM_gar	---MK-IALLVLCFLGTSLAFV	I----VPGAFIDHE--	-----AKMOOYRWYG-APQOQRO	
ENAM_zebrafish	---MK-AVALLMCLLGSLSAAP	-----APDGGNE--	QAIAASHANTALQLMELYRMLGQLRQQ	
ENAM_trout	---MI-SVLLMCLLGFSLAAP	-----TPDGGDE--	Q-VAAHANEALRWEMELYRMYGSL---	
ENAM_fugu	---MVTILVALMVLLGTAWSAF	-----ALGEEAEEN--	---VAAHANALRWMMHYRLY---QOO	
ENAM_medaka	---MK--LFVFLYLIISASAAF	-----APDGGSE--	---VAAHANAALRLMELFRMY---QOO	
ENAM_tilapia	---MK-QLVFFICLLVSTLAAP	-----APAEISE--	---IAAHANEALRWEMERYLY---QOO	
	exon6			
ENAM_human	M-AHL-GPFFGNG--LPQQFPQYQM--		PMWPPPPNT-WHPRKSS-----APKRHNKTD-	
ENAM_pig	M-AHL-GTLYGNMQLPQFFPQYQM--		PMWPPPPNK-KHPKPS-----ASKQSKTD-	
ENAM_platypus	M-AQL-GPLYGYLQLPQFPQYQM--		PPWPPPPNG-WPAQNGGPPQ-----GGKPAKPG-	
ENAM_Anolis	L-TQLAASLYGYRSGFPQMVFPQQ--		PTFPLQRFYL-WPQMPVHQS-----ARPPQKAH-	
ENAM_Xenopus	M-PGI-GSMYGLG--FPHYSMIQL--		QCAFGRQPS-WLQNIARTQIHKETIPLNLPRFHLQPV-	
ENAM_coelacanth	QVMQL-RSLYGNAPQRL-MYEQQLQQQQQRKLLVARLLAQALKQTKQSRQAQNAAPQLQVQKQQLQPLQEPQVQQ-----KQKQQQLLQ			
ENAM_bichir				
ENAM_gar				
ENAM_zebrafish				
ENAM_trout				
ENAM_fugu				
ENAM_medaka				
ENAM_tilapia				
	exon6 (continued)			
ENAM_human	QTQETQKPNQ-----TQSKKPP-QKRFL--		KQPSHNOQOPE-----EEAQPPQ-	
ENAM_pig	PAPESQKPNQ-----PQPKTPT-PKQPL--		NEPSPPTQOPE-----EETQTPQ-	
ENAM_platypus	QSNTKQPNP-----SQPKKPP-QQPPF--		KQAAKQPAQPSKET-----EETRQPPQ-	
ENAM_Anolis	QTFVPKKPNSRPRLKPRLVFPQPR:QQPQPKIHLPAKQPF		---VATHRQOQPTPIQPPKGEKQPOQ-	
ENAM_Xenopus	QSLSQLPAP-----FVQQQLNPPQSKSNYVTKQF		HQPLQLPPAFD-----QQAQAPM-	
ENAM_coelacanth	QAQNLKNVLP-----QQPQRLQKQSPQQLQHQKQKPPQQAQKQEQSRQR--		FSRKQPPQKDLQQL	
ENAM_bichir				
ENAM_gar				
ENAM_zebrafish				
ENAM_trout				
ENAM_fugu				
ENAM_medaka				
ENAM_tilapia				
	exon7	exon8		
ENAM_human	AFPPFGNGLFVYQPPWQI--PQ	RLPPPQY-----GRPPIQNEE--GG		
ENAM_pig	AFPPFGNGLFVYQPLWHV--PH	RIPPGY-----GRDPTQNEE--GG		
ENAM_platypus	MYPPFGNGFFPQLOPWQI--PP	QMPPGY-----GRPPGQNEE--GG		
ENAM_Anolis	AFPP-----HQQPWQF--PQ	RMPP-----GRPPNEE--G		
ENAM_Xenopus	MFPFSSNIMNRYQYVW--QQ	PFMHGF-----GRPPNEE--GN		
ENAM_coelacanth	VYPSQGYIRMLPQMLWQQRQQQ	PV-----GRRKQGGYEEQGMG		
ENAM_bichir	GNYP-----KVVYIQLSPPE-MAG	MGNR-----AAPAEQE--GG		
ENAM_gar	QDFP-----KVVYVHLPP--MAG	MGPNT-----AAPAEQEA--GA		
ENAM_zebrafish	GY-GA-----VAG-PQL--P	GFFFNPAFA-----PTGDGDEE--VA		
ENAM_trout	GQ-LA-----ABA--QP-PLMN	KFFYPSPMN-----DEE--A		
ENAM_fugu	AMLQN-----PFV--FPAGRAQ	VNAAPVQPLAVAPQAAVAHPAAVAHPAAVGDDEE--AQ		
ENAM_medaka	GLA-N-----PFL--PAAGAH	INV-PVQPA-----AATPAGDADEE--AE		
ENAM_tilapia	GIVQN-----PFV--PSANSF	VDAAPACPADVPE-----PAHVQVGDQEE--TE		

C (continued)

ENAM_human exon9
 ENAM_pig
 ENAM_platypus
 ENAM_Anolis
 ENAM_Xenopus
 ENAM_coelacanth
 ENAM_bichir
 ENAM_gar
 ENAM_zebrafish
 ENAM_trout
 ENAM_fugu
 ENAM_medaka
 ENAM_tilapia

ENAM_human exon9 (continued)
 ENAM_pig
 ENAM_platypus
 ENAM_Anolis
 ENAM_Xenopus
 ENAM_coelacanth
 ENAM_bichir
 ENAM_gar
 ENAM_zebrafish
 ENAM_trout
 ENAM_fugu
 ENAM_medaka
 ENAM_tilapia

ENAM_human exon9 (continued)
 ENAM_pig
 ENAM_platypus
 ENAM_Anolis
 ENAM_Xenopus
 ENAM_coelacanth
 ENAM_bichir
 ENAM_gar
 ENAM_zebrafish
 ENAM_trout
 ENAM_fugu
 ENAM_medaka
 ENAM_tilapia

ENAM_human exon9 (continued)
 ENAM_pig
 ENAM_platypus
 ENAM_Anolis
 ENAM_Xenopus
 ENAM_coelacanth
 ENAM_bichir
 ENAM_gar
 ENAM_zebrafish
 ENAM_trout
 ENAM_fugu
 ENAM_medaka
 ENAM_tilapia

C (continued)

exon9 (continued)

ENAM_human KHSSYQPAVYPEEIPSP-A---KEHFPAGRNTW-----DHQEISPP---FKEDPGRQEEHLPHPSHGSRGSV-FYPEY
 ENAM_pig KHSAYQPV-YTEGIPSP-A---KEHFPAGRNTW-----NQOEISPP---FKEDPGRQEEHLPHLSHGSRVHV-YYPDY
 ENAM_platypus KQKPFHPV-TSERTPR-----EPSPTKPKQR-----TSPEPPPP---LEEGPGRQDKSLTYPPISPRSGI-PYPDH
 ENAM_Anolis REQPFNHI-DQKR-----GQFLSAQTPTW-----NNREASQL---FQEAPPRQSSMYSSGSFNQREMP-TYS GK
 ENAM_Xenopus KLSVFNNG-DNSGN-INTAHIKSEDLFRGGEFFQHNGENAALYQKEGTASDIDGTASDIDGTAYDIGTAYDIGTATTGQQLES IYLVKHFPTNHFQVYGDN
 ENAM_coelacanth SGQESISPNSRYNPSGQEIHPNSRYNPSGQENIPSNPNYNPSDQGRMLPHHEVNLPNKKS IQPNSGYHPSGQKNLSPRHMYPNSDHGSPNQNH EIDPL
 ENAM_bichir -----VTSSPSGNNNAVLNIPYYPSNGG-----NINPV
 ENAM_gar -----TTPAPMGKSPALNIPFSPSEGA-----KRNPS
 ENAM_zebrafish -----
 ENAM_trout -----
 ENAM_fugu -----
 ENAM_medaka -----
 ENAM_tilapia -----

exon9 (continued)

ENAM_human N-----PYDPRENSPYLRGNTWDERDDSPNTM--GQKESPL---YPI-NTP-DQKEIVPYNEEDPVDPTGDEVFPQNRWG-EELSFKG-----
 ENAM_pig N-----PYDPRENSPYLRNTWYERDDSPNTM--GQPENPH---YPM-NTP-DPKETI PYNEEDPIDPTGDEHFPQSRWDMEELS FKE-----
 ENAM_platypus H-----PYDPK-----GTWDEGNEPTPSS--GPAEQFGRQSPCSPRM--GQRGEVYNEEDPVDPTGDEPNLGGPWE-ETANGKE-----
 ENAM_Anolis KSYNQYTHPPFP-----RAWEDREHSPTISPSDQRESSP---YPP-VSPSDQIQRNYY--RRIQPE-YEPYSRQDPWT-REQHLHD-----
 ENAM_Xenopus N-----PKNPS-----HLLSNTWQKKPDQ-----LESFR--N---NLL-GQKEIYLYPDTELYHQTNNELPLGKHWD-NEIDFSAVQFENPGS
 ENAM_coelacanth GQRHQSSPVNTLVQEDMSHYPRSNPLRQGGNSPNHETFSHGQKYSHYDRSNHVRGKGNSPCHGSHSPSQVIHHKQPKGNSQIH DENPWGQEGQMS
 ENAM_bichir GMRKTP-----VKNGC-----
 ENAM_gar GIRKPP-----VQVPC-----
 ENAM_zebrafish -----
 ENAM_trout -----
 ENAM_fugu -----
 ENAM_medaka -----
 ENAM_tilapia -----

exon9 (continued)

ENAM_human -----GPTVR--HYEGEQYT---SNQPKEYLPYSLDNPSKPREDFYSEFYWPSPDENFPPSYNTASTMPPPIESRGYYVNN AAGPEESTLF
 ENAM_pig -----DPTVR--HYEGEQYT---SNQPKEYLPYSLDNPSKPREDFLYGEFYWNPEENFPPSYNTAPTSSPVESRGYYANNAV GQEESTM
 ENAM_platypus -----DPS-R--YHKSPOYPQSRATEPQDYPPYPEGGLEK-HRDFPYGDFYPRGPEDNLPYNGADQ--LPVEGRGYYTHDAV GQGGPLQF
 ENAM_Anolis -----PD-N--QYSNSPYN---PTHHQTYQKYTTENPPSEISNLPYRKINQWQEEHS PVH-TAGHL-RQMNVPYRTNSKFEQGE-RKF
 ENAM_Xenopus STYYMKDSQRQKQWPTASNNRVELDRQP---TDQPKYLLISKLD-----PAKNTDVS--TFTTNEPMAQNGYITNQEAY---LQS
 ENAM_coelacanth NAGRIPSNRGDSSPYPLFLSENRESSPYLASKPVNQGQISPNHEVYSSAQGGNRPYTGMIPTDKI I SPNHGPPSSIQGDL SQNYRGYQSGKERNQPYT
 ENAM_bichir -----NMG-----NHGPTTV-----
 ENAM_gar -----NKG P-----HGPY-----
 ENAM_zebrafish -----
 ENAM_trout -----
 ENAM_fugu -----
 ENAM_medaka -----
 ENAM_tilapia -----

exon9 (continued)

ENAM_human PSRNSWDHRIQAQGRERRPYFNRNIDWQATHLQKAPARPDPQKGNQPPYNTPAGLQKNPIWHHEG-E-NLNYGMQITRMNSPEREH-----SSFPNF
 ENAM_pig PSWSWDPRIAQGGQKEGRPYLNRNFWDQSTNLYKTP TSSPHQKENQPSNNSPAGLQKNPTWHEG-E-NLNYGMQITRLNSPERDH-----LAFPDL
 ENAM_platypus PSRDSWDWGLRSAAQERRAPYDERDSWDHAINS---PKSTAGATETLSPSHSVPAGIRGKPHHEESV-D-WSYRGGQVSDLGVP EAE-----PALMEP
 ENAM_Anolis QNVGSGSPQQRNTFQEE-AQYAERTWVPQRV-----DPSAQKETAPYYNIYSTDFRGNPTHVEDRERI IHMNAPISTENAPRRRH-----YLD RMG
 ENAM_Xenopus PNN--KSPFIFNKPSTDGILYIPIDELHDS-----QNNPVHLKLT P-----YSKAHSENHSPGVFRSLQEV
 ENAM_coelacanth EVIPRGQKSIYSAPSGQRGENSENLRINPSGENSENLFYPERKPSGKESDLPSYRVYVKGQGENSPYSQPV-PSSQRIMTSDPLL NQSTQKGTMPYFTLNS
 ENAM_bichir -----EPVAPSAGNSEIPSFGE-----GILMA-----
 ENAM_gar -----GNGE-----QPGFVDPMNPDAEG IILA-----
 ENAM_zebrafish -----
 ENAM_trout -----
 ENAM_fugu -----
 ENAM_medaka -----
 ENAM_tilapia -----

C (continued)

exon9 (continued)

ENAM_human IPPSYPSGQKEAHL-----FHLS-QRG-SCCAGSSTGPKDNPLAL-----
 ENAM_pig IPPDYPGGQKESHV-----FHLS-QRG-PCCAGGSMWPKNNPLAL-----
 ENAM_platypus VARSDPADPREASS-----YPVAVARS-SCCAGGSPGPRDVLAPPDYPLALPGYLPLDPLDYPPLDRPDYLPAPRDYPPP
 ENAM_Anolis YSDDYPKERRMIT-----SQST-ANQ-LCCADDSFMPRENRLAP-----
 ENAM_Xenopus NFNLYPSSLRRQSLNPRKTGHYSCITPLTGVDPTLEHSN-ALILVCCKLNDM-GQSHPSAN-----
 ENAM_coelacanth PGQKTNSPYISENPSDQRGTSFKNRIYPAGQRRARLPYPASSVFSQRDNHPIHGTFNPSGQEDIPPNQDTNLLSKIGFTTPTLENPSEQRQSGFCPRNNQL
 ENAM_bichir -----AAEAGSDGEGRT-----
 ENAM_gar -----AADA-----
 ENAM_zebrafish -----
 ENAM_trout -----
 ENAM_fugu -----
 ENAM_medaka -----
 ENAM_tilapia -----

exon9 (continued)

ENAM_human -----QDYTPSYGLAPGENQDTSPLYTDGSHTKQTRDII-SPTSILPGQRNSSE----KRESQNPFRDGVSTLRRNTPCSI
 ENAM_pig -----QDYTQSFGLAPGENPDTSIGYAEDSHIKYARQTV-SPTSIVPGQRNSSEKILPGESQNPSPFKDDVSTLRRSTPCSV
 ENAM_platypus DTPNYSPLDTPDYPPHYPLAPQDYSSPLGLSLWELQVGGPRSPGGSRSHVQQA-SPVGYPAIRRNGETEAPTRREEPAPSRDGLA-PGRSPPCSL
 ENAM_Anolis -----LRSAPQFRHALWEAKESST-YPDGSHANYVRHAH-SPAGIQA--NNPLKNGGREQEELGAFRVENAGSEKSLPSCSN
 ENAM_Xenopus -----EDNVHHLKVVVLENEYKESVP-HQDQTHRQHTRDVIESPKFSI--Q-NNVMTVRPNSSNNHDNIQSSRGKSSDSVPCTS
 ENAM_coelacanth CQRNSNPDFKGNSPITERKSLGWRERTHSVGATPIHGNDHPDSFSPRKSQFPADNFPYQAGNPETPRRNQFVDRENMPITRDSPLGSEGNMMPKPSLSE
 ENAM_bichir -----
 ENAM_gar -----IHGDS-----
 ENAM_zebrafish -----
 ENAM_trout -----
 ENAM_fugu -----
 ENAM_medaka -----
 ENAM_tilapia -----

exon9 (continued)

ENAM_human KNQLGQKEI-MPFPEASSLQSKNTPCLKNDLGGDGNVLEQVFEFN-QLNERTVDLTPPEQLVIGTPEDEGSNPEGIQSQVQENESERQQQRPSN--ILHL
 ENAM_pig KSQLSQRGI-MPLPEANSLQSKNTPCLTSDLGGDGNVLEQIFEEN-QLNERTVDLTPPEQLVFGTDPKPEPRPEGIPNEMQGNESERQQQRQSS--ILQL
 ENAM_platypus RPRASPRAG-LAFPEAGLPPAKPAPCFKSRRLRGDVSVEQIILDPD-PPVAGNDGPAPGDLAVVPERDPGPGVIRGDP---EPGPEAGAPG--FQWV
 ENAM_Anolis SH-LSQDSKQEANLQRLFKFRNMPCHGNSNIRGDRHNPLAHLVGTGQSFGRGTLNLFPEQ-----FPQPHGIQSEALVSEDDGKEYAALG--AKRI
 ENAM_Xenopus KPSYEQNSE-YPY-----AFPKKQCS--YRGDADSYLPPSQDESISLKK--RFLASENVYML-ANTPEPDL-PDRRI--KDDGKVKKGIKLEIL
 ENAM_coelacanth -----RVNLFDRDNPALFHSHTTANNFEEGISTTSPKEMPPPGVDLTRHATKSWIPAAAGDMLPAPIKKA
 ENAM_bichir -----
 ENAM_gar -----RV-----
 ENAM_zebrafish -----
 ENAM_trout -----
 ENAM_fugu -----
 ENAM_medaka -----
 ENAM_tilapia -----

exon9 (continued)

ENAM_human PCFGSKLAKHHSSTT-----GTPS-----SDGRQSP-----FDGDSI--TPT-----NPNTLVELA--TEEQFKS-----INVDP-
 ENAM_pig PCFGSKLANYHTSSI-----GTPS-----SLGRQDS-----FDGDI--MPT-----TPNSLAGLA--TGAQFQN-----INVDP-
 ENAM_platypus PCFGSETRDSK-SLL-----GMDGRSPWQREAGPIRAGVHRTRRDPGRGLWASDRGAFRGPQP--SPFIRFTALQDPPVGSRR--SGTTGSR-----LLVAP-
 ENAM_Anolis PCFGSWLKQYL-SNT-----EVPS-----DDQQRDP-----FYGENP--VPT-----RPNSIPPKPEPISSSFLS-----NGVEE-
 ENAM_Xenopus TCPQDQEIHTNKKATNIATRPTA-----SLLNHDK-----LSADAIP-----VLNTEQ
 ENAM_coelacanth PCFGDRTSPTS RDKIMSFGGSATLAGVRKAPPNGGDSMVLAVKPNLVPPASKTTVATDPKNDMALTSTLNKFPGTEKDALSAPPIAVNEDPRENRPDCVLLRD-
 ENAM_bichir PCRVHDTATAGKESNFYY-----
 ENAM_gar PCVVHDSPAQAEENHYYY-----
 ENAM_zebrafish -----
 ENAM_trout -----
 ENAM_fugu -----
 ENAM_medaka -----
 ENAM_tilapia -----

Figure S1. Multiply aligned amino acid sequences encoded by matrix SCPP genes, Related to Figure 1.

(A) Multiply aligned amino acid sequences encoded by *AMEL* and *SCPP5*, their chromosomal locations, and expression of bichir *scpp5* in the jaw and skin. Aromatic residues, negatively charged residues, and positively charged residues are shown in sky blue, blue, and brown, respectively, while Pro, Gln, and potentially phosphorylated Ser residues (potential pSer residues: Ser residues in Ser-Xaa-Glu or Ser-Xaa-pSer sequences, where Xaa represents any amino acids, are known to be phosphorylated in SCPPs.) (Kawasaki and Amemiya, 2014) are highlighted in yellow, blue, and green, respectively. Uninterrupted Pro-Xaa-Yaa (Yaa also represents any amino acid; PXY) repeats are shown with dashed underlines. The three conserved modules, the aromatic residue-rich region (YY), the Pro/Gln-rich region containing uninterrupted PXY repeats (P/Q), and the hydrophilic C-terminus (DE) are shown at the bottom. Unaligned sequence gaps are shown by dashes. Amino acid sequences were retrieved from GenBank (accession numbers for *AMEL*: NP_873632.1 for humans, NP_998965.1 for pig, reconstructed from GCA_00002275 for platypus, XP_003228746.1 for *Anolis*, AAC78133.1 for caiman, AAZ23149.1 for axolotl, NP_001107153.1 for *Xenopus*, AYU58914.1 for lungfish, and reconstructed from GCA_000225785.1 for coelacanth; accession numbers for *SCPP5*: AYU58917.1 for bichir, XP_015200896.1 for gar, XP_035273951.1 for eel, NP_001138708.1 for zebrafish, reconstructed from GCA_000180675.1 for stickleback, NP_001032946.1 for fugu, reconstructed from GCA_000188235.2 for tilapia, and reconstructed from GCA_002234675.1 for medaka). Note that all teleost species used for multiple amino acid sequence alignments (A, B, and C) belong to different orders. In sarcopterygians, *AMEL* is located within intron 1 of *ARHGAP6*, while in actinopterygians *SCPP5* is located within a P/Q-rich SCPP gene cluster (Kawasaki et al., 2017). In *L. chalumnae*, locations of genes are shown as contig names (JHxxxxxx). Expression of bichir *scpp5* was detected in the jaw and skin that secretes the ganoin matrix on scales, but not in the skeletal muscle (negative control). Bichir *gapdh* (glyceraldehyde 3-phosphate dehydrogenase) was used as the positive control.

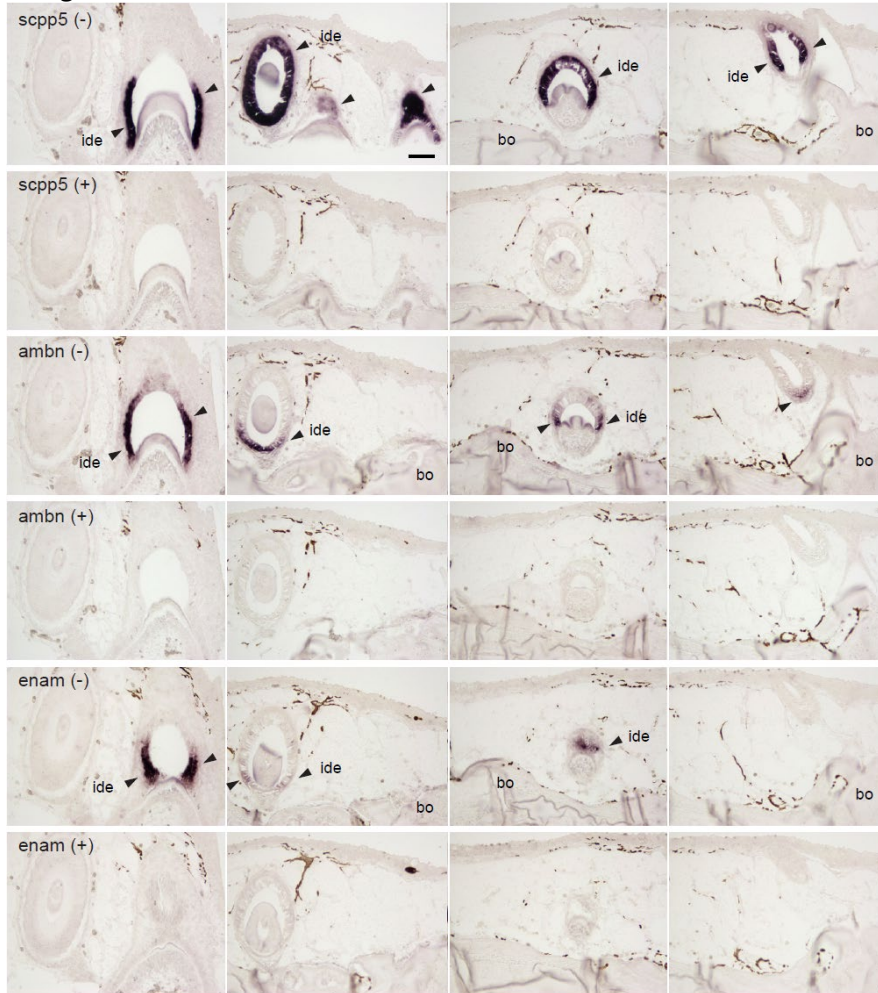
(B) Multiply aligned amino acid sequences encoded by *AMBN*. Cys residues is shown in red. See the legend above. *AMBN* in sturgeon, medaka, and tilapia has a small last exon, and the C-terminus is shown by a period. The amino acid sequences shown under exon 6 of human *AMBN* are encoded by two different exons in bichir, sturgeon, and gar (separated by an underscore). In a previous study, *ambn* was named *scpp6* in teleosts (Kawasaki, 2009). However, conservation of the amino acid sequences encoded by exons 11 and 12 (corresponds to exon 12 of human *AMBN*) between *AMBN* and *SCPP6*, the absence of the entirely untranslated last exon in both *AMBN* and *SCPP6*, the presence and locations of four potential pSer residues, and an equivalent chromosomal location of gar *ambn* and zebrafish *scpp6* (between *scpp3* genes and *scpp7*) implies the orthology of *AMBN* and *SCPP6* (Qu et al., 2015; Braasch et al., 2016; Kawasaki et al., 2017). In the present study, the orthology of *AMBN* and *SCPP6* was confirmed by a similar expression pattern during tooth development. Amino acid sequences were retrieved from GenBank (AAG35772.1 for humans, XP_007667389.2 for platypus,

AAK92227.1 for caiman, XP_016846623.1 for *Anolis*, XP_002938667.2 for *Xenopus*, AYU58913.1 for lungfish, XP_006011890.1 for coelacanth, AYU58915.1 for bichir, reconstructed from SRX424534 for sturgeon, AMD08894.1 for gar, NP_001138709.1 for zebrafish, reconstructed from GCA_002234675.1 for medaka, reconstructed from GCA_000188235.2 for tilapia, and NP_001032945.1 for fugu).

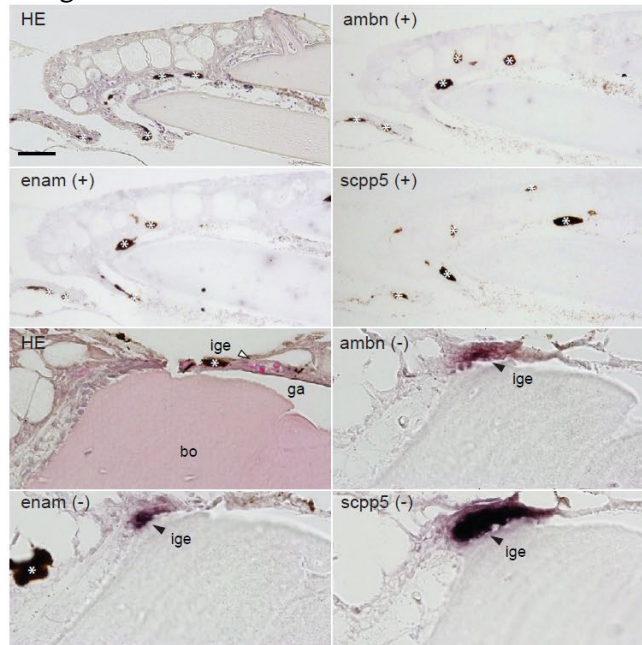
(C) Multiply aligned amino acid sequences encoded by exons 3, 4, 5, 6, 7, 8, and a 5' half of exon 9 of *ENAM*. The amino acid sequence of porcine 32-kDa enamelin (Tanabe et al., 1990; Yamakoshi et al., 1998) was underlined. Small amino acids (Ala and Gly) in the 32-kDa enamelin and corresponding regions are highlighted in grey. In the region corresponding to exon 6 of human *ENAM*, many small duplicate exons were identified in gar and bichir *enam* genes but are not shown in the figure. Amino acid sequence similarities in the portion located C-terminal to the 32-kDa enamelin (not highlighted or not shown in color) are low or undetectable between phylogenetically distant vertebrates (e.g., tetrapods and teleosts). In teleosts, *enam* was originally referred to as *fa93e10* (Goldsmith et al., 2003). However, sequence similarities in the 32-kDa enamelin and its N-terminal region encoded by *ENAM* and *FA93E10*, the absence of the entirely untranslated last exon in both *ENAM* and *FA93E10*, the presence and locations of four pSer residues, and an equivalent chromosomal location of far *enam* and zebrafish *enam* (immediately downstream of *scpp5*) imply the orthology of *ENAM* and *FA93E10* (Qu et al., 2015; Braasch et al., 2016; Kawasaki et al., 2017). In the present study, the orthology of *ENAM* and *FA93E10* was confirmed by a similar expression pattern during tooth development. Amino acid sequences were retrieved from GenBank (AAG43242.1 for humans, NP_999406.1 for pig, reconstructed from GCA_000002275.2 for platypus, ADJ67842.1 for *Anolis*, NP_001139215.1 for *Xenopus*, reconstructed from GCA_000225785.1 for coelacanth, AYU58916.1 for bichir, AMD08897.1 for gar, NP_001139028.1 for zebrafish, CDQ64219.1 for trout, reconstructed from GCA_000180735.1 for fugu, reconstructed from GCA_002234675.1 for medaka, and XP_003454248.1 for tilapia).

A

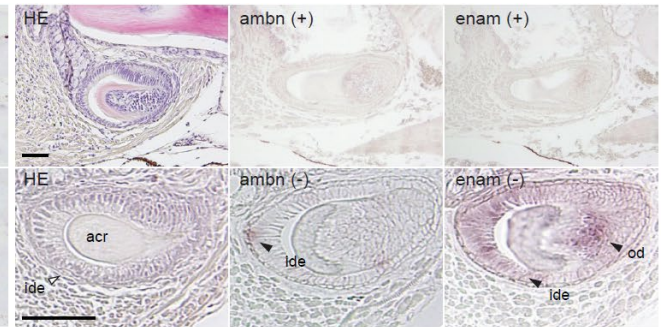
ISH: gar teeth



ISH: gar scales

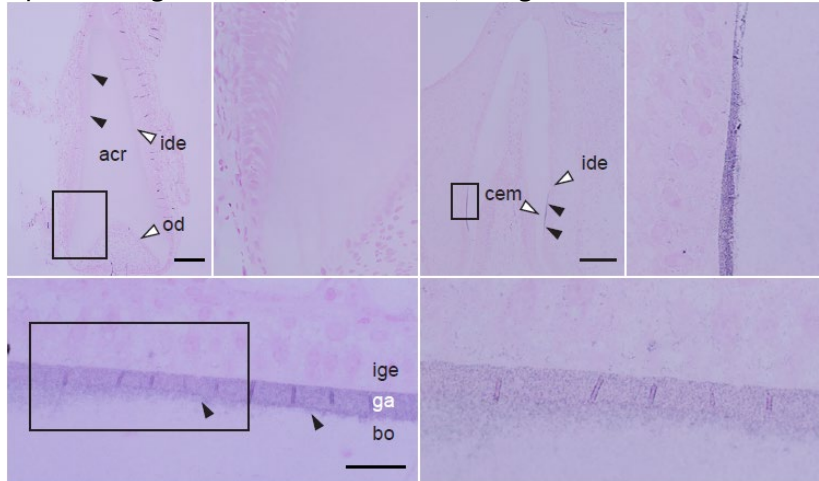


ISH: zebrafish teeth



B

Optical IHC: gar acroдин, collar enamel, and ganoin



C

TEM IHC: gar acroдин, collar enamel, and ganoin

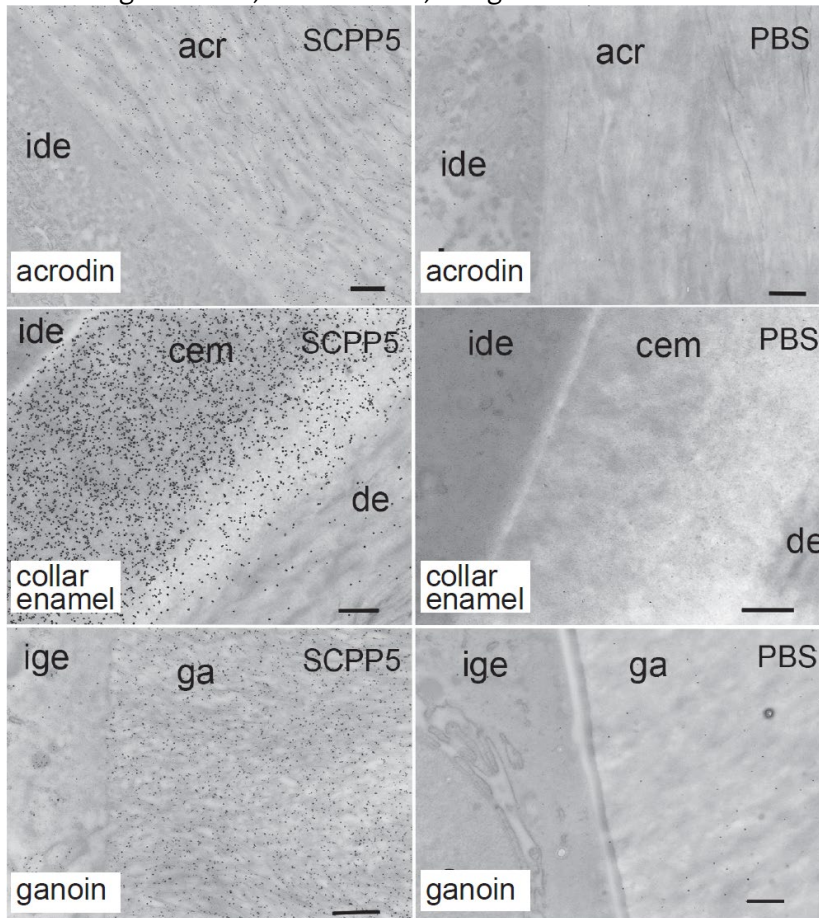


Figure S2. Expression of gar and zebrafish *scpp5*, *ambn*, and *enam* genes in teeth and/or scales, and distribution of gar *Scpp5* in teeth and scales, Related to Figures 2-4.

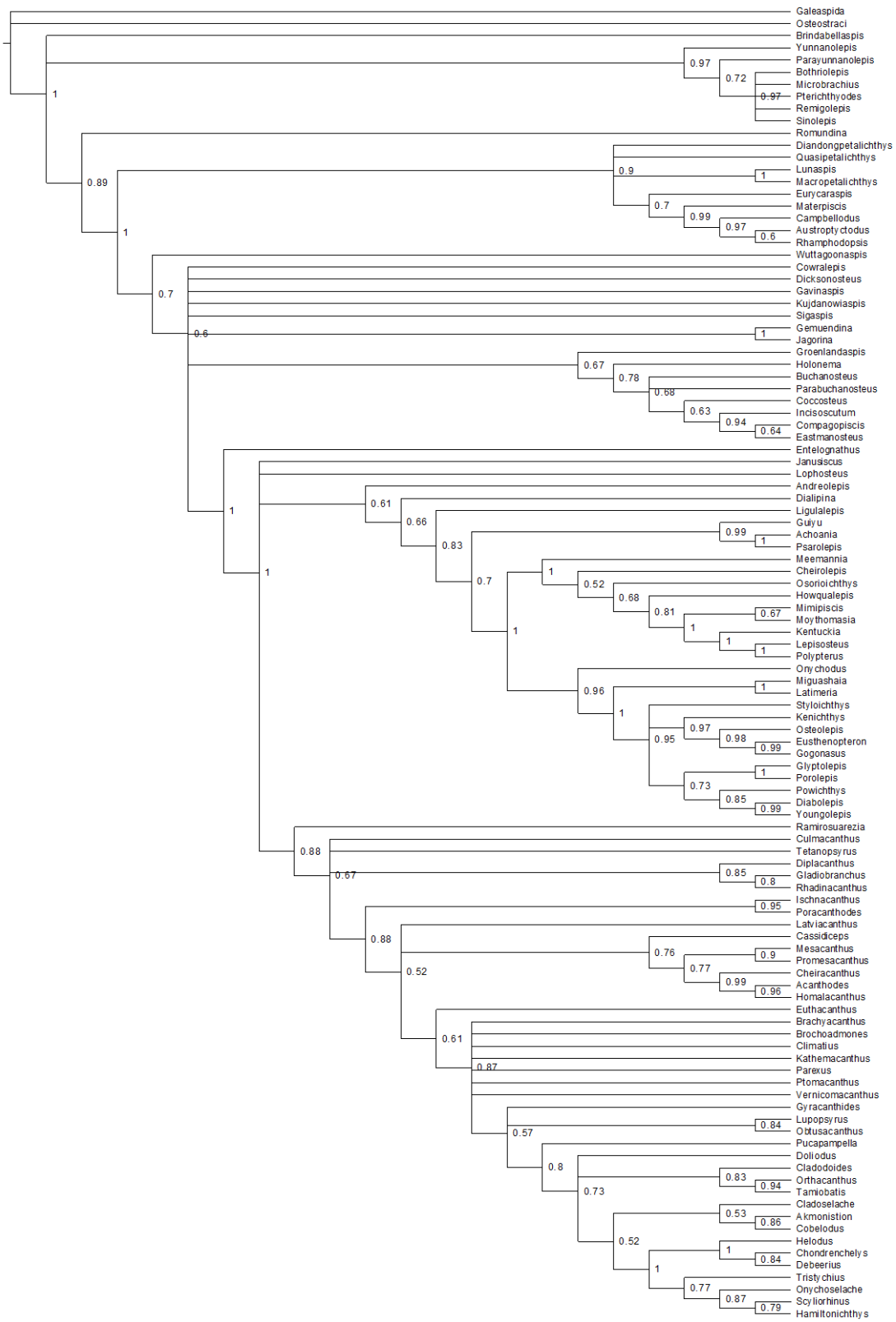
(A) ISH analysis of gar teeth, gar scales, and zebrafish teeth. Expression of *scpp5*, *ambn*, and *enam* genes were detected using antisense (-) probes, but not using sense (+) probes, which serve as negative controls. Images of HE staining are shown for gar scales and zebrafish teeth. All our ISH results are summarized in Figure 2P.

Abbreviations: acr, acrodin; bo, bone; ga, ganoin; ide, inner dental epithelial cells; ige, inner ganoin epithelial cells; od, odontoblasts. Scale bar, 50 μm .

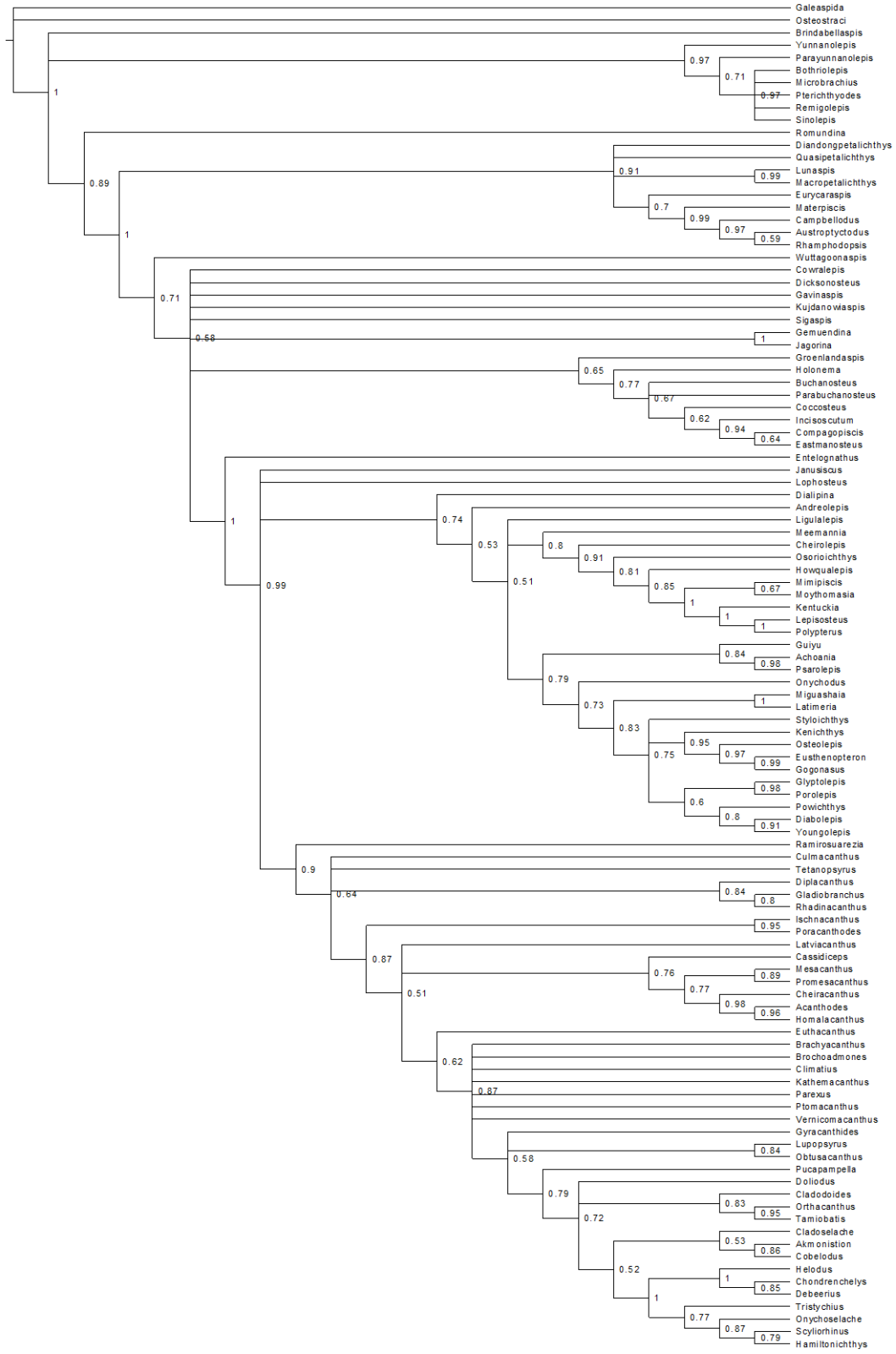
(B) Original images for optical IHC analysis (Figures 3A-3F) without enhancing image contrast. See the legend of Figure 3. Scale bar, 100 μm (upper row) or 20 μm (lower row).

(C) TEM IHC analysis of gar *Scpp5* in acrodin, collar enamel, and ganoin. PBS was used as negative controls.

Abbreviations: acr, acrodin, cem, collar enamel; de, dentine; ga, ganoin; ide, inner dental epithelial cells; ige, inner ganoin epithelial cells. Scale bar, 500 nm.

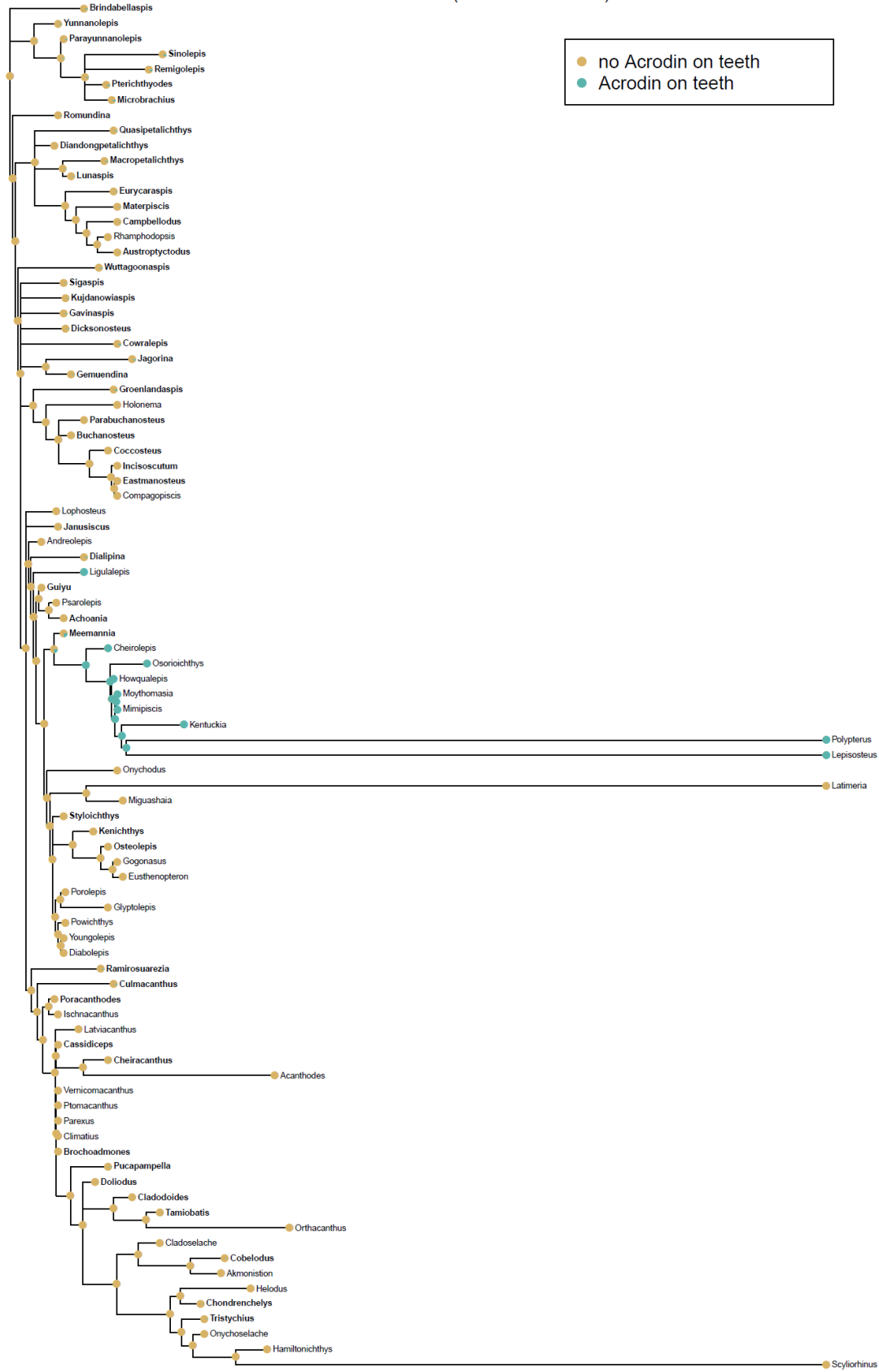
A

B



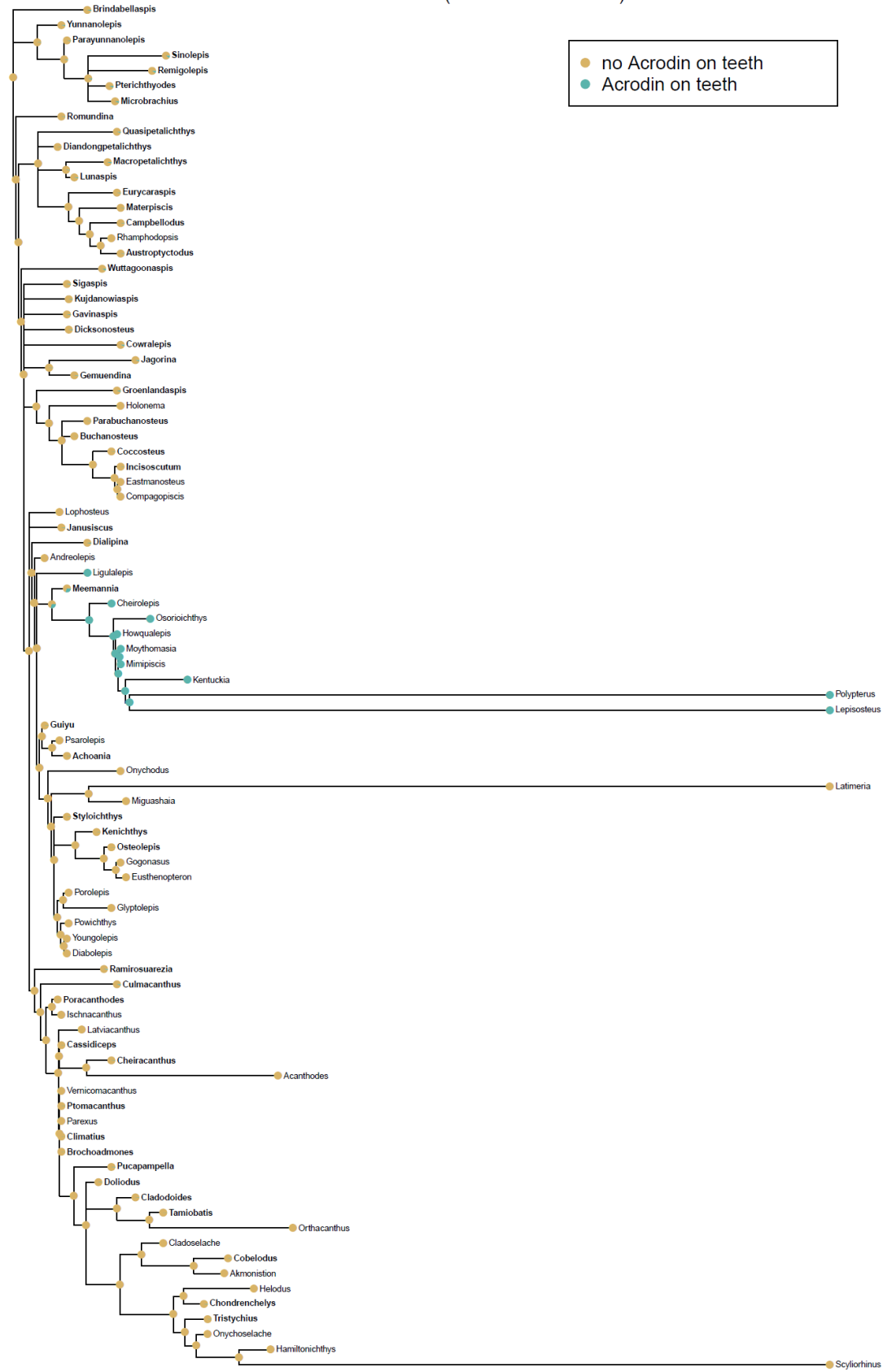
C

Dental Acrodin (constrained tree)



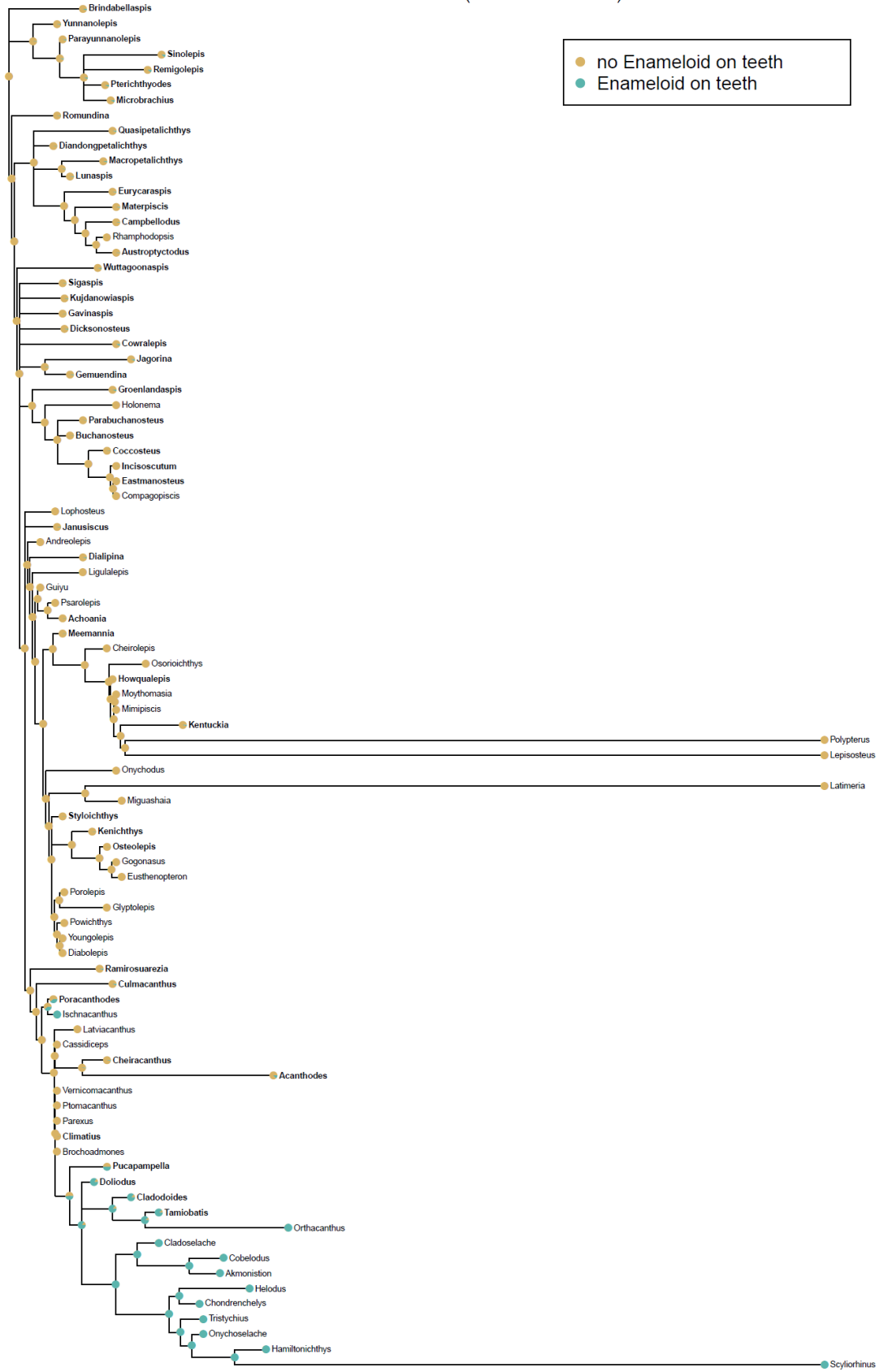
D

Dental Acrodin (unconstrained tree)



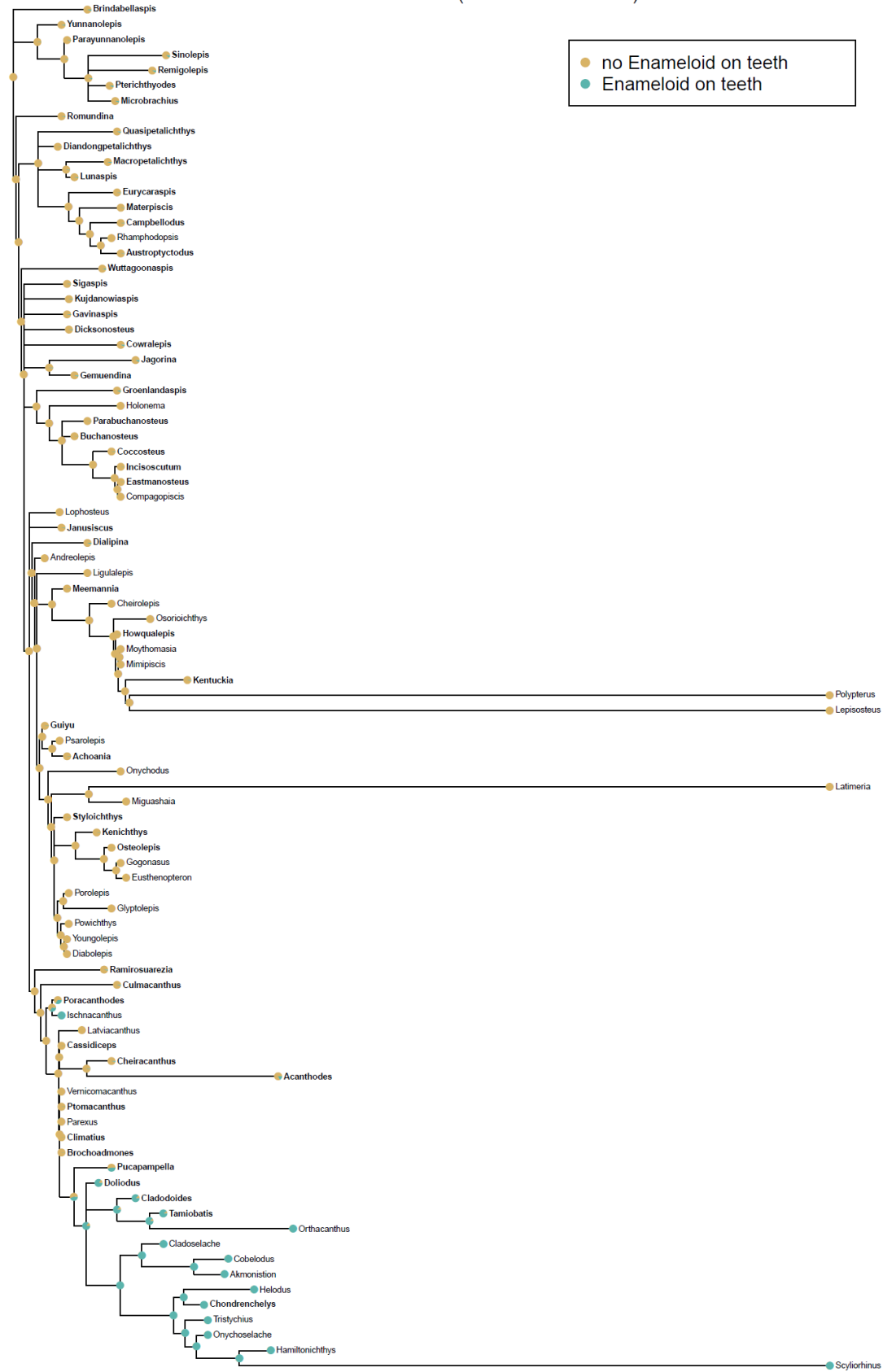
E

Dental Enameloid (constrained tree)



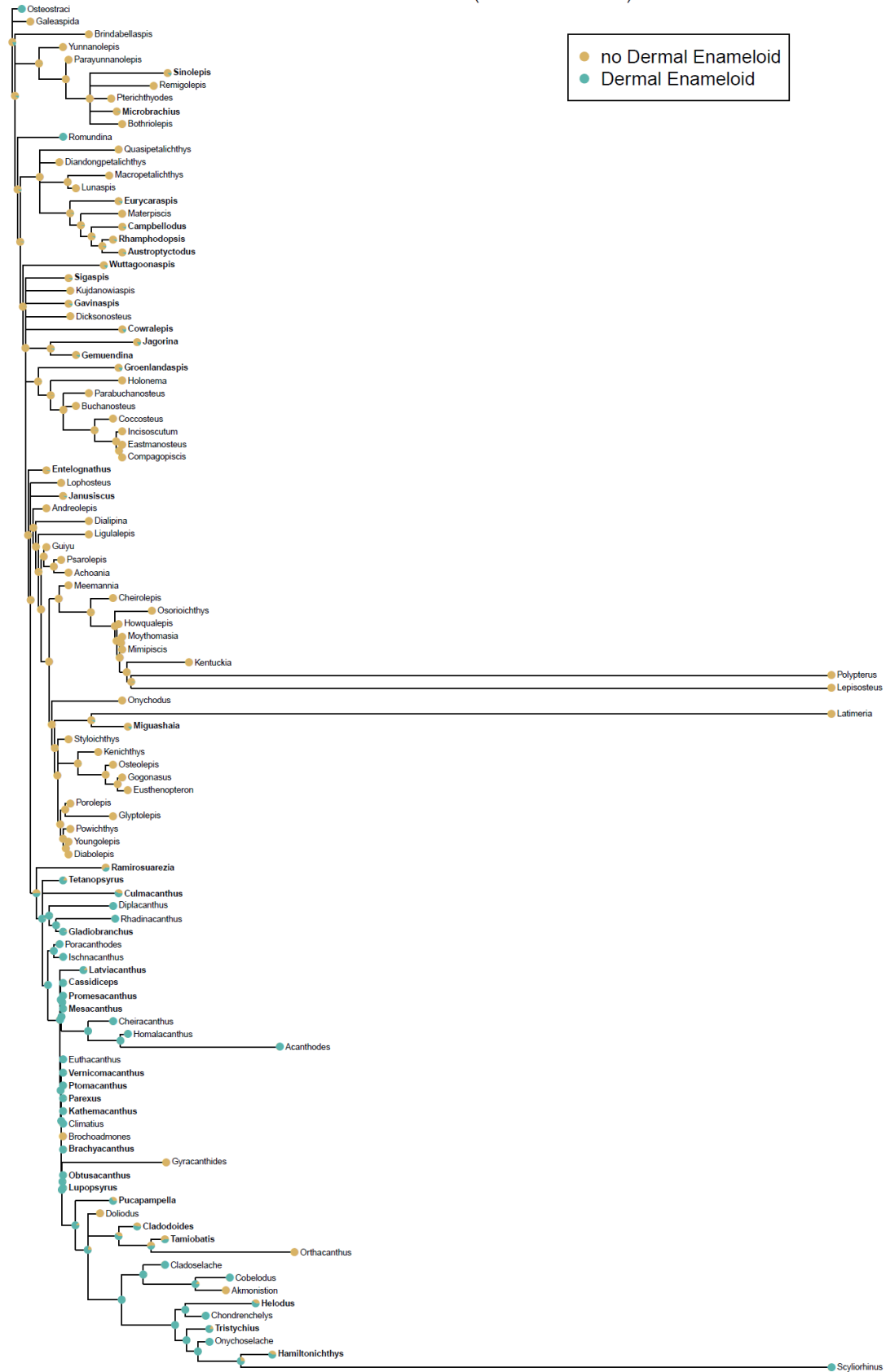
F

Dental Enameloid (unconstrained tree)



G

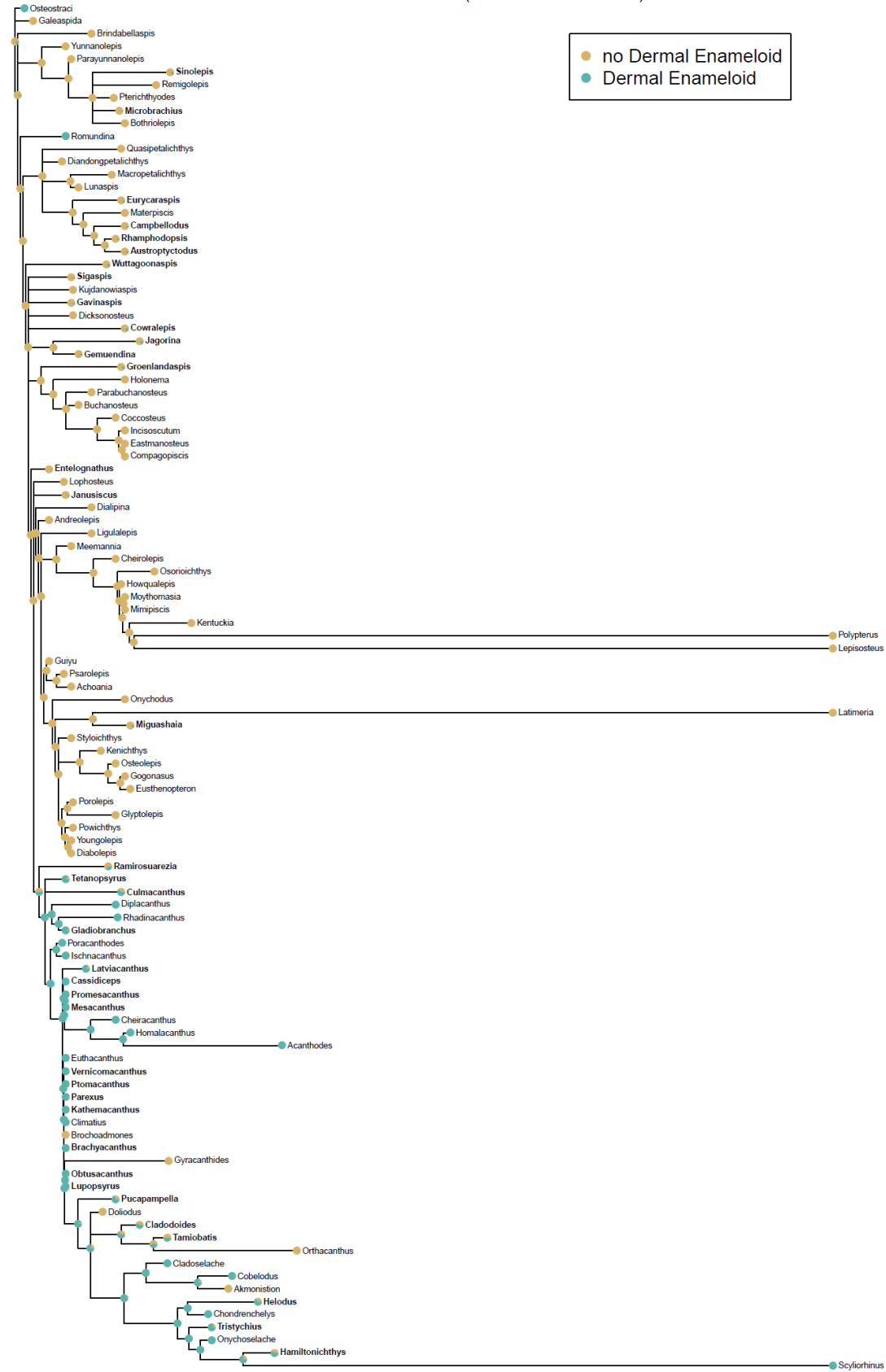
Dermal Enameloid (constrained tree)



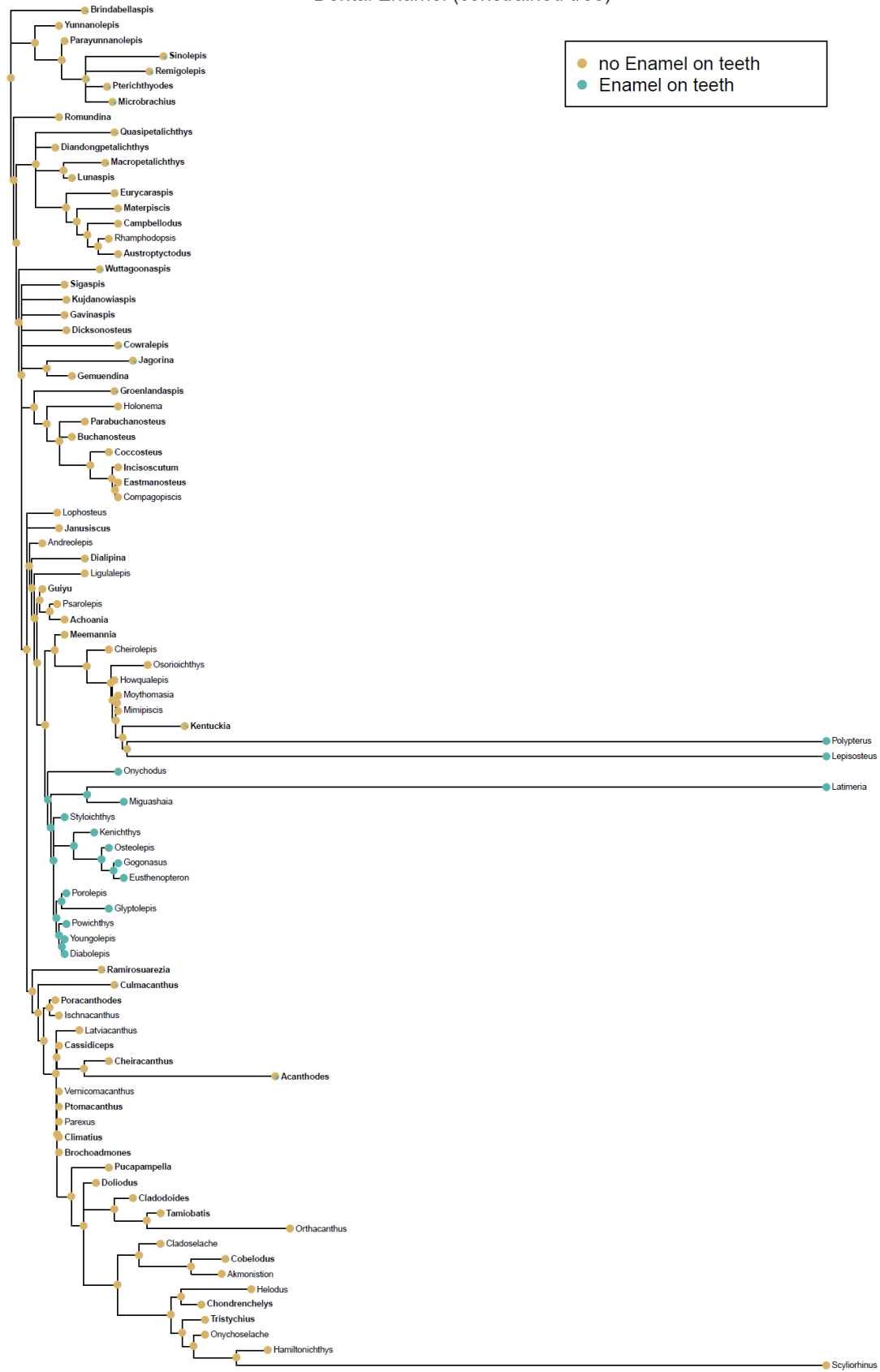
H

Dermal Enameloid (unconstrained tree)

● no Dermal Enameloid
● Dermal Enameloid

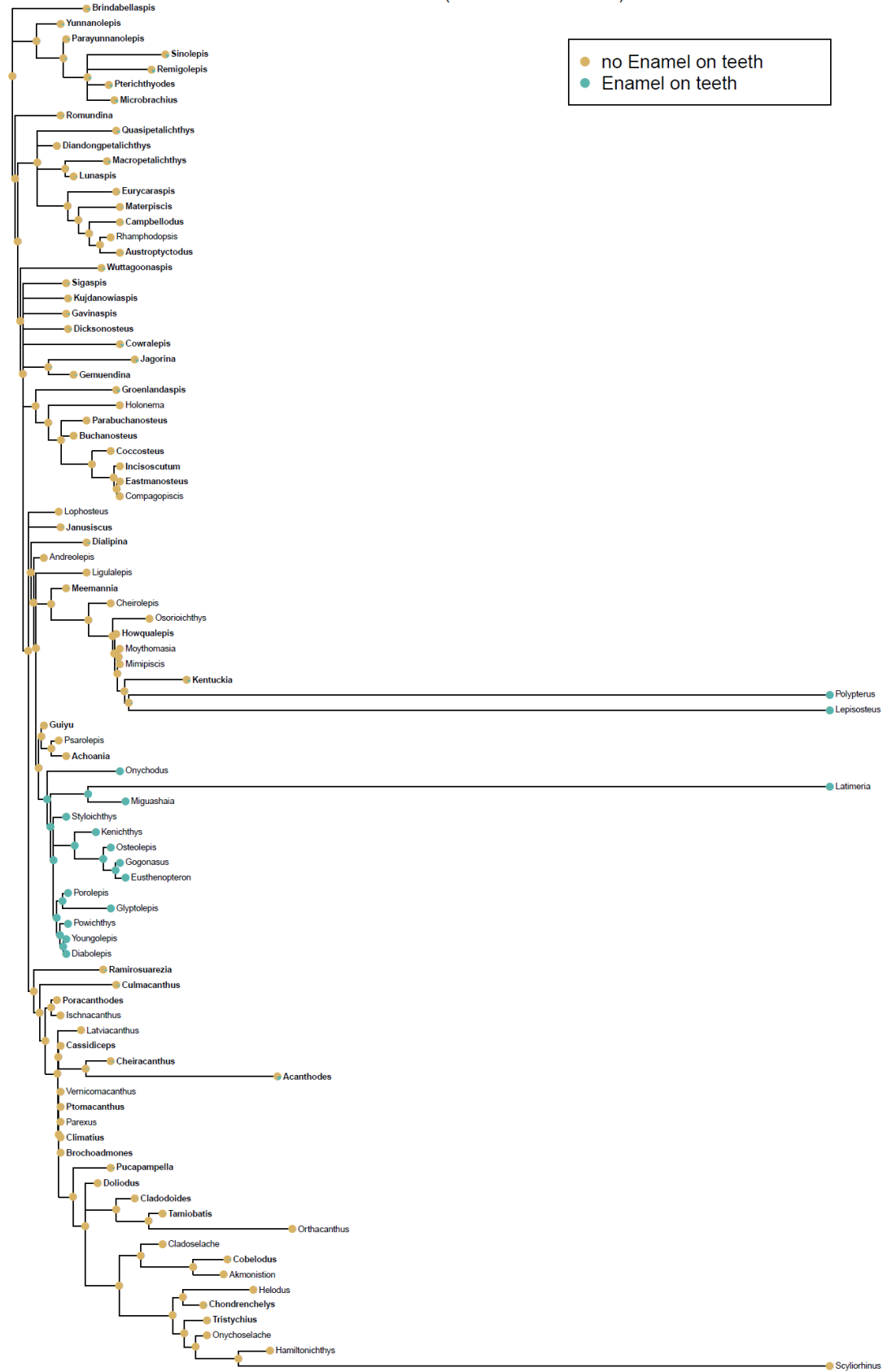


Dental Enamel (constrained tree)



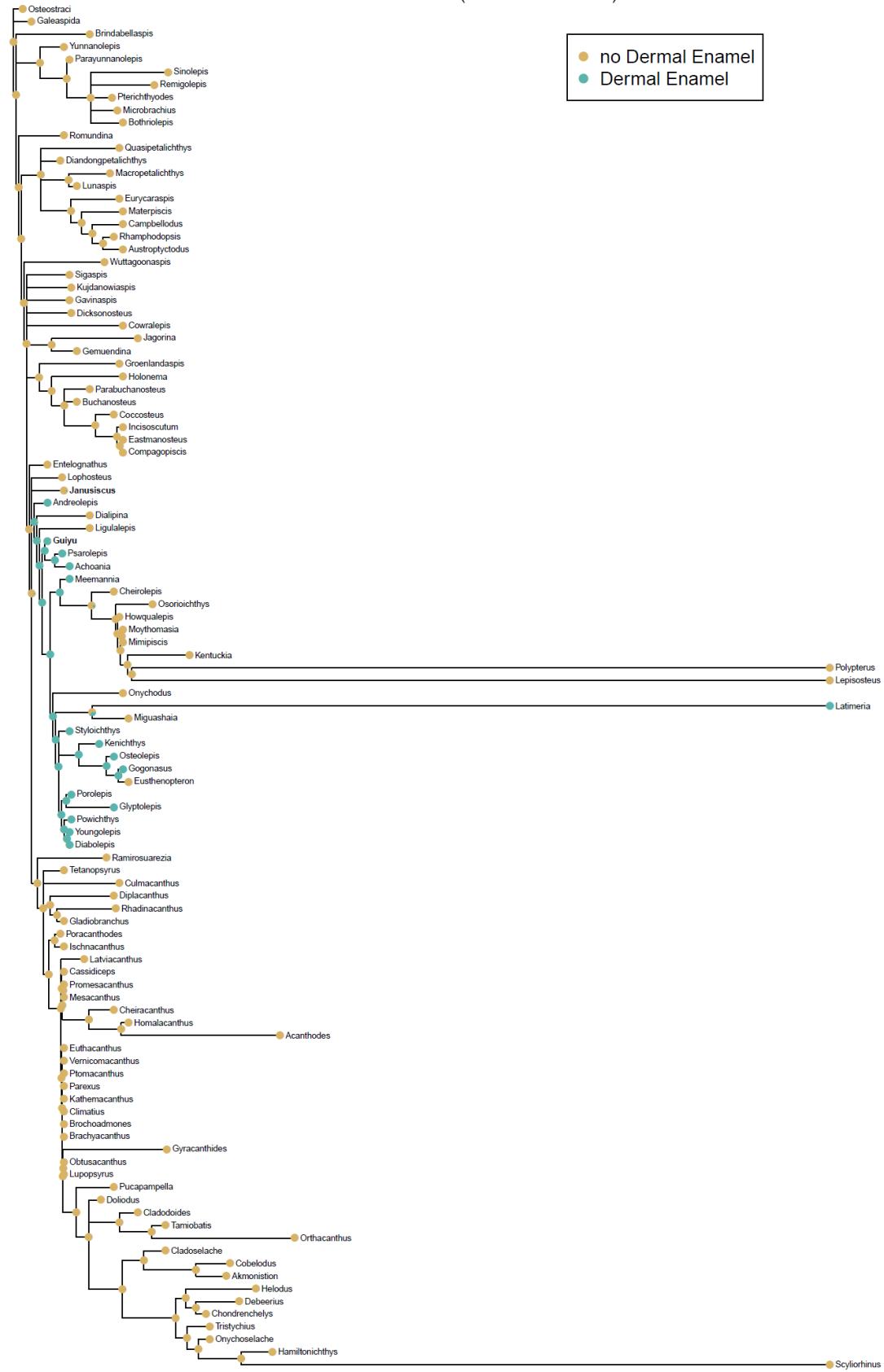
J

Dental Enamel (unconstrained tree)



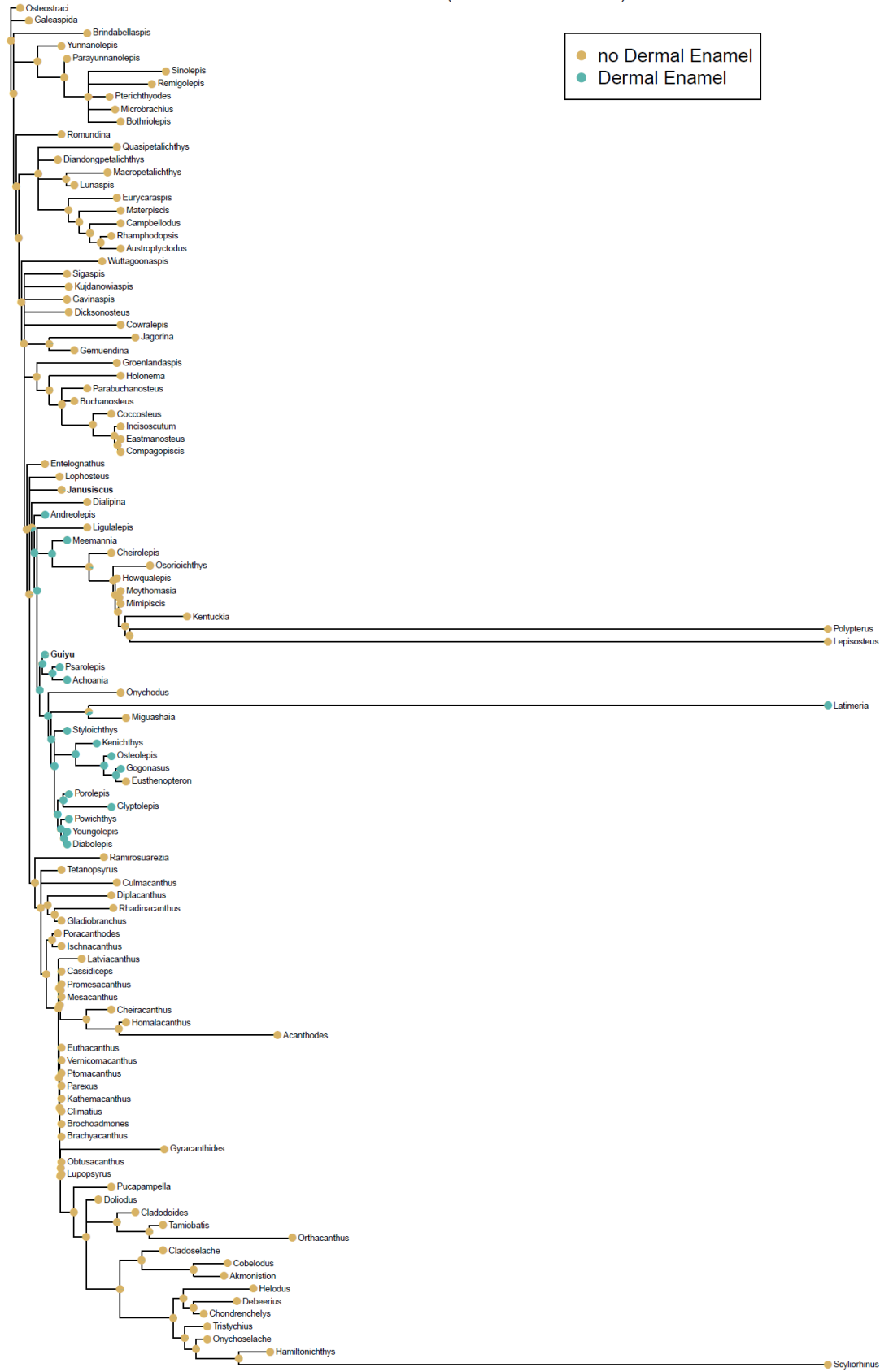
K

Dermal Enamel (constrained tree)



L

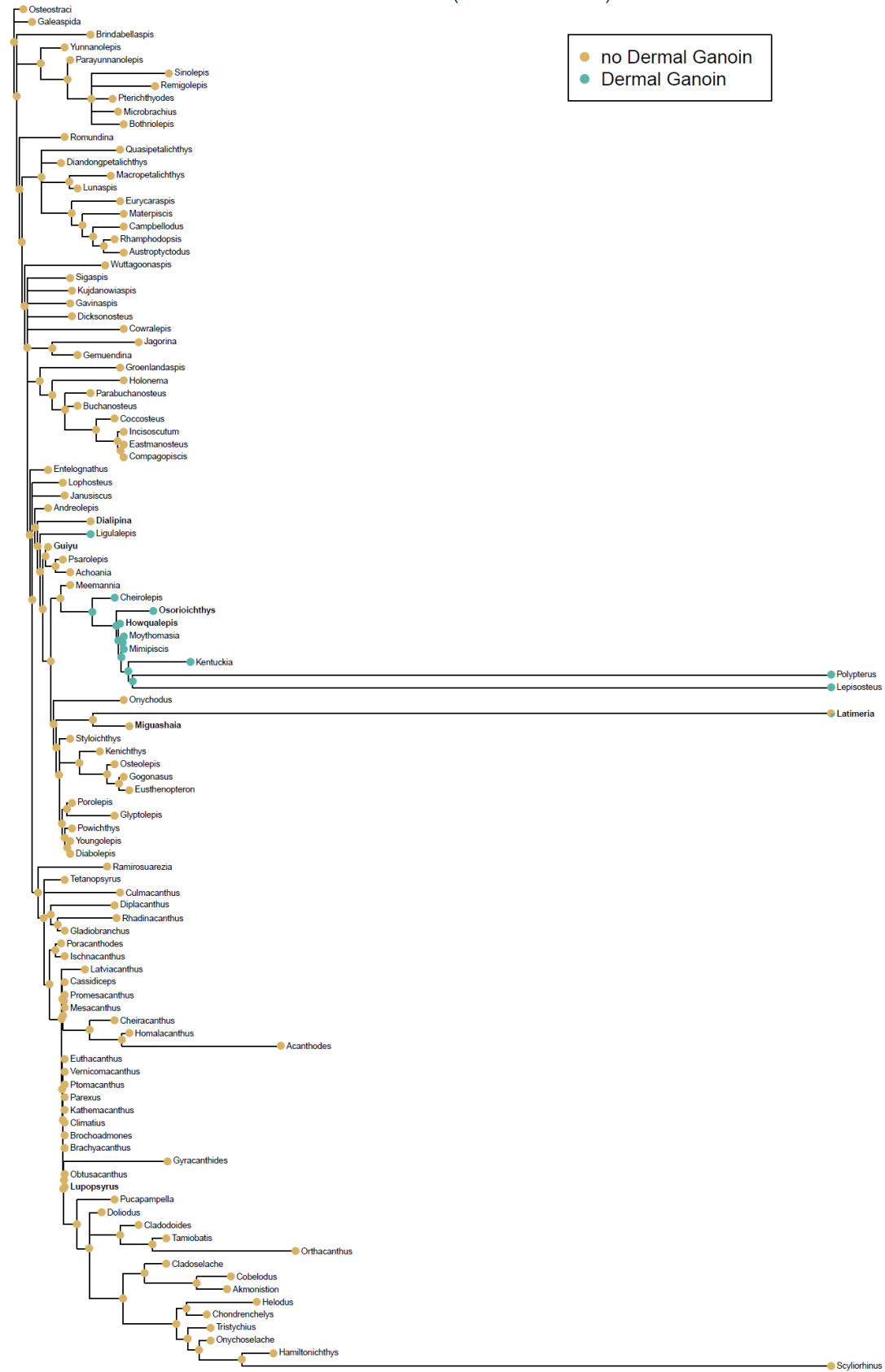
Dermal Enamel (unconstrained tree)



M

Dermal Ganoin (constrained tree)

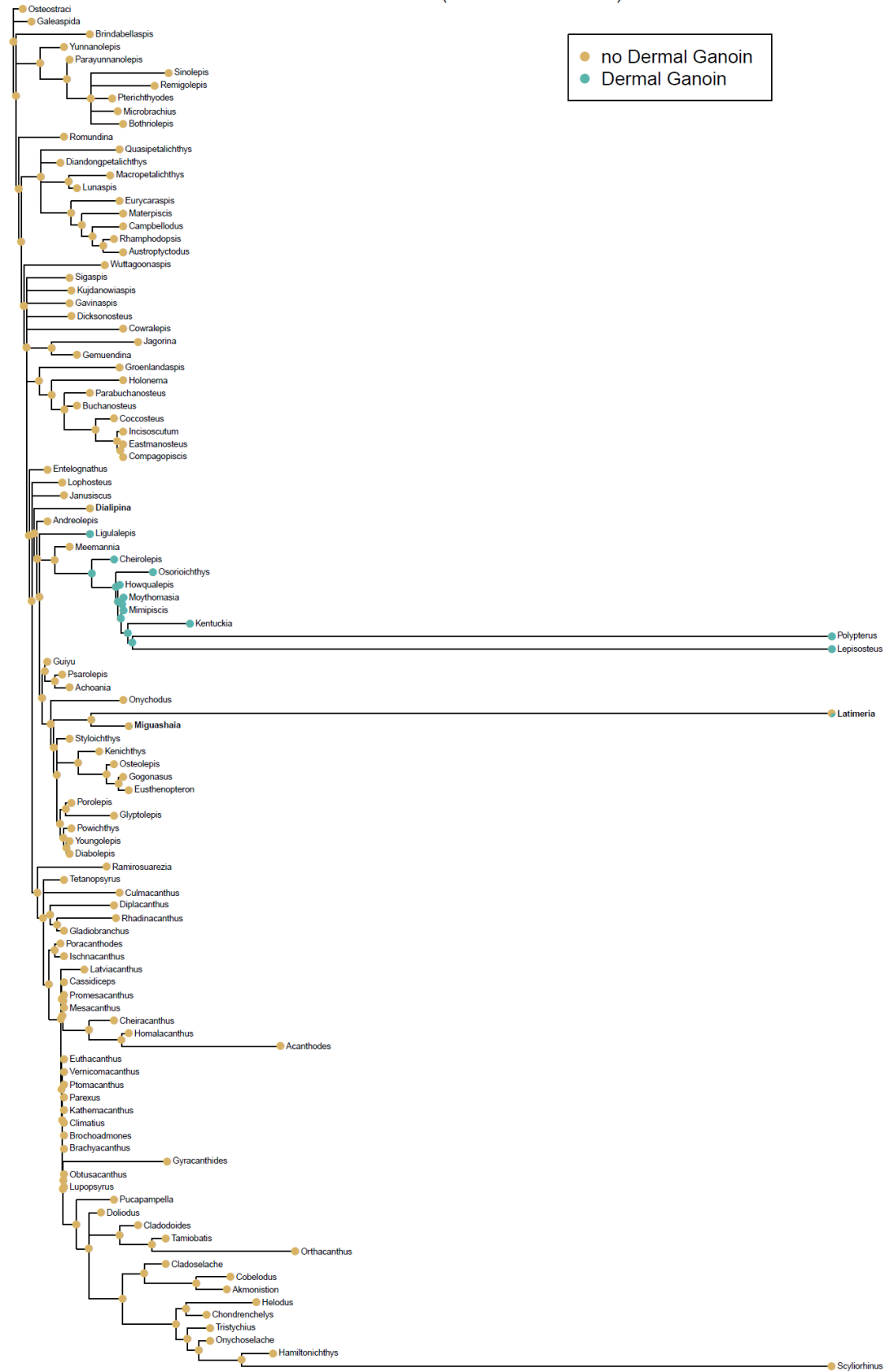
● no Dermal Ganoin
● Dermal Ganoin



N

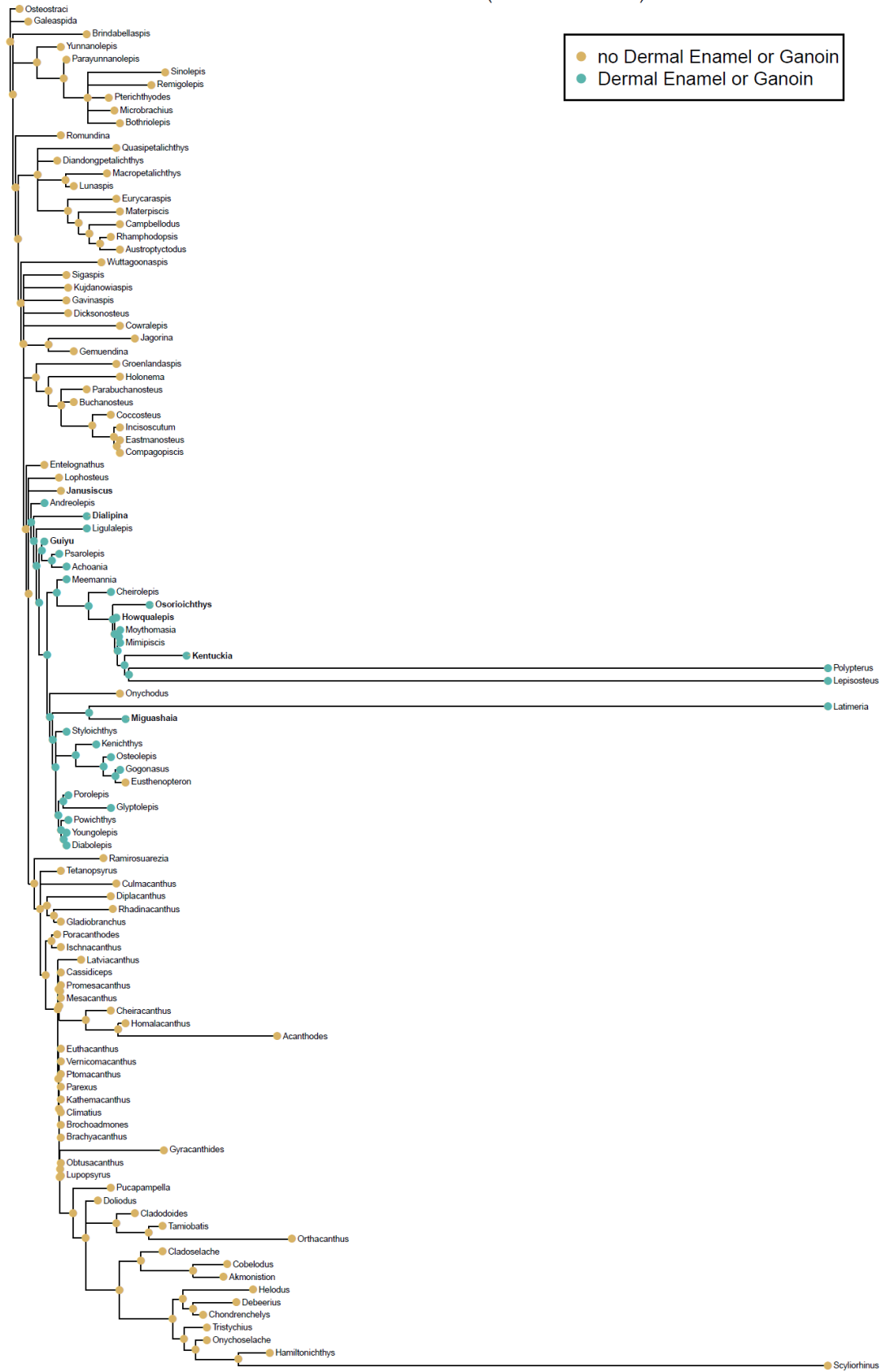
Dermal Ganoin (unconstrained tree)

● no Dermal Ganoin
● Dermal Ganoin



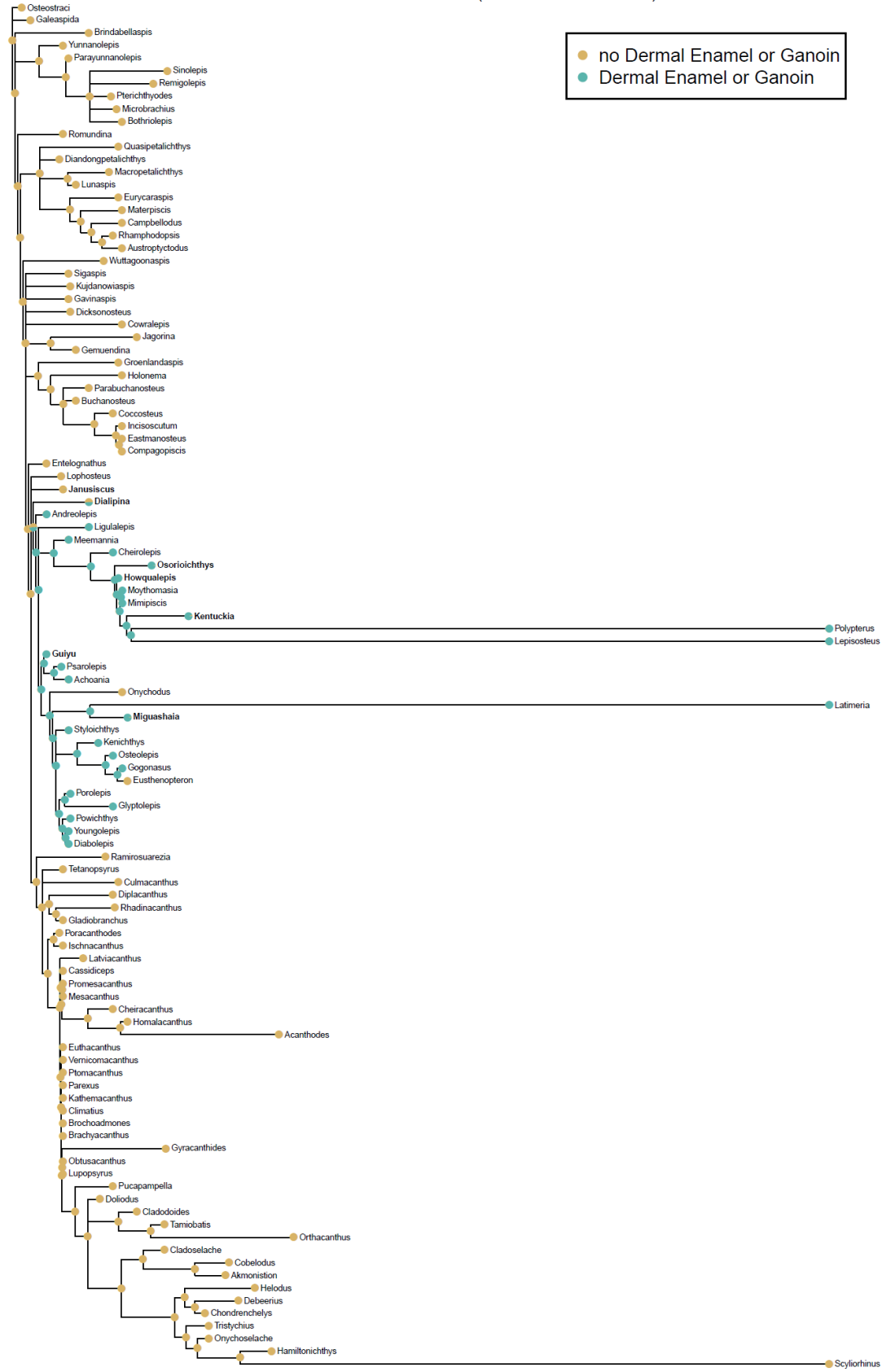
O

Dermal Enamel/Ganoin (constrained tree)



P

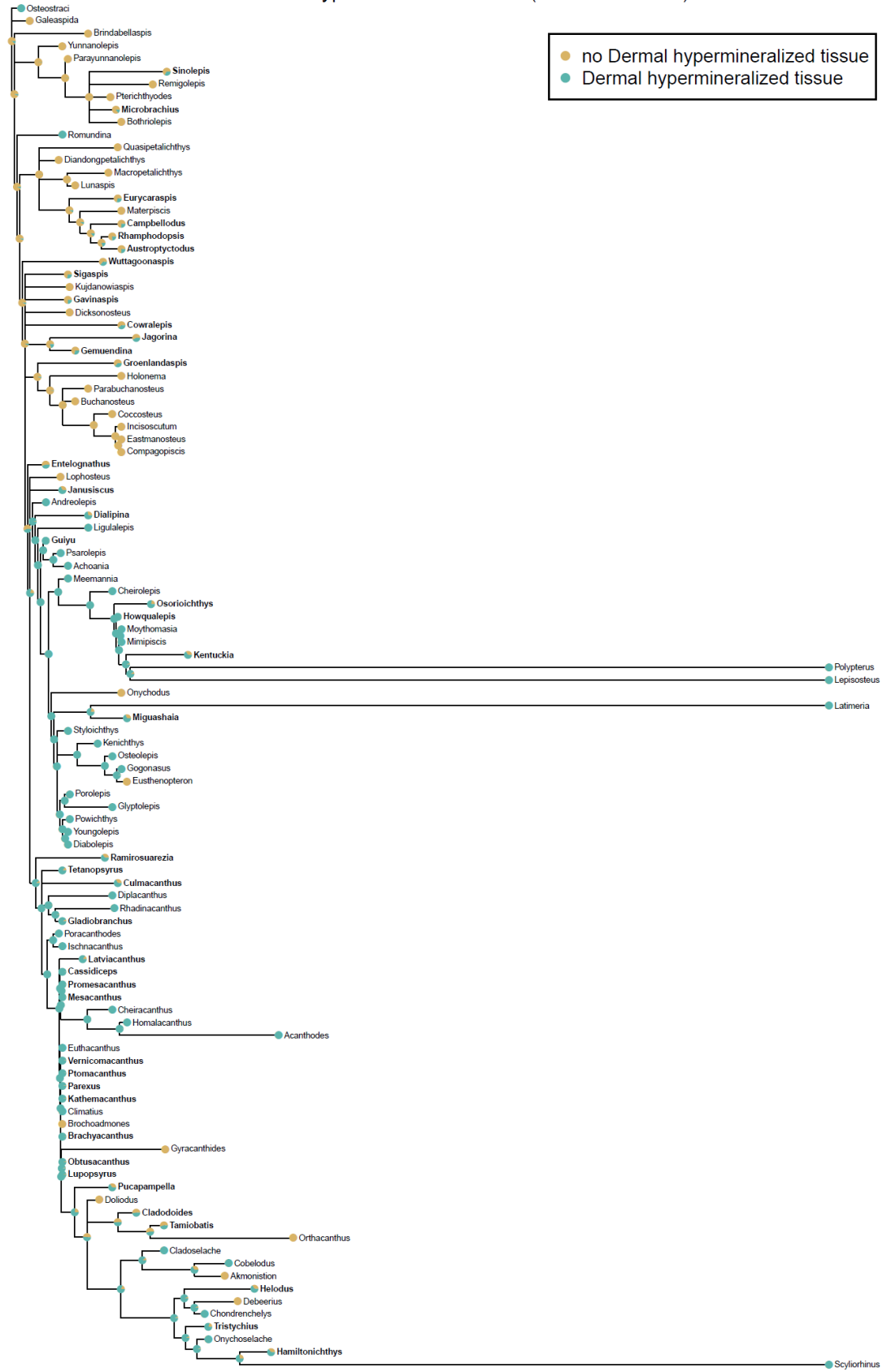
Dermal Enamel/Ganoin (unconstrained tree)



Q

Dermal hypermineralized tissue (constrained tree)

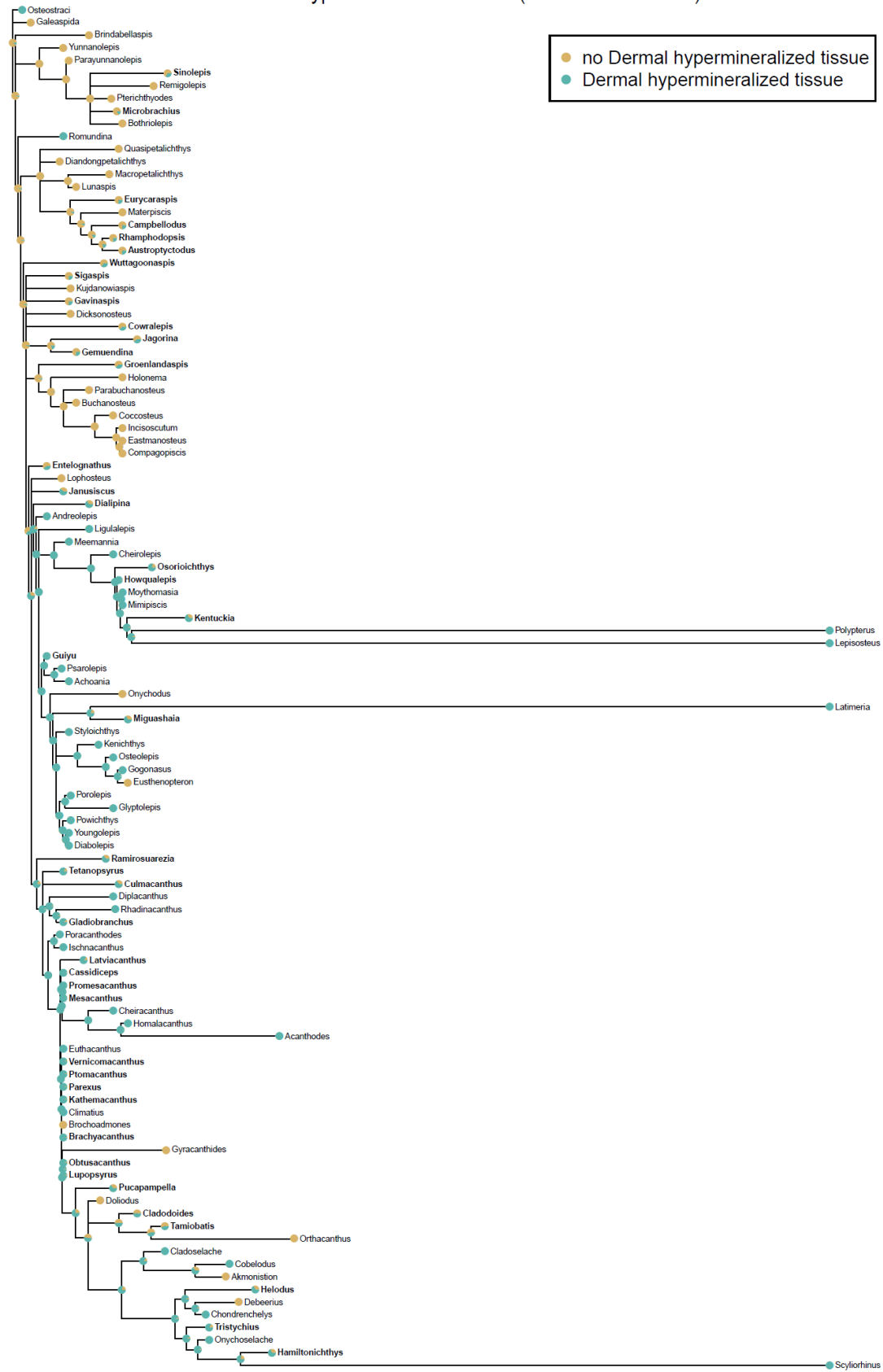
● no Dermal hypermineralized tissue
● Dermal hypermineralized tissue



R

Dermal hypermineralized tissue (unconstrained tree)

● no Dermal hypermineralized tissue
● Dermal hypermineralized tissue



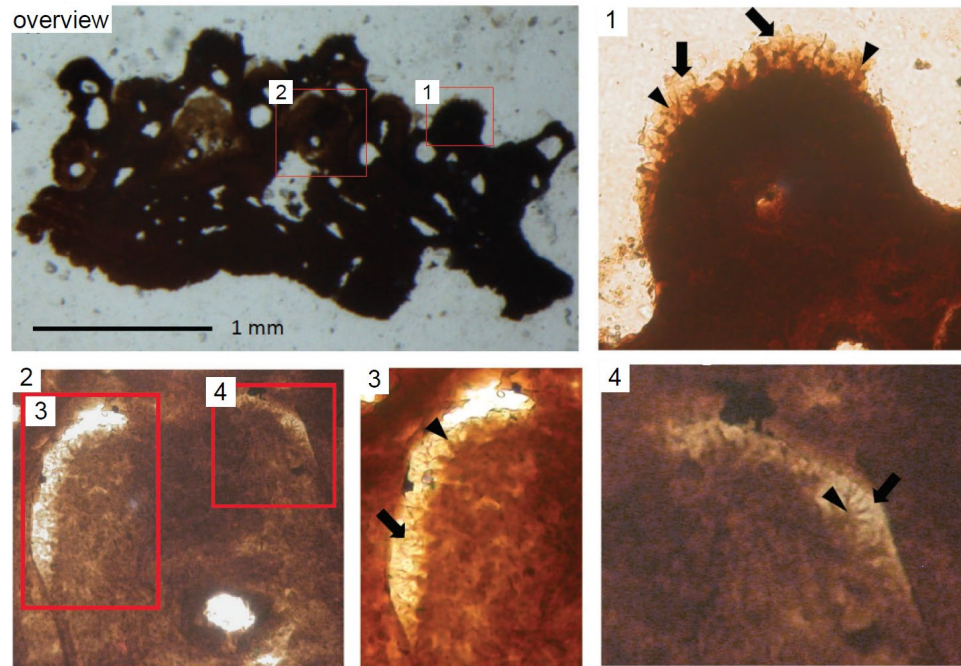
S

Figure S3. Reconstructing ancestral states of hypermineralized tissues, Related to Figure 5.

(A) Constrained tree used in ancestral state estimation. The GAP clade is constrained to a stem-osteichthyan affinity. Reliabilities of branches are shown at nodes as posterior probabilities.

(B) Unconstrained tree used in ancestral state estimation. The affinity of the GAP clade was unconstrained in the analysis and is resolved to a stem-sarcopterygian affinity. Reliabilities of branches are shown at nodes as posterior probabilities.

(C) Prediction of ancestral states of dental acrodin - GAP clade stem-osteichthyans, shown as pie charts.

(D) Prediction of ancestral states of dental acrodin - GAP clade unconstrained, shown as pie charts.

(E) Prediction of ancestral states of dental enameloid - GAP clade stem-osteichthyans, shown as pie charts.

(F) Prediction of ancestral states of dental enameloid - GAP clade unconstrained, shown as pie charts.

(G) Prediction of ancestral states of dermal enameloid - GAP clade stem-osteichthyans, shown as pie charts.

(H) Prediction of ancestral states of dermal enameloid - GAP clade unconstrained, shown as pie charts.

(I) Prediction of ancestral states of dental enamel - GAP clade stem-osteichthyans, shown as pie charts.

(J) Prediction of ancestral states of dental enamel - GAP clade unconstrained, shown as pie charts.

(K) Prediction of ancestral states of dermal enamel - GAP clade stem-osteichthyans, shown as pie charts.

(L) Prediction of ancestral states of dermal enamel - GAP clade unconstrained, shown as pie charts.

(M) Prediction of ancestral states of dermal ganoin - GAP clade stem-osteichthyans, shown as pie charts.

(N) Prediction of ancestral states of dermal ganoin - GAP clade unconstrained, shown as pie charts.

(O) Prediction of ancestral states of dermal enamel+ganoin - GAP clade stem-osteichthyans, shown as pie charts.

(P) Prediction of ancestral states of dermal enamel or ganoin - GAP clade unconstrained, shown as pie charts.

(Q) Prediction of ancestral states of dermal enamel, ganoin, or enameloid - GAP clade stem-osteichthyans, shown as pie charts.

(R) Prediction of ancestral states of dermal enamel, ganoin, or enameloid - GAP clade unconstrained, shown as pie charts.

(S) Cladoselache skin denticles [specimen P.9294 (NHMUK)]. Detail of boxed areas (1-4) in overview is enlarged. The dentin tubules (arrowheads), protruding through the enameloid layer (arrow), are clearly visible.

	lpq20	lpq8	lpq7	enam	scpp5	scpp7	lpq6	ambn	lpq5	lpq4	scpp3dl	scpp3cl	scpp3bl	scpp3al	odam	lpq3	scpp9	lpq2	lpq1
Coverage ¹	173	16	5	1761	6385	1768	3689	825	12	4	0	0	189	237	209	1119	69	336	2755
FPKM	54	6	2	548	2204	547	1118	262	4	2	0	0	101	106	66	338	21	102	1181
Conf ²	4	3	1	8	35	31	45	7	1	1	0	0	45	22	5	12	4	9	62

(continued)

	dmp1	scpp1	dspp1	ibsp	mepe1	mepe2	Spp1	lpq17	lpq16b	lpq16a	lpq15	lpq14	lpq13	lpq12	lpq19	lpq11	lpq18	lpq1	lpq9
Coverage ¹	33	237	33	632	513	991	1089	66	305	264	1010	79	126	951	386	1043	1201	3180	108
FPKM	12	72	11	211	193	313	326	20	125	101	341	23	38	317	117	331	383	1037	35
Conf ²	1	5	1	7	22	35	11	2	21	16	8	9	5	6	4	9	7	13	7

Table S1. Expression levels of SCPP genes in the skin, Related to Figure 2. Relative expression levels of SCPP genes in the skin were estimated as fragments per kilobase of transcript per million mapped reads (FPKM values) (Trapnell et al., 2010). ¹Coverage shows the estimate for the absolute depth of read coverage across the whole transcript. ²Conf represents the 95% confidence interval.

Taxa	Dermal characters					Dental characters		
	Enameloid	Ganoin	Enamel	Enamel+Ganoin	Enamel+Ganoin+Enameloid	Acrodin	Enameloid	Enamel
Galeaspida	0	0	0	0	0	-	-	-
Osteostraci	1	0	0	0	1	-	-	-
Acanthodes	1	0	0	0	1	0	0/1	0/1
Achoania	0	0	1	1	1	0/1	0/1	0/1
Akmonistion	0	0	0	0	0	0	1	0
Austroptyctodus	0/1	0	0	0	0/1	0/1	0/1	0/1
Bothriolepis	0	0	0	0	0	-	-	-
Brachyacanthus	0/1	0	0	0	0/1	-	-	-
Brindabellaspis	0	0	0	0	0	0/1	0/1	0/1
Brochoadmones	0	0	0	0	0	0/1	0/1	0/1
Buchanosteus	0	0	0	0	0	0/1	0/1	0/1
Campbellodus	0/1	0	0	0	0/1	0/1	0/1	0/1
Cassidiceps	0/1	0	0	0	0/1	0/1	0/1	0/1
Cheiracanthus	1	0	0	0	1	0/1	0/1	0/1
Cheirolepis	0	1	0	1	1	1	0	0
Chondrenchelys	1	0	0	0	1	0/1	0/1	0/1
Cladodoides	0/1	0	0	0	0/1	0/1	0/1	0/1
Cladoselache	1	0	0	0	1	0	1	0
Climatius	1	0	0	0	1	0/1	0/1	0/1
Cobelodus	1	0	0	0	1	0/1	0/1	0/1
Cocosteus	0	0	0	0	0	0/1	0/1	0/1
Compagopiscis	0	0	0	0	0	0	0	0
Cowralepis	0/1	0	0	0	0/1	0/1	0/1	0/1
Culmacanthus	0/1	0	0	0	0/1	0/1	0/1	0/1
Debeerius	-	-	0	0	0	-	-	-
Diabolepis	0	0	1	1	1	0	0	1
Dialipina	0	0/1	0	0/1	0/1	0/1	0/1	0/1
Diandongpetalichthys	0	0	0	0	0	0/1	0/1	0/1
Dicksonosteus	0	0	0	0	0	0/1	0/1	0/1
Diplacanthus	1	0	0	0	1	-	-	-
Doliodus	0	0	0	0	0	0/1	0/1	0/1
Eastmanosteus	0	0	0	0	0	0/1	0/1	0/1
Entelognathus	0/1	0	0	0	0/1	-	-	-
Eurycaraspis	0/1	0	0	0	0/1	0/1	0/1	0/1
Eusthenopteron	0	0	0	0	0	0	0	1
Euthacanthus	1	0	0	0	1	-	-	-
Gavinaspis	0/1	0	0	0	0/1	0/1	0/1	0/1
Gemuendina	0/1	0	0	0	0/1	0/1	0/1	0/1
Gladiobranchus,	0/1	0	0	0	0/1	-	-	-
Glyptolepis	0	0	1	1	1	0	0	1
Gogonaspis	0	0	1	1	1	0	0	1
Groenlandaspis	0/1	0	0	0	0/1	0/1	0/1	0/1
Guiyu	0	0/1	0/1	0/1	0/1	0/1	0/1	0/1
Gyracanthides	0	0	0	0	0	-	-	-
Scyliorhinus	1	0	0	0	1	0	1	0
Hamiltonichthys	0/1	0	0	0	0/1	0	1	0
Helodus	0/1	0	0	0	0/1	0	1	0
Holonema	0	0	0	0	0	0	0	0
Homalacanthus	1	0	0	0	1	-	-	-
Howqualepis	0	0/1	0	0/1	0/1	0/1	0/1	0/1
Incisoscutum	0	0	0	0	0	0/1	0/1	0/1
Ischnacanthus	1	0	0	0	1	0	1	0
Jagorina	0/1	0	0	0	0/1	0/1	0/1	0/1
Janusiscus	0/1	0	0/1	0/1	0/1	0/1	0/1	0/1

Kathemacanthus	0/1	0	0	0	0/1	-	-	-
Kenichthys	0	0	1	1	1	0/1	0/1	0/1
Kentuckia	0	0/1	0	0/1	0/1	0/1	0/1	0/1
Kujdanowiaspis	0	0	0	0	0	0/1	0/1	0/1
Latviacanthus	0/1	0	0	0	0/1	0	0	0
Ligulalepis	0	1	0	1	1	1	0	0
Lophosteus	0	0	0	0	0	0	0	0
Lunaspis	0	0	0	0	0	0/1	0/1	0/1
Lupopsyrus	0/1	0/1	0	0/1	0/1	-	-	-
Macropetalichthys	0	0	0	0	0	0/1	0/1	0/1
Materpiscis	0	0	0	0	0	0/1	0/1	0/1
Meemannia	0	0	1	1	1	0/1	0/1	0/1
Mesacanthus	0/1	0	0	0	0/1	-	-	-
Microbrachius	0/1	0	0	0	0/1	0/1	0/1	0/1
Miguashaia	0/1	0/1	0	0/1	0/1	0	0	1
Lepisosteus	0	1	0	1	1	1	0	1
Polypterus	0	1	0	1	1	1	0	1
Mimipiscis	0	1	0	1	1	1	0	0
Moythomasia	0	1	0	1	1	1	0	0
Obtusacanthus	0/1	0	0	0	0/1	-	-	-
Onychodus	0	0	0	0	0	0	0	1
Onychoselache	1	0	0	0	1	0	1	0
Orthacanthus	0	0	0	0	0	0	1	0
Osorioichthys	0	0/1	0	0/1	0/1	1	0	0
Osteolepis	0	0	1	1	1	0/1	0/1	0/1
Parabuchanosteus	0	0	0	0	0	0/1	0/1	0/1
Parayunnanolepis	0	0	0	0	0	0/1	0/1	0/1
Parexus	0/1	0	0	0	0/1	0	0	0
Poracanthodes	1	0	0	0	1	0/1	0/1	0/1
Porolepis	0	0	1	1	1	0	0	1
Powichthys	0	0	1	1	1	0	0	1
Promesacanthus	0/1	0	0	0	0/1	-	-	-
Psarolepis	0	0	1	1	1	0	0	0
Pterichthyodes	0	0	0	0	0	0/1	0/1	0/1
Ptomacanthus	0/1	0	0	0	0/1	0/1	0/1	0/1
Pucapampella	0/1	0	0	0	0/1	0/1	0/1	0/1
Quasipetalichthys	0	0	0	0	0	0/1	0/1	0/1
Ramirosuarezia	0/1	0	0	0	0/1	0/1	0/1	0/1
Remigolepis	0	0	0	0	0	0/1	0/1	0/1
Rhadinacanthus	1	0	0	0	1	-	-	-
Rhamphodopsis	0/1	0	0	0	0/1	0	0	0
Romundina	1	0	0	0	1	0/1	0/1	0/1
Sigaspis	0/1	0	0	0	0/1	0/1	0/1	0/1
Sinolepis	0/1	0	0	0	0/1	0/1	0/1	0/1
Styloichthys	0	0	1	1	1	0/1	0/1	0/1
Latimeria	0	0/1	1	1	1	0	0	1
Tamiobatis	0/1	0	0	0	0/1	0/1	0/1	0/1
Tetanopsyrus	0/1	0	0	0	0/1	-	-	-
Tristychius	0/1	0	0	0	0/1	0/1	0/1	0/1
Vernicomacanthus	0/1	0	0	0	0/1	0	0	0
Wuttagoonaspis	0/1	0	0	0	0/1	0/1	0/1	0/1
Youngolepis	0	0	1	1	1	0	0	1
Yunnanolepis	0	0	0	0	0	0/1	0/1	0/1
Andreolepis	0	0	1	1	1	0	0	0

Table S2. Presence or absence of enameloid, ganoin, enamel, enamel or ganoin, enamel, ganoin, or enameloid in the dermal skeleton, and acrodin, enameloid, and enamel on teeth, Related to Figure 5. 0, 1, and 0/1 represent the absence, presence, and unknown status of the tissue, respectively. “-” in dental characters represents the absence of teeth.

Taxa	Minimum age (Mya)	Reference
Galeaspida	432.6	
Osteostraci	437.4	
Acanthodes	298	King et al. 2016
Achoania	412	King et al. 2016
Akmonistion	327	King et al. 2016
Austroptyctodus	383	King et al. 2016
Bothriolepis	383	King et al. 2016
Brachyacanthus	415	King et al. 2016
Brindabellaspis	401	King et al. 2016
Brochoadmones	415	King et al. 2016
Buchanosteus	408	King et al. 2016
Campbellodus	383	King et al. 2016
Cassidiceps	415	King et al. 2016
Cheiracanthus	388	King et al. 2016
Cheirolepis	388	King et al. 2016
Chondrenchelys	338	King et al. 2016
Cladodoides	375	King et al. 2016
Cladoselache	360	King et al. 2016
Climatius	415	King et al. 2016
Cobelodus	325	King et al. 2016
Coccosteus	388	King et al. 2016
Compagopiscis	383	King et al. 2016
Cowralepis	383	King et al. 2016
Culmacanthus	385	King et al. 2016
Debeerius	320	King et al. 2016
Diabolepis	412	King et al. 2016
Dialipina	401	King et al. 2016
Diangongpetalichthys	417	King et al. 2016
Dicksonosteus	411	King et al. 2016
Diplacanthus	388	King et al. 2016
Doliodus	395	King et al. 2016
Eastmanosteus	383	King et al. 2016
Entelognathus	424	King et al. 2016
Eurycaraspis	385	King et al. 2016
Eusthenopteron	380	King et al. 2016
Euthacanthus	415	King et al. 2016
Gavinaspis	412	King et al. 2016
Gemuendina	408	King et al. 2016
Gladiobranchus	415	King et al. 2016
Glyptolepis	388	King et al. 2016
Gogonaspis	383	King et al. 2016
Groenlandaspis	385	King et al. 2016
Guiyu	424	King et al. 2016
Gyracanthides	359.3	Warren et al. 2000
Scylliorhinus	0	
Hamiltonichthys	302	King et al. 2016
Helodus	311	King et al. 2016
Holonema	383	King et al. 2016
Homalacanthus	380	King et al. 2016
Howqualepis	385	King et al. 2016
Incisoscutum	383	King et al. 2016
Ischnacanthus	415	King et al. 2016
Jagorina	375	King et al. 2016
Janusiscus	415	King et al. 2016
Kathemacanthus	415	King et al. 2016
Kenichthys	396	King et al. 2016
Kentuckia	347	King et al. 2016
Kujdanowiaspis	411	King et al. 2016
Latviacanthus	404	King et al. 2016
Ligulalepis	401	King et al. 2016
Lophosteus	416	Cunningham et al. 2012
Lunaspis	408	King et al. 2016
Lupopsyrus	415	King et al. 2016
Macropetalichthys	390	King et al. 2016
Materpiscis	383	King et al. 2016
Meemannia	412	King et al. 2016
Mesacanthus	415	King et al. 2016
Microbrachius	386	King et al. 2016
Miguashaia	380	King et al. 2016
Lepisosteus	0	
Polypterus	0	
Mimipiscis	383	King et al. 2016
Moythomasia	383	King et al. 2016

Obtusacanthus	415	King et al. 2016
Onychodus	383	King et al. 2016
Onychoselache	336	King et al. 2016
Orthacanthus	290	King et al. 2016
Osorioichthys	367	King et al. 2016
Osteolepis	388	King et al. 2016
Parabuchanosteus	401	King et al. 2016
Parayunnanolepis	412	King et al. 2016
Parexus	415	King et al. 2016
Poracanthodes	417	King et al. 2016
Porolepis	411	King et al. 2016
Powichthys	411	King et al. 2016
Promesacanthus	415	King et al. 2016
Psarolepis	416	King et al. 2016
Andreolepis	424	Chen et al. 2016
Pterichthyodes	389	King et al. 2016
Ptomacanthus	415	King et al. 2016
Pucapampella	388	King et al. 2016
Quasipetalichthys	385	King et al. 2016
Ramirosuarezia	392	King et al. 2016
Remigolepis	366	King et al. 2016
Rhadinacanthus	386.9	Lukševičs et al. 2010
Rhamphodopsis	388	King et al. 2016
Romundina	415	King et al. 2016
Sigaspis	412	King et al. 2016
Sinolepis	358.5	Zhu et al. 2000
Styloichthys	412	King et al. 2016
Latimeria	0	
Tamiobatis	360	King et al. 2016
Tetanopsyrus	415	King et al. 2016
Tristychius	336	King et al. 2016
Vernicomacanthus	415	King et al. 2016
Wuttagoonaspis	393	King et al. 2016
Youngolepis	412	King et al. 2016
Yunnanolepis	415	King et al. 2016

Table S3. Tip ages used for the ancestral state estimation, Related to Figure 5.

Transparent Methods

Bioinformatic analyses

We searched for SSCP genes in RNA-seq datasets (GenBank accession numbers: SRX796494 and SRX1016233-SRX1016241 for lungfish, SRX796491 and SRX1386644- SRX1386647 for bichir, and SRX424533 and SRX424534 for sturgeon) by tblstn (<http://www.ncbi.nlm.nih.gov/>) (Altschul et al., 1990) using amino acid sequences of gar orthologs as queries (Kawasaki et al., 2017).

Relative expression levels of gar SSCP genes were estimated as FPKM values (Trapnell et al., 2010), calculated for a dataset of gar skin (GenBank accession number, SRP042013) (Braasch et al., 2016) using Galaxy (<https://usegalaxy.org/>) (Afgan et al., 2016). The dataset was retrieved from the EBI SRA database, trimmed using Trimmomatic (Bolger et al., 2014), aligned with the gar genome sequence (LepOcu1) using TopHat (Kim et al., 2013). FPKM values and confidence intervals were calculated using Cufflinks (Trapnell et al., 2010) based on BAM files obtained by the TopHat analysis. All default conditions were used in Galaxy analysis, except additional options in Trimmomatic (ILLUMINACLIP and MINLEN=50) and Cufflinks (multi-read correction). Genomic coordinates of SSCP genes were determined using Splign ([http://www.ncbi.nlm.gov./](http://www.ncbi.nlm.gov/)) (Kapustin et al., 2008), and used for the Cufflinks analysis.

Molecular analysis

All animals used in our study (gar, 16-55 cm in total length; zebrafish, 3.0 cm in total length; bichir, 20 cm in total length; and lungfish, 20 cm in total length; sex not determined for these animals) were sacrificed according to the guidelines issued by the Ministry of Justice in Japan. SSCP genes identified in bichir and lungfish RNA-seq datasets were confirmed by PCR using cDNA libraries made from a tooth plate (lungfish) and tooth germs (bichirs), as described (Braasch et al., 2016). These libraries were made using SMART cDNA library construction kit (Takara Bio), and PCR products were size fractionated by agarose gel electrophoresis, purified from the agarose gel using the FastGene Gel/PCR Extraction Kit (Nippon Genetics), and ligated into T-vector pMD20 (Takara Bio). The ligation mix was then used to transform *E.*

coli HSTo8 Premium Competent Cells (Takara Bio). Exon-intron borders were also determined by PCR using genomic DNA as the substrate. For expression analysis of bichir *scpp5*, total RNA molecules were isolated using RNAiso Plus (Takara Bio) and cDNA was synthesized using the PrimerScript II 1st strand cDNA synthesis kit (Takara bio). Primer sequences used for RT-PCR are as follows: 5'-GAGACTTGCGATCTTCTCTTCTG-3' and 5'-GTGAATTGACCTGAGGCAGGA-3' for *scpp5*; and 5'-CACAGTTTGCCAGATGGTCC-3' and 5'-CACCACCAATTGCCTTGCTC-3' for *gapdh*.

ISH analysis

For ISH analysis, jaws and skin of gar and zebrafish were fixed with neutralized 4% paraformaldehyde, decalcified with Morse's solution (10% w/v sodium citrate and 22.5% v/v formic acid), dehydrated through a graded ethanol series and xylene, embedded in paraffin, sectioned in the coronal plane at 4 µm in thickness, and mounted on glass slides. These glass slides were deparaffined, treated with Proteinase K (20 mg/ml) for 10 min, and used for hybridization (50% formamide, 10 mM Tris pH7.6, 1xDenhart's solution, 5% dextran sulfate, 600 mM NaCl, 0.25% SDS, 1 mM EDTA, and 100µg/ml *Escherichia coli* tRNA) at 70C (Nakatomi et al., 2006). A specific portion of *scpp5*, *ambn*, and *enam* were amplified from the cDNA libraries (primer sequences: 5'-GTTGGTGCTACAGCAGGAAGT-3' and 5'-GTTGTGCTTCCCTGAACTG-3' for gar *ambn*; 5'-AAGGCCTCAGCTTCGTCCAG-3' and 5'-ACTCCTTCTCGTTGACTTCGT-3' for gar *enam*; 5'-TATTCTGAGGAGTCACCAA-3' and 5'-ATTCTGACTTGGCTGGACG-3' for gar *scpp5*; 5'-ACTTCATCAACAGGTGCCCAATC-3' and 5'-TGAAAGCTCCGTGACCTGAATCT-3' for zebrafish *ambn*; and 5'-CTGCCCCTGACAGTGGCAGTAATG-3' and 5'-CATCAGGCCCAACAACAGTGGTGT-3' for zebrafish *enam*), cloned using the pGEM-T Easy Vector (Promega), and labeled with digoxigenin using the DIG RNA Labeling Kit (Roche), and used for hybridization. After hybridization, glass slides were washed with 2xSSC. Hybridization signals were detected using Anti-Digoxigenin-AP, Fab fragments (Roche) and NBT/BCIP Solution (Roche), as specified.

IHC analysis

For IHC analysis, jaws and scales were fixed (4% paraformaldehyde-0.2% glutaraldehyde, 0.05M HEPES buffer pH7.4), dehydrated, and embedded in LR-White resin (LR-W, London Resin), and processed for the Protein A-gold (PAG) method (Sasagawa et al., 2012). For light microscopic analysis, semi-thin sections were made from LR-W resin block, mounted on glass slides, masked with 5% goat serum in phosphate buffered saline (PBS), and reacted with the antibody diluted 1:100 or 1:200 in PBS containing 0.5% bovine serum albumin (PBS-BSA) at 4C. These sections were washed with PBS, incubated with the PAG conjugate (gold nanoparticles 5nm, BBI) diluted 1:100 with PBS-BSA, washed with PBS, treated with the silver enhancer solution (50 mM citrate buffer, 0.85% hydroquinone, 0.1% maleic acid, 0.11% silver lactate, and 1% acacia powder) (Uchida et al., 1991) for 10 min in a dark box, and rinsed with water. These sections were then immersed in photographic fixatives, and stained with fast red or toluidine blue (Sasagawa et al., 2012).

For TEM analysis, ultrathin sections obtained from LR-W resin block were mounted on nickel grids, floated on a drop of 1% goat serum and then the antibody diluted 1:200-1:400 in PBS-BSA. These sections were subsequently washed with PBS-BSA, treated with 1% goat serum and then with the PAG conjugate diluted 1:10 in PBS-BSA. These sections were stained with platinum blue and lead citrate (additionally stained with phosphotungstic acid for some samples), and examined using TEM (JEM-1010, JEOL) (Sasagawa et al., 2016). In some sections, immunoreactions were enhanced using the silver enhancer solution (Uchida et al., 1991) or the Silver Enhancer Kit (Kirkegaard & Perry Laboratories). Polyclonal antibodies to gar Scpp5 were raised against YRQQPQQN (SIGMA-ALDRICH).

Phylogenetic analysis

We augmented an existing phylogenetic dataset (Qiao et al., 2016) with codings for *Scyliorhinus*, *Lepisosteus*, *Polypterus*, and *Latimeria*, representing the three major living clades of jawed vertebrates. We also coded *Andreolepis*, a putative stem-osteichthyan (Qu et al., 2015), and coded *Ligulalepis* as

present for dental acrodin (Schultze, 2016; Schultze, 2018). Morphological data were analyzed using the Mkv+G model in MrBayes 3.2.7 (Ronquist et al., 2012). We constrained the positions of the extant taxa to mitigate long branch effects: *Scyliorhinus* with *Helodus*, *Chondrenchelys*, *Debeerius*, *Tristychius*, *Hamiltonichthys*, *Onychoselache* (Qiao et al., 2016); *Latimeria* with *Miguashaia* (Arratia and Schultze, 2015); and *Polypterus*, *Lepisosteus*, *Kentuckia* with *Moythomasia*, *Mimipiscis* (Giles et al., 2017). We conducted two phylogenetic analyses of this dataset, one in which the GAP clade were constrained to be stem-osteichthyans (King et al., 2017; Lu et al., 2017), and the other in which these taxa were topologically unconstrained, yielding a stem-sarcopterygian affinity (Lu et al., 2016; Qiao et al., 2016; Choo et al., 2017). Each analysis used four independent runs of four chains over 10,000,000 generations, sampling every 10,000 generations, with 25% of burnin. Convergence was assessed using Tracer 1.7 (Rambaut et al., 2018). Results were summarized using majority rule consensus trees (Figures S3A and S3B).

Ancestral state estimation

We compiled data on the distribution of five dermal (enameloid, ganoin, enamel, ganoin/enamel, hypermineralized tissue) and three dental (acrodin, enameloid, enamel) skeletal characters (Table S2); we confirmed the presence of dental enameloid in *Cladoselache* (Figure S3S).

Branch lengths were estimated using the function *timePaleoPhy* in the R package (<https://www.R-project.org/>) *paleotree* (Bapst, 2012) using the 'equal' method (Brusatte et al., 2008), with the root age increased by 5 million years. Tip ages are from King et al. (King et al., 2017) (Table S3) but the root (Tinn and Märss, 2018), and the crown clades of chondrichthyans, sarcopterygians, actinopterygians, osteichthyans, and gnathostomes (Benton et al., 2015) were calibrated to be minimally as old as oldest fossil representative. Using these time-scaled topologies, we estimated ancestral states using Stochastic Character Mapping (Huelsenbeck et al., 2003) in the R package *phytools* (Revell, 2012). Each character was coded as a binary presence or absence state for all taxa; uncertain tip states were assigned an equal prior probability of 0.5. Taxa were excluded from an analysis when the character was

inapplicable. By running all separate analyses for each character, there is zero modelled co-variance between each character. Stochastic character mapping was run using the *make.simmap* function (Bollback, 2006) in *phytools* but with a single Q matrix for all simulations under the 'all rates different' and a naïve equal prior probability for absence or presence at the root. Each character model ran for 1000 simulations and each iteration used the same 'empirical' Q matrix with the highest likelihood transition probabilities. Results were summarized using the *describe.simmap* function and the posterior probabilities of node and tip states were plotting on each tree.

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