MIND bomb 2 prevents RIPK1 kinase activity-dependent and -independent apoptosis through ubiquitylation of cFLIP_L

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Supplementary Materials

Supplementary Figures 1~20

Supplementary Data Files 1~8



Supplementary Fig. 1 Immunoprecipitates with anti-cFLIP antibody exhibit multiple bands of cFLIP_L. WT HeLa cells were transfected with non-target (NT) or *CFLAR* siRNAs. Knockdown of cFLIP_L by *CFLAR* siRNA was verified by immunoblotting (a). After transfection, cells were immunoprecipitated with control IgGs (C) or anti-cFLIP antibody, and the immunoprecipitates were analyzed by anticFLIP antibody (b). The upper and lower arrowheads indicate modified and unmodified cFLIP_L, respectively. Asterisks indicate degraded bands of cFLIP_L. Results are representative of two independent experiments.



Supplementary Fig. 2 MIB2 colocalizes with cFLIPL, but not cFLIPs or caspase 8. a, b HEK293 cells were co-transfected with GFP-MIB2 along with DsRed-cFLIPL, DsRed-cFLIPs, or DsRed-C8C/S. Cellular localization of the indicated proteins was analyzed by DeltaVision (**a**). C8C/S indicates a protease-inactive mutant of caspase 8. Relative fluorescence intensities on the dotted white lines (merged images) were analyzed by Image J and are expressed as cyan (MIB2) and magenta lines (cFLIPL, C8C/S, and cFLIPs), respectively (**b**). At least 5 cell images per transfection were collected. Scale bar, 10 µm. Results are representative of two or three independent experiments.



Supplementary Fig. 3 MIB2 ubiquitylates multiple sites of $cFLIP_L$. a HEK293 cells were transfected with $cFLIP_L$ or deletion mutants of $cFLIP_L$ containing indicated amino acids along with or without MIB2 and HA-Ub. At 24 hours after transfection, cell lysates were immunoprecipitated with anti-FLAG antibody, ubiquitylation of $cFLIP_L$

and its mutants was analyzed by immunoblotting with the indicated antibodies. Protein expression was verified by immunoblotting. **b** Diagrams of cFLIP_L and its deletion mutants, and their binding to MIB2 and ubiquitylation. A deletion mutant containing the indicated amino acids was expressed as GST fusion protein with N-terminal FLAG tag (G193-260), since small polypeptides might be easily degraded in the cells. DED, death effector domain; DM, a docking site for MIB2; CLD, caspase-like domain; GST, glutathione S-transferase. D196 and D376 indicate aspartic acids cleaved by activated caspase 8. **c** MIB2 ubiquitylates multiple lysines in the C-terminal portion of cFLIP_L. Δ N193-480 and its lysine to arginine (KR) mutants at the indicated positions were transfected and analyzed as in **a**. Each number indicates lysine residues' positions, and lysines replaced with arginines are highlighted with red characters. MT indicates the mutants of Δ N193-480. Results are representative of two or three independent experiments.





Supplementary Fig. 4 MIB2 ubiquitylates multiple sites of cFLIP_L. a Alignment of C-terminal caspase-like domain of cFLIP_L in various species. Red boxes highlight conserved amino acids among species. Each number indicates the position of the amino acids of human cFLIP_L. b The dimerization interface of C-terminal domains of cFLIP_L and caspase 8 (adopted from PDB ID codes 3H11) is visualised by RasMol (http://www.openrasmol.org). β 6-strand of cFLIP_L is indicated by orange. Red characters indicate four ubiquitylated lysine residues and Q468.

а

b

а



Supplementary Fig. 5 TNF-induces complex I formation is not altered in WT and MIB2 KO HCT 116 cells. a Generation of *MIB2* KO HCT116 cells. Expression of MIB2 was verified by immunoblotting. b WT and *MIB2* KO HCT116 cells were stimulated with GST-TNF (1 μ g/ml) for the indicated times, and TNFR-containing complex I precipitated with glutathione-Sepharose. Precipitated proteins were analysed by immunoblotting with the indicated antibodies. Protein expression was verified by immunoblotting with the indicated antibodies. c WT and *MIB2* KO HCT116 cells were stimulated with TNF for the indicated times, and phosphorylation and degradation of I κ B α were analysed by immunoblotting with the indicated times, and phosphorylation and the results are representative of two or three independent experiments.



Supplementary Fig. 6 TNF induces phosphorylation of RIPK1 in *MIB2* KO cells in the presence of IAP inhibitor. WT and *MIB2* KO HCT116 cells were stimulated with GST-TNF (1 μ g/ml)/BV6 (0.1 μ M) for the indicated times, and TNFR-containing complex was precipitated and analysed as in Fig. 6f. Asterisks indicated cross-reacted bands. All results are representative of two or three independent experiments.



Supplementary Fig. 7 cFLIPL does not bind to MIB1. a HEK293 cells were transfected with the indicated expression vectors. Cell lysates were immunoprecipitated with control Ig (C) or anti-Myc (M) antibodies, co-immunoprecipitated proteins were analyzed by immunoblotting with anti-FLAG antibody. Protein expression was verified by the indicated antibodies using cell lysates. C8C/S indicates a protease-inactive mutant of caspase 8. b WT and MIB2 KO HeLa cells were immunoprecipitated with control Ig (C) or anti-cFLIP (F) antibody. Immunoprecipitates were analyzed by immunoblotting with anti-MIB1 or anti-MIB2 antibodies. Protein expression was verified by the indicated antibodies. The upper and lower arrowheads indicate modified and unmodified cFLIP_L, respectively. Asterisks indicate degraded bands of cFLIP_L. c WT and MIB2 KO HeLa cells were transfected with non-target (NT) or MIB1 siRNAs. Protein expression was verified by immunoblotting with the indicated antibodies. d WT and MIB2 KO HeLa cells were treated with the indicated siRNAs. Sixteen hours after transfection, cells were stimulated with TNF (10 ng/ml) and BV6 (0.1 μ M) for 8 hours. Cell death was determined by LDH release assay. Results are mean \pm SD of triplicate samples. Unpaired two-tailed Student *t*-test. ns, not significant. All results are representative of two independent experiments.



Supplementary Fig. 8 MIB2 does not promote degradation but rather increases the stability of cFLIP_L. a-d WT and *MIB2* KO HeLa were stimulated with TNF (1 ng/ml) (a), CHX (2.5 μ g/ml) (b), TNF (1 ng/ml)/CHX (2.5 μ g/ml) (c), or TNF (10 ng/ml)/BV6 (0.1 μ M) (d) for the indicated times, and expression of cFLIP_L and cFLIPs was analyzed by immunoblotting with anti-cFLIP antibody. Signal intensities of cFLIP_L, cFLIPs, and tubulin at the indicated times were calculated by Image J and normalized by those of tubulin. Relative intensities of cFLIP_L and cFLIPs at the indicated times compared to those at time 0 (100%) are plotted. e-h WT and *MIB2* KO HCT116 were stimulated with TNF (1 ng/ml) (e), CHX (2.5 μ g/ml) (f), TNF (1 ng/ml)/CHX (2.5 μ g/ml) (g), or TNF (10 ng/ml)/BV6 (0.1 μ M) (h) for the indicated times, and analyzed as in (a). Results are representative of two independent experiments.



Supplementary Fig. 9 Complex IIb formation is facilitated in *MIB2* KO HCT116 cells. a, b WT and *MIB2* KO HCT116 cells were stimulated with TNF (10 ng/ml) and BV6 (0.1 μ M) for the indicated times, and cell lysates were immunoprecipitated with anti-FADD (a) or anti-caspase 8 (b) antibodies. Immunoprecipitated proteins were analyzed by immunoblotting with the indicated antibodies. Protein expression was verified by immunoblotting with the indicated antibody. *Cross-reacted band; C-Casp. 8, cleaved caspase 8; C-cFLIP_L, cleaved cFLIP_L. P and C indicate proform and cleaved form, respectively. All results are representative of two or three independent experiments.



Supplementary Fig. 10 TNF induces dissociation of the complex of MIB2 and cFLIP_L. **a** HeLa or HCT116 cells were stimulated with TNF (10 ng/ml) or IL-1 β (10 ng/ml) for the indicated times, and cell lysates immunoprecipitated with anti-cFLIP antibody. Immunoprecipitated MIB2 and cFLIPL were analysed by immunoblotting with anti-MIB2 and anti-cFLIP antibodies, respectively. The upper and lower arrowheads indicate modified and unmodified cFLIP_L, respectively. Asterisks indicate degraded bands of cFLIP_L. Protein expression was verified by immunoblotting with the indicated antibody. **b** WT and *MIB2* KO HeLa cells were pretreated or not with TNF (10 ng/ml) for 12 hours and then stimulated with the indicated concentrations of TNF (ng/ml) and BV6 (0.1 μ M) for 8 hours. Cell death was determined by the LDH release assay. Results are mean \pm SD of triplicate samples. Unpaired two-tailed Student *t*-test. ns, not significant.





Supplementary Figure 11 | Uncropped blots for Figs. 1 and 2.



Supplementary Figure 12 | Uncropped blots for Figs. 3 and 4.



Supplementary Figure 13 | Uncropped blots for Figs. 5 and 6.

Fig7a



Supplementary Figure 14 | Uncropped blots for Figs. 7 and 8.

Fig9a



Supplementary Figure 15 | Uncropped blots for Fig. 9.





Supplementary Fig3a



Supplementary Fig3c



Supplementary Figure 17 | Uncropped blots for Supplementary Fig. 3.

Supplementary Fig5a



Supplementary Fig5b

35



Supplementary Figure 18 | Uncropped blots for Supplementary Figs. 5 and 6.

Supplementary Fig7a



Supplementary Fig7b



(kD)

150

100

75

50

37

Supplementary Fig7c



Supplementary Fig8a-h



Supplementary Figure 19 | Uncropped blots for Supplementary Figs. 7 and 8.

Supplementary Fig9a







IP-WB: cFLIPL





Input: MIB2

-	(kD)
	100 75
	50 37
======	

S	upp	lem	entai	ry F	ig9	b
				-	-	







IP-WB: CASPASE8

(kD)	
190	
60	
45	
35	
25	

Input: RIPK1 (kD) 100 75 50 37 (kD) 100 75 50 37 25



50

37

25



Input: FADD





Input: cIAP1 (kD) 150 100 75 50

37

25

Input: Tubulin (kD) 100 75 50 37 25

Supplementary Fig10a



Supplementary Figure 20 | Uncropped blots for Supplementary Figs. 9 and 10.