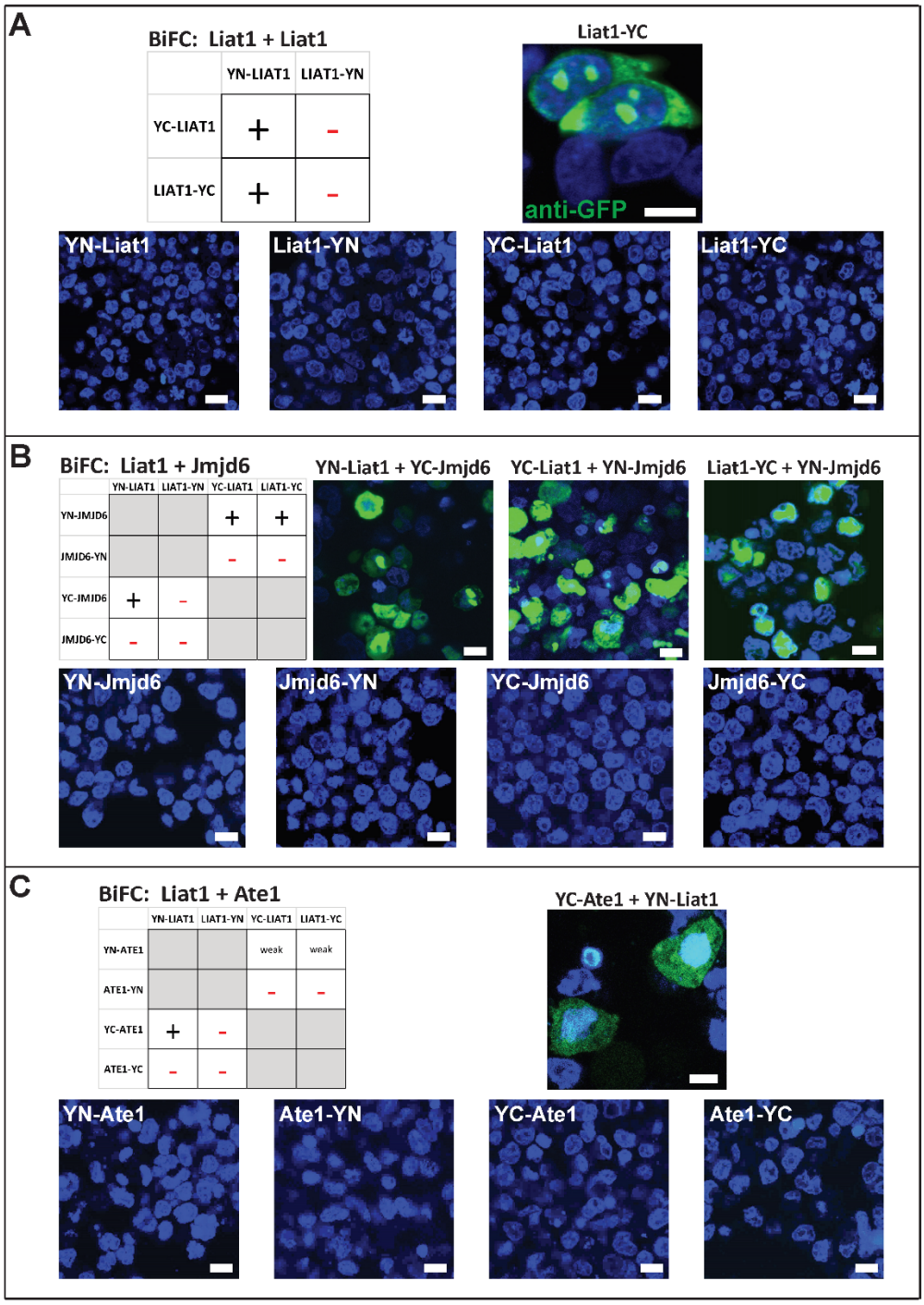

Supplemental Fig. Bimolecular Fluorescence Complementation (BiFC) platforms. (A) *Upper left panel*, table of BiFC detected by Liat1 self-interaction using indicated plasmid combinations. "+" indicates reconstituted fluorescence and "–" indicates absence of fluorescence. *Upper right panel*, Liat1-YC expression detected with an anti-GFP antibody. *Lower panels*, lack of BiFC detected with the individual indicated plasmids. (B) *Upper left panel*, table of Liat1-Jmjd6 BiFC detected. *Upper right panel*, Liat1-Jmjd6 BiFC detected with indicated plasmid combinations. *Lower panels*, lack of BiFC detected with the individual indicated plasmids. (C) Same as in B except with Liat1-Ate1 BiFC. *White bars, 10 µm. DAPI, 4'-6-diamidino-2-phenylindole.*

Supplemental Table S1. Plasmids used in this study.

Plasmid	Description	Source or Reference
pCDNA3.0-Neo	Amp^R; Neo^R; Expression vector for cloning your gene of interest from the CMV promoter.	Invitrogen
pcDNA3.1-Hyg	Amp^R; Hyg^R; Expression vector for cloning a gene of interest.	Invitrogen
pACT2	Amp^R; *LEU2* selectable marker in yeast. Y2H expression vector with P_{ADH1} promoter. Produces ha-tagged Gal4-AD fusion.	Clontech
pAS2	Amp^R; (also called pAS1-CYH2); *CYH* selectable marker in yeast. Y2H expression vector with P_{ADH1} promoter. Produces ha-tagged Gal4-DBD fusion.	Clontech
pEGFP-N1	Kan^R; Neo^R; eGFP expression vector under the control of mammalian P_{CMV} promoter.	Clontech
pEYFP-N1	Kan^R; Neo^R; eYFP expression vector under the control of mammalian P_{CMV} promoter.	Clontech
LentiCRISPR v2	Amp^R; Pur^R; lentivirus CRISPR mammalian expression vector. Cas9 is expressed by the $P_{EF\text{-}1\alpha}$ promoter; the single-guide RNA is expressed by the P_{U6} promoter.	(1)
pAA01	Amp^R; pcDNA3.1-based plasmid encoding mouse $^{3xHA}Liat1\Delta E^{(\Delta 14\text{-}23)}$ under the control of mammalian P_{CMV} promoter.	Present study
pAA02	Amp^R; pcDNA3.1-based plasmid encoding mouse $^{3xHA}Liat1\Delta K^{(\Delta 53\text{-}60)}$ under the control of mammalian P_{CMV} promoter.	Present study
pAA03	Kan^R; pEGFP-N1-based vector containing the IDR of mouse $Liat1^{(1\text{-}118)}$ fused to the eGFP cDNA to express ^{3xHA}IDR-GFP under the control of mammalian P_{CMV} promoter.	Present study
pAA05	Amp^R; LentiCRISPR v2-based plasmid expressing *S. pygenes* Cas9 and $CRISPR^{Liat1}$-A.	Present study

pAA06	Amp^R; LentiCRISPR v2-based plasmid expressing *S. pygenes* Cas9 and $CRISPR^{Liat1}$-B.	Present study
pCER020	Amp^R; pACT2-based plasmid encoding Gal4-AD fused to $^{ha}Liat1^{(\Delta 1\text{-}124)}$ under the control of the yeast P_{ADH1} promoter.	(2)
pCER021	Amp^R; pACT2-based plasmid encoding Gal4-AD fused to $^{ha}Liat1^{(113\text{-}165)}$ under the control of the yeast P_{ADH1} promoter.	(2)
pCER022	Amp^R; pACT2-based plasmid encoding Gal4-AD fused to $^{ha}Liat1^{(\Delta 153–228)}$ under the control of the yeast P_{ADH1} promoter.	(2)
pCB179	Amp^R; pcDNA3.1-based plasmid encoding mouse ^{3xHA}Liat1 under the control of the mammalian P_{CMV} promoter.	(2)
pCB239	Amp^R; pAS2-based plasmid encoding Gal4-DBD domain fused to $^{ha}Liat1^{(1\text{-}228)}$ under the control of the yeast P_{ADH1} promoter.	(2)
pCB409	Amp^R; pH10UE-based vector used for the bacterial expression of a fusion between His_{10}-ubiquitin and $Ate1^{1A7A}$.	(2)
pCB432	Amp^R; pACT2-based plasmid encoding Gal4-AD fused to $^{ha}Liat1^{(1\text{-}228)}$ under the control of the yeast P_{ADH1} promoter.	(2)
pCB433	Amp^R; pACT2-based plasmid encoding Gal4-AD fused to $^{ha}Liat1^{(61\text{-}228)}$ under the control of the yeast P_{ADH1} promoter.	(2)
pCB434	Amp^R; pACT2-based plasmid encoding Gal4-AD fused to $^{ha}Liat1^{(113\text{-}228)}$ under the control of the yeast P_{ADH1} promoter.	(2)
pCB435	Amp^R; pACT2-based plasmid encoding Gal4-AD fused to $^{ha}Liat1^{(135\text{-}228)}$ under the control of the yeast P_{ADH1} promoter.	(2)

pCB436	Amp^R; pACT2-based plasmid encoding Gal4-AD fused to $^{ha}Liat1^{(135-165)}$ under the control of the yeast P_{ADH1} promoter.	(2)
pCB437	Amp^R; pACT2-based plasmid encoding Gal4-AD fused to $^{ha}Liat1^{(1-165)}$ under the control of the yeast P_{ADH1} promoter.	(2)
pCB439	Amp^R; pACT2-based plasmid encoding Gal4-AD fused to $^{ha}Liat1^{(95-228)}$ under the control of the yeast P_{ADH1} promoter.	(2)
pCB458	Amp^R; pcDNA-Neo-based plasmid encoding $Jmjd6^{3xflag}$ under the control of the mammalian P_{CMV} promoter.	(2)
pCB481	Amp^R; Neo^R; pcDNA3.0-based plasmid expressing the N-terminal portion of eYFP (amino acids 1-154) and linker upstream of the multiple cloning site.	Present study
pCB482	Amp^R; Neo^R; pcDNA3.0-based plasmid expressing the C-terminal portion of eYFP (amino acids 154-238) and linker upstream of the multiple cloning site.	Present study
pCB483	Amp^R; Neo^R; pcDNA3.0-based plasmid expressing the linker and N-terminal portion of eYFP (amino acids 1-154) downstream of the multiple cloning site.	Present study
pCB484	Amp^R; Neo^R; pcDNA3.0-based plasmid expressing the linker and C-terminal portion of eYFP (amino acids 154-238) downstream of the multiple cloning site.	Present study
pCB486	Amp^R; pCB481-based plasmid expressing the N-terminal half of eYFP (amino acids 1-154) fused to $Ate1^{1A7A}$ (YN-ATE1) from the P_{CMV} promoter.	Present study
pCB487	Amp^R; pCB482-based plasmid expressing the C-terminal half of eYFP (amino acids 154-238) fused to $Ate1^{1A7A}$ (YC-ATE1) from the P_{CMV} promoter.	Present study

pCB488	Amp^R; pCB483-based plasmid expressing $Ate1^{1A7A}$ fused to the N-terminal half of eYFP (amino acids 1-154) (ATE1-YN) from the P_{CMV} promoter.	Present study
pCB489	Amp^R; pCB484-based plasmid expressing $Ate1^{1A7A}$ fused to the C-terminal half of eYFP (amino acids 154-238) (ATE1-YC) from the P_{CMV} promoter.	Present study
pCB490	Amp^R; pCB481-based plasmid expressing the N-terminal half of eYFP (amino acids 1-154) fused to Liat1 (YN-Liat1) from the P_{CMV} promoter.	Present study
pCB491	Amp^R; pCB482-based plasmid expressing the C-terminal half of eYFP (amino acids 154-238) fused to Liat1 (YC-Liat1) from the P_{CMV} promoter.	Present study
pCB492	Amp^R; pCB483-based plasmid expressing Liat1 fused to the N-terminal half of eYFP (amino acids 1-154) (Liat1-YN) from the P_{CMV} promoter.	Present study
pCB493	Amp^R; pCB484-based plasmid expressing Liat1 fused to the C-terminal half of eYFP (amino acids 154-238) (Liat1-YC) from the P_{CMV} promoter.	Present study
pCB494	Amp^R; pCB481-based plasmid expressing the N-terminal half of eYFP (amino acids 1-154) fused to Jmjd6 (YN-Jmjd6) from the P_{CMV} promoter.	Present study
pCB495	Amp^R; pCB482-based plasmid expressing the C-terminal half of eYFP (amino acids 154-238) fused to Jmjd6 (YC-Jmjd6) from the P_{CMV} promoter.	Present study
pCB496	Amp^R; pCB483-based plasmid expressing Jmjd6 fused to the N-terminal half of eYFP (amino acids 1-154) (Jmjd6-YN) from the P_{CMV} promoter.	Present study
pCB497	Amp^R; pCB484-based plasmid expressing Jmjd6 fused to the C-terminal half of eYFP (amino acids 154-238) (Jmjd6-YC) from the P_{CMV} promoter.	Present study

pCB500	Amp^R; pcDNA-Neo-based plasmid expressing C-terminally triple flag-tagged $Jmjd6^{N277A}$ under the control of the mammalian P_{CMV} promoter.	Present study
pCB501	Amp^R; pcDNA-Neo-based plasmid expressing C-terminally triple flag-tagged $Jmjd6^{1-334}$ under the control of the mammalian P_{CMV} promoter.	Present study
pCB502	Amp^R; pcDNA-Neo-based plasmid expressing C-terminally triple flag-tagged $Jmjd6^{1-290}$ under the control of the mammalian P_{CMV} promoter.	Present study
pCB531	Amp^R; pACT2-based plasmid encoding Gal4-AD fused to $^{ha}Liat1^{(41-228)}$ under the control of the yeast P_{ADH1} promoter.	(2)

Supplemental Table S2. Primers used in this study

Primer	Primer's sequence (5' to 3')
CB200F	GATCGCTAGCGCCACCATGTACCCATACGATGTTCCTGAC
CB443F	GATCAAGCTTGCCACCATGGTGAGCAAGGGCGAGG
CB444R	GATCGGTACCAGTAGCAATCGATCGCATGATATAGACGTTGTGGCTG
CB445F	GATCAAGCTTGCCACCATGGCCGACAAGCAGAAGAACGGC
CB446R	GATCGGTACCAGTAGCAATCGATCGCTTGTACAGCTCGTCCATGCC
CB447F	GATCTCTAGACGATCGATTGCTACAATGGTGAGCAAGGGCGAGG
CB448R	GATCGGGCCCTCACATGATATAGACGTTGTGGCTG
CB449F	GATCTCTAGACGATCGATTGCTACAATGGCCGACAAGCAGAAGAACGGC
CB450R	GATCGGGCCCTCACTTGTACAGCTCGTCCATGCC
CB451F	GATCGAATTCTGATGGCCTCGGTGGTGGAATAC
CB452R	GATCTCTAGATCAGTGTCTGAACAGCAGCATCC
CB453F	GATCAAGCTTGGCGCCATGGCCTCGGTGGTGGAATAC
CB454R	GATCCTCGAGGTGTCTGAACAGCAGCATCC
CB455F	GATCGAATTCTGATGGCCGGCCGTGGTGGG
CB456R	GATCTCTAGATCACTCTGCTACAGTGGTGGCTAATTCTGGG
CB457F	GATCAAGCTTGGCGCGATGGCCGGCCGTGGTGGG
CB458R	GATCCTCGAGCTCTGCTACAGTGGTGGCTAATTCTGGG
CB459F	GATCGGTACCATGAACCACAAGAGCAAGAAGCGC
CB460R	GATCTCTAGATCACCTGGAGGAGCTGCGCTC
CB461F	GATCAAGCTTGGCACCATGAACCACAAGAGCAAGAAGCGC
CB462R	GATCTCTAGACCTGGAGGAGCTGCGCTC
CB475F	GTTGTCCTCGCCCTTGACACCACC
CB476R	ATGCCACCATCCCCCTGGTACAAAGAC
CB477F	TCCACAGGCATTGCCTCTGAC
CB478R	CTCCTGGAGGTCAACTGCGTC
CB479F	AGCACCAACTTCCCTGTTGTG
CB480R	GCTGGCAAAGTTCTGGGTGAT
AA01F	GCGCGGGAAGGCGGAGC
AA02R	ACCATACTCCGCCGCACCGG
AA03F	ACTAAAGGATCGGGCAAGGGCGAC

AA04R	CACCTTCCGTTTGGCCAGCTCG
AA05R	ATCGGTACCGAATAGGGAAGGGTTTGCCTATTTTCCTCTTTGTCTCTGGGC
JD10	CACCGTGAATCGGAGAAAACGGTAC
JD11	AAACGTACCGTTTTCTCCGATTCAC
JD12	CACCGGTCGGGGTGGAAGCCCTTGA
JD13	AAACTCAAGGGCTTCCACCCCGACC

References

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