## nature research

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## **Reporting Summary**

Nature Research wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Research policies, see our Editorial Policies and the Editorial Policy Checklist.

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FOLS	an statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.			
n/a	Confirmed			
	$oxed{x}$ The exact sample size $(n)$ for each experimental group/condition, given as a discrete number and unit of measurement			
	🕱 A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly			
	The statistical test(s) used AND whether they are one- or two-sided Only common tests should be described solely by name; describe more complex techniques in the Methods section.			
×	A description of all covariates tested			
×	A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons			
	A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)			
×	For null hypothesis testing, the test statistic (e.g. <i>F</i> , <i>t</i> , <i>r</i> ) with confidence intervals, effect sizes, degrees of freedom and <i>P</i> value noted Give <i>P</i> values as exact values whenever suitable.			
x	For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings			
×	For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes			
×	Estimates of effect sizes (e.g. Cohen's d, Pearson's r), indicating how they were calculated			
·	Our web collection on <u>statistics for biologists</u> contains articles on many of the points above.			
Software and code				
Policy information about availability of computer code				

Data collection NIS Elements 4.13.4 (Nikon), Utrack 2.2.1

Data analysis

NIS Elements 4.13.4 (Nikon), SMTracker, BLASTP online tool, Phobius online tool, Raptorx online tool, Coiled-coils online tool, SWISS-MODEL

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Research guidelines for submitting code & software for further information.

Policy information about availability of data

All manuscripts must include a data availability statement. This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A list of figures that have associated raw data
- A description of any restrictions on data availability

Numerical data underlying Figs. 2A, 2C and 4B, Supplementary Figs.4B-D, 18 and 24, as well as Supplementary Tables 1-8 are provided as Source Data file, The authors declare that the main data supporting the findings of this study are available within the article and its Supplementary information files. All other data are available from the corresponding author upon request.

Field-specific reporting					
Please select the one below that is the best fit for your research. If you are not sure, read the appropriate sections before making your selection.					
Life sciences  Behavioural & social sciences  Ecological, evolutionary & environmental sciences					
For a reference copy of t	the document with all sections, see <a href="mailto:nature.com/documents/nr-reporting-summary-flat.pdf">nature.com/documents/nr-reporting-summary-flat.pdf</a>				
Life sciences study design					
All studies must disclose on these points even when the disclosure is negative.					
Sample size	For single cell fluorescence microscopy analyses of exponentially growing cultures, two to four independent cultivations and microscopy analyses were performed. In case of numeric evaluation of fluorescence patterns in cells in exponentially growing cultures, the cell number per replicate was aimed at the minimum of 300 and in case of time-lapse analysis, at the minimum of 100 cells. In most cases, the actual numbers of analyzed cells substantially exceeded the minimum. For growth curve analysis and Western blot analysis, either three or four replicates were used.				
Data exclusions	No data was excluded from the analysis				
Replication	All attempts at replication were successful. The experiments were replicated at least once.				
Randomization	Not relevant to this study, since no experimental groups were generated				
Blinding	Not relevant to this study, since no experimental groups were generated				
Reporting for specific materials, systems and methods					
We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.					
Materials & experimental systems Methods					
n/a Involved in th	n/a Involved in the study				
Antibodies	ChIP-seq				
<b>x</b> Eukaryotic	cell lines Flow cytometry				
	ogy and archaeology MRI-based neuroimaging				
<b>=</b> 1 <b>=</b>	Human research participants				
Dual use research of concern					

## **Antibodies**

Antibodies used

Monoclonal ANTI-FLAG® M2-Peroxidase (HRP) antibody produced in mouse (Sigma-Aldrich)

Validation

Commercial antibody against the FLAG tag, species-independent. The Monoclonal ANTI-FLAG M2-Peroxidase is a mouse IgG antibody covalently conjugated to horseradish peroxidase (HRP). The antibody binds to FLAG fusion proteins and recognizes the FLAG epitope at FLAG peptides. ANTI-FLAG is a registered trademark of Sigma-Aldrich Co. LLC