

Figure S1. Transsulfuration cannot support NSCLC cysteine pools (related to Figure 1). (A) A representative image of Sytox Green (SG) stained A549 cells following cystine starvation for 1 or 49 hours in the presence of Vehicle (0.1% DMSO), Fer-1 (10 μ M), or DFO (100 μ M). Images show the same well position at the two different time points (1 or 49 hrs). A representative Sytox Green positive (SG⁺) cell is indicated with a white arrow. (B) A representative image of non-stained A549 cells following 49 hours of cystine starvation treated with Vehicle (0.1% DMSO), Fer-1 (10 μ M), or DFO (100 μ M). Representative ferroptotic cells are indicated with white arrows. (C) Measurement of A549 cell lipid peroxidation under cystine starved (0 μ M) or replete (200 μ M) conditions treated with Vehicle (0.1% DMSO) or Fer-1 (10 µM) for 10 hrs (N=3). The median fluorescence intensity (median FI) of oxidized BODIPY-C11 signal was normalized by mean value of vehicle treated cystine replete conditions. (D) Incucyte time course measurement of NSCLC cell death under cystine starved (0 µM) or replete (200 µM) conditions treated with Vehicle (0.1% DMSO), Ferrostatin-1 (Fer-1, 10 μ M) or DFO (100 μ M) (N=4). Cell death was determined by Sytox Green every 3 hours over 73 hours and normalized to cell density. For C and D, data are shown as mean \pm SD. N is the number of biological replicates. *P<0.05 and ****P<0.0001. For C, a one-way ANOVA with Bonferroni's multiple comparison test was used for statistical analyses.



Figure S2. Cystine starvation induces glutamate-derived γ -glutamyl-peptide accumulation (related to Figure 2). (A) A549 cell 2, 3, 3-²H₃-Ser tracing under cystine replete or starved conditions in the presence and absence of SHIN-1 (0.5 µM) for 12 hrs (N=3). (B-C) Intracellular (B) cysteine and (C) γ -Glu-peptides levels in A549 cells under cystine replete or starved conditions in the presence and absence of erastin (10 µM) for 12 hrs (N=3). For A-C, data are presented as mean ± SD. N is number of biological replicates. *P<0.05, **P<0.01, ***P<0.001, and ****P<0.0001; #P<0.05, ##P<0.01. For A-C, a one-way ANOVA with Bonferroni's multiple comparison test was used for statistical analyses.



Figure S3. GCLC mediates γ -glutamyl-peptide synthesis in cell culture (related to Figure 3). (A) Representative immunoblots of GCLC and GSS expression from H1299^{Cas9} cells transduced with sgRNAs (sgCon, sgGCLC or sgGSS), followed by reconstitution with GFP (+GFP), sgRNA-resistant GCLC (+GCLC_{Res}), or sgRNA-resistant GSS (+GSS_{Res}). β -actin was used as a loading control. (B-C) Intracellular γ -Glu-tripeptides (B) and γ -Glu-dipeptides (C) in the cells from (A) under cystine replete or starved conditions for 3 hrs (N=3). For B-C, data were normalized to the mean value of sgCon + GFP cells under cystine replete conditions. (D-E) Intracellular GSH (D) and γ -Glu-peptide levels (E) in A549 cells treated with erastin (10 μ M) and/or BSO (100 μ M) for 12 hrs (N=3). (F) Intracellular y-Glu-peptides levels in four NSCLC cell lines under cystine replete or starved conditions in the presence and absence of BSO (100 µM) for 12 hrs (N=3). (G) Schematics depicting the effect of cystine availability and GCLC activity on γ -Glu-peptide production. For B - F, data are presented as mean + SD. N is number of biological replicates. n.d. not detected. **P<0.01, ***P<0.001, and ****P<0.0001; ####P<0.0001. For GSH in B, one-way ANOVA with Bonferroni's multiple comparison test was used for the statistical comparison between sqCon + GFP, sqGCLC + GFP, and sqGSS + GFP. For the statistical comparison between sgGCLC + GFP and sgGCLC + GCLC_{Res} or sgGSS + GFP and sgGSS + GSS_{Res}, an unpaired two-tailed t test was used. For γ -Glu-Ala-Gly and γ -Glu-2AB-Gly in B and C, an unpaired two-tailed t test was used for the comparison with parental cells under cystine replete conditions [+(Cys)₂]. For D-F, a one-way ANOVA with Bonferroni's multiple comparison test was used for statistical analyses.



Figure S4. GCLC mediates γ-glutamyl-peptide synthesis in vivo (related to Figure **4**). (A-C) Analysis of lung cysteine (A), GSH (B), and γ-Glu-peptides levels (C) in mice treated with Cyst(e)inase or PBS, together with BSO or saline. (D-F) Analysis of kidney cysteine (D), GSH (E), and γ-Glu-peptides levels (F) in the mice from (A-C). The metabolite levels were normalized to the mean value of PBS/saline treated mice (N=5). (G-H) Analysis of lung GSH (G) and γ-Glu-peptides levels (H) in Gclc^{f/f} and Gclc^{-/-} mice. (I-J) Analysis of kidney GSH (I) and γ-Glu-peptides levels (J) in the mice from (G-H). The metabolite levels are normalized to the mean value of Gclc^{f/f} mouse tissues (N=4). For A-J, data are presented as mean ± SD. N is number of biological replicates. n.d., not detected; *P<0.05, **P<0.01, ***P<0.001, and ****P<0.0001. For A-F, a one-way ANOVA with Bonferroni's multiple comparison test was used for statistical analyses. For G-J, an unpaired two-tailed t test was used for the statistical comparisons.







γ-Glu-Ala



γ-Glu-Leu

γ-Glu-2AB-Gly

KEAP1^{WT}

KEAP1^{WT}

LANG CINA POCKANA

KEAP1^{MUT}

KEAP1^{MUT}

KEAP1^{MUT}

V CONTROL

KEAP1^{MUT}

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ЗĬ?

KEAP1^{WT}

🔲 (Cys)₂ 200 μM

KEAP1^{WT}



KEAP1^{MUT}

Figure S5. NRF2 promotes γ -glutamyl-peptide synthesis via GCLC (related to Figure 5). (A) Analysis of relative γ -Glu-dipeptide levels in KEAP1^{WT} (N=7) and KEAP^{MUT} (N=6) NSCLC cell lines. The γ -Glu-peptides were analyzed following culture in cystine replete or starved conditions for 12 hrs. The γ -Glu-peptide amount of each NSCLC cell line was normalized to the mean value of H1581 cells under cystine replete conditions.



Figure S6. Dipeptide synthesis protects KEAP1 mutant cells from ferroptosis (related to Figure 6). (A) Determination of the ability of exogenous γ -Glu-dipeptides to elevate intracellular γ -Glu-dipeptide levels. A549 cells were treated with the indicated concentration of γ -Glu-dipeptides for 12 hours under cystine replete and starved conditions and intracellular levels were assayed. γ -Glu-peptide amounts were normalized by the mean value of cystine replete conditions (N=3). (B) NSCLC cell death was monitored using Sytox Green (SG) every 3 hours with the Incucyte system. Cells were cultured under cystine starved or replete conditions and treated with vehicle (0.1% DMSO) or BSO (100 μ M) for 73 hours (N=4). Vehicle-treated cystine replete and starved data are from Figure S1D. (C) Representative immunoblot of GPX4 expression in NRF2^{LOW} and NRF2^{HIGH} NSCLC cell lines. This blot is a reprobing of the GCLM blot shown in Figure 5A and shares the loading control (HSP90). For A and B, data are presented as mean \pm SD. N is number of biological replicates.



Figure S7. Dipeptide synthesis scavenges glutamate (related to Figure 7). (A) Representative immunoblot of xCT, GOT1, and SLC1A5 in NRF2^{LOW} and NRF2^{HIGH} NSCLC cell lines. The xCT and GOT1 blots share a membrane with the NRF2 blot shown in Figure 5A and share the same loading control (HSP90). The SLC1A5 blot is a reprobing of the same membrane as the GCLC and GSS blots shown in Figure 5A and shares the loading control (HSP90). (B) (Left) Determination of the Glu export rate in NSCLC cell lines under cystine replete and starved conditions for 12 hrs (N=3). (Right) Comparison of the Glu export rate between KEAP1 WT (N=7) and KEAP1 MUT NSCLC (N=6) cells. (C) (Left) Analysis of intracellular Gly levels in NSCLC cell lines under cystine replete and starved conditions for 12 hrs (N=3). (Right) Comparison of Gly levels between KEAP1 WT (N=7) and KEAP1 ^{MUT} NSCLC (N=6) cells. (D) Analysis of intracellular Glu concentrations in A549 cells treated with erastin (10 µM) in the presence and absence of 100 µM BSO for 12 hrs (N=3). (E) Analysis of relative intracellular Glu levels in the H1299^{Cas9} transduced with sgRNAs (sgCon, sgGCLC or sgGSS), followed by reconstitution with GFP (+GFP), sgRNA-resistant GCLC (+GCLC_{Res}), or sgRNA-resistant GSS (+GSS_{Res}) under cystine replete or starved conditions for 3 hrs (N=3). (F) Analysis of intracellular Gln, Glu, α -ketoglutarate (α KG), succinic acid (Suc), Fumaric acid (Fum), Malic acid (Mal), and Citric acid (Cit) under cystine replete and starved conditions for 12 hrs in the presence and absence of GluEE (5 mM), AOA (0.5 mM), or Gln starvation. (N=3). (G) Analysis of intracellular A549, H1944, Calu3, and H1975 cell Glu levels in the presence of GluEE (5 mM) for 12 hrs (N=3). (H) Analysis of intracellular A549 cell Cys (left) and Glu (right) levels following culture in cystine replete and starved conditions in the presence of GluEE (5 mM), Erastin (0.5 µM), or Glu (5 mM) for 12 hours (N=3). (I) Analysis of the death of NSCLC cell lines treated with GluEE (5 mM), AOA (0.5 mM) or starved of Gln. Cells were starved of cystine and cell death was monitored by Sytox Green every 3 hours for 49 hours, followed by AUC calculation. (N=4 except H1299, cystine 0 µM + GluEE: N=3). (J) Analysis of the death of GCLC KO A549 cells reconstituted with GCLC (+GCLC_{Res}), treated with Fer-1 (10 µM), or starved of Gln with or without Glu. Cells were starved of cystine for the indicated time points and death was monitored by Cytotox Red (CR) every 3 hours, and counts were normalized to density (N=3). (K) Analysis of A549 cell mitochondrial superoxide levels with MitoSOX under cystine starved and replete

conditions in the presence of AOA (0.5 mM), GluEE (5 mM), or Gln starvation for 8.5 hrs (N=3). MFI = mean fluorescence intensity. For B-K, data is presented as mean \pm SD. N is number of biological replicates. *P<0.05, **P<0.01, ***P<0.001, and ****P<0.0001. For B (left), C (left), E, and G, unpaired two-tailed t tests were used for the statistical comparisons. For E, * is for the comparison with sgCon + GFP under cystine replete conditions [+(Cys)₂] and # is for the comparison between sg Con + GFP under cystine starved conditions [-(Cys)₂]. For B (right), C (right), D, F, H, I, and K, one-way ANOVA with Bonferroni's multiple comparison tests were used for statistical analyses.