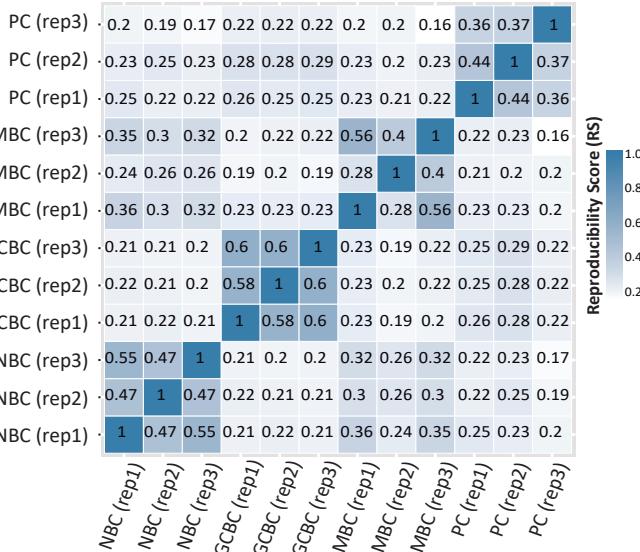


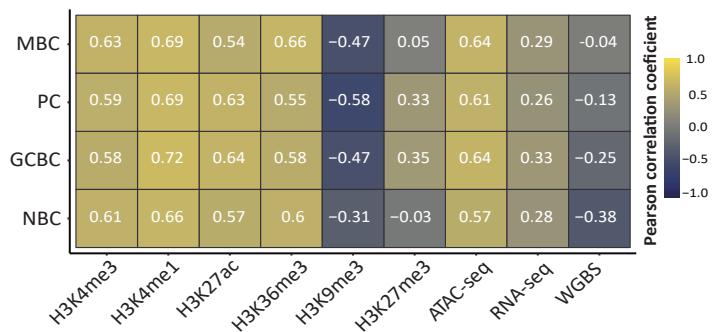
Supplementary Figures

Supplementary Fig. 1

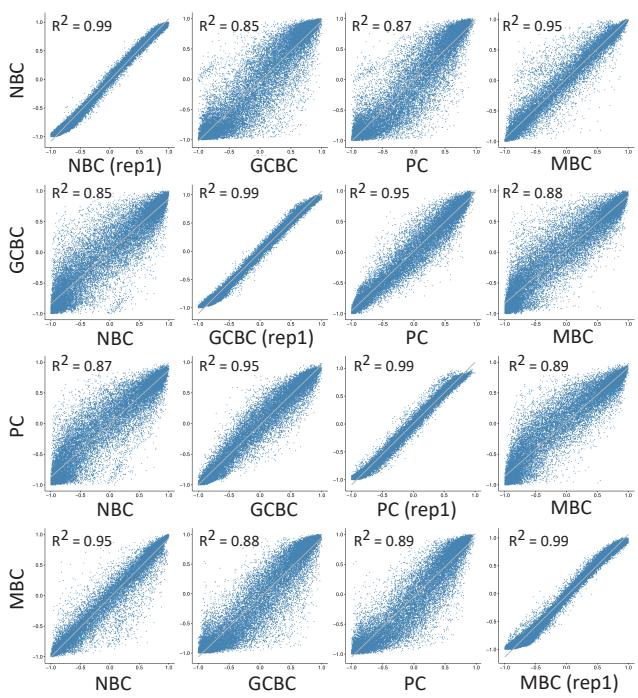
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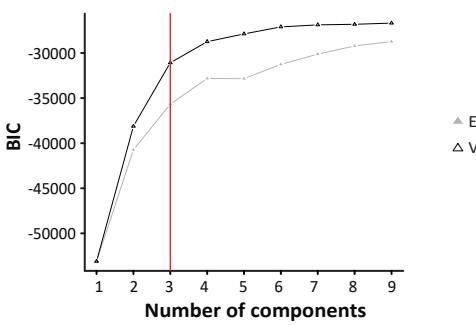
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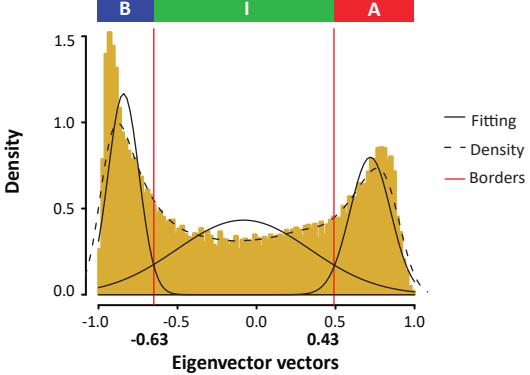
c



d



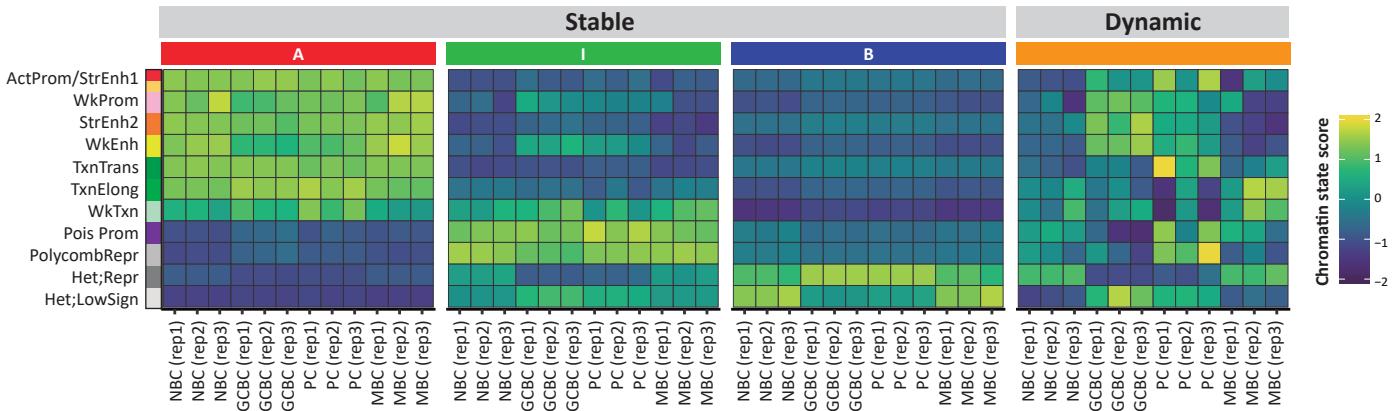
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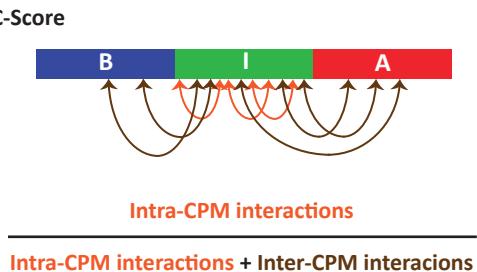
Supplementary Fig. 1. **a** Average genome-wide reproducibility score matrix of each B cell subpopulation replicate at 100 kb resolution. The reproducibility score ranging between 0 (totally different matrices) and 1 (identical matrices). **b** Pearson correlation between the eigenvector coefficients with six histone marks, chromatin accessibility (ATAC-seq), gene expression (RNA-seq) and DNA methylation (WGBS). Positive values of the eigenvector show higher correlation with H3K4me1 (enhancer mark) and chromatin accessibility. **c** Genome-wide scatterplots of coefficients from the first eigenvectors between pairs of B cell subpopulations at 100 kb resolution. The coefficient of determination (R^2) is indicated. **d** Bayesian Information Criterion (BIC) plot for the equal (E) and unequal (V) variance model parameterization ranged from 1 to 10 clusters. **e** Compartment definition model. The x-axis shows the distribution of the eigenvector coefficients and the y-axis indicates the density. The fitting model proposed is highlighted using solid black lines. The red lines mark the intersection points ($IV_1 = -0.63$ and $IV_2 = +0.43$) used to distinguish the three different compartments (A-type, I-type, B-type). Sample sizes were for NBC, GCBC, PC and MBC: $n = 3$ merged biologically independent samples.

Supplementary Fig. 2

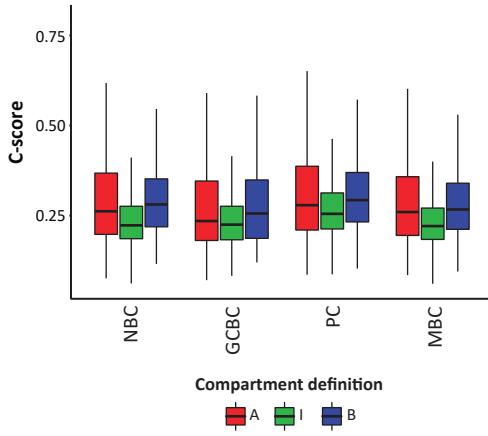
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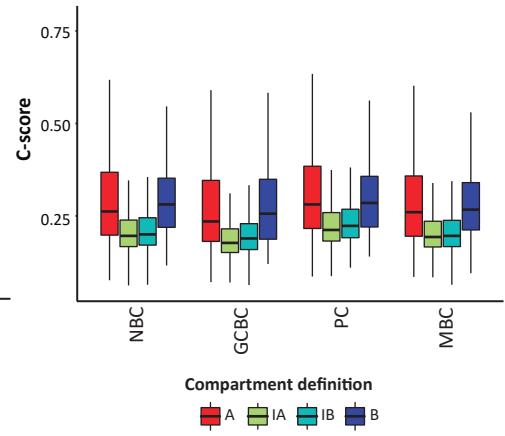
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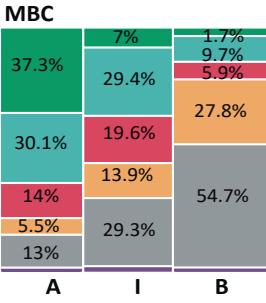
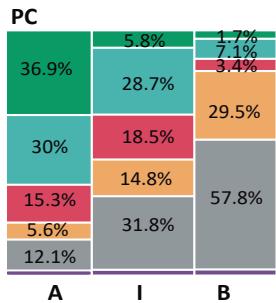
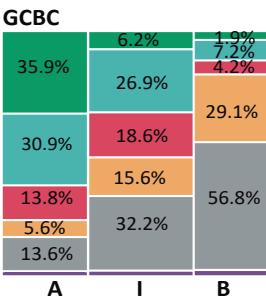
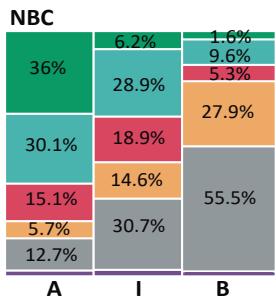
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d

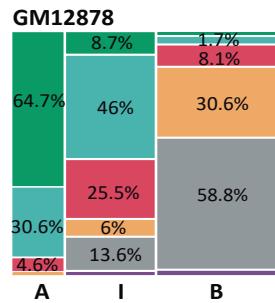


e



Subcompartments defined
by Rao *et al.*, 2014

A1
A2
B1
B2
B3
B4



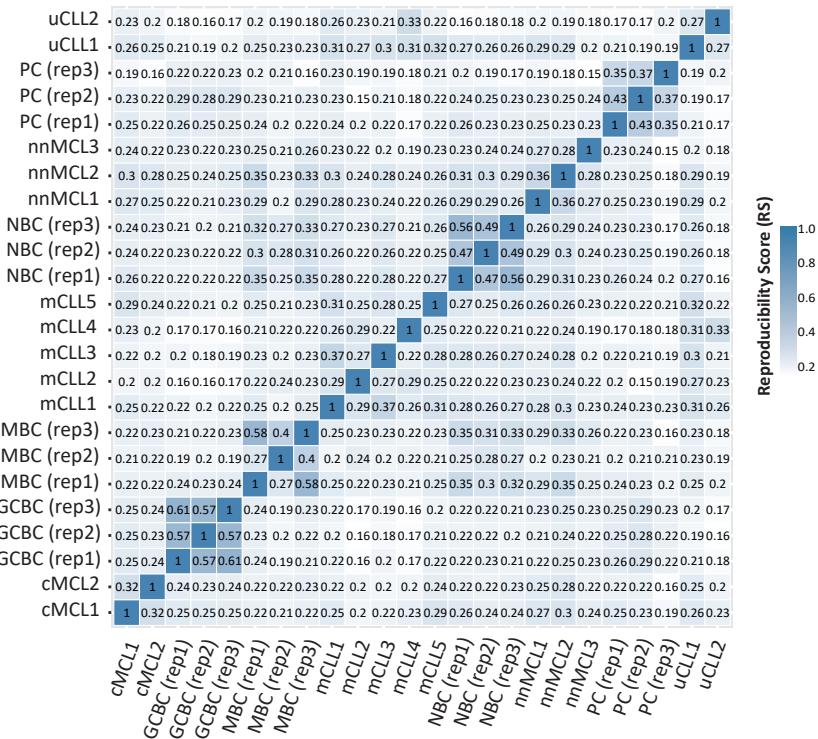
Subcompartments defined
by Rao *et al.*, 2014

A1
A2
B1
B2
B3
B4

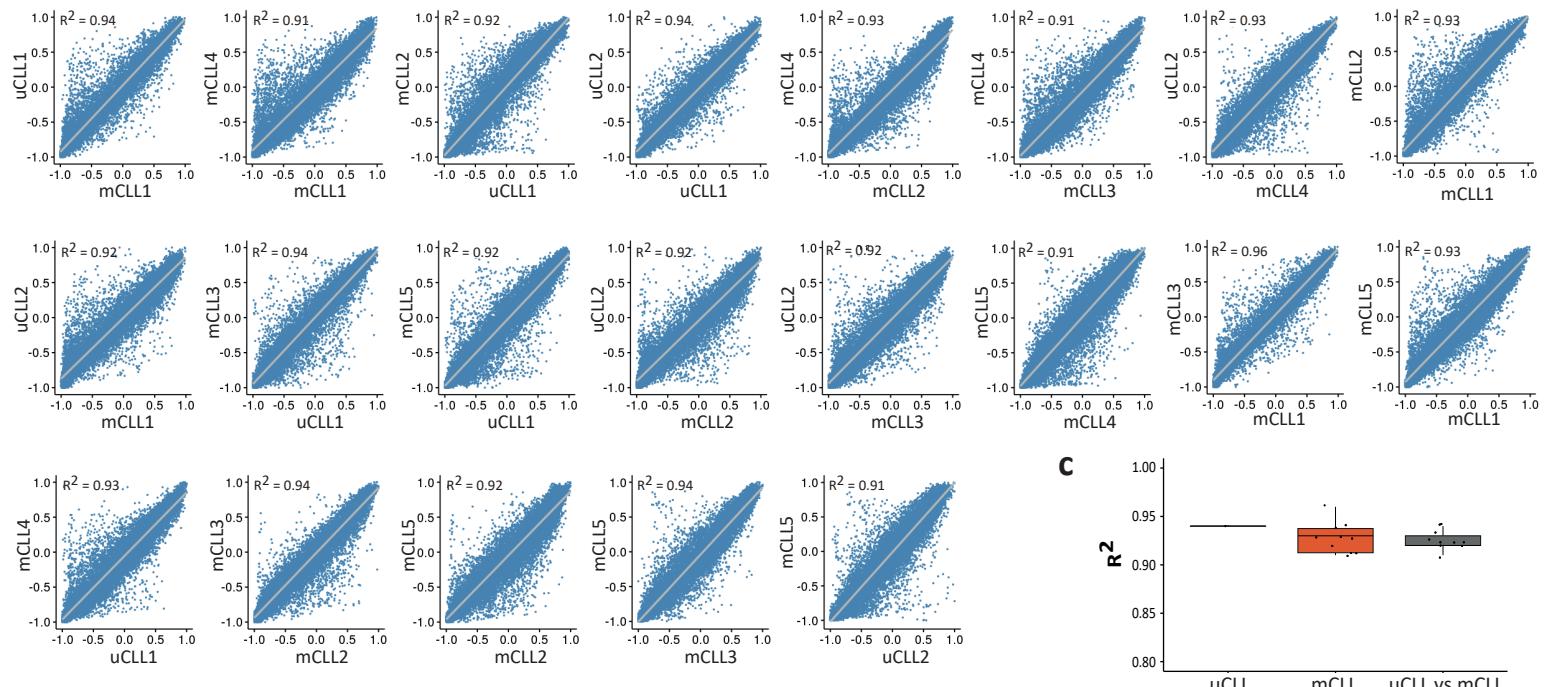
Supplementary Fig. 2. **a** Functional validation of the stable (A-type, I-type and B-type) and dynamic compartments in all B cell subpopulation replicates using eleven different chromatin states. The chromatin state score is normalized by sample and chromatin state. **b** C-score, a method defined by the ratio of contacts between regions within the same compartment (intra-compartment contacts) over the total chromosomal contacts per compartments (intra- and inter-chromosomal interactions). **c/d** C-score distributions at the three defined compartments A-type, I-type and B-type (**c**) and segmenting the I-type compartment onto positive (IA) or negative (IB) eigenvector coefficients (**d**). **e/f** Correlation between the three compartment segmentation and the six subcompartments defined by Rao et al.¹² in normal B cell subpopulations (**e**) and GM12878 (**f**). The percentage (>1%) of each subcompartment is indicated. Sample sizes were for NBC, GCBC, PC and MBC: $n = 3$ merged biologically independent samples. For the boxplots, centerline, box limits and whiskers represent the median, 25th and 75th percentiles and 1.5x interquartile range, respectively.

Supplementary Fig. 3

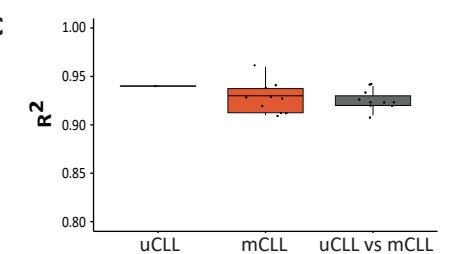
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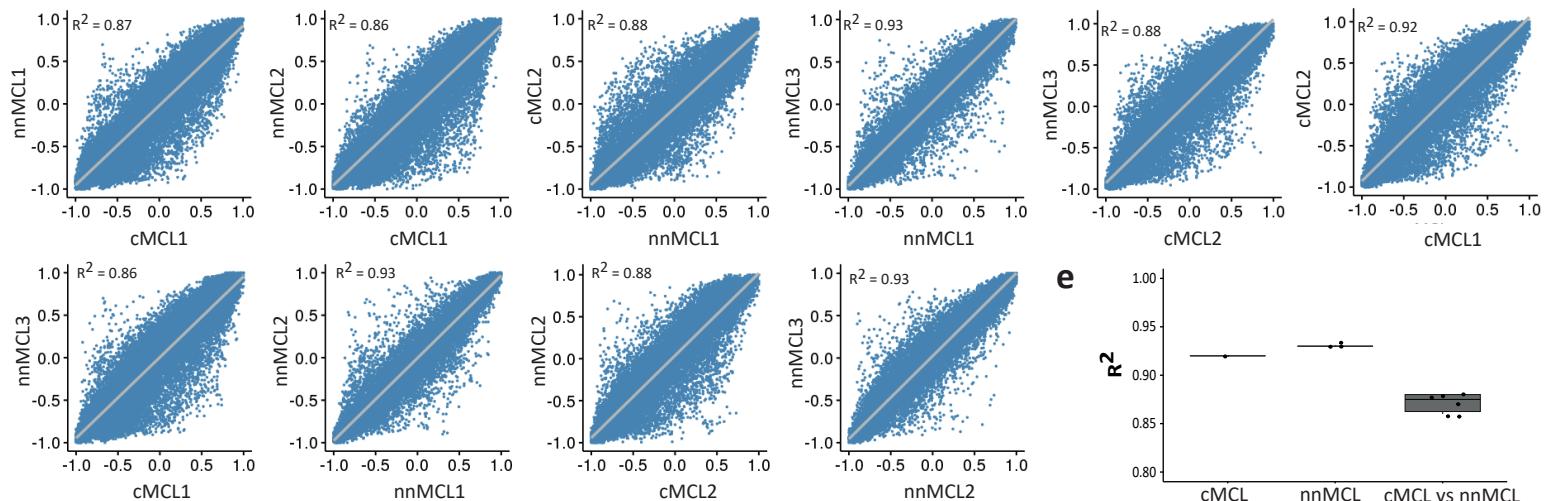
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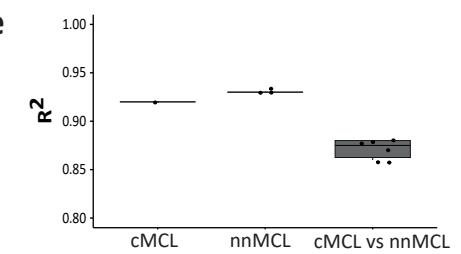
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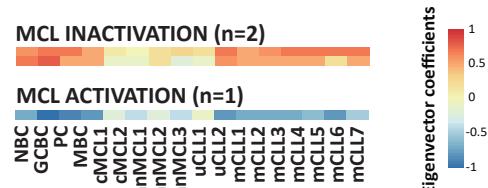
e



Supplementary Fig. 3. **a** Average genome-wide reproducibility score matrix of each B cell subpopulation replicate and B cell neoplasia at 100 kb. The reproducibility score ranging between 0 (totally different matrices) and 1 (identical matrices). **b/d** Genome-wide scatterplots of the first eigenvector showing the correlation between pairs of each B cell malignancy samples at 100 kb resolution. CLL cases are shown in panel **b** and MCL cases in panel **d**. The coefficient of determination (R^2) is indicated. **c/e** Boxplot of the coefficients of determination calculated intra- or inter- each neoplasia subtype. CLL cases are shown in panel **c** and MCL cases in panel **e**. Samples sizes were for uCLL: $n = 2$ biologically independent samples, for mCLL: $n = 5$ biologically independent samples, for cMCL: $n = 2$ biologically independent samples and for nnMCL: $n = 3$ biologically independent samples. For the boxplots, centerline, box limits and whiskers represent the median, 25th and 75th percentiles and 1.5x interquartile range, respectively.

Supplementary Fig. 4

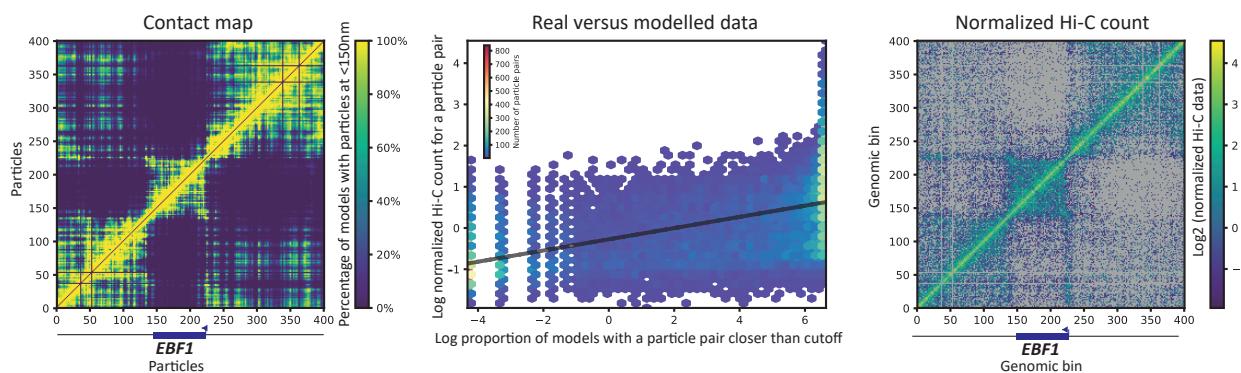
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b

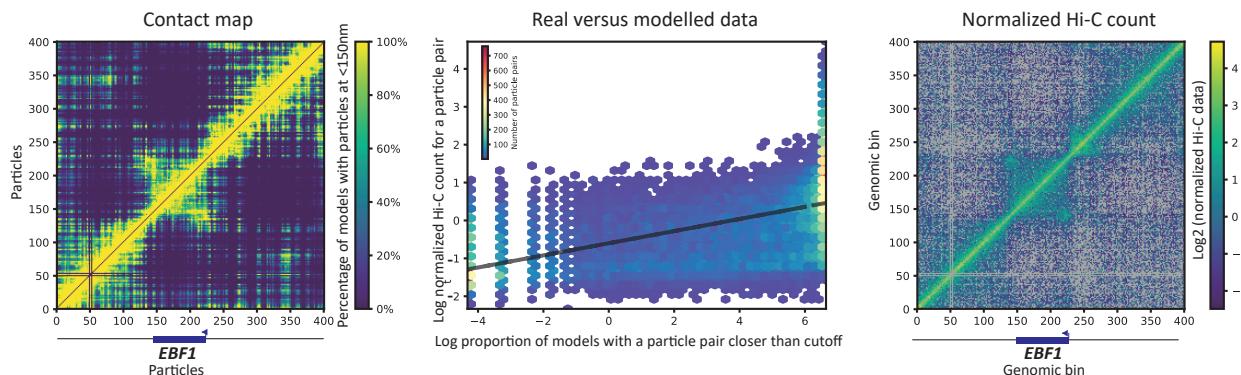
NBC

Correlation between normalized-real and modelled contact maps (cor = 0.6075)

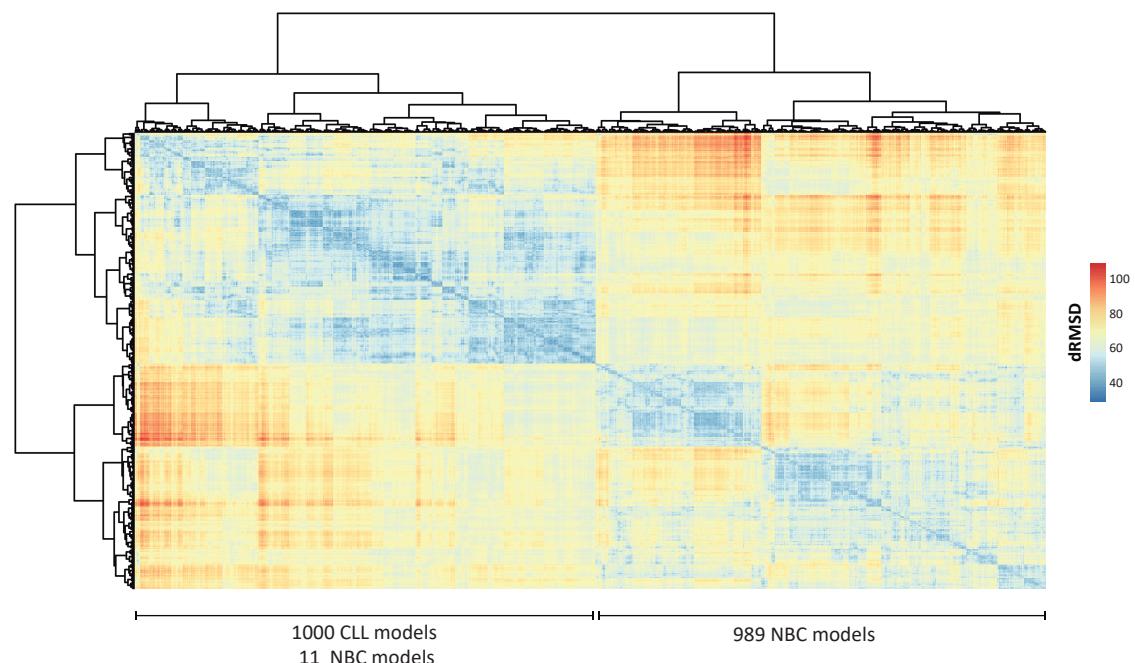


CLL

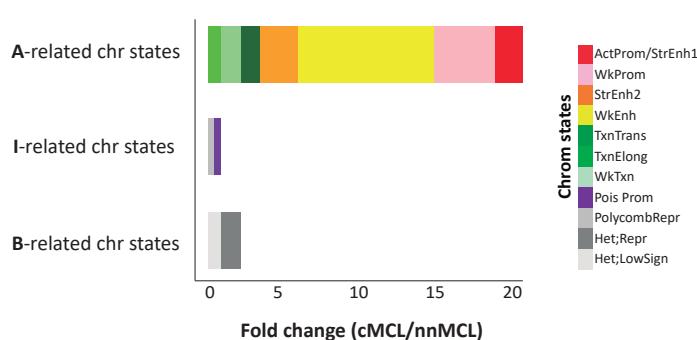
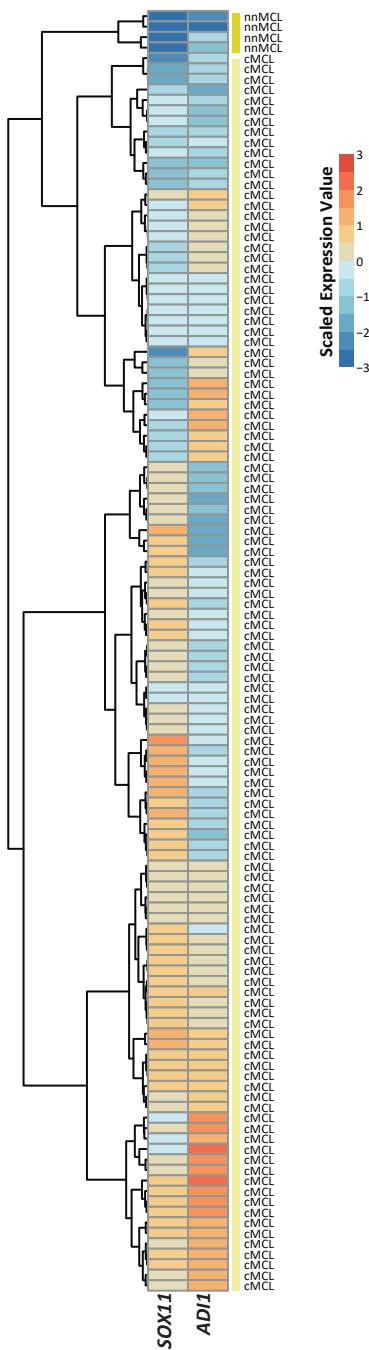
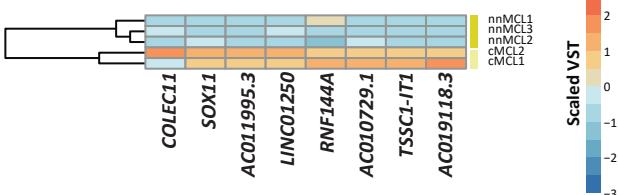
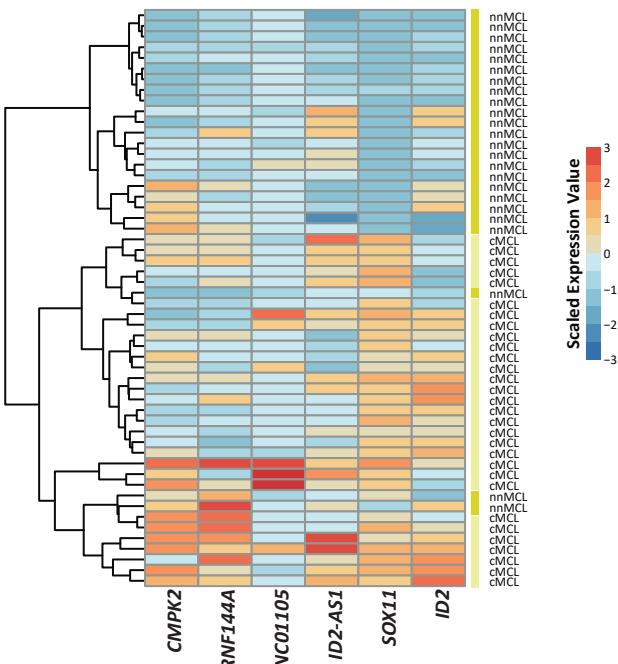
Correlation between normalized-real and modelled contact maps (cor = 0.6315)



c



Supplementary Fig. 4. **a** Heatmap showing the eigenvector coefficients of the MCL specific compartments. *Top*: Losing activation. *Bottom*: Gaining activation. **b** Correlation between normalized Hi-C and modeled contact maps in *EBF1* regulatory landscape. *Top*: NBC. *Bottom*: CLL. *Left*: Contact map computed from the restraint-based model. *Middle*: Scatterplot of Hi-C normalized map vs. modeled contact data with linear regression. *Right*: Normalized Hi-C data. The position of *EBF1* is indicated in blue at the bottom of the matrix plots. **c** Heatmap of the hierarchical clustering of the dRMSD values computed for all the possible pairs of generated models (1,000 in NBC and 1,000 in CLL).

Supplementary Fig. 5**a****d****LLMPP cohort****b****BLUEPRINT cohort****c****Peripheral blood cohort**

Supplementary Fig. 5. **a** Bar graphs represent the fold change between cMCL and nnMCL of each three groups of chromatin states (arranged by their relationship to the A-type, I-type and B-type compartments). **b-d** Heatmaps of the overexpressed genes located within 6.1 Mb region in each cohort: BLUEPRINT (FDR < 0.05) (**b**), peripheral blood (adjusted *p* value < 0.1) (**c**) and/or LLMPP (adjusted *p* value < 0.1) (**d**), with MCL samples classified as cMCL (light yellow) and nnMCL (dark yellow) subtypes. The VST and expression values were scaled by genes. ActProm-StrEnh1 active promoter-strong enhancer 1, WkProm weak promoter, StrEnh2 strong enhancer 2, WkEnh weak enhancer, TxnTrans transcription transition, TxnElong transcription elongation, WkTxn weak transcription, PoisProm poised promoter, PolycombRepr polycomb-repressed, Het;Repr heterochromatin-repressed, Het;LowSign heterochromatin-low signal.