

## SUPPLEMENTARY INFORMATION

### Translational control of enzyme scavenger expression with toxin-induced micro RNA switches

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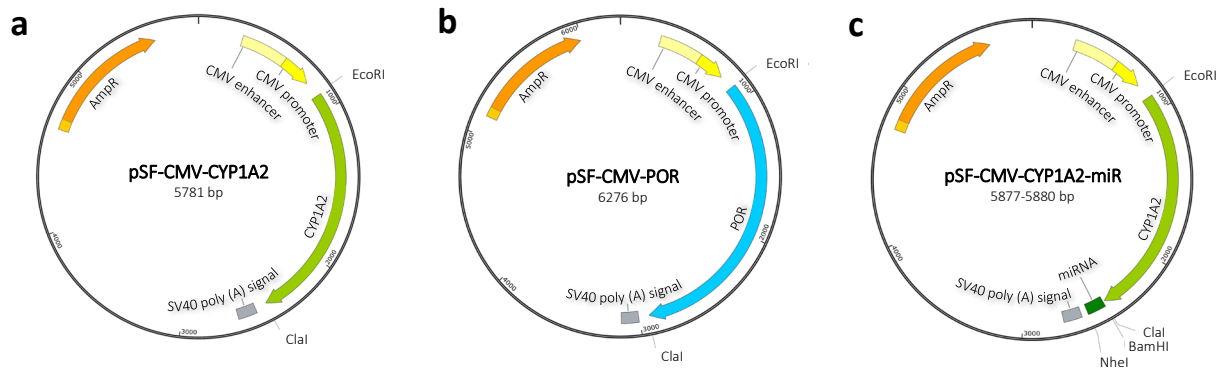
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**Figure S1:** Plasmid maps.

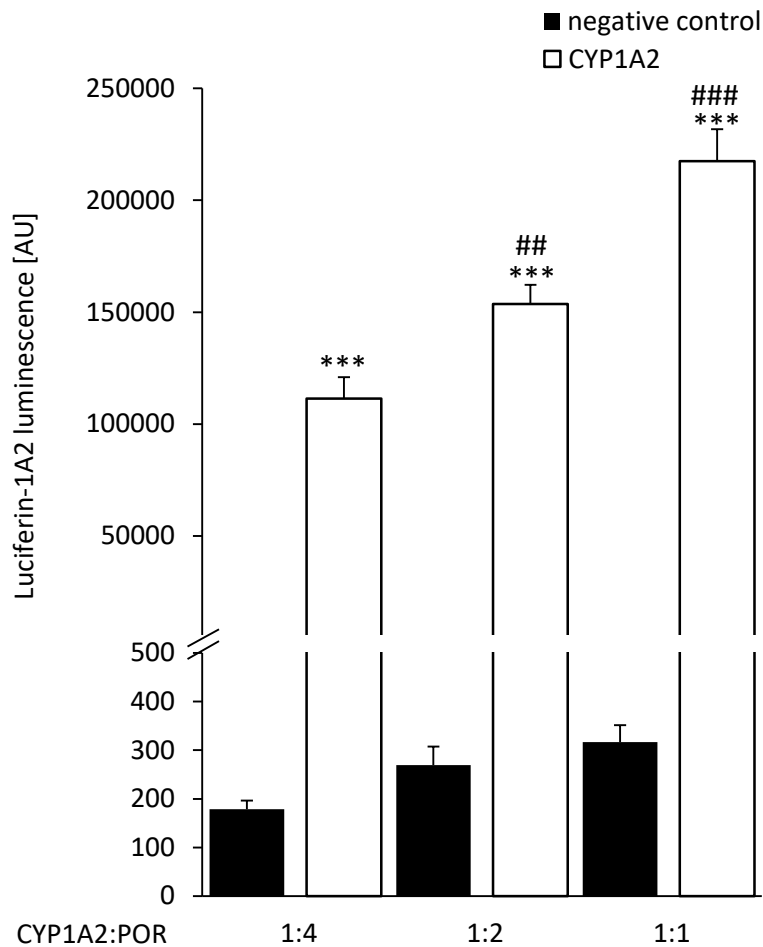
**Figure S2:** Optimization of CYP1A2 enzyme activity by co-expression of POR, the principal redox partner of Cytochrome P450.

**Figure S3:** CYP1A2 luminescence assay calibration curve with recombinant human CYP1A2.

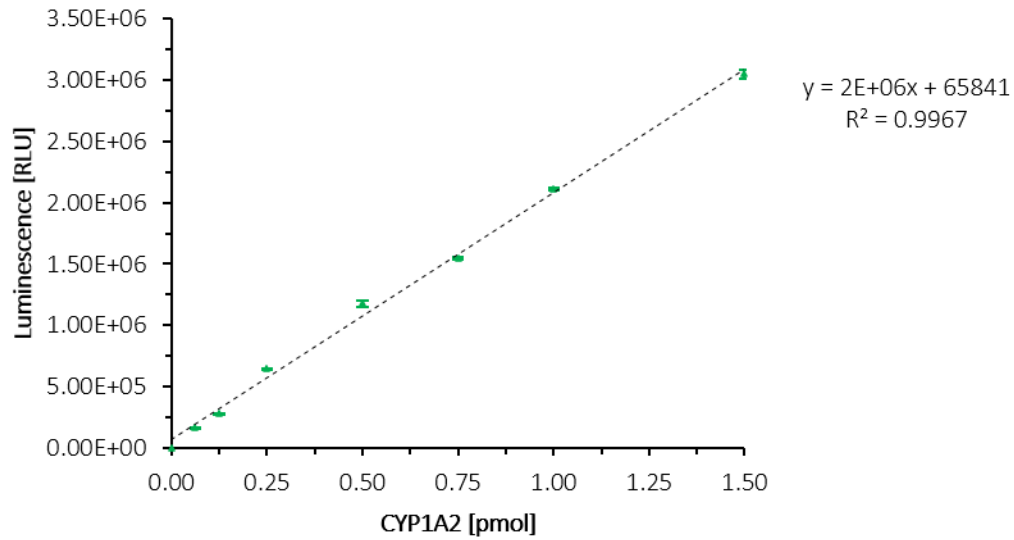
**Figure S4:** Increased CYP1A2 expression in HEK-293 cells transfected with miRNA switches in presence of varying concentrations of theophylline.



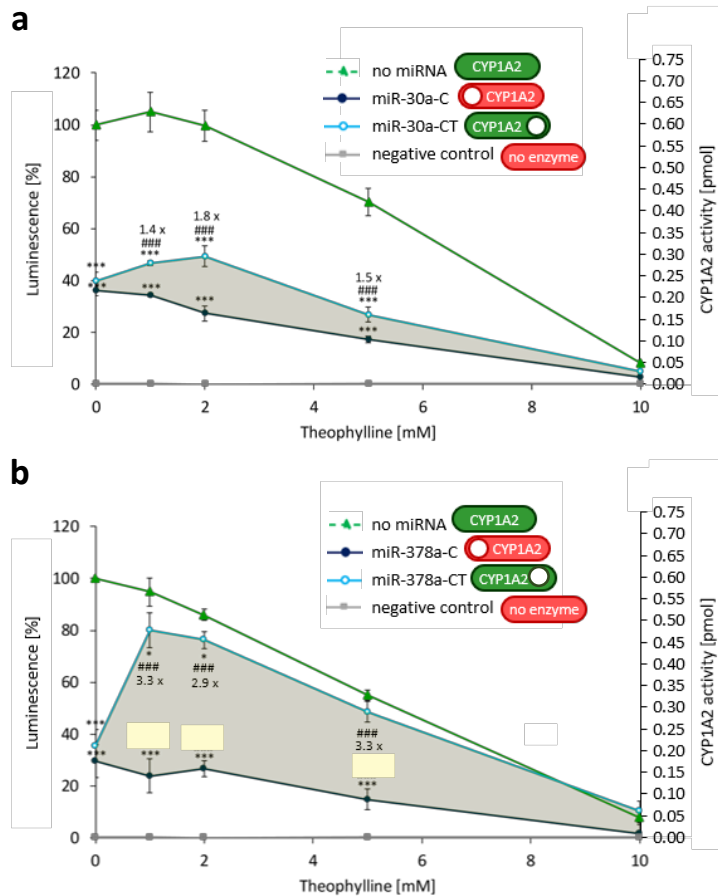
**Figure S1:** Plasmid maps. To obtain (a) pSF-CMV-CYP1A2 and (b) pSF-CMV-POR the mouse gene CYP1A2 or POR, respectively, were cloned into the EcoRI/Clal restriction sites of pSF-CMV-FLuc with a consensus Kozak sequence (GCCACC) immediately upstream of the start codon. CYP1A2 or POR are constitutively expressed from the mammalian Cytomegalovirus (CMV) immediate early promoter, which has the CMV enhancer upstream. (c) To generate the 12 versions of pSF-CMV-CYP1A2-miR, six miRNAs with and six miRNAs without theophylline aptamer in the basal segment of the primary miRNA sequence were cloned into BamHI/NheI restriction sites located in the 3' untranslated region of pSF-CMV-CYP1A2 and upstream of the SV40 poly (A) signal. Plasmid maps were produced using SnapGene software (GSL Biotech; available at <https://www.snapgene.com>). Abbreviations: AmpR, ampicillin resistance; CMV, Cytomegalovirus; CYP1A2, Cytochrome P450 1A2; POR, cytochrome P450 oxidoreductase; miR or miRNA, micro RNA.



**Figure S2:** Optimization of CYP1A2 enzyme activity by co-expression of POR, the principal redox partner of Cytochrome P450. Luciferin-1A2 luminescence levels were determined at different ratios of CYP1A2 to POR, changing the POR amount. Corresponding wells were co-transfected with the fixed amount of 100 ng CYP1A2, which was mixed with either 100 ng POR + 300 ng negative control, 200 ng POR + 200 ng negative control or 400 ng POR. Luminescence was measured 72-hours post transient transfection with indicated constructs (n = 3; \*\*\* P < 0.001 versus negative control; ## P < 0.01 and ### P < 0.001 versus same CYP1A2:POR ratio). Abbreviations: AU, arbitrary units; CYP1A2, Cytochrome P450 1A2; POR, P450 oxidoreductase.



**Figure S3:** CYP1A2 luminescence assay calibration curve with recombinant human CYP1A2. P450-Glo CYP1A2 Assay (Promega, Wisconsin, United States) was performed with a range of CYP1A2 concentrations. Substrate concentration (6  $\mu$ M Luciferin-1A2) and incubation time (10 min, 37°C) were as recommended by the manufacturer. Recombinant human CYP1A2 was purchased (Abcam, Cambridge, United Kingdom).



**Figure S4:** Increased CYP1A2 expression in HEK-293 cells transfected with miR-30a-CT (a) and miR-378a-CT (b) in presence of theophylline, compared to not theophylline-responsive miRNAs (miR30a-C and miR378a-C), and compared to no miRNA control, at varying concentrations of theophylline. Percent luminescence (%) and CYP1A2 activity (pmol) for each construct is shown as mean  $\pm$  standard deviation ( $n = 3$ ). CYP1A2 enzyme activity (pmol) was calculated using a calibration curve (Figure S3) using recombinant CYP1A2. Firefly luciferase is used as negative control (no CYP1A2 enzyme). Legend: \*  $P < 0.05$ , \*\*\*  $P < 0.001$  miR-C and miR-CT compared to no miRNA; ####  $P < 0.001$  miR-CT compared to miR-C). The expected CYP1A2 enzyme expression status for each construct is illustrated with switches: green corresponds to 'ON', red depicts 'OFF' and white circle depicts switching capability.