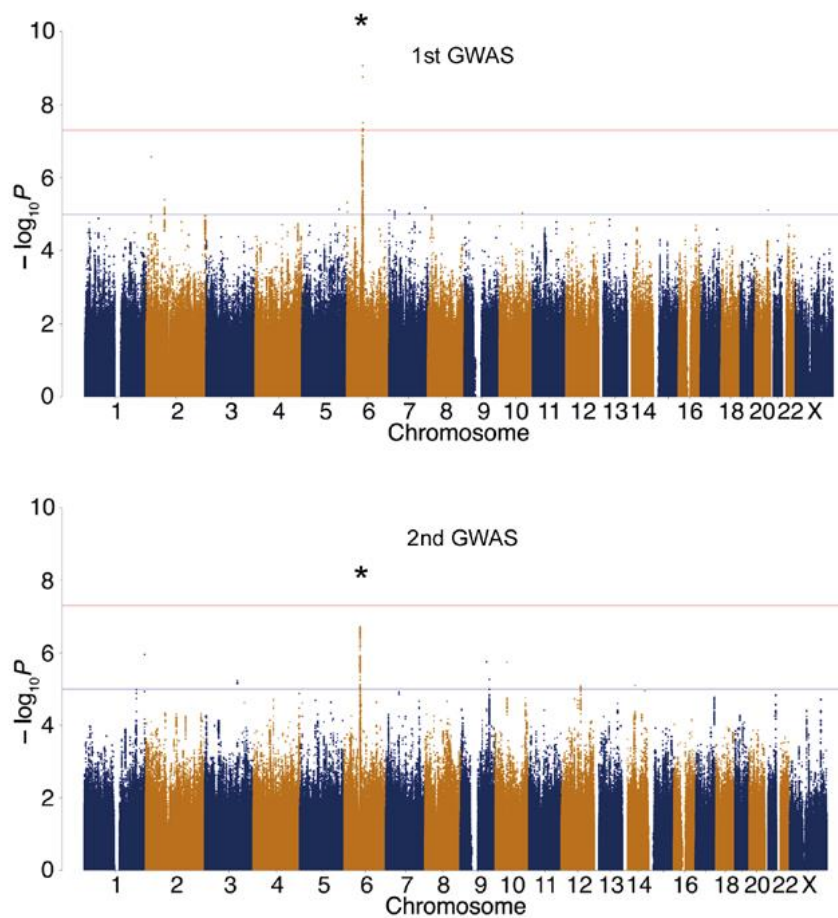


1 **A hypomorphic variant in *EYS* detected by genome-wide association study**  
2 **contributes toward retinitis pigmentosa**  
3 Nishiguchi KM et al.,  
4

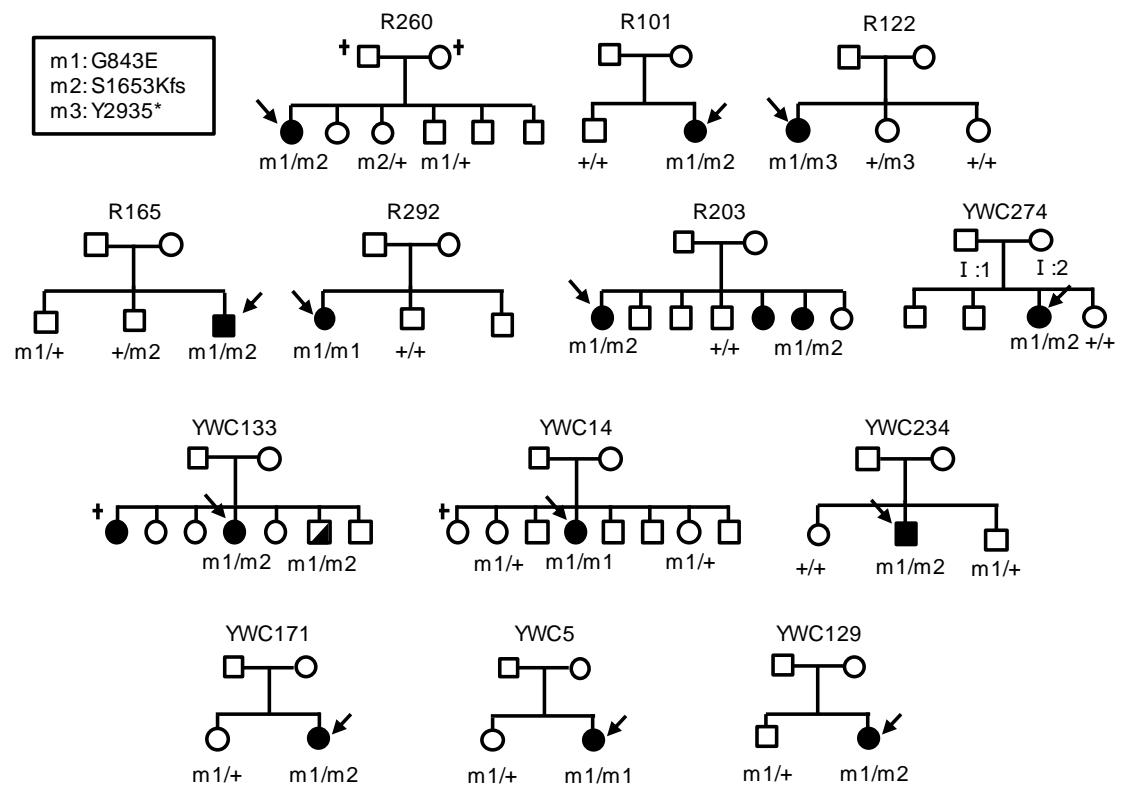
SUPPLEMENTARY INFORMATION



5  
6 **Supplementary Figure 1. Manhattan plots of 1<sup>st</sup> GWAS and 2<sup>nd</sup> GWAS**

7 The *EYS* locus is labeled with an asterisk. GWAS, genome-wide association study. Genome-  
8 wide significance ( $P = 5.0 \times 10^{-8}$ ) and possible significance ( $P = 1.0 \times 10^{-5}$ ) are marked with red  
9 and blue lines, respectively.

10



11

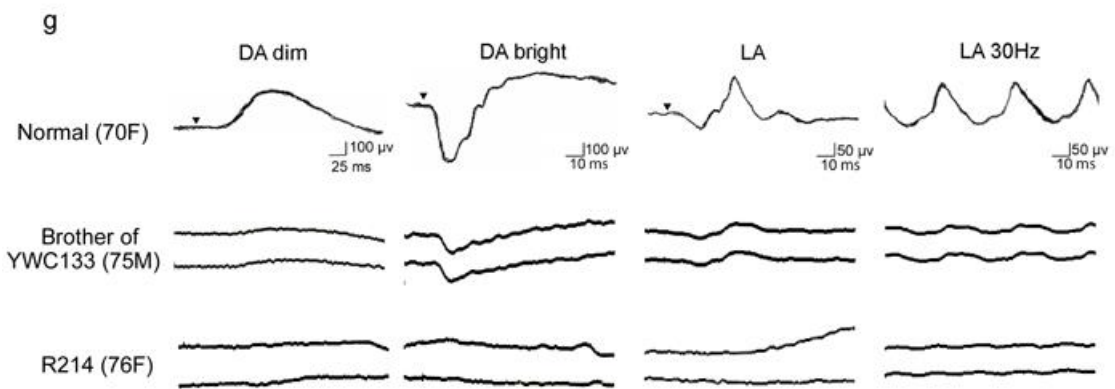
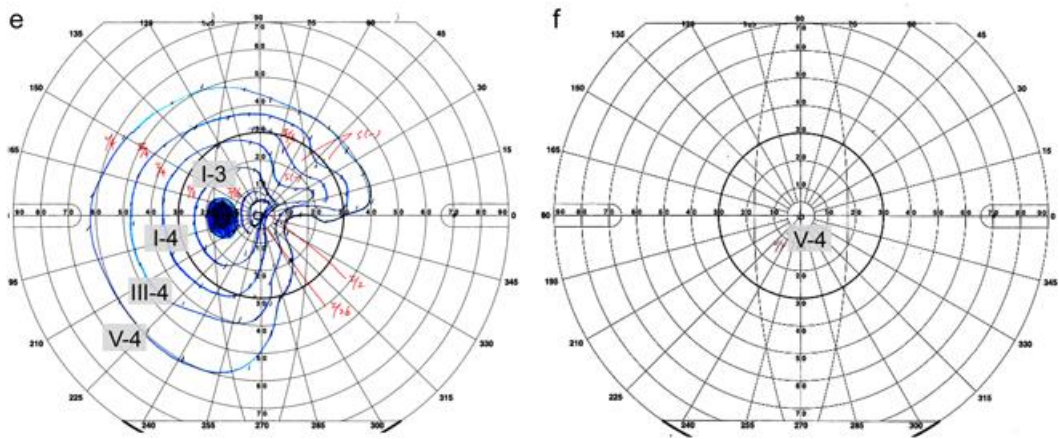
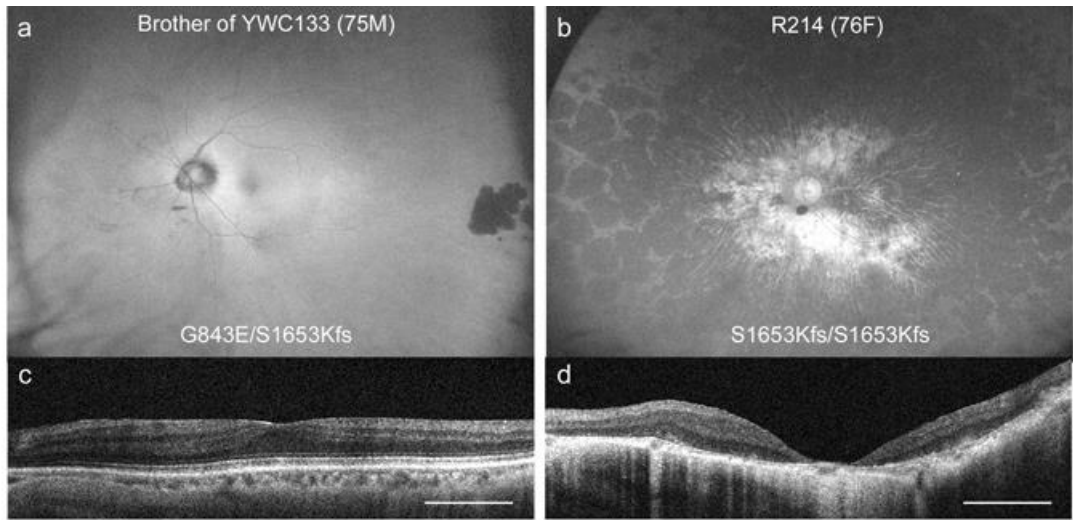
12 **Supplementary Figure 2. Co-segregation of the G843E genotype with phenotype as an**

13 **ARRP allele**

14 The arrows indicate index patients. The half-filled symbol indicates an asymptomatic retinitis

15 pigmentosa patient initially diagnosed as unaffected.

16

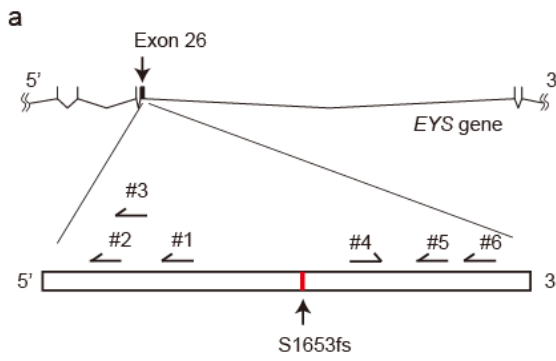


21 tomography (c,d), and Goldman perimetry (e,f) in a 75 years-old asymptomatic brother of  
22 YWC133 with G843E and S1653Kfs (a, c, e) and a 75 years-old female (R214) with  
23 homozygous S1653Kfs. The former had a normal vision (20/20) in both eyes whereas the  
24 latter had severely reduced acuity (20/400 OD and hand motion OS). Note abnormalities in  
25 fundus autofluorescence, optical coherence tomography, and Goldman perimetry are much  
26 milder in the brother of YWC133 compared to R214.

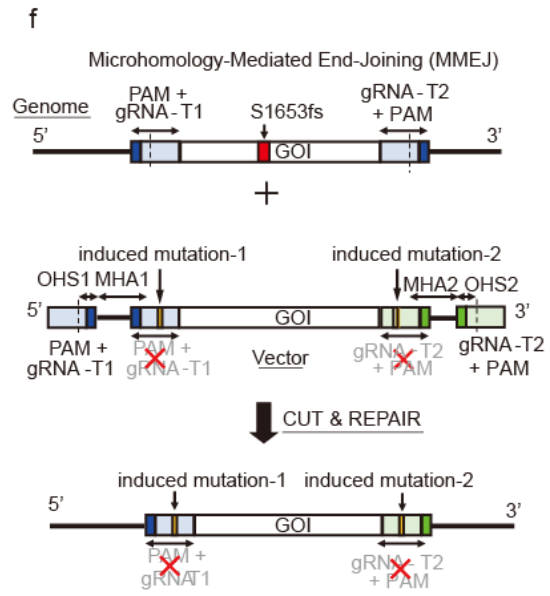
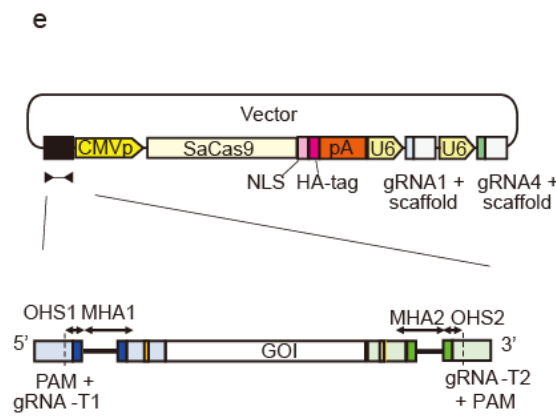
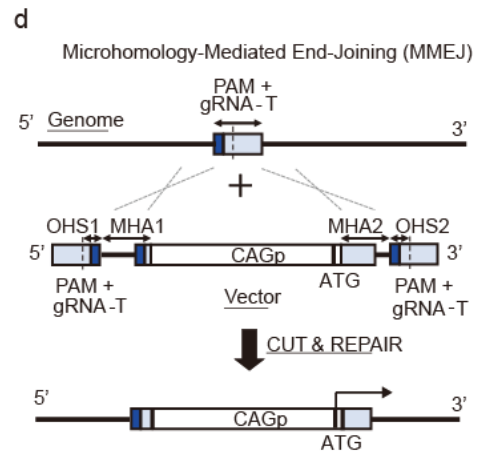
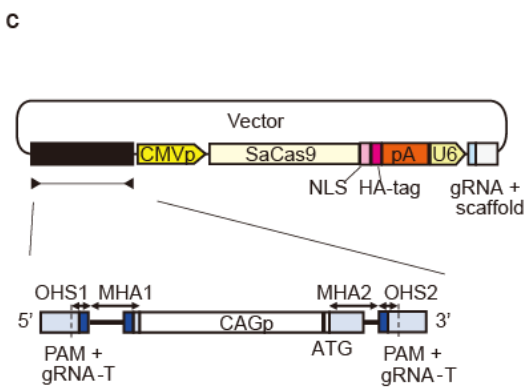
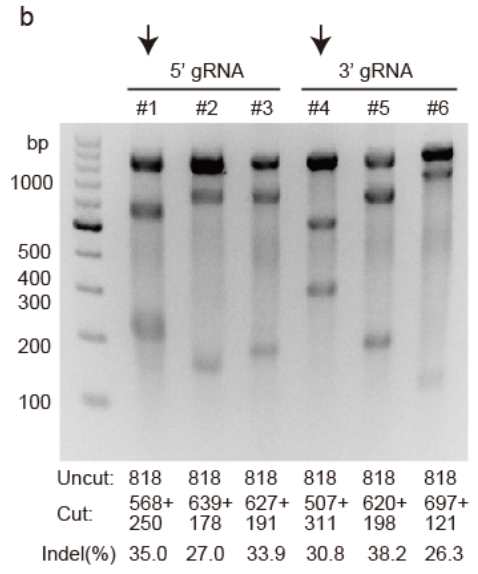
27 **g.** Electroretinogram. Electroretinogram recorded following a standard protocol outlined by  
28 the International Society for Clinical Electrophysiology of Vision showed severely reduced  
29 responses in both patients. DA, dark-adapted; LA, light-adapted.

30 Scale bar: 1.0 mm.

31

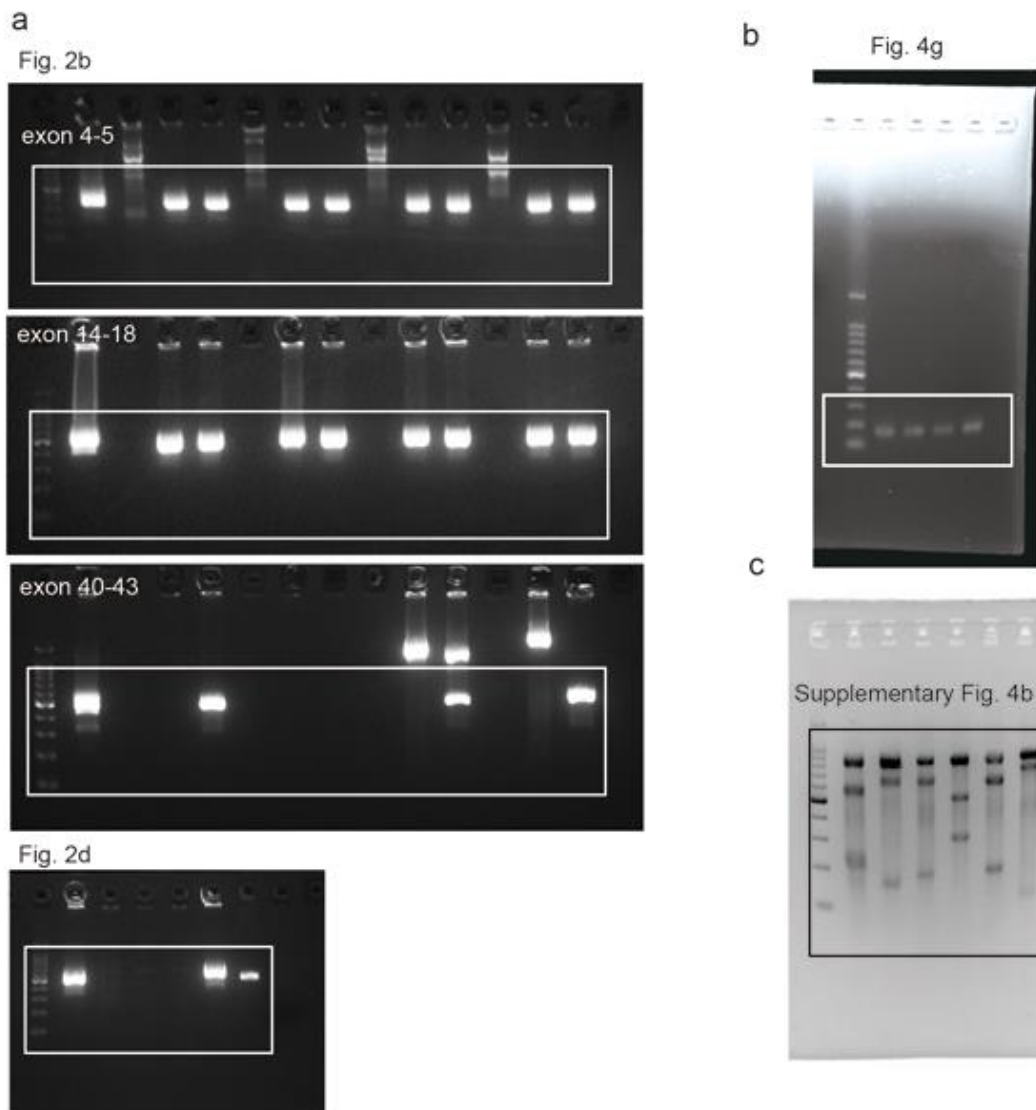


	sequence (21bp+PAM)	locus (GRCh38/hg38)
#1	CATACCAGCTGGCTAATATCGCTGAGT	chr6:64591074-64591101
#2	ACTTGATCTGAGAATTCACGAGAGGT	chr6:64591145-64591172
#3	TTGCTATGCAAACTTGATCTGAGAAT	chr6:64591133-64591160
#4	CAAACATCTCTTCAGACTTGATGAAT	chr6:64590826-64590853
#5	CTCTCTTGATTTAGTACCTCAGTGGGT	chr6:64590704-64590731
#6	CAAATCCAGAGAACTATCACTTGGGT	chr6:64590627-64590654



33 **Supplementary Figure 4. Genome editing strategy to insert CAG promoter or correct**  
34 **S1653Kfs mutation in lymphoblastoid cell lines (LCLs)**

35 **a.** Design of targeting plasmid to insert a CAG promoter immediately upstream of the initiation  
36 codon of *EYS* gene through genome editing. **b.** Illustration of the microhomology-mediated  
37 end-joining CAG promoter knock-in strategy. The DNA sequence of the LCL genome and  
38 plasmid vector were excised at the gRNA target sites (gRNA-T; dotted line) by two gRNAs  
39 and SaCas9. A CAG promoter was inserted into the genome using micro homology arms  
40 (MHA). **c.** Schematic map of gRNA designed inside exon 26 and list of their sequences. **d.**  
41 T7E1 assay for each gRNAs. Expected DNA size and quantified editing efficiency are  
42 displayed under the representative gel image. gRNA-1 and -4 were selected for the  
43 downstream mutation replacement genome editing experiment. **e.** Design of targeting  
44 plasmid to correct S1653Kfs mutation in *EYS* gene through in genome editing. **f.** Illustration  
45 of microhomology-mediated end-joining mutation replacement strategy. Genome of interest  
46 (GOI) with and without the S1653Kfs mutation are excised at the flanking gRNA target sites  
47 (gRNA-T; dotted line) from LCL genome and plasmid vector, respectively, by two gRNAs (1  
48 and 4) and SaCas9. GOI without S1653Kfs mutation is inserted into the genome using MHA.  
49 gRNA-T, guide RNA target; PAM, protospacer adjacent motif; OHS, over-hanging sequence;  
50 NLS, nuclear localizing signal; pA, ploy A; U6, human U6 promoter.



51

52 **Supplementary Figure 5. Uncropped images of the gel electrophoresis presented in**  
 53 **the main and supplementary figures.**

54 **a. Uncropped gels from Figure 2. b. Uncropped gel from Figure 4g. c. Uncropped gel from**  
 55 **supplementary Figure 4b.**

56

57

Gene_name	Genomic location	Chr	Coordinate	Ref	Alt	ToMMo AF	Odds ratio	P-value
NT5C1B-								
RDH14(dist=58909),LINC01376(dist=337974)	intergenic	2	18829755	A	T	0.073	0.128	2.70E-07
EGR4(dist=3282),ALMS1(dist=88775)	intergenic	2	73524111	G	GGT	ND	1.723	3.93E-06
CAMK2A	intronic	5	149610967	AAAT	A	ND	0.558	7.37E-06
EYS	intronic	6	65728469	C	T	ND	4.005	8.61E-10
DNAAF5	intronic	7	791207	ATT	A	ND	1.621	7.71E-06
STK31(dist=421431),NPY(dist=30246)	intergenic	7	24293561	G	A	0.140	1.914	8.27E-06
SEMA3A(dist=231370),LOC101927378(dist=106205)	intergenic	7	84055587	T	TATA G	ND	0.494	9.55E-06
NOC3L	exonic (:c.A1415C:p.E472A)	10	96104665	T	G	0.305	0.632	9.12E-06

**Supplementary Table 1. List of candidate loci at  $P < 1.0 \times 10^{-5}$  after 1st GWAS**

Chr, chromosome; Ref, reference sequence; Alt, alternative sequence; AF, allele frequency.



Gene_name	Genomic location	Chr	Coordinate	Ref	Alt	ToMMo AF	Odds ratio	<i>P</i> -value
CPNE4	intronic	3	131630949	G	GGTTTTA	0.2976	0.480	5.83E-06
EYS	intronic	6	64595973	G	A	0.5567	0.480	1.93E-07

**Supplementary Table 2. List of candidate loci at  $P < 1.0 \times 10^{-5}$  after 2nd GWAS**

Chr, chromosome; Ref, reference sequence; Alt, alternative sequence; AF, allele frequency.

Gene_name	Genomic location	Chr	Coordinate	Ref	Alt	ToMMo AF	Odds ratio	P-value
LINC01767(dist=29496),PLPP3(dist=49285) GS1-	intergenic	1	56911134	A	T	0.0263	1.97	5.73E-06
279B7.1(dist=108903),LINC01350(dist=114438)	intergenic	1	185413074	C	T	0.3195	0.68	6.67E-06
HAAO(dist=134357),LINC01819(dist=100884)	intergenic	2	43154108	G	A	0.3156	0.63	1.90E-06
LINC02233(dist=357406),FSTL5(dist=1248702) STC2	intergenic	4	161056342	C	CA	ND	0.59	7.89E-07
EYS	intronic	5	172750120	C	A	0.3541	1.60	8.30E-06
	intronic	6	65700352	A	G	0.0414	3.95	1.18E-13
C8orf87(dist=113319),LINC00535(dist=66297) XKR4	intergenic	8	94292398	C	A	0.0001	5.18	1.42E-06
SGCZ	intronic	8	56130779	G	T	0.0042	4.00	5.46E-06
LINC02451	intronic	8	14999045	G	C	0.0693	2.12	9.28E-06
	ncRNA_intronic	12	43047088	G	T	0.3846	0.66	2.11E-06
OAS2(dist=19943),DTX1(dist=26191) ITPR2	intergenic	12	113469471	G	A	0.4733	0.71	7.76E-06
	intronic	12	26860830	CA	C	ND	0.65	9.88E-06
RPL28(NM_001136134:c.*1285A>G, NM_000991:c.*1169A>G)	UTR3	19	55900869	A	G	0.8789	0.55	9.43E-06

**Supplementary Table 3. List of candidate loci at  $P < 1.0 \times 10^{-5}$  after meta GWAS**

Chr, chromosome; Ref, reference sequence; Alt, alternative sequence; AF, allele frequency.

$R^2$  values

	G843E (chr6:65622490)	S1653fs (chr6:65300802)	G2186E (chr6:64791763)	Y2935X (chr6:64431122)
Peak 1 (chr6:65700352)	<b>0.6817</b>	0.0133	0.0021	< 0.0001
Peak 2 (chr6:64602534)	< 0.0001	0.0004	< 0.0001	0.0031
Peak 3 (chr6:65320870)	0.0186	<b>0.7750</b>	0.0001	0.0011

$D'$  values

	G843E (chr6:65622490)	S1653fs (chr6:65300802)	G2186E (chr6:64791763)	Y2935X (chr6:64431122)
Peak 1 (chr6:65700352)	<b>0.9131</b>	0.1730	0.1541	nd
Peak 2 (chr6:64602534)	nd	0.1181	0.0021	0.2691
Peak 3 (chr6:65320870)	0.1978	<b>0.9397</b>	1.0000	1.0000

**Supplementary Table 4. Evaluation of linkage disequilibrium of top SNPs for *EYS***

**peaks in retinitis pigmentosa patients**

Primer	Sequence (5'>3')	Use
Ex4-F	GTGGCTGAGTGTTGGGACAC	RT-PCR (Fig. 2)
Ex5-R	ATGGAAACAGACATGTGGTTGA	RT-PCR (Fig. 2)
Ex14-F	GGACATTGATGACTGCATCC	RT-PCR (Fig. 2)
Ex18-R	AGAGATCCAGAAAACCCAGG	RT-PCR (Fig. 2)
Ex40-F	GTTGGCCAGTGTCATGCTTC	RT-PCR (Fig. 2)
Ex43-R	CGCCAAGGTTGTAGCGAAGT	RT-PCR (Fig. 2)
ATGMO	CTCATGTTTGTCTTGGCTCGACTGG	Morpholino
SPMO1	TTGACTTACCCTTAAATCCTGGTG	Morpholino
SPMO2	AAAGTTCCTTCACTGTGAATGGAGC	Morpholino
SPMO3	CAGAGCAGTTGACACCTATGATGTG	Morpholino
Standard control	CCTCTTACCTCAGTTACAATTTATA	Morpholino
Eys201F	ACGTTGACGGATGCTATGAGCAG	RT/qRT-PCR (Fig. 4)
Eys201R	AGCAAGCTCCTTCTTTGCACC	RT/qRT-PCR (Fig. 4)
DrGAPDHF1	TCACACCAAGTGTCAGGACG	RT/qRT-PCR (Fig. 4)
DrGAPDHR1	CGCCTTCTGCCTTAACCTCA	RT/qRT-PCR (Fig. 4)

**Supplementary Table 5 Primers used for RT-PCR**