## **Supplemental Materials**

Molecular Biology of the Cell

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1 Figure S1. Acute ER stress does not inhibit insulin-dependent AKT activation. (A) Serum 2 starvation does not affect activation of XBP1 splicing in WT, ire1a<sup>-/-</sup>, or traf2<sup>-/-</sup> MEFs, 3T3-3 F442A cells, C<sub>2</sub>C<sub>12</sub> cells, or Hep G2 cells. Cells were serum-starved for 18 h, where indicated, 4 and then treated with 1  $\mu$ M thapsigargin for 1 h. (B) 3T3-F442A adipocytes, C<sub>2</sub>C<sub>12</sub> myotubes, 5 and Hep G2 cells were serum-starved for 18 h and treated with the indicated concentrations of 6 thapsigargin, tunicamycin, 1 µg/ml SubAB or SubAA272B during the last 30 min of serum 7 starvation and then stimulated with 100 nM insulin for 15 min, where indicated, in the continued 8 presence of thapsigargin, tunicamycin, SubAB, or SubAA272B. Cell lysates were analysed by 9 Western blotting. (C) Quantification of the phosphorylation of AKT on S473. Bars represent 10 standard errors (n = 6 for unstressed, insulin-stimulated cells and n = 3 for all other samples). 11 p values for comparison of ER-stressed samples and samples not stimulated with 100 nM 12 insulin to samples stimulated with 100 nM insulin were calculated by ordinary two-way 13 ANOVA with Dunnett's multiple comparisons test.

14 Figure S2. Acute ER stress does not inhibit insulin-stimulated phosphorylation of IRS1 at four 15 specific tyrosine phosphorylation sites in 3T3-F442A cells. Tyrosine phosphorylation at (A) 16 Y608, (B) Y628, (C) Y891, and (D) Y935 was analysed by Western blotting. 3T3-F442A cells 17 were serum-starved for 12 h before exposure to 1 µM thapsigargin for 10, 20, or 30 min (left 18 side of figure panels) or to 0.1, 1.0, or 10 µg/ml tunicamycin for 30 min (right side of figure 19 panels), followed by stimulation with the indicated concentrations of insulin for 5 min in the 20 continued presence of thapsigargin or tunicamycin. Bars represent standard errors (n = 3 for 21 phosphorylation at Y935, n = 4 for phosphorylation at Y608, Y628, and Y891). p values for 22 comparison of effects of thapsigargin or tunicamycin within one insulin concentration and for 23 comparison of effects of different insulin concentrations were calculated by ordinary two-way 24 ANOVA with Tukey's multiple comparisons test. Data for effects of thapsigargin on Y628 25 phosphorylation of IRS1 were square root-transformed before statistical analyses.

Figure S3. Acute ER stress does not inhibit insulin-stimulated phosphorylation of IRS1 at four
specific tyrosine phosphorylation sites in Hep G2 cells. Tyrosine phosphorylation at (A) Y612,
(B) Y632, (C) Y896, and (D) Y941 was analysed by Western blotting. Hep G2 cells were

29 serum-starved for 18 h before exposure to 1  $\mu$ M thapsigargin for 10, 20, or 30 min (left side of 30 figure panels) or to 0.1, 1.0, or 10  $\mu$ g/ml tunicamycin for 30 min (right side of figure panels), 31 followed by stimulation with the indicated concentrations of insulin for 5 min in the continued 32 presence of thapsigargin or tunicamycin. Bars represent standard errors (n = 4 for effects of thapsigargin on Y612 phosphorylation, n = 5 for Y632 phosphorylation and effects of 33 34 thapsigargin on Y896 and Y941 phosphorylation, n = 6 for effects of tunicamycin on Y612 and 35 Y896 phosphorylation, and n = 7 for effects of tunicamycin on Y941 phosphorylation). p values 36 for comparison of effects of thapsigargin or tunicamycin within one insulin concentration and 37 for comparison of effects of different insulin concentrations were calculated by ordinary two-38 way ANOVA with Tukey's multiple comparisons test. Data for Y896 and Y941 39 phosphorylation of IRS1 were square root-transformed before statistical analyses.

40 Figure S4. Prolonged ER stress does not affect viability of (A-C)  $C_2C_{12}$  and (D-F) Hep G2 cells.  $C_2C_{12}$  and Hep G2 cells were exposed to the indicated ER stressors for 24 h or 36 h, 41 42 respectively. (A, D) Residual cell staining with crystal violet. (B, E) Determination of cell 43 viability by measurement of the activity of the mitochondrial redox chain with MTT. (C, F) 44 Standardisation of mitochondrial activity measured by reduction of MTT to N-[4,5-45 dimethylthiazol-2-yl]-3, N<sup>\*\*</sup>-diphenylformazan to cell number determined by crystal violet 46 staining. Bars represent standard errors (n = 6). p values for comparison to the 0.1% (v/v) 47 DMSO control (labelled 'DMSO') were calculated by ordinary one-way ANOVA with 48 Dunnett's multiple comparisons test. Abbreviation: PBS/G - PBS + 50% (v/v) glycerol, added 49 to a final concentration of 0.13% (v/v) to culture medium.

**Figure S5.** Depletion of insulin receptors correlates with decreased AKT S473 phosphorylation in ER-stressed Hep G2 cells. (A) Serum-starved Hep G2 cells were treated with the indicated concentrations of thapsigargin, tunicamycin, 1 µg/ml SubAB or 1 µg/ml SubA<sub>A272</sub>B for 18 - 36 h before stimulation with 100 nM insulin for 15 min. Western blots for pS473-AKT, total AKT, the insulin receptor (INSR), and GAPDH. (B) Quantification of the phosphorylation of AKT on S473 ('pS473-AKT') and of the relative abundance of  $\beta$  chains of the insulin receptor (' $\beta$ chains'). Bars represent standard errors (*n* = 6 for cells stimulated with 100 nM insulin, *n* = 3

| 57 | for all other samples). Phosphorylation of AKT at S473 and the relative abundance of $\beta$ chains              |
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| 58 | are expressed relative to unstressed cells that were stimulated with 100 nM insulin for 15 min.                  |
| 59 | p values for comparison of ER-stressed to unstressed samples were calculated using ordinary                      |
| 60 | two-way ANOVA with Dunnett's multiple comparisons test. (C) Correlation of insulin-                              |
| 61 | stimulated AKT phosphorylation with insulin receptor $\beta$ chains ( $r^2 = 0.74$ , two-tailed $p < 0.0001$     |
| 62 | for a significantly non-zero slope, and $p > 0.05$ for deviation from linearity calculated by a runs             |
| 63 | test, $n = 27$ ). Dotted lines represent the 95% confidence interval of the linear regression line.              |
| 64 | The relative phosphorylation of AKT at S473 shown in panel B was plotted against the relative                    |
| 65 | abundance of $\beta$ chains shown in panel B. (D) Quantification of the relative abundance of $\alpha$ - $\beta$ |
| 66 | precursors of the insulin receptor in Hep G2 cells exposed to the indicated concentrations of                    |
| 67 | thapsigargin, tunicamycin, 1 $\mu$ g/ml SubAB, or SubA <sub>A272</sub> B for 36 h. Bars represent standard       |
| 68 | errors ( $n = 6$ for unstressed, insulin-stimulated cells and $n = 3$ for all other samples). $p$ values         |
| 69 | for comparison of ER-stressed samples and samples not stimulated with 100 nM insulin to the                      |
| 70 | sample stimulated with 100 nM insulin were calculated using ordinary one-way ANOVA with                          |
| 71 | Dunnett's multiple comparisons test.   |

72 Figure S6. JNKi VIII and XVI inhibit phosphorylation of c-Jun at S63 after activation of JNKs 73 in a dose-dependent manner. (A) Dose-response curves for JNKi VIII and (B) JNKi XVI. Hep 74 G2 cells were treated with the indicated concentrations of the JNK inhibitors for 30 min and 75 then stimulated with UV light (400 J/m<sup>2</sup>,  $\lambda = 254$  nm) or treated with 5 µg/ml anisomycin (ANI) 76 for 30 min in the continued presence of the corresponding concentrations of JNKi VIII or XVI. 77 After UV exposure, cells were cultured in the presence of the corresponding concentrations of 78 JNKi VIII or XVI for 30 min. Cell lysates were analysed by Western blotting for pS63-c-Jun, 79 c-Jun, and GAPDH. Relative pS63-c-Jun phosphorylation data were fitted to the equation

80 in GraphPad Prism to calculate  $EC_{50}$  values and their 95% confidence intervals (CI).