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Supplementary Materials for

Peptide-protein coassembling matrices as a biomimetic 3D model of ovarian cancer

Clara Louise Hedegaard, Carlos Redondo-Gómez, Bee Yi Tan, Kee Woei Ng, Daniela Loessner, Alvaro Mata*

*Corresponding author. Email: a.mata@nottingham.ac.uk

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Figs. S1 to S10



Fig. S1. Peptide purity of in-house synthesized sequences PA-H, PA-VH, PA-RGDS, and PA-GHK. (A) Reverse-phase high-performance liquid chromatography (RP-HPLC) traces. (B) Electrospray ionization mass spectra (ESI-MS).



Fig. S2. Immunohistochemical (IHC) staining for Keratin type I (KRT32) and Keratin type II (KRT82). (A) IHC staining for KRT82 and KRT32 in a confluent monolayer of epithelial ovarian cancer cells. (B) IHC controls of the monolayers stained with only secondary antibody. Scale bar = $100 \mu m$.



Fig. S3. Hydrogel (PA/KN) characterization at the nanoscale. (A) Circular dichroism (CD) Spectra of pure KN and **PA-VH**, as well as, **PA-VH/KN** (1:1) mixed in equal concentration (0.1 mg/mL in HEPES; right) and equimolar (0.02 mmol/dm³ in HEPES; left). (B) CD-Spectra of pure **PA-RGDS** and **PA-GHK**, and in mixtures with **PA-H** and with **PA-VH** (All PA solutions were pre-prepared at 0.1 mg/mL in HEPES). (C) Transmission electron microscopy (TEM) micrographs of pure peptide **PA-H**, **PA-RGDS**, and **PA-GHK**. (D) Representative TEM micrograph of a mixture of two peptides, in this case **PA-VH/PA-RGDS**, mixed 1:1 at equal concentration (0.1 mg/mL in HEPES). (E) TEM micrograph of pure keratin (KN) prepared in HEPES (0.1 mg/mL). Scale bars = 500 nm.



Fig. S4. Hydrogel (PA/KN) stiffness and characterization at the macroscale. (A) G' (storage modulus) and G'' (loss modulus) were measured at 25°C at a constant frequency of 1 Hz in the 0.01-10% strain during the amplitude sweep, while the oscillation frequency experiments were carried out at a 0.1% fixed strain along 0.1-100 Hz, using a dynamic rheology (DHR-3, TA Instruments, USA). Graphical plot shows data for PA-VH/KN hydrogels after 1 day in HEPES buffer and after 1 and 15 days in cell medium at 37°C. (B) Graphical plot showing PA/KN hydrogel stability in HEPES 10 mM over a period of 3 months. The hydrogel mass was measured after a 4-hour hydrogel formation period, and recorded as 'M₀', with all subsequent timepoints (t) reported as 'M₀-M_t/M₀'. Data shown for PA-VH/KN (n=4) and PA-H/KN (n=4).



Monocultures of ovarian cancer cells PA-VH/ in KN hydrogels and Matrigel. (A) Brightfield image of PA-VH/KN а hydrogel after 21 days of incubation in cell medium, but without cells. Image used as reference to identify cells in PA/KN hydrogel (**B**) Live matrix. (green) / Dead (red) staining of ovarian cancer cells detached from Matrigel and now adhering to the well bottom 21 days after encapsulation. IF image staining for red) and nuclei cancer mono-culture fragmentations at day 21. Scale bars = 100Readout (**C**) μm. values fluorescence from the AlamarBlue assay of monocultures ovarian cancer cells in PA-VH/KN hydrogels. represented Data graphically and in a table with Mean, SD, n (number of and samples). (D) Spheroid size distribution plot, shown as the crosssectional area against number of tumours, in PA-H/KN hydrogels at 1, 7, 14 and 21. IF images from (D) were used to determine the cross-sectional tumour each time point. (E) IF images stained for nuclei (DAPI, blue), of ovarian cancer spheroids growing in PA-H/KN and PA-VH/KN hydrogels. bars = $100 \,\mu m$.

S5.

Fig.



Fig. S6. Hydrogel (PA/KN) compatibility with stromal cells. (A) Live (green) / Dead (red, pseudo colored magenta) fluorescent assay images of HUVECs or MSCs encapsulated in **PA-VH/KN** hydrogels. Live/Dead assay was carried out after 1 and 7 days of incubation. Scale bars = 100 μ m. (**B**) Graphical plot of the cell viability in percentage, as determined by analysis of the Live/Dead assay images shown in (A). (**C**) Brightfield images of HUVECs encapsulated in Matrigel at three time-pointes post-encapsulation; day 1, 2, and 4. (**D**) Scanning electron microscopy (SEM) images of encapsulated HUVECs interacting with the nanofibrous hydrogel network in **PA-VH/KN** hydrogels (left) and with Matrigel (right). (**E**) Brightfield image of a HUVEC network formed on top of a Matrigel at day 8 post cell seeding.



B

	Matrigel			PA-VH/PA-RGDS/KN/FN		
	Mean	SD	n	Mean	SD	n
D1	27694	612	3	19135	1759	3
D4	20610	2140	3	14159	1070	3
D7	29422	6035	3	26070	8246	3
D10	23986	3349	3	27281	4890	3
D14	18536	3127	3	24424	2151	3



Fig. S7. Tricultures of ovarian cancer cells, HUVECs, and MSCs in PA-VH/PA-RGDS/KN/FN hydrogels and Matrigel. (A) Analysis of F-actin network coverage in tri-cultures within PA/KN hydrogels using Angiotool and IF stained images. Red stain is F-actin. Scale bars = $100 \mu m$. (B) Readout fluorescence values from the AlamarBlue assay for treatment study of tricultures in PA-VH/PA-RGDS/KN/FN hydrogels and Matrigel. Data shown as Mean, SD, and n (number of samples) before correcting for background (AB+Media+hydrogel, no cells). Graphical plot in Manuscript Fig. 5B shows values after correcting for the background. (C) Brightfield and Live (green) / Dead (red) assay images of tri-cultures in Matrigel and PA-VH/PA-RGDS/KN and Matrigel. Scale bars = $100 \mu m$.

А



Staining of PA-VH+PA-RGDS/KN hydrogels at day 5



Staining of PA-VH+PA-RGDS/KN hydrogels at day 4



D



Staining of PA-VH/PA-RGDS/KN hydrogels at day 7

CD-31 + DAPI + F-actin

Volume







Fig. S8. Immunofluorescent (IF) staining of tricultures (560 Ovarian cancer cells/µL, 6000 HUVECs/µL, and 600 **MSCs/µL) in PA/KN hydrogels and Matrigel.** (A) IF staining for F-actin (Phalloidin, red, left) and α -SMA (magenta, right) in a PA-VH/PA-RGDS/KN hydrogel at day 5 post encapsulation. (B) IF staining for CD-31 (green) and F-actin (Phalloidin, red pseudo coloured magenta) in a PA-VH/PA-RGDS/KN hydrogel and Matrigel, at day 4 post encapsulation. (C) Still image sequence showing IF staining for CD-31 (green), nuclei (DAPI, blue), and F-actin (red) in a PA-VH/PA-RGDS/KN hydrogel at day 7 post encapsulation. Still images taken from 3D-reconstructions of stacked IF imaging. Far right, a volume reconstruction based on the IF images showing the spherical tumour formations with cell-cell networks between adjacent tumours. (D) Fluorescent and brightfield image of HUVECs tagged with green cell-tracker encapsulated in a PA/KN hydrogel and Matrigel at day 4 post encapsulation. Merged brightfield and fluorescent image show HUVECs spreading. (E) IF staining for nuclei (DAPI, blue), F-actin (Phalloidin, red), and HUVECs (green, cell-tracker dye) at day 14 post-encapsulation in a tri-culture PA-VH/PA-RGDS/KN hydrogel.



Fig. S9. Treatment study with cytotoxic drugs and inhibitor (I). (A) IF and brightfield image from day 1 post encapsulation, and prior to drug treatment of ovarian cancer cells in **PA-VH/KN** hydrogel. (B) Metabolic activity of ovarian cancer cells in **PA-VH/KN** hydrogels (values normalised to day 1), measured using an AlamarBlue assay. Values shown for the control (no treatment) and GM6001 (treatment with inhibitor) at three timepoints, day 1, 7, and 14 post cell-encapsulation. (C) Brightfield images of ovarian cancer cells in Matrigel at day 14 for four treatment conditions; control, GM6001, Carboplatin, and Paclitaxel. (D) Metabolic activity of a triculture (Ovarian cancer cells, HUVECs, and MSCs) in a **PA-H/PA-RGDS/KN/FN** hydrogel (values normalised to day 1), measured using an AlamarBlue assay. Values shown for four conditions; control (no treatment), GM6001 (treatment with inhibitor), TXL (treatment with Paclitaxel) and PLT (treatment with Carboplatin), at three timepoints, day 1, 7 and 14 post cell-encapsulation.



Fig. S10. Treatment study with cytotoxic drugs and inhibitor (II). (A) Readout fluorescence values from the AlamarBlue assay for treatment study of monocultures in **PA-VH/KN** hydrogels. Data represented graphically and in a table with Mean, SD, and n (number of samples). (B) Readout fluorescence values from the AlamarBlue assay for treatment study of monocultures in Matrigel. (C) Readout fluorescence values from the AlamarBlue assay for treatment study of tricultures in **PA-H/KN** hydrogels.