

Figure S1

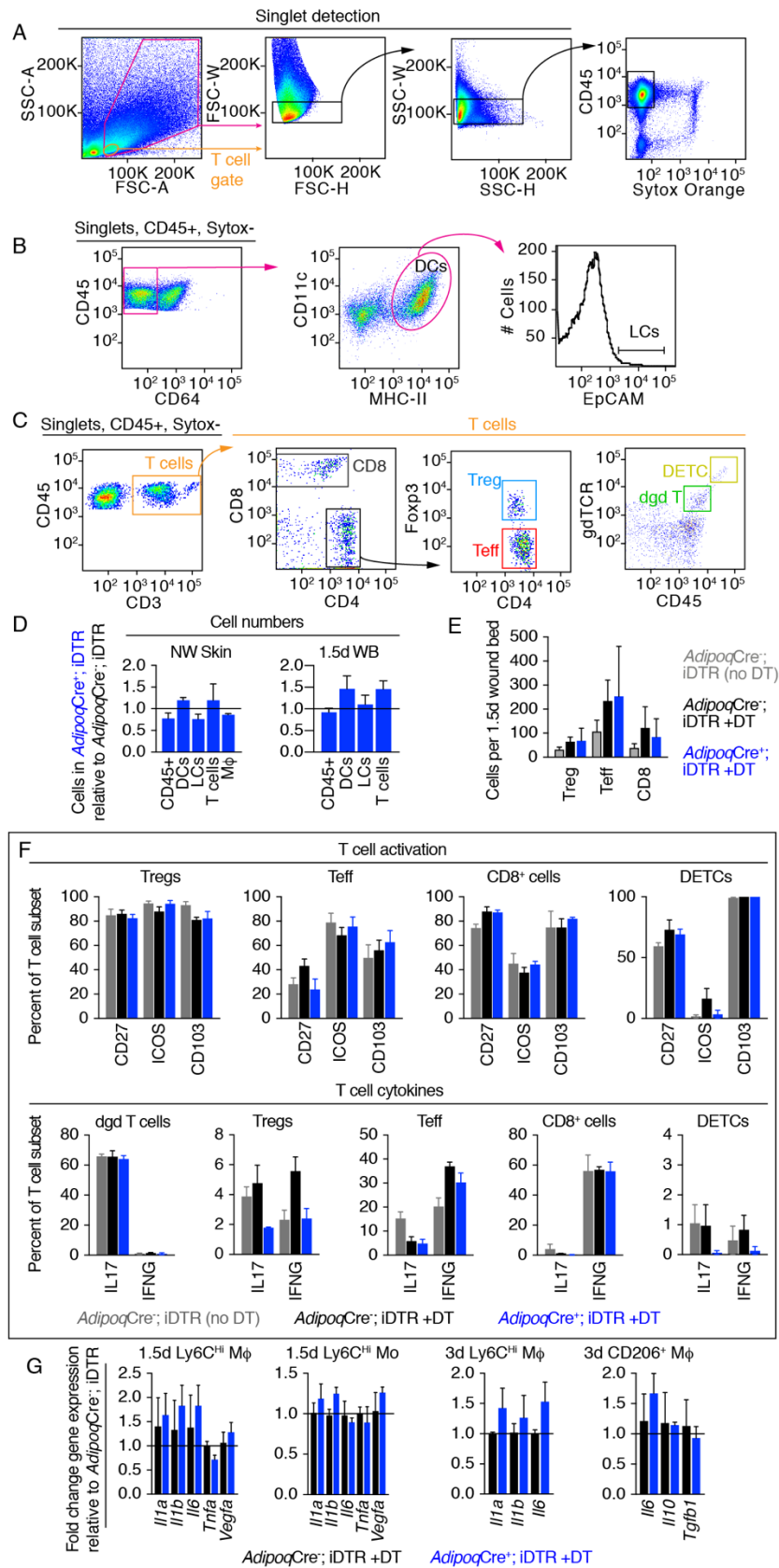


Figure S1. Immune cell characterization in wound beds from dermal adipocyte depleted mice. Related to Figure 1.

(A-C) Flow cytometry plots to detect non-dead CD45⁺ immune cell singlets (A), dendritic cells (DC) and Langerhans cells (LC) (B), and different T cell subsets (C). (D) Quantification of immune cell subsets in nonwounded (NW) skin and 1.5-day wound beds from *Adipoq*Cre⁺; iDTR mice relative to the same *Adipoq*Cre⁻; iDTR mice ($n = 3-4$ mice for each condition and time point). (E-F) Quantification of T cell subsets (E) and the percentage of each subset that is positive for different activation/phenotype markers (F) in 1.5-day wound beds ($n = 3-5$ mice for each condition). (G) Quantitative real-time PCR of inflammatory gene expression in macrophages and monocytes from day 1.5 and 3 wound beds of *Adipoq*Cre⁺; iDTR mice relative to time point matched wound beds of *Adipoq*Cre⁻; iDTR mice ($n = 3$ mice for each time point). Error bars indicate mean \pm SEM. M ϕ , macrophage; Mo, monocyte; DETC, dendritic epidermal T cell; dgd T, dermal gamma-delta T cell; WB, wound bed; DT, diphtheria toxin.

Figure S2

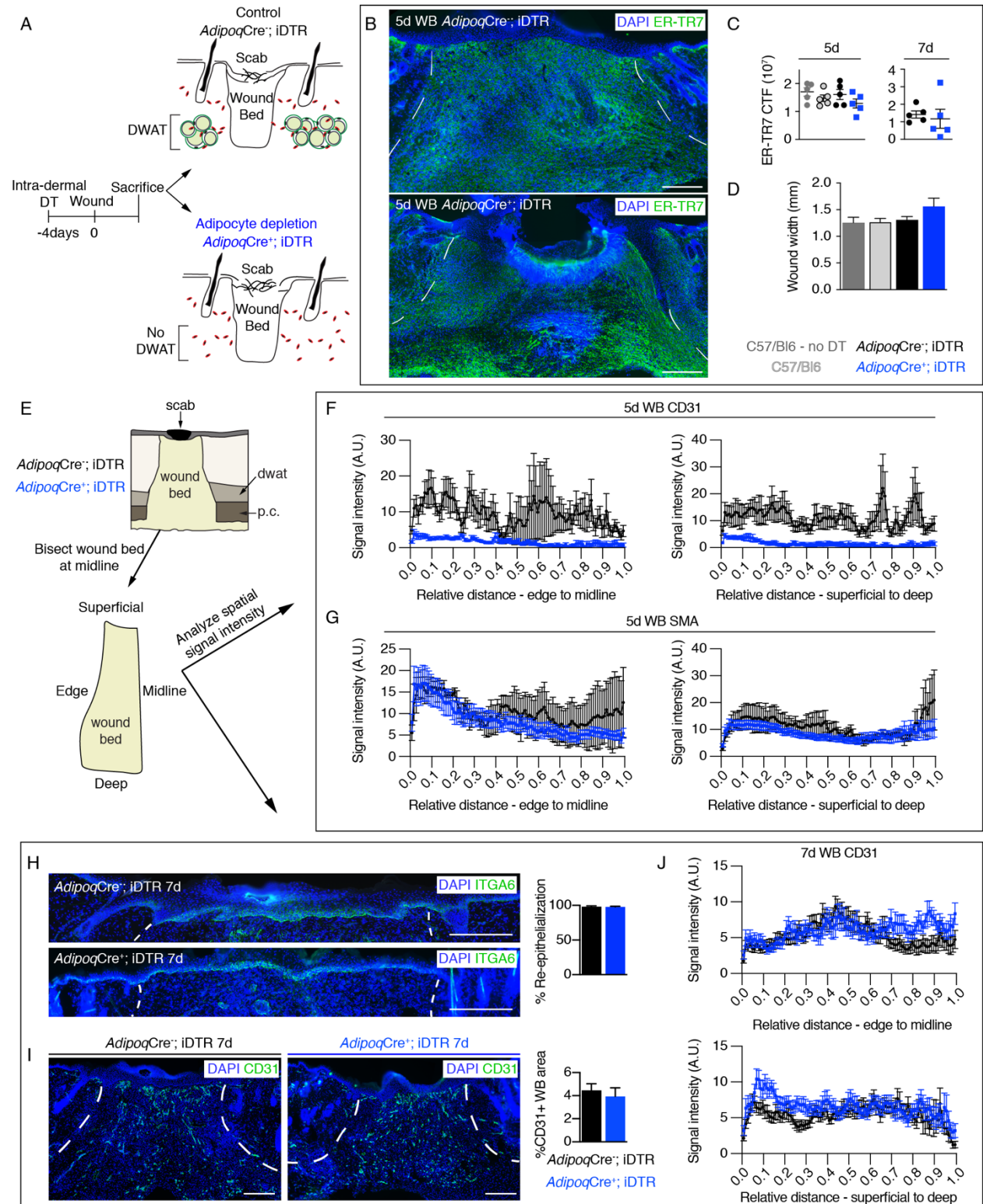


Figure S2. Delayed revascularization and re-epithelialization are corrected in wound beds from dermal adipocyte depleted mice. Related to Figure 1.

(A) Strategy to ablate dermal adipocytes and schematic of skin following dermal adipocyte depletion. (B-D) Composite images from ER-TR7 immunostained sections (B) and quantification of corrected total fluorescence (fibroblast repopulation) in wound beds (C) and calculated wound width (D) ($n \geq 5$ mice for each condition). (E) Pipeline for processing immunostained tissue sections to quantify the spatial distribution of fluorescent signal. (F-G) Spatial distribution of CD31 (F) and SMA (G) signal in 5-day wound beds ($n \geq 6$ mice for each condition). (H) Composite ITGA6 immunostained images and quantification of re-epithelialization in 7-day wound beds. (I-J) Composite CD31 immunostained images (I) and quantification of the total CD31+ area in wound beds (I) and spatial distribution of CD31 (J) in wound beds 7 days after injury ($n = 5$ mice for each condition). White lines delineate wound edges. All scale bars, 250 μ m. Error bars indicate mean \pm SEM.

Figure S3

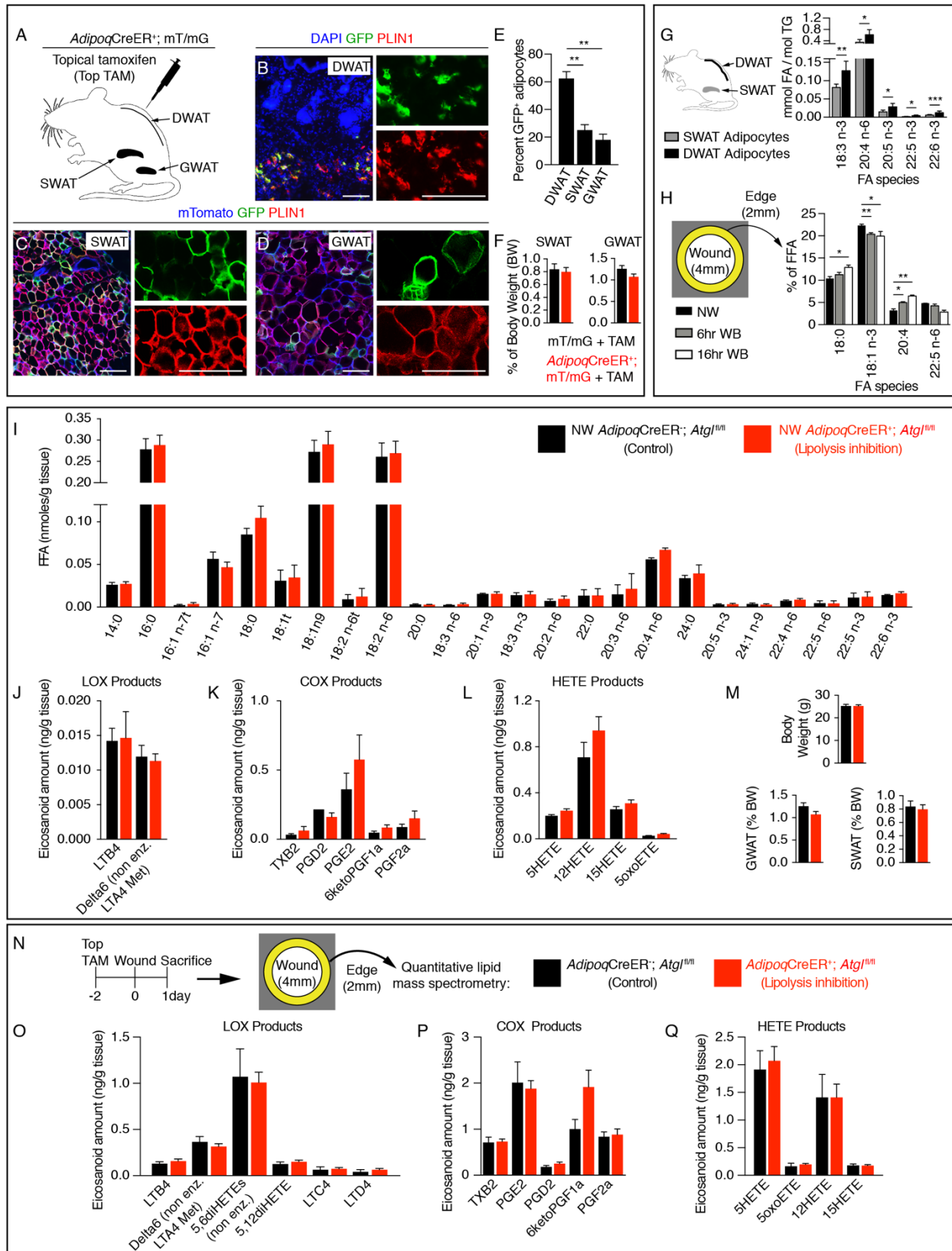


Figure S3. Dermal adipocyte-derived lipids are not required for eicosanoid production during early inflammation. Related to Figure 2.

(A) Schematic of topical tamoxifen administration and location of adipose depots analyzed in *Adipoq*CreER; mT/mG mice. (B-D) GFP and PLIN1 immunostained sections from dermal (DWAT), subcutaneous (SWAT) and gonadal (GWAT) white adipose tissue. Scale bars, 100 μ m. (E) Quantification of the number of PLIN1+ cells that are GFP+ in *Adipoq*CreER; mT/mG mice receiving topical tamoxifen onto back skin ($n = 4$ mice). (F) Quantification of the percentage of total mouse weight that is SWAT and GWAT ($n = 4-5$ mice for each condition). (G) Quantitative lipid mass spectrometry of fatty acid species stored as triglycerides within adipocytes from dermal white adipose tissue (DWAT) and subcutaneous white adipose tissue (SWAT) ($n = 6$ mice for each depot). (H) Quantification of inflammatory free fatty acids at the wound edge following injury ($n = 3$ mice for each time point). (I-L) Quantitative lipid mass spectrometry of free fatty acids (I) and various eicosanoids (J-L) in uninjured skin from *Adipoq*CreER-; *Atgl*^{fl/fl} and *Adipoq*CreER+; *Atgl*^{fl/fl} mice ($n = 4$ mice for each condition). (M) Quantification of total body weight and the percentage of body weight that is GWAT and SWAT from uninjured *Adipoq*CreER-; *Atgl*^{fl/fl} and *Adipoq*CreER+; *Atgl*^{fl/fl} mice ($n \geq 5$ mice for each condition). (N-Q) Experimental paradigm (N) and quantitative lipid mass spectrometry various eicosanoids (O-Q) in the tissue surrounding wound beds 24 hours after injury ($n = 4-5$ mice for each time point). Error bars indicate mean \pm SEM. *, $p < 0.05$; **, $p < 0.01$; ***, $p < 0.001$.

Figure S4

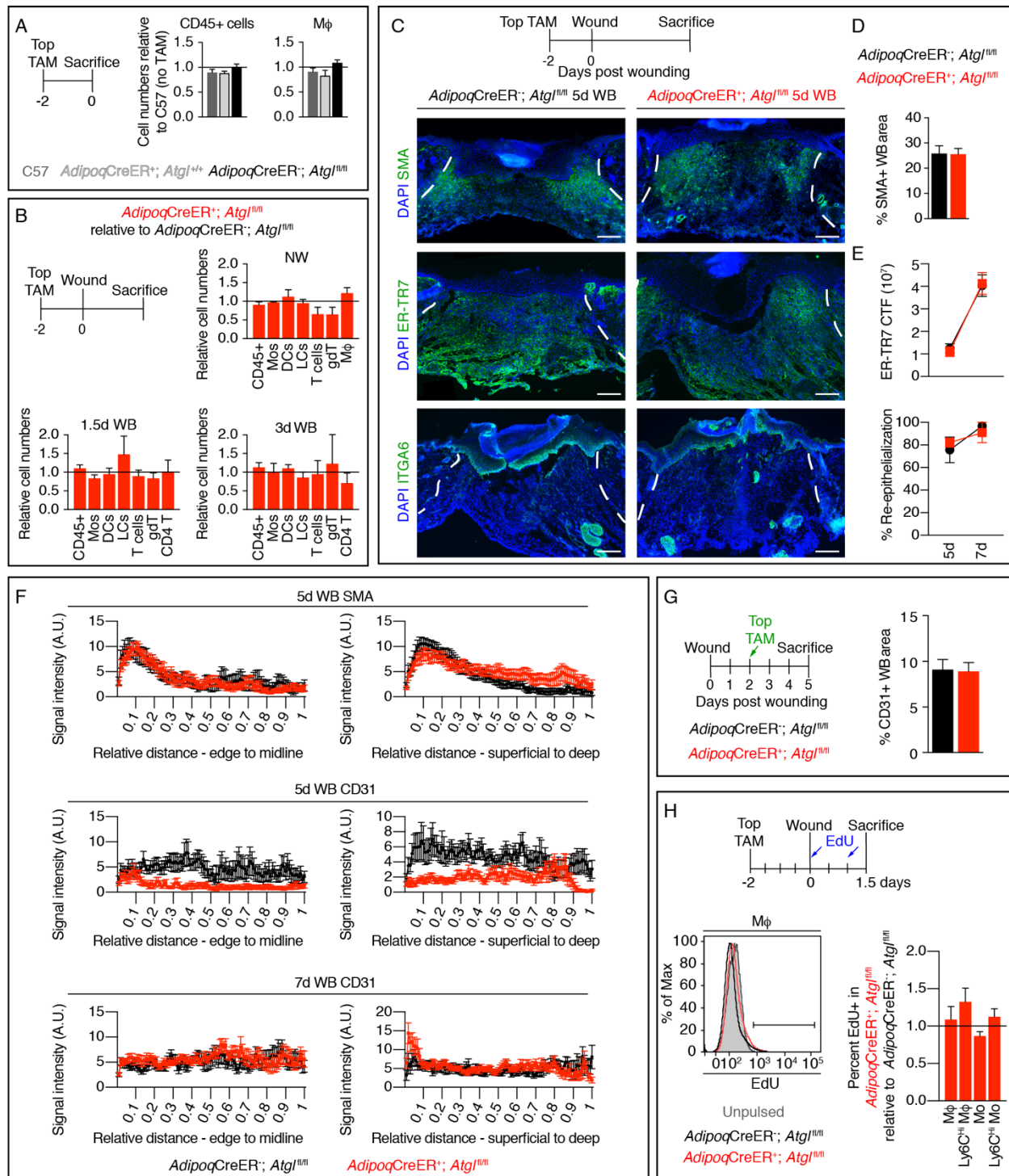


Figure S4. Fibroblast repopulation and re-epithelialization are not delayed when dermal adipocyte lipolysis is inhibited. Related to Figure 3.

(A) Normalized quantification of immune cells and macrophages in uninjured skin treated with tamoxifen, relative to untreated C57 skin ($n = 3-4$ mice for each condition). (B) Normalized quantification of immune cell subsets in non-wounded skin (NW) ($n = 6$ mice for each condition), and wound beds 1.5 days ($n = 6$ mice for each condition) and 3 days ($n = 4$ mice for each condition) after injury. (C) Composite images from tissue sections immunostained for SMA, ER-TR7 and ITGA6 in day 5 wound beds. Scar bar, 250 μ m. White lines delineate wound edges. (D) Quantification of SMA in 5 day wound beds and (E) Quantification of ER-TR7 intensity and ITGA6 in 5- and 7-day wound beds of control (*AdipoqCreER-; Atg^{fl/fl}*) and dermal adipocyte lipolysis inhibited (*AdipoqCreER+; Atg^{fl/fl}*) mice ($n = 6$ mice for 5 day wounds; $n = 3$ mice for 7 day wounds). (F) Mean relative intensity of SMA in 5-day wound beds and CD31 in 5- and 7-day wound beds of *AdipoqCreER-; Atg^{fl/fl}* and *AdipoqCreER+; Atg^{fl/fl}* mice ($n \geq 6$ mice for each condition and time point). (G) Quantification of CD31 immunostained tissue sections from day 5 wound beds or mice treated with tamoxifen 2 days after injury ($n \geq 5$ mice for each condition). (H) Schematic of tamoxifen and EdU treatment. Flow cytometry plot and quantification of EdU-incorporating macrophages and monocytes in 1.5-day wound beds of *AdipoqCreER-; Atg^{fl/fl}* and *AdipoqCreER+; Atg^{fl/fl}* mice ($n = 4$ mice for each condition). Error bars indicate mean \pm SEM. M ϕ , macrophage; Mo, monocyte; DC, dendritic cell; LC, Langerhans cell; gd T, gamma-delta T cell; NW, non-wounded; WB, wound bed.

Figure S5

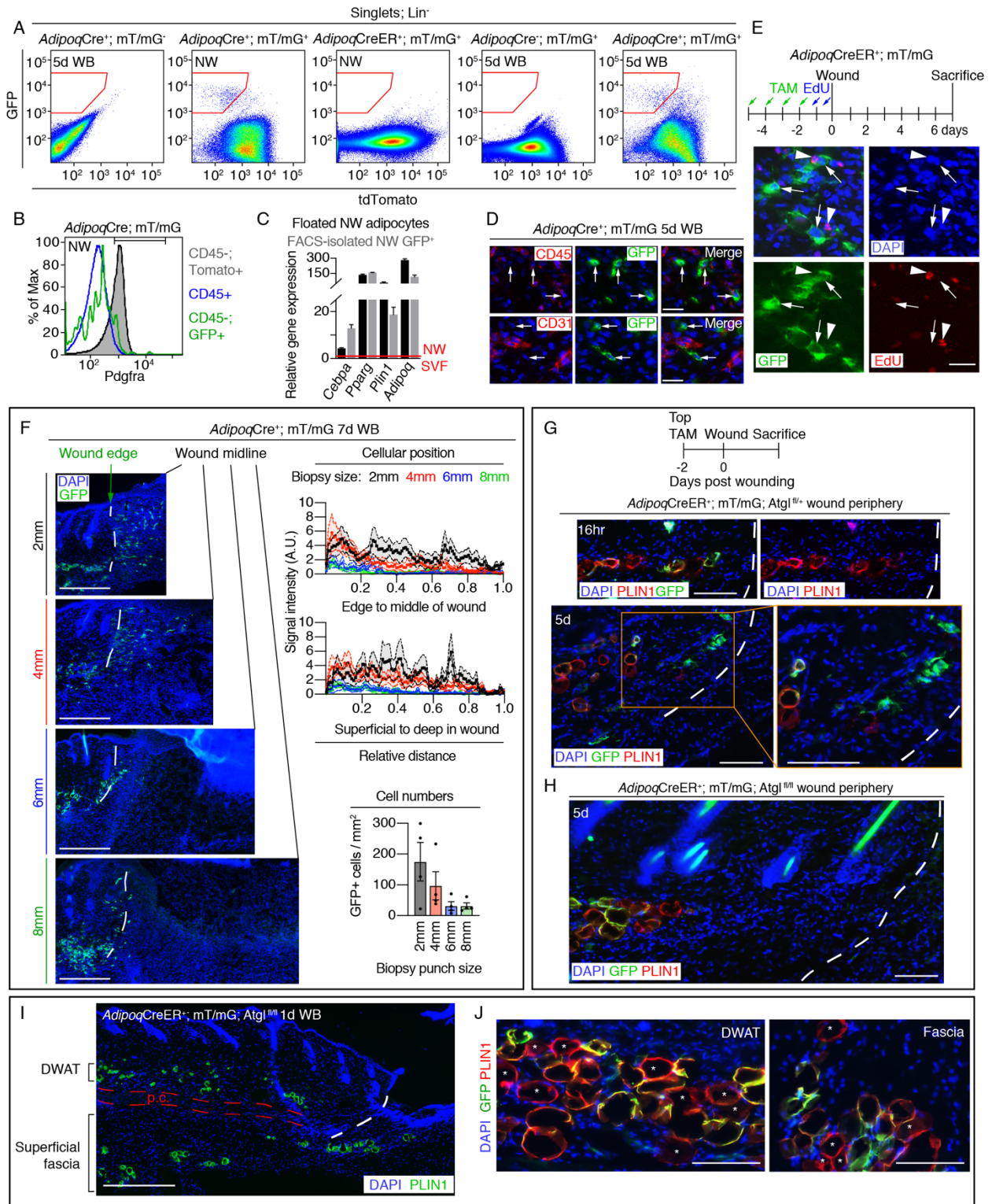


Figure S5. Wound edge dermal adipocyte-derived cells migrate into wound beds following lipid mobilization. Related to Figure 4 and Figure 5.

(A) Flow cytometry plots of GFP⁺ cells in back skin (NW) and wound beds from *Adipoq*Cre⁺; mT/mG and *Adipoq*CreER⁺; mT/mG mice given tamoxifen 7 days before analysis. (B) Flow cytometry plot of *Pdgfra* colocalization in cell populations from *Adipoq*Cre⁺; mT/mG mouse skin. (C) Quantitative real-time PCR of adipocyte-related genes in adipocytes and GFP⁺ cells isolated from *Adipoq*Cre⁺; mT/mG back skin. Red bar indicates gene expression of the dermal stromal vascular fraction ($n = 3$ mice). (D) Immunostained images of GFP and lineage markers (CD31 and CD45) in day 5 wound beds. White arrows indicate GFP⁺ cells. Scale bars, 25 μ m. (E) Labeling strategy and immunostained section showing GFP⁺ cells in the wound bed of *Adipoq*CreER⁺; mT/mG mice given EdU prior to wounding. Arrows indicate GFP⁺ cells and arrowheads indicate EdU⁺ cells. Scale bar, 25 μ m. (F) Composite images and quantification of the distribution and numbers of GFP⁺ cells in 7-day wounds of *Adipoq*Cre⁺; mT/mG mice. Original wounds were made with either a 2, 4, 6, or 8mm biopsy punch ($n = 4$ mice for each wound size). Scale bars, 500 μ m. (G-H) Images of GFP and PLIN1 immunostaining at the wound edge of 16-hour and 5-day wound beds from *Adipoq*CreER⁺; *Atg*^{fl/+} mice (G) and 5-day *Adipoq*CreER⁺; *Atg*^{fl/fl} mice (H). Scale bars, 100 μ m. (I-J) Low magnification image of PLIN1 immunostained tissue at the wound edge (I) and higher magnification images of GFP and PLIN1 colocalization in the DWAT and adipocytes located in the deeper fascia (J). Red dotted line delineates the panniculus carnosus (p.c.). Asterisks indicates GFP⁻; PLIN1⁺ cells. Scale bars, 500 μ m (I) and 100 μ m (J). White dotted lines delineate wound edges. Error bars indicate mean \pm SEM. SVF, stromal vascular fraction; WB, wound bed.

Figure S6

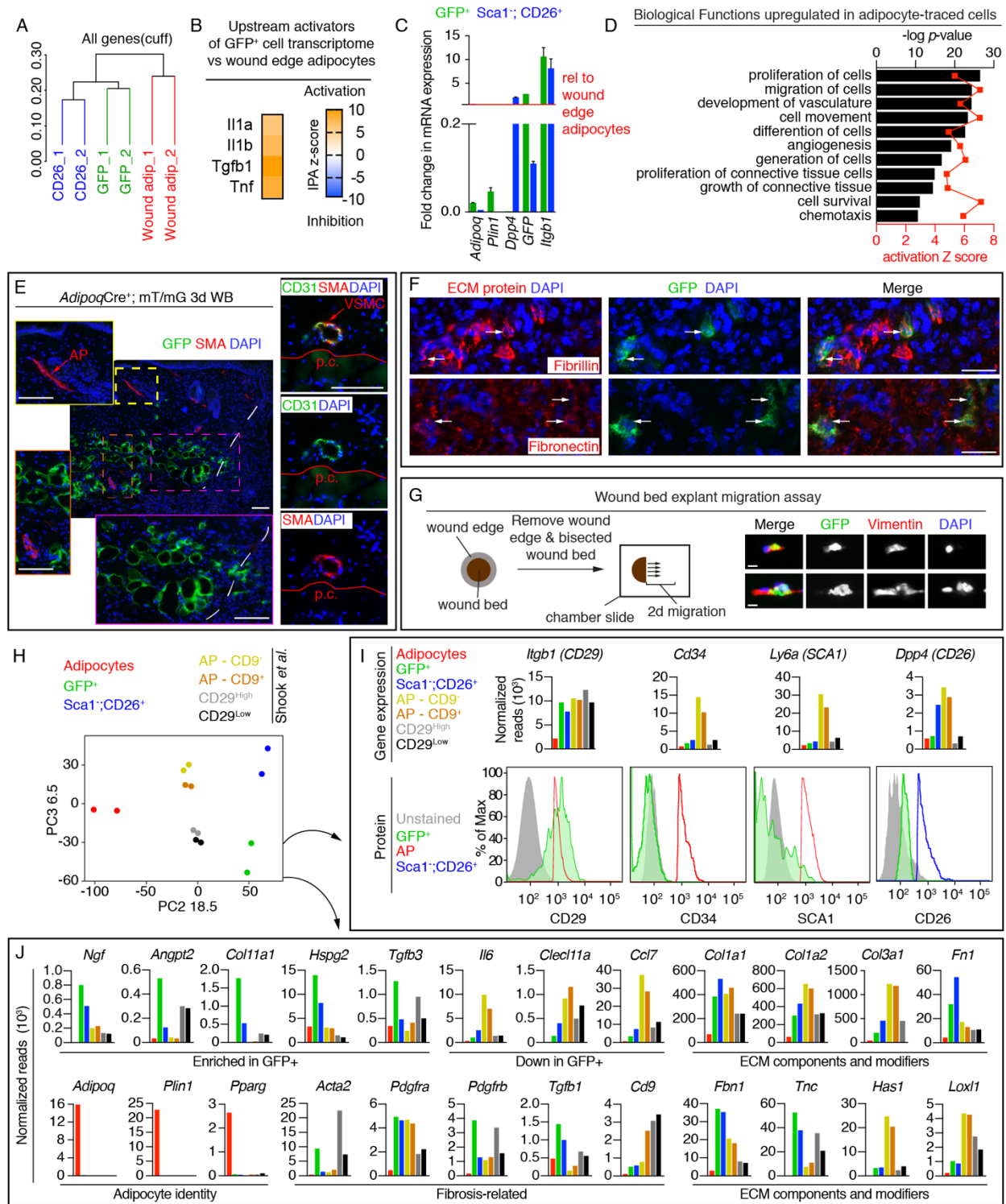


Figure S6. Adipocyte-derived cells have a unique myofibroblast gene expression profile. Related to Figure 6.

(A) Hierarchical clustering of cell populations from RNA-sequencing. (B) Ingenuity Pathway Analysis of the predicted upstream activators for the genes enriched in GFP+ cells relative to adipocytes. (C) Quantitative RT-PCR in cell populations isolated for RNA-seq experiments ($n = 3$ mice). (D) Predicted biofunctions upregulated by the gene expression profile of adiponectin-traced cells relative to mature adipocytes at the wound periphery. (E) SMA immunostained sections at the periphery of day 3 wound beds co-stained with GFP or CD31 in *AdipoqCre+*; mT/mG mice. Colored dashed boxes show the area of higher magnification. Scale bars, 100 μ m. Red line indicates panniculus carnosus (p.c.). (F) Images from GFP immunostained sections from the center of 5-day wound beds from *AdipoqCre+*; mT/mG mice co-stained with fibrillin or fibronectin. Arrow indicate GFP+ cells that colocalize with ECM proteins. Scale bars, 100 μ m. (G) Schematic and images of GFP+; Vimentin+ cells that migrated out day 5 wound beds from *AdipoqCre+*; mT/mG mice (72/78 GFP+ cells were vimentin+, $n = 4$ wounds). Scale bars, 10 μ m. (H) Principal component (PC) analysis of wound-associated populations from Figure 6 with previously reported myofibroblast subsets. (I) Normalized read counts from RNA-seq experiments (top) and flow cytometry plots (bottom) of CD34, CD29, SCA1 and CD26 of GFP+ cells in 5-day wound beds. (J) Normalized read counts comparing gene expression of GFP+ cells to wound edge adipocytes and different subsets of day 5 wound bed myofibroblasts. Error bars indicate mean \pm SEM. AP, arrector pili; VSMC, vascular smooth muscle cell.

Figure S7

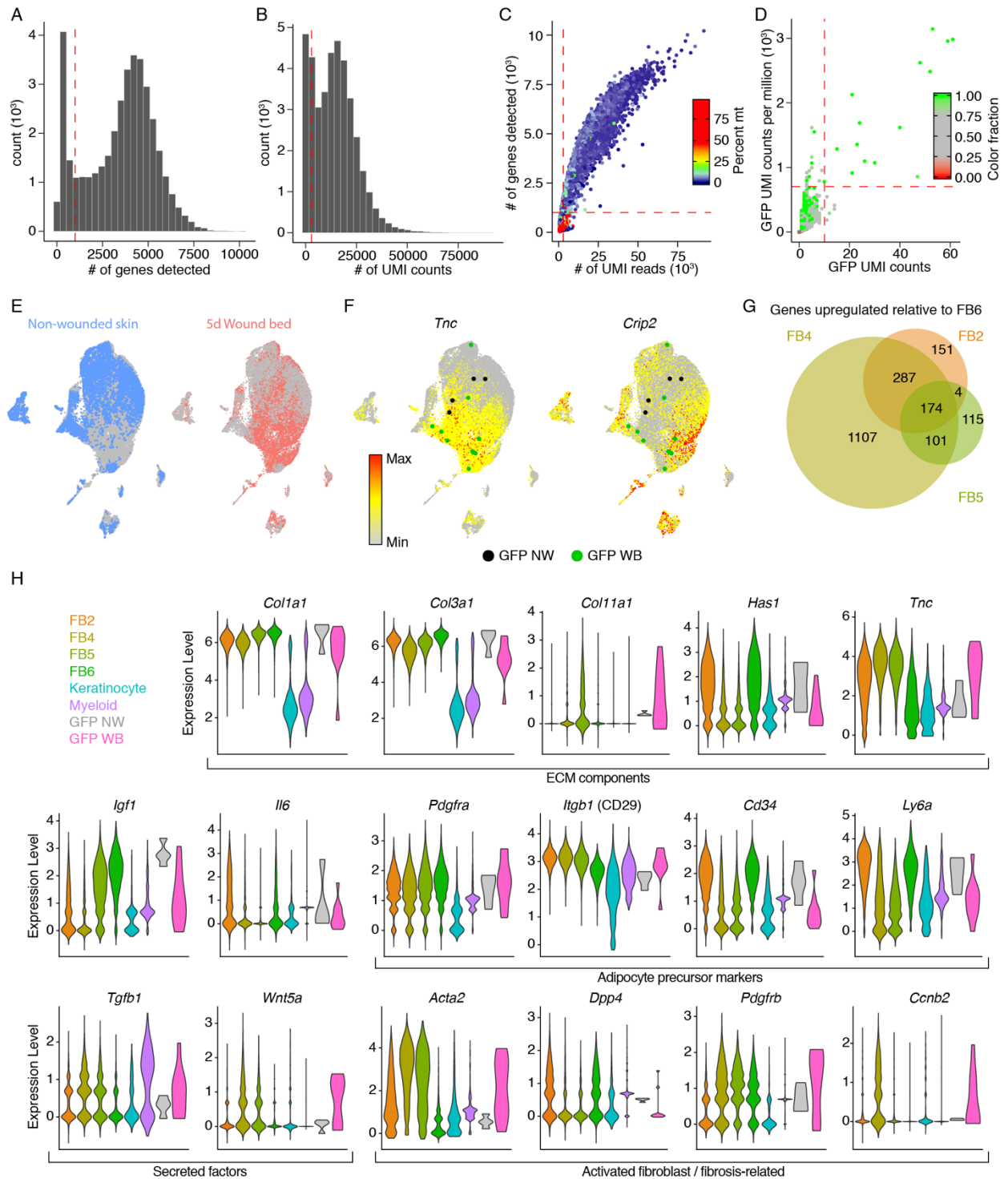


Figure S7. Single cell RNA-seq analysis of wound bed myfibroblasts and *Adipoq*-traced cells. Related to Figure 7, Table S1 and Table S2.

(A,B) Histogram of the number of genes detected per barcode (A) and number of UMI counts per barcode (B) with cutoffs for inclusion for future analysis shown as a red vertical lines. Barcodes with fewer than 100 genes detected were excluded from this analysis. (C) Scatterplot of UMI reads versus number of genes detected per barcode, with the percentage of reads mapping to mitochondrial genes indicated on a color scale. Cutoffs for cell inclusion are indicated as dashed lines. (D) Scatterplot of the number of UMI counts mapping to GFP and the fraction of all UMIs mapping to GFP (expressed in counts per million). Cells are colored by the fraction of reads that map either to eGFP (green) or tdTomato (red). Cutoffs to identify confidently GFP+ cells are indicated as dashed lines. (E) UMAP plots showing the location of cells from non-wounded skin and wound beds. (F) Gene expression plots showing distribution of *Tnc* and *Crip2*. Expression levels for each cell are shown as Pearson residuals and displayed using a color scale, overlaid onto the UMAP plot. GFP+ cells from NW and WB samples are overlaid to emphasize their location. (G) Venn diagrams displaying the number of differentially expressed genes in selected fibroblast clusters relative to FB6, using the Wilcoxon rank sum test implemented through Seurat. (H) Violin plots of genes that functionally discriminate subsets of wound-associated fibroblast populations.