

Figure S1. Related to Figure 1. Enumeration of epidermal cells by flow cytometry is an underestimate

Thy1.1⁺ OT-I cells were adaptively transferred to naïve WT mice followed by VV-OVA skin infection (flank). At least 60 days post infection, LC and OT-I Trm cells were enumerated by either flow cytometry or immunofluorescence microscopy. LC are defined as live CD45⁺MHCII⁺ cells by flowcytometry and MHCII⁺ cells by immunofluorescence microscopy. OT-I cells were identifies as live CD45⁺Thy1.1⁺ in flow cytometry and Thy1.1⁺ by immunofluorescence microscopy. Data are representative from three separate experiments. ****p < 0.0001. Each symbol represents data from an individual animal and lines indicate data from the same animal.

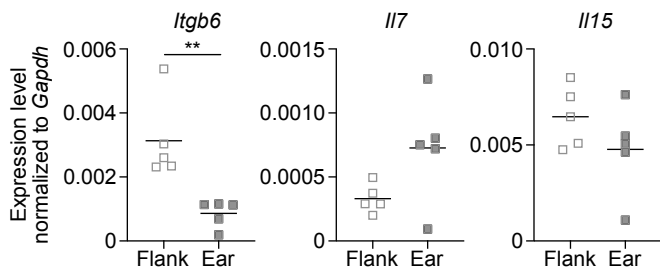
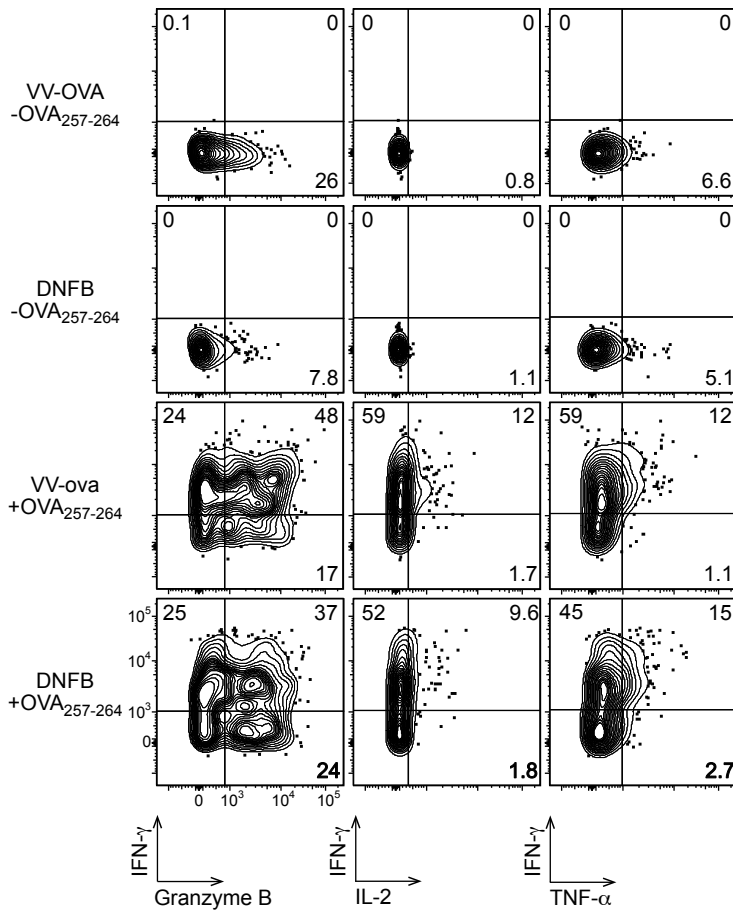


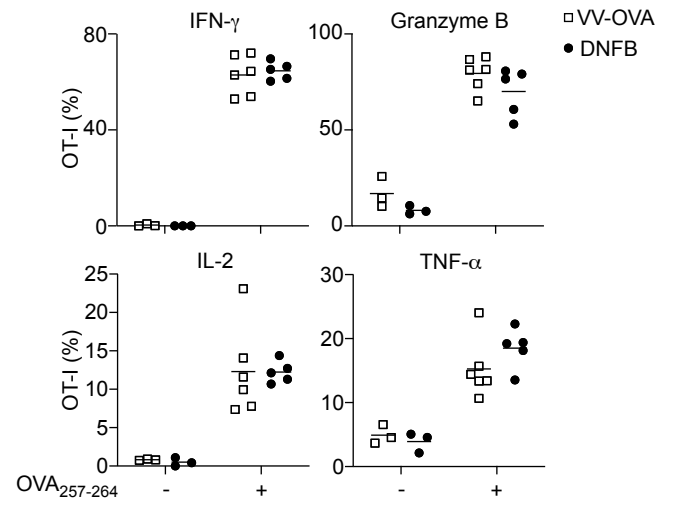
Figure S2. Related to Figure 2. Gene expressions in flank and ear epidermis

Itgb6, *I17*, and *I115* mRNA relative expression normalized to *Gapdh* in epidermis isolated from flank or ear skin are shown. Data are representative from two separate experiments. **p < 0.01. Each symbol represents data from an individual animal.

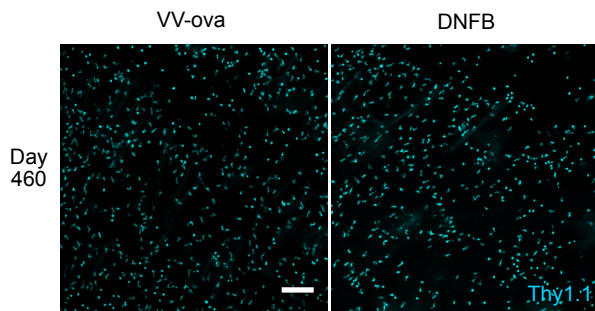
A



B



C



D

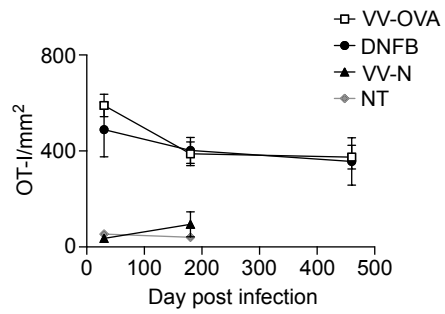


Figure S3. Related to Figure 4. Phenotype of antigen-specific Trm cells and bystander Trm cells

(A-B) Thy1.1⁺ OT-I cells were adoptively transferred into naïve WT mice followed by skin VV-OVA infection on left flank skin. On day 5 post infection, the right flank was painted with 0.15% DNFB. Either left flank or right flank were epicutaneously challenged OVA₂₅₇₋₂₆₄ day 70 post infection. The mice were injected i.p. with brefeldin A 6 hour after the challenge and the epidermis were harvested for analyzing recall responses 12 hour after the challenge. The representative flow plots gated on live CD45⁺Thy1.1⁺ OT-I cells (A), and the frequency of OT-I cells (B). (C-D) Thy1.1⁺ OT-I cells were adoptively transferred into naïve WT mice followed by skin VV-OVA infection on left flank skin. At the same day, some mice were also infected VV-N on the right flank. The mice not infected with VV-N were epicutaneously treated with DNFB on the right flank 5 days post infection. The epidermis from each site were harvested day 30, 180, 460 post infection. The representative immunofluorescence images of epidermal whole mount for Thy1.1⁺ OT-I cells (C) and the quantification (D) are shown. Data are representative from two separate experiments. Scale bar represents 100 μ m.

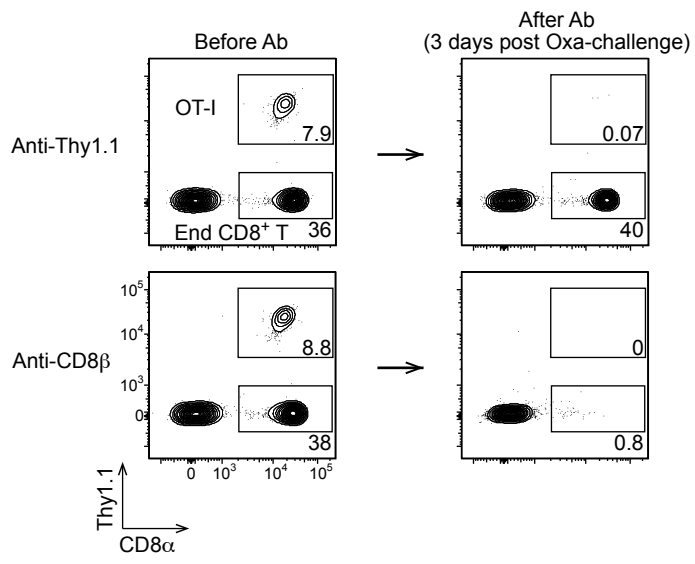


Figure S4. Related to Figure 6 and 7. Depletion of circulating CD8⁺ T cells

Thy1.1⁺ OT-I cells were adoptively transferred into naïve WT mice followed by skin VV-OVA infection on left flank. On day 5 post infection, the right flank was painted with 0.15% DNFB. More than 35 days after DNFB treatment, mice were sensitized at abdomen with oxazolone twice with 5 days interval. The mice were treated with titrated anti-Thy1.1 or anti-CD8b mAbs and then both flanks were challenged by oxazolone at day 5 after the second sensitization. Blood samples were collected one day before mAbs treatment and 3 days after oxazolone challenge. Representative flow plots gated on live CD3⁺TCRβ⁺ T cells in blood are shown.

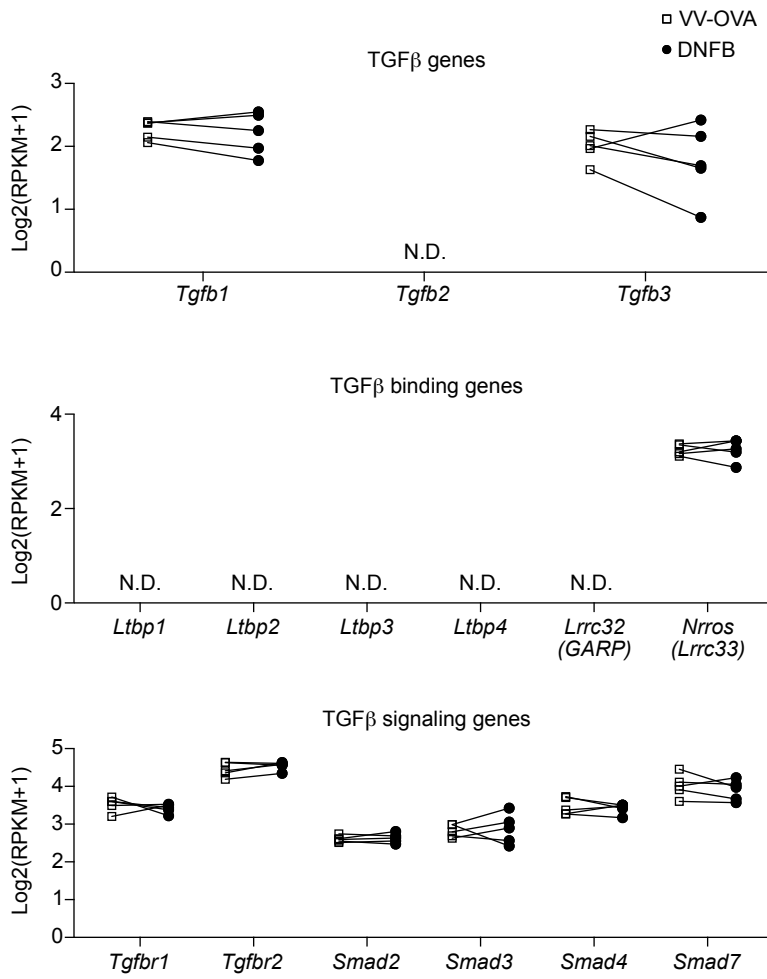


Figure S5. Related to Figure 7. TGF β -related gene expression in antigen-specific Trm and bystander Trm

Thy1.1⁺ OT-I cells were adoptively transferred into naïve WT mice followed by skin VV-OVA infection on left flank. On day 5 post infection, the right flank was painted with 0.15% DNFB. At day 57-70 after infection, liveCD45⁺Thy1.2⁻ Thy1.1⁺OT-I Trm from each flank from the same animals were FACS sorted and analyzed by RNA-seq. Mice from 2 individual animals were pooled for each repeat. N.D. is not detected if RPKM of all samples were less than 4.