

Table S1. Bacterial strains, plasmids and primers used in this study.

Strain	Description	Source
<i>E. coli</i>		
	Cloning strain; F ⁻ Φ80 <i>lacZ</i> ΔM15 Δ(<i>lacZYA-argF</i>)	
DH5α	U169 <i>recA1 endA1 hsdR17</i> (r _K ⁻ , m _K ⁺) <i>phoA supE44 λ</i> ⁻ <i>thi-1 gyrA96 relA1</i>	Invitrogen
SM10	Bi-parental mating strain; <i>thi thr leu tonA lacY supE</i> <i>recA::RP4-2-Tc::Mu K_m λpir</i> , Kan ^R , Tet ^R	(1)
BTH101	BACTH strain; F ⁻ <i>cya-99 araD139 galE15 galk16</i> <i>rpsL1</i> (Str ^R) <i>hsdR2 mcrA1 mcrB1</i>	Euromedex
<i>P. aeruginosa</i>		
PAO1	Wild-type strain	M. Parsek
PAO1 Δ <i>wspF</i> Δ <i>psl</i>	PAO1 Δ <i>wspF</i> (in-frame); Δ <i>pslBCD</i> (polar); <i>araC</i> -P _{BAD}	(2)
P _{BAD} <i>pel</i>	inserted upstream of <i>pelABCDEFGHI</i>	
GBW1	PAO1 Δ <i>wspF</i> Δ <i>psl</i> P _{BAD} <i>pel</i> Δ <i>rmcA</i>	This study
GBW2	PAO1 Δ <i>wspF</i> Δ <i>psl</i> P _{BAD} <i>pel</i> Δ <i>morA</i>	This study
GBW3	PAO1 Δ <i>wspF</i> Δ <i>psl</i> P _{BAD} <i>pel</i> Δ <i>rmcA</i> <i>attTn7::miniTn7T</i> - Gm:: <i>araC</i> -P _{BAD} :: <i>rmcA</i> , Gen ^R	This study
GBW4	PAO1 Δ <i>wspF</i> Δ <i>psl</i> P _{BAD} <i>pel</i> Δ <i>morA</i> <i>attTn7::miniTn7T</i> - Gm:: <i>araC</i> -P _{BAD} :: <i>morA</i> , Gen ^R	This study
GBW5	PAO1 Δ <i>wspF</i> Δ <i>psl</i> P _{BAD} <i>pel</i> Δ <i>rmcA</i> <i>attTn7::miniTn7T</i> - Gm:: <i>araC</i> -P _{BAD} :: <i>rmcA</i> ^{C-VSV-G} , Gen ^R	This study
GBW6	PAO1 Δ <i>wspF</i> Δ <i>psl</i> P _{BAD} <i>pel</i> Δ <i>morA</i> <i>attTn7::miniTn7T</i> - Gm:: <i>araC</i> -P _{BAD} :: <i>morA</i> ^{C-VSV-G} , Gen ^R	This study
SMC232	UCBPP-PA14	(3)

SMC6279	PA14 $\Delta rmcA$ (in-frame)	(4)
SMC6289	PA14 $\Delta morA$ (in-frame)	(4)
SMC7006	PA14 <i>att::DMS3₄₂</i>	(5)
SMC2893	PelA KO	(6, 7)
SMC6607	Pel inducible, P _{pel} replaced with P _{bad}	(8)
SMC8808	PA14 $\Delta rmcA$ <i>att::DMS3₄₂</i>	This study
SMC8809	PA14 $\Delta morA$ <i>att::DMS3₄₂</i>	This study
SMC8810	PA14 pKNT25 GcbC R363E	This study
SMC8811	PA14 $\Delta rmcA$ pKNT25 GcbC R363E	This study
SMC8812	PA14 $\Delta morA$ pKNT25 GcbC R363E	This study
SMC8813	PA14 $\Delta rmcA$ <i>attTn7::miniTn7T-Gm::araC-P_{BAD}::rmcA</i> , Gen ^R	This study
SMC8814	PA14 $\Delta morA$ <i>attTn7::miniTn7T-Gm::araC-P_{BAD}::morA</i> , Gen ^R	This study
SMC8815	PA14 $\Delta rmcA$:P _{tac} - <i>mKO</i> -κ	This study
SMC8816	PA14 $\Delta morA$:P _{tac} - <i>mKO</i> -κ	This study
SMC8817	PA14 $\Delta rmcA$:P _{smc21} - <i>gfp</i>	This study
SMC8818	PA14 $\Delta morA$:P _{smc21} - <i>gfp</i>	This study
SMC8819	PA14 $\Delta rmcA$ $\Delta pelA$	This study
SMC8820	PA14 $\Delta morA$ $\Delta pelA$	This study
SMC8821	PA14 $\Delta rmcA$, inducible Pel under the control of P _{BAD}	This study
SMC8822	PA14 $\Delta morA$ inducible Pel under the control of P _{BAD}	This study

Plasmid	Description	Source
Two-step allelic exchange		
pEX18Gm	Suicide vector for allelic exchange in <i>P. aeruginosa</i> , encodes SacB, Gen ^R	(1)
pEX18Gm:: <i>ΔrmcA</i>	pEX18Gm with <i>P. aeruginosa</i> PAO1 <i>ΔrmcA</i> allele cloned between the EcoRI and HindIII sites, Gen ^R	This study
pEX18Gm:: <i>ΔmorA</i>	pEX18Gm with <i>P. aeruginosa</i> PAO1 <i>ΔmorA</i> allele cloned between the SacI and HindIII sites, Gen ^R	This study
pSMC21	pUCP-based plasmid containing <i>gfpmut2</i> , Ap ^r , Cb ^r , Km ^r	(9)
pMQ30	Suicide vector; Gm ^r , <i>sacB</i> , <i>URA3</i> , <i>CEN6/ARSH4</i> , <i>lacZα</i>	(10)
pKO PA14_07500	PA14_07500 knockout construct in pMQ30	(4)
pKO PA14_07500	PA14_60870 (<i>morA</i>) knockout construct in pMQ30	(4)
pKNT25 GcbC-R363E	GcbC protein with an I-site mutation	(11)
Complementation analysis		
pUC18T-miniTn7T-Gm	<i>aacC1</i> on miniTn7-based vector with transcriptional terminators at the right end of the Tn7 transposon; Amp ^R , Gen ^R	(12)
pUC18T-miniTn7T-Gm-pBAD	pUC18T-miniTn7T-Gm containing <i>araC</i> -P _{BAD} and a downstream MCS (SmaI-NotI-PstI-NcoI) cloned between the HindIII and SacI sites; Amp ^R , Gen ^R	This study

pTNS2	Helper plasmid encoding <i>tnsABCD</i> , Amp ^R	(13)
	pUC18T-miniTn7T-Gm-pBAD with <i>P. aeruginosa</i>	
p-miniTn7- <i>rmcA</i>	PAO1 <i>rmcA</i> fused to an upstream synthetic ribosome binding site, cloned between the SmaI and NotI sites; Amp ^R , Gen ^R	This study
	pUC18T-miniTn7T-Gm-pBAD with <i>P. aeruginosa</i>	
p-miniTn7- <i>morA</i>	PAO1 <i>morA</i> fused to an upstream synthetic ribosome binding site, cloned between the NotI and Mph1103I sites; Amp ^R , Gen ^R	This study
p-miniTn7- <i>rmcA</i> ^{C-VSV-G}	p-miniTn7- <i>rmcA</i> fused to a downstream VSV-G tag; Amp ^R , Gen ^R	This study
p-miniTn7- <i>morA</i> ^{C-VSV-G}	p-miniTn7- <i>morA</i> fused to a downstream VSV-G tag; Amp ^R , Gen ^R	This study

BACTH assays

pUT18C	Encodes T18 fragment of <i>B. pertussis</i> adenylate cyclase toxin on N-terminus, Amp ^R	(14)
pKT25	Encodes T25 fragment of <i>B. pertussis</i> adenylate cyclase toxin on N-terminus, Kan ^R	(14)
pUT18C::zip	BACTH positive control, pUT18C containing sequence coding for the leucine zipper region of yeast GCN4 protein, Amp ^R	(14)
pKT25::zip	BACTH positive control, pKT25 containing sequence coding for the leucine zipper region of yeast GCN4 protein, Kan ^R	(14)

pKT25:: <i>pelD</i>	<i>P. aeruginosa</i> PAO1 <i>pelD</i> cloned into the XbaI and BamHI sites of pKT25, Kan ^R	(15)
pUT18C:: <i>rncA</i>	<i>P. aeruginosa</i> PAO1 <i>rncA</i> cloned into the XbaI and SmaI sites of pUT18C, Amp ^R	This study
pUT18C:: <i>morA</i>	<i>P. aeruginosa</i> PAO1 <i>morA</i> cloned into the XbaI and KpnI sites of pUT18C, Amp ^R	This study
pBADGr	For expression of third protein in the BACTH assay, <i>ori araC-P_{BAD} dhfr::Gm^R mob⁺, Gen^R</i>	(16)
pBADGr:: <i>pelE</i>	<i>P. aeruginosa</i> PAO1 <i>pelE</i> cloned into the EcoRI and HindIII sites of pBADGr with synthetic RBS (GAGGAGGATATTC), Gen ^R	(15)
pBADGr:: <i>pelG</i>	<i>P. aeruginosa</i> PAO1 <i>pelG</i> cloned into the EcoRI and HindIII sites of pBADGr with synthetic RBS (GAGGAGGATATTC), Gen ^R	(15)
pBADGr:: <i>pelEG</i>	<i>P. aeruginosa</i> PAO1 <i>pelE</i> and <i>pelG</i> ORFs separated by a clean, unmarked deletion of <i>pelF</i> cloned into the EcoRI and HindIII sites of pBADGr with synthetic RBS (GAGGAGGATATTC), Gen ^R	This study

Abbreviations: Amp, ampicillin; Kan, kanamycin; Gen, gentamicin; Str, streptomycin; Tet, tetracycline

Primers	Sequence*
Allelic exchange vectors	
<i>rnc</i> APAO1upF	GGG GAA TTC <u>GAA CCA CGA GAC CGA CGA GA</u>
<i>rnc</i> APAO1upR	<u>TAC CGG CGC TTC TCG GGG AGC ACA CAG GAG AAG</u> CAC GGC CTG
<i>rnc</i> APAO1downF	<u>GCT CCC CGA GAA GCG CCG GTA</u>
<i>rnc</i> APAO1downR	AGG AAG CTT <u>GGA ATA ACC AAC GGT GCC AGG T</u>
<i>mor</i> APAO1upF	AAT GAG CTC <u>TCC TAC CTG GAA GCG CTG GA</u>
<i>mor</i> APAO1upR	<u>TCA GCC CTC GTT GAA CAT GAA CGA CGG GGT TAG</u> CGA GGG GGT
<i>mor</i> APAO1downF	<u>TTC ATG TTC AAC GAG GGC TGA</u>
<i>mor</i> APAO1downR	GGG AAG CTT <u>AGG TAG ACC GCG GCG TTG GC</u>
miniTn7 complementation vectors	
miniTn7-pBAD-F	GGG AAG CTT <u>TTA TGA CAA CTT GAC GGC TA</u>
miniTn7-pBAD-R	GGG GAG CTC <u>CCA TGG CTG CAG GCG GCC GCC CCG</u> GGC <u>AAA AAA ACG GGT ATG GAG AAA CAG TA</u>
<i>rncA</i> -miniTn7-F	GAG CCC GGG AGG AGG ATA TTC <u>ATG CAG CGT CTG</u> CAG GCC GT
<i>rncA</i> -miniTn7-R	GGG GCG GCC GCT <u>TAT ACC GGC GCT TCT CGG GG</u>
<i>morA</i> -miniTn7-F	GGG GCG GCC GCA GGA GGA TAT TCG <u>TGT CGA CCC</u> CCT CGC TAA C
<i>morA</i> -miniTn7-R	GGG ATG CAT <u>TCA GCC CTC GTT GAA CAT GAA CA</u>
<i>rncA</i> -C-VSV-G-F	TGA ATA GAT TAG GAA AAT <u>AAG CGG CCG CCT GCA GC</u>
<i>rncA</i> -C-VSV-G-R	TTT CAA TAT CTG TAT <u>ATA CCG GCG CTT CTC GGG GA</u>

<i>morA</i> -C-VSV-G-F	ATA GAT TAG GAA AAT <u>GAA TGC ATG ATC GAA TTA GCT</u> <u>TCA A</u>
<i>morA</i> -C-VSV-G-R	TCA TTT CAA TAT CTG TAT <u>AGC CCT CGT TGA ACA TGA</u> <u>ACA G</u>

BACTH vectors

<i>pelD</i> -BACTH-F	GGG TCT AGA <u>GAT GTC CGC GCA CAA GGA TTT CA</u>
<i>pelD</i> -BACTH-R	GTT GGA TCC <u>CCT AAA TAG CCA CTT GCT GAT CAT TC</u>
<i>rmcA</i> -BACTH-F	GAT TCT AGA <u>GAT GCA GCG TCT GCA GGC CG</u>
<i>rmcA</i> -BACTH-R	GAG CCC GGG <u>TTA TAC CGG CGC TTC TCG GG</u>
<i>morA</i> -BACTH-F	GAG TCT AGA <u>GGT GTC GAC CCC CTC GCT AA</u>
<i>morA</i> -BACTH-R	GTG GGT ACC <u>TCA GCC CTC GTT GAA CAT GAA C</u>
<i>pelE</i> -BADGr-F	GGG GAA TTC GAG GAG GAT ATT <u>CAT GAT CAG CAA</u> <u>GTG GCT ATT TAG C</u>
<i>pelE</i> -BADGr-R	GAG AAG CTT <u>TCA TGT CCA GTA TCT CGC CAG</u>
<i>pelG</i> -BADGr-F	GGG GAA TTC GAG GAG GAT ATT <u>CAT GGC CGG CAT</u> <u>CGG CTTC</u>
<i>pelG</i> -BADGr-R	GAG AAG CTT <u>TCA GCG ATT GAG CAT GAA GGT</u>
<i>pelF</i> -del-BADGr-upR	<u>TCA TGC AAT CTC CGT GGC TTC CGG AGC GGT GTG</u> <u>TTC GGT CAT</u>
<i>pelF</i> -del-BADGr-downF	<u>GAA GCC ACG GAG ATT GCA TGA</u>

Sequencing primers

M13F	<u>GTA AAA CGA CGG CCA G</u>
M13R	<u>CAG GAA ACA GCT ATG AC</u>
<i>rmcA</i> -SEQ-F	<u>TGG ACA TGC CTG AGG ACA AG</u>
<i>rmcA</i> -SEQ-R	<u>CGC CTG TTT CAG CAC GTC AAA A</u>

<i>morA</i> -SEQ-F	<u>CGG ACT TCC TCG ACC CG</u>
<i>morA</i> -SEQ-R	<u>CGC CAT GGG CCA GGC T</u>
miniTn7-SEQ-F	<u>GCG GAT CCT ACC TGA CGC TT</u>
miniTn7-SEQ-R	<u>AAC TTC AGA GCG CTT TTG AA</u>
pUT18C-SEQ-F	<u>CTC GCC GGA TGT ACT GGA AAC</u>
pUT18C-SEQ-R	<u>CGG GGC TGG CTT AAC TAT GC</u>
pKT25-SEQ-F	<u>GGC GCG CAG TTC GGT GAC CAG CGG C</u>
pKT25-SEQ-R	<u>GGG ATG TGC TGC AAG GCG ATT AAG</u>
pBADGr-SEQ-F	<u>ATG CCA TAG CAT TTT TAT CC</u>
pBADGr-SEQ-R	<u>GAT TTA ATC TGT ATC AGG</u>
<i>rncA</i> -SEQ-int1	<u>GGA GAT CGA GCG ACT CCA GG</u>
<i>rncA</i> -SEQ-int2	<u>CGA GCA GCA GAT CGG CAT CG</u>
<i>rncA</i> -SEQ-int3	<u>TCA ACC GCG CCT TCA GCG AG</u>
<i>rncA</i> -SEQ-int4	<u>ATC GCC CTG TTC CCC AAG GA</u>
<i>morA</i> -SEQ-int1	<u>TGC TCG ACG ACA TGC AGA CC</u>
<i>morA</i> -SEQ-int2	<u>TGC GCC AGG CTC TTG TGG AG</u>
<i>morA</i> -SEQ-int3	<u>TGG CAC TGG TTC GAC ATC CG</u>
<i>morA</i> -SEQ-int4	<u>ACC GCC AGG AAG CCA ACC AG</u>

*Restriction sites and Gateway *att* sequences are bolded; regions of complementary to the target amplicon are underlined; regions of reverse complementarity (to facilitate splicing) are italicized; synthetic ribosomal binding sites are in bold italics

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