

Biochemical and cellular characterisation of the *Plasmodium falciparum* M1 alanyl aminopeptidase (PfM1AAP) and M17 leucyl aminopeptidase (PfM17LAP).

Rency Mathew, Juliane Wunderlich, Karine Thivierge, Krystyna Cwiklinski, Claire Dumont,
Leann Tilley, Petra Rohrbach, John P. Dalton

Supplementary Information

Supplementary Figure 1. Preparation of antibodies to N-terminal, middle Domain 1 and C-terminal domain of PfM1AAP. (A) Antibodies were prepared against 14-mer peptide sequences at positions 140 – 153 (anti-PepA, yellow) within the N-terminal extension, 366-380 (anti- PepB, magenta) within Domain 1 and 873 – 889 (anti-PepC, green) within C-terminal Domain 4. Glutaminic acid (E) 195 is the first residue in the recombinant form of PfM1AAP. The start and end of the p68 processed form (~70 kDa) of PfM1AAP according to the paper by Azimzadeh *et al.* [27] are indicated by the boxed and in bold **VKKNE** and **PVNAQ** sequences, respectively. (B) Schematic representation of the sequence and position of PepA, PepB and PepC within the PfM1AAP.

Supplementary Figure 2. Incubation of recombinant PfM1AAP with *P. falciparum* cytosolic fraction (C1). Recombinant PfM1AAP (1 µg) was incubated overnight (16 h) with cytosolic C1 fractions (5 µg) that were prepared without (-) or with (+) a cocktail of proteinase inhibitors. Samples taken before (lanes 2 and 3) and after (lanes 4 and 5) the overnight incubation were analysed by immunoblots probed with anti-His-tag mouse monoclonal antibodies. The 100-kDa recombinant PfM1AAP enzyme is resistant to hydrolysis by the cytosolic C1 fractions.

Supplementary Figure 3. IFA using paraformaldehyde/glutaraldehyde fixed cells malaria-infected erythrocytes. Concanavalin A (0.5 mg/ml) was coated onto a coverslip in humid chamber at 37°C for 30 minutes. iRBCs were harvested, washed in PBS and resuspended in PBS at 3% hematocrit and incubated for 15 minutes at RT on the coverslip. Cells were fixed with 2% PFA and 0.008% glutaraldehyde for 20 minutes at RT. Cells were permeabilised by adding PBS 0.1% Triton X-100 for 10 minutes at RT. Then, 3% BSA in PBS was added and incubated for 1 hour at RT as blocking step before adding the primary antibody diluted in PBS/3% BSA and

incubate for 1 hour at RT. The secondary antibody was diluted in PBS/3% BSA, and incubated for 1 hour at RT. DAPI (10 µg/ml in PBS) was added. Parasite nuclei were visualized using DAPI (blue staining; 4,6-diamidino-2-phenylindole). Red staining highlights the (A) cells probed with anti-*PfM1AAP* antibodies and (B) cells probed with anti-*PfM17LAP* antibodies. Panel A, differential interference contrast (DIC) is shown for reference; Panel B, cells probed with antibodies; Panel C, merged.

Supplementary Figure 4. IFA using acetone-fixed malaria-infected erythrocytes. iRBCs were harvested, washed in PBS and resuspend in PBS at 3% haematocrit. The iRBC were smeared on a coverslip, air-dried at RT and fixed in acetone (100%) for 10 minutes at RT. PBS/3% BSA was added and incubated for 1 hour at RT as blocking step before adding the primary antibody diluted in PBS/3% BSA and incubating for 1 hour at RT. The secondary antibody was diluted in PBS/ 3% BSA, and incubated for 1 hour at RT. The nucleus was stained with DAPI (10 µg/ml) diluted in PBS. Parasite nuclei were visualized using DAPI (blue staining; 4,6-diamidino-2-phenylindole). Red staining highlights the (A) cells probed with anti-*PfM1AAP* antibodies and (B) cells probed with anti-*PfM17LAP* antibodies. Panel A, differential interference contrast (DIC) is shown for reference; Panel B, cells probed with antibodies; Panel C, merged.

Supplementary Figure 5. pH profile of recombinant *PfM17LAP*. The pH profile of recombinant *PfM17LAP* (1 µg) was obtained by incubating the enzyme in a series of pH-controlled buffers that were also employed for the studies of the *PfM1AAP* in this report; these were 0.1M sodium acetate pH 4.0 – 5.5, 0.1M sodium citrate pH 5.5 – 6.0, 0.1 M sodium phosphate pH 6.0 – 8.0, 0.1 M Tris buffer pH 7.5 – 9.0. Reactions were initiated by the the addition of the fluorogenic substrate H-Leu-NHMec at 37°C and the release of the fluorophore NHMec monitored in a fluorimetre set with excitation at 360 nm and emission at 460 nm.

Supplementary Figure 6. Recombinant production of ~70 kDa fragment of *PfM1AAP*. (A) *E. coli*-expressed recombinant ~70 kDa form of *PfM1AAP* was partially purified by Ni-Chelate affinity chromatography. The partially purified protein migrated at 70 kDa when analyzed on 10% SDS-PAGE gels. Lane 1: molecular weight standards. Lane 2: insoluble *E. coli* lysate fraction. Lane 3: soluble *E. coli* lysate fraction. Lane 4: column run-through. Lane 5: column

wash. Lane 6: eluted recombinant *PfM1AAP*. **(B)** Immunodetection of recombinant ~70-kDa form of the enzyme using anti-His tag mouse monoclonal antibody (lane 3) and anti-Pep B antibody (lane 4). Molecular weight standards are shown in lane 1.

Supplementary Figure 7. Specificity of antibody against *PfM1AAP* and *PfM17LAP*.

Antibodies prepared in rabbits against *PfM1AAP* and *PfM17LAP* were adsorbed against freeze-thaw/sonicated extracts of *E. coli* expressing a recombinant His-tagged *Schistosoma mansoni* peroxiredoxin (SmPrx) as described in Materials and Methods. Immunoblots were prepared with molecular size markers (lane 1), recombinant *PfM1AAP* (lane 2) and recombinant *PfM17LAP* (lane 3) and probed with the adsorbed anti-*PfM1AAP* (Panel A) or anti-*PfM17LAP* (Panel B). These adsorbed antibodies were employed in both immunoblot and cytochemical studies in this report.

Supplementary Figure 8. Relative activity of *PfM1AAP*, *PfM17LAP* and *P. falciparum* 3D7 parasite cytosolic extract towards several fluorogenic NHMec substrates (20 μ M) at pH 7.3.

Enzyme assays were performed as described in Materials and Methods. Results shown are the average of three replicates in a typical substrate screening. The substrates used were Leu, H-Leu-NHMec; Glu, H-Glu-NHMec; Arg, H-Arg-NHMec; Pro, H-Pro-NHMec; PR, Pro-Arg-NHMec; ZPR, Z-Pro-Arg-NHMec; TGPR; Tosyl-Gly-Pro-Arg-NHMec; TGPK, Tosyl-Gly-Pro-Lys-NHMec; FR, Phe-Arg-NHMec; ZFR, Z-Phe-Arg-NHMec; ZLR, Z-Leu-Arg-NHMec; ZR, Z-Arg-NHMec.

Supplementary Figure 9. Full length gel photos. (A) Fig. 1B. Immunoblot of the recombinant *PfM1AAP* and *PfM17LAP* proteins and *P. falciparum* TPE and cellular fractions (C1, C2, V1 and V2) probed with (i) anti-*PfM1AAP*, (ii) anti-*PfM17LAP*, and (iii) anti-plasmeepsin 1 antibodies. **(B)** Fig. 3A. Recombinant *PfM1AAP* was probed with antibodies prepared against 14-mer peptides derived from the N-terminal extension (PepA), spanning domain 1 (PepB) and domain 4 (PepC) and Anti-*PfM1AAP*. The light grey box denotes lanes that contain proteins not relevant to this study. **(C)** Fig. 3B. Cytosolic fractions of malaria parasites prepared without (-) and with (+) a protease inhibitor cocktail were probed with anti-PepA, anti-PepB and anti-PepC antibodies. The light grey box denotes lanes that contain proteins not relevant to this study.

Supplementary Fig. 1

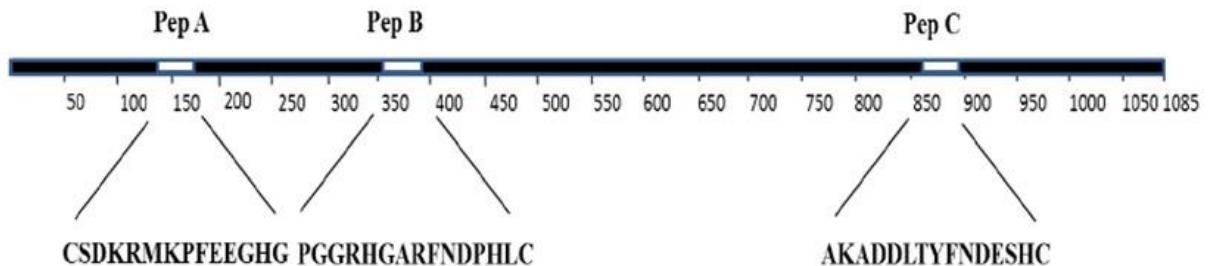
A

```

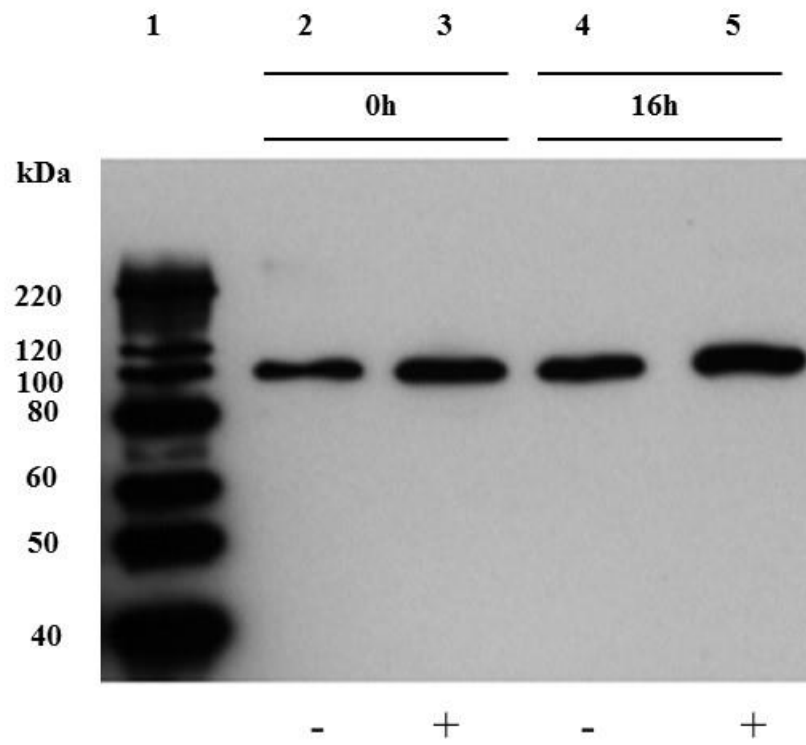
5   10  15  20  25  30  35  40  45  50  55  60  65  70  75  80
MKLTK GCAYK YIIPT VLILA NILYD NKKRC MIKRN LRISS CGIIS RLLKS NSNYN SFNKN YNFTS AISEL QFSNF WNLDI
85  90  95  100 105 110 115 120 125 130 135 140 145 150 155 160
LQKDI FSNIH NKNK PQSYI IHKRL MSEKG DNNNN NHQNN NGNDN KKRLG SVVNN EENTC SDKRM KPFE EGHG LT QVDRM
165 170 175 180 185 190 195 200 205 210 215 220 225 230 235 240
NNSD HLQQN GVMNL NSNNV ENNNN NNSVV VKKNE PKIHY RKDYK PSGFI INNVT LNINI HDNET IVRSV LDMDI SKHNV
245 250 255 260 265 270 275 280 285 290 295 300 305 310 315 320
GEDLV FDGVG LKINE ISINN KKLVE GEEYT YDNEF LTIFS KFVPK SKFAF SSEVI IHPET NYALT GLYKS KNIIV SQCEA
325 330 335 340 345 350 355 360 365 370 375 380 385 390 395 400
TGFRR ITFFI DRPDM MAKYD VVTVA DREKY FVLLS NGDKV NEFEI PGGRH GARFN DPHLK PCYLF AVVAG DLKHL SATYI
405 410 415 420 425 430 435 440 445 450 455 460 465 470 475 480
TKYTK KRVEL YVFE EKYVS KQWA LECLK KSMF DEDYF GLEYD LSRLN LVAVS DFNVG AMENK GLNIF NANSL LASKK
485 490 495 500 505 510 515 520 525 530 535 540 545 550 555 560
NSIDF SYARI LTVVG HEYFH NYTGN RVTLR DWFQL TLKEG LTVHR ENLFS EEMTK TVTTR LSHVD LLRSV QFLED SSPLS
565 570 575 580 585 590 595 600 605 610 615 620 625 630 635 640
HPIRP ESYVS MENFY TTVY DKGSE VMRMY LTI LG EEYK KGF DI YIKKN DGNTA TCEDF NYAME QAYKM KKADN SANLN
645 650 655 660 665 670 675 680 685 690 695 700 705 710 715 720
QYLLW FQSG TPHVS FKYNY DAERK QYSH VNOYT KPDEN QKEKK PLFIP ISVGL INPEN GKEMI SQTTL ELTKE SDTFV
725 730 735 740 745 750 755 760 765 770 775 780 785 790 795 800
FNNIA VKPIP SLFRG FSAPV YIEDN LTDEE RILLL KYDSD AFVRY NSCTN IYMKQ ILMNY NEFLK ARNEK LESFN LTPVN
805 810 815 820 825 830 835 840 845 850 855 860 865 870 875 880
AQFID AIKYL LEDPH ADAGF KSYIV SLPQD RYIIN FVSNL DTDVL ADTKE YIYKQ IGDKL NDVYY KMFKS LEAKA DDLTY
885 890 895 900 905 910 915 920 925 930 935 940 945 950 955 960
FNDES HVDFD QNMNR TLRNT LLSLL SKAQY FNILN EIIH SKSPY PSNWL TSLSV SAYFD KYFEL YDKTY KLSKD DELLL
965 970 975 980 985 990 995 1000 1005 1010 1015 1020 1025 1030 1035 1040
QEWLK TVSRD DRKDI YEILK KLENE VLKDS KNPND IRAVY LPFTN NLRRF HDISG KGYKL IAEVI TKTDK FNPV ATQLC
1045 1050 1055 1060 1065 1070 1075 1080 1085
EPPFL WNKLD TRQRE LMLNE MNTML QEPNI SNNLK EYLLR LTNKL

```

B

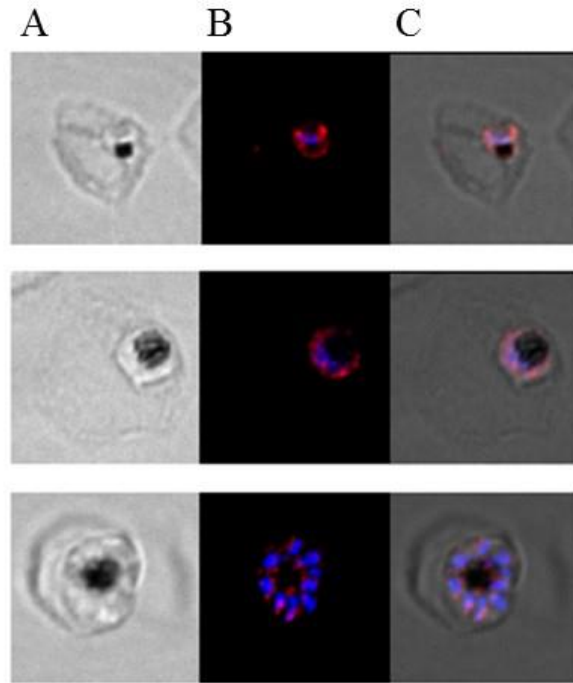


Supplementary Fig. 2

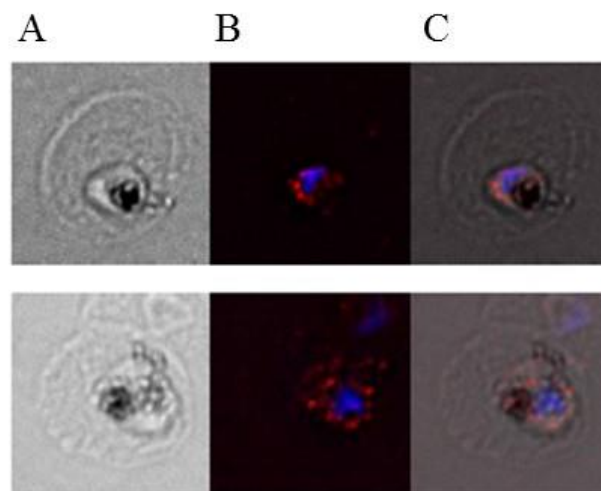


Supplementary Fig. 3

A. Cells probed with anti-*Pf*M1AAP antibodies

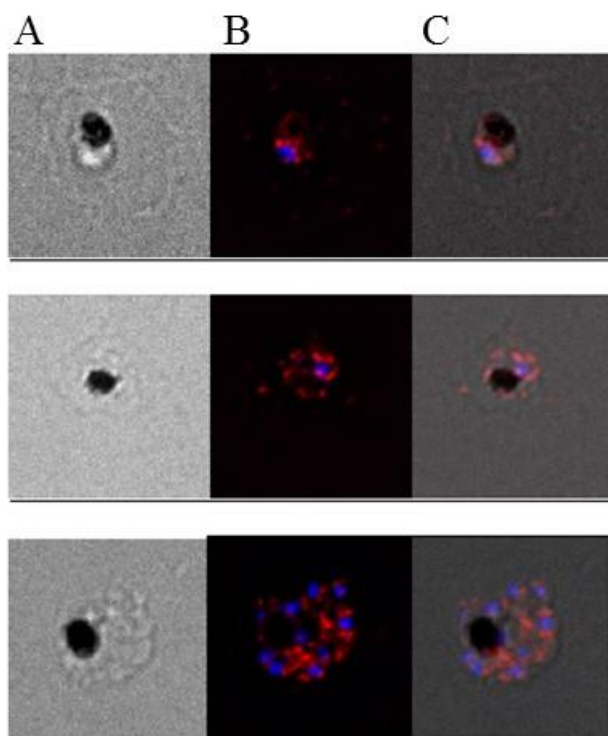


B. Cells probed with anti-*Pf*M17LAP antibodies

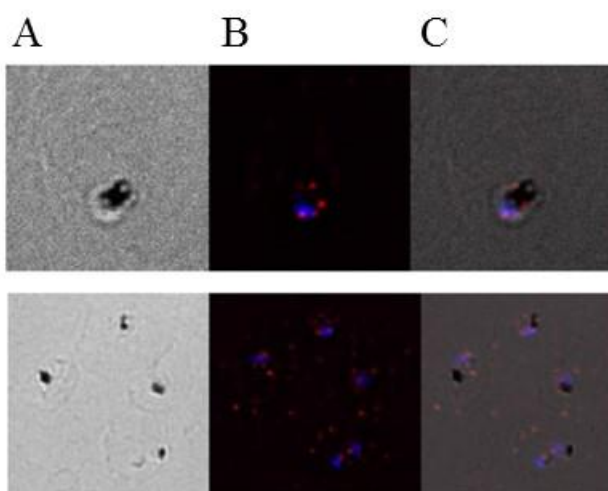


Supplementary Fig. 4

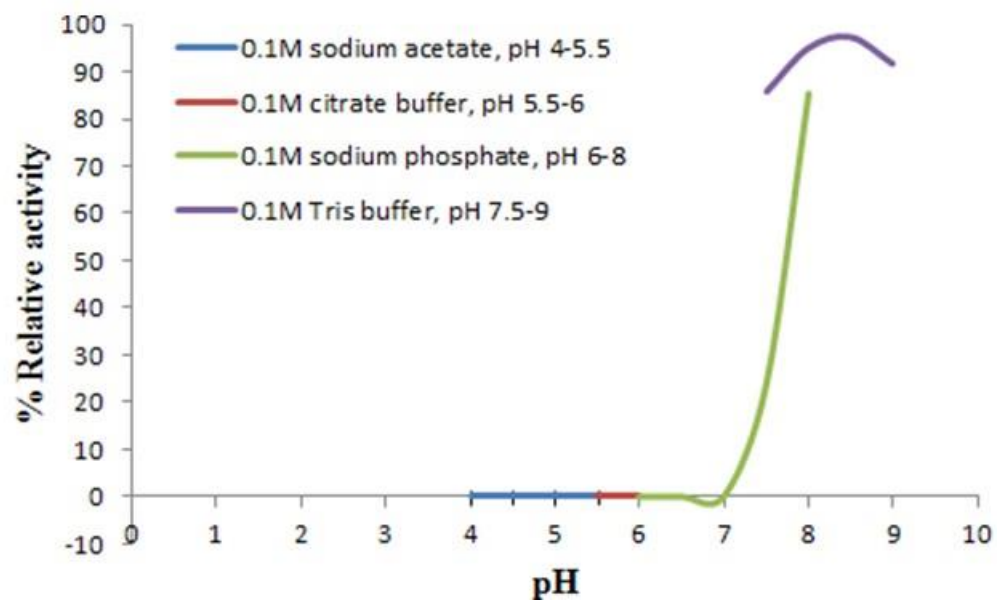
A. Cells probed with anti-*Pf*M1AAP antibodies



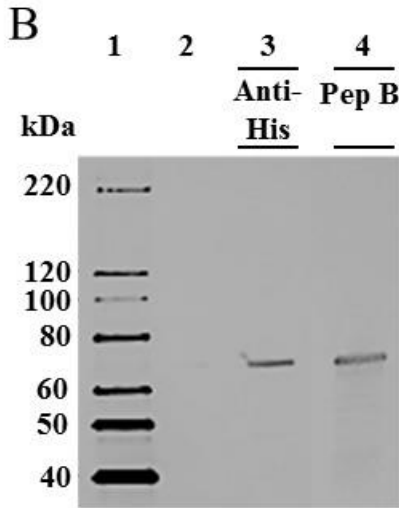
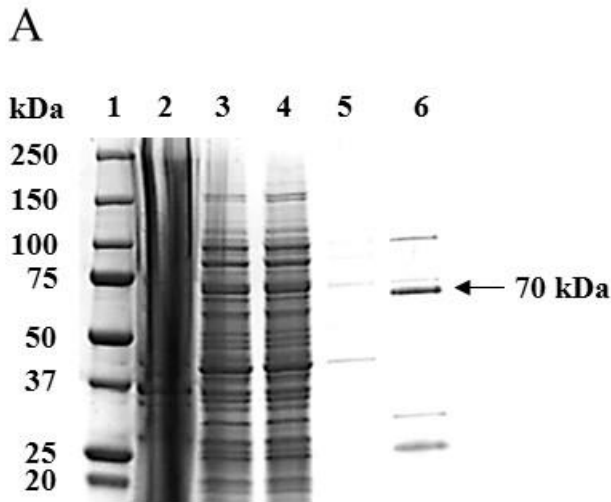
B. Cells probed with anti-*Pf*M17LAP antibodies



Supplementary Fig. 5

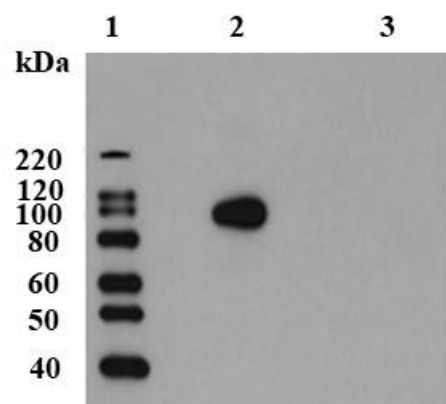


Supplementary Fig. 6

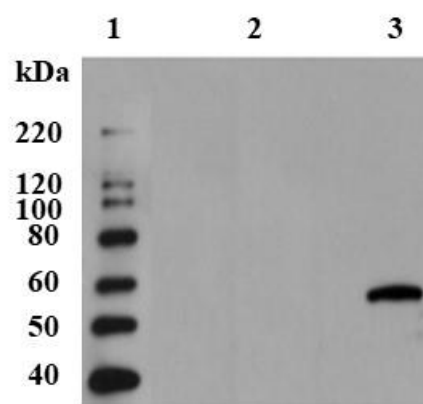


Supplementary Fig. 7

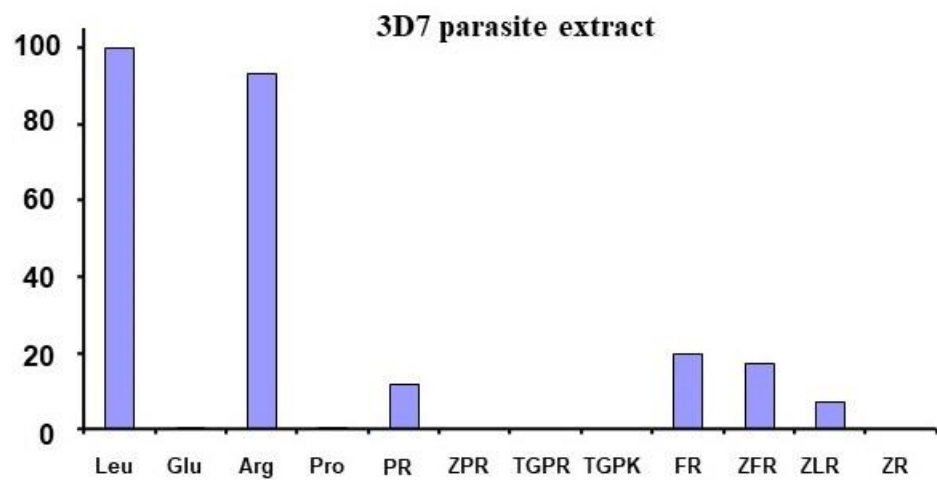
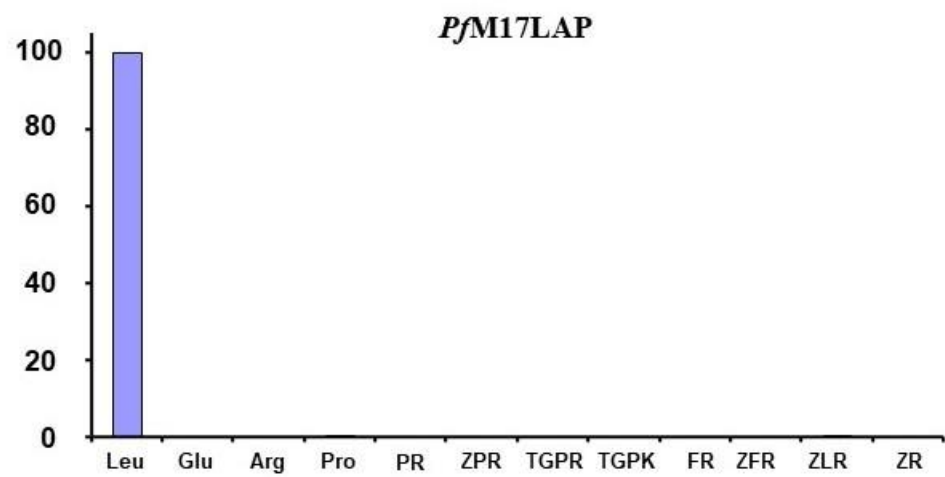
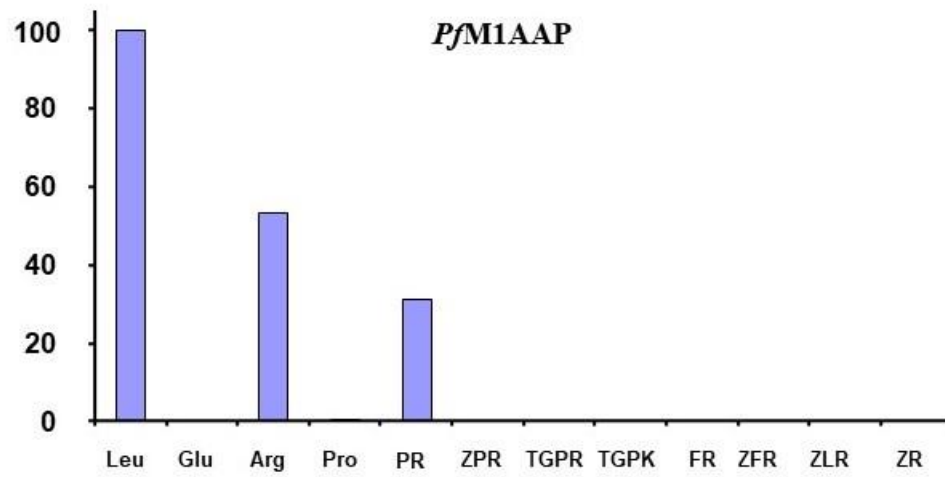
A



B

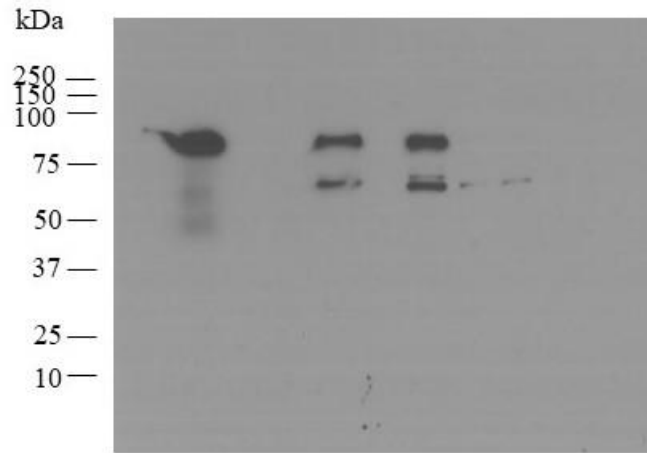


Supplementary Fig. 8

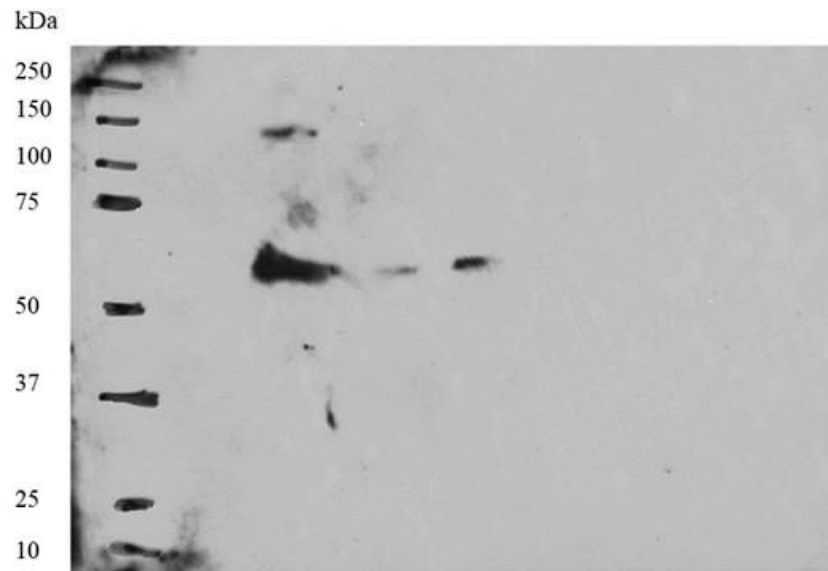


Supplementary Fig. 9A

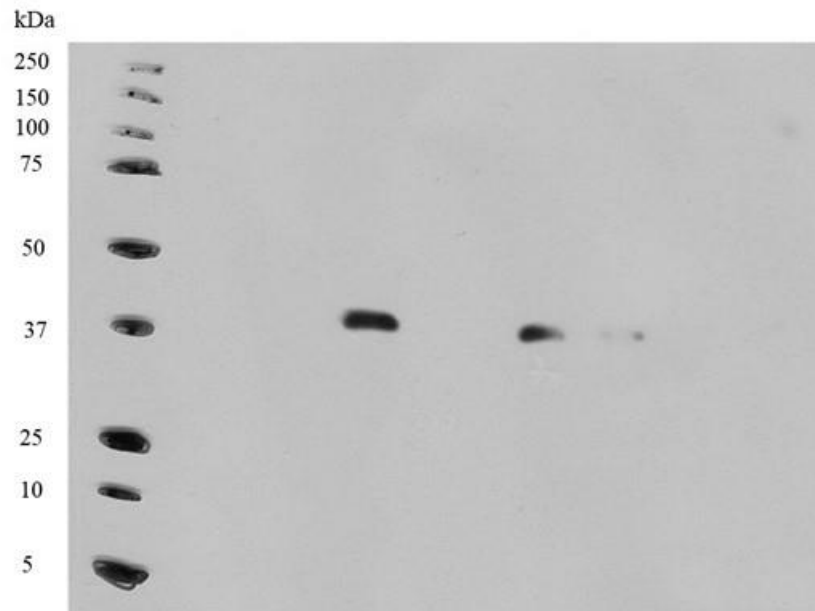
anti-*Pf*M1AAP



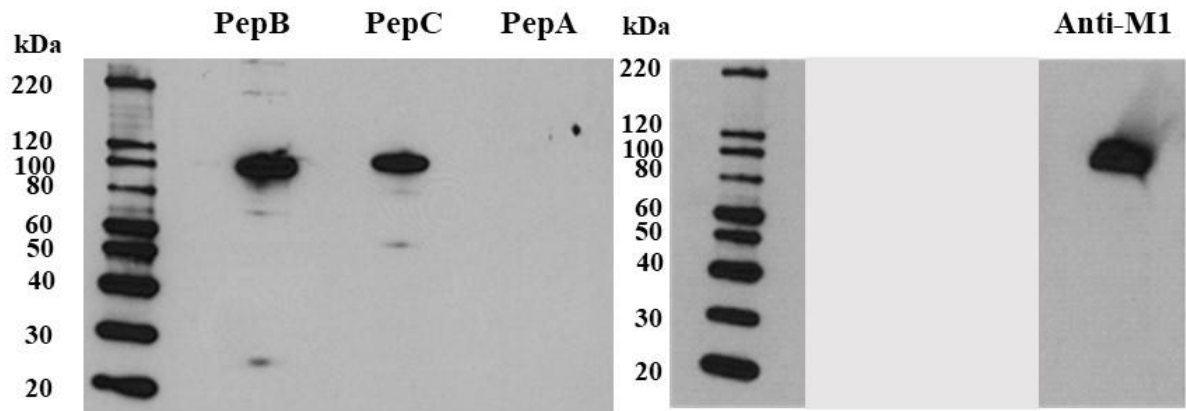
anti-*Pf*M17LAP



anti-plasmeprin 1



Supplementary Fig. 9B



Supplementary Fig. 9C

