Supplementary Material

Supplementary Table 1: Known cancer predisposition genes and potential chordoma genes included for germline pathogenic variant evaluation.

ABCB11	CYLD	GPC3	PMS2	SERPINA1	VHL	FLNA	PLXNB2
ALK	DDB2	HFE	POLD1	SETD2	WAS	GATA3	POLE
APC	DICER1	HMBS	POLE	SH2D1A	WRN	GNAS	POLRMT
ARHGAP26	DIS3L2	HRAS	POLH	SLC25A13	WT1	HGF	PTEN
ARID1A	DKC1	ΙΤΚ	PRKAR1A	SMAD4	XPA	HIST1H1E	PTPRC
ATM	DOCK8	KIT	PRSS1	SMARCA4	XPC	HLA-A	RPL22
AXIN2	EGFR	LYST	PTCH1	SMARCB1	STAB1	IDH1	RUNX1
BAP1	ELANE	MAX	PTEN	SMARCE1	AC007461.1	INPPL1	SETD2
BLM	ERBB4	MEN1	PTPN11	SOS1	UBBP4	IRF2	SF3B1
BMPR1A	ERCC2	MET	PTPRD	SRY	EIF5AL1	JAK1	SPTA1
BRCA1	ERCC3	MLH1	RAD51C	STAT3	APOB	KDM5C	STK11
BRCA2	ERCC4	MSH2	RAD51D	STK11	ARID2	KMT2A	TP53
BRIP1	ERCC5	MSH6	RB1	SUFU	B2M	LATS2	TXNIP
BUB1B	EXT1	MTAP	RECQL4	Т	BCOR	MAP3K4	USP9X
CBL	EXT2	MUTYH	RET	TERT	CACNA1A	MAX	VHL
CDC27	FAH	NBN	RHBDF2	TGFBR1	CDKN2A	MED12	ZFHX3
CDC73	FANCA	NF1	RMRP	TMEM127	CDKN2C	MGA	ZNF750
CDH1	FANCC	NF2	RUNX1	TNFRSF6	CSDE1	MLH1	
CDK4	FANCG	PALB2	SBDS	TP53	DAZAP1	NIPBL	
CDKN1B	FH	PBRM1	SDHA	TRIM37	EGFR	NUP133	
CDKN2A	FLCN	PDGFRA	SDHAF2	TSC1	EP300	PBRM1	
CEBPA	GATA2	РНОХ2В	SDHB	TSC2	EPAS1	PDGFRA	
CHEK2	GBA	<i>РІКЗСА</i>	SDHC	UROD	ERBB4	PGR	
COL7A1	GJB2	PIK3R1	SDHD	USP9X	FAT1	PLCG1	



Supplementary Figure 1: Average sequencing depth for normal and tumor (80 primary + 11 recurrent) samples among 80 chordoma patients

SBS							
signature	Mean	Range					
SBS1	0.10	0.04	0.20				
SBS2	0.01	0	0.15				
SBS5	0.68	0	0.95				
SBS8	0.10	0	0.32				
SBS13	0.01	0	0.17				
SBS40	0.05	0	0.70				
SBS44	0.01	0	0.29				

Supplementary Table 2A: The distribution of single-base substitutions (SBS) observed in 80 primary tumors

Supplementary Table 2B: The distribution of *de novo* indel (ID) signatures observed in 80 primary tumors

ID									
signature	Mean		Range						
А	0.27	0	1						
В	0.20	0	0.74						
С	0.08	0	1						
D	0.11	0	1						
E	0.34	0	0.88						

Supplementary Figure 2: Germline *TBXT* duplication in one chordoma patient. *TBXT* gene location: chr6:166571146-166582157 (hg19). Red indicates the duplication region.



Supplementary Table 3: Comparison of mutational profiles of chordoma patients to those of multiple other cancer types using MutaGene. Somatic mutations for each chordoma tumor sample were input to the program separately and compared to those of 9,450 cancer samples included in MutaGene. Ranking indicates the level of similarity between the input chordoma sample and each MutaGene cancer type. Numbers in the table reflect the number of chordoma samples (out of 80) in each ranking category for each cancer type.

Cancer type	1	2	3	4	5	6	7	8	9	10
Bladder Urothelial Carcinoma					1		1			
Bone Cancer	2	6	5	11	25	20	5	4	1	1
Brain Glioblastoma Multiforme							1			2
Brain Lower Grade Glioma		1	1		1				5	42
Breast Cancer	2						1		1	1
Chronic Lymphocytic Leukemia	21	11	11	18	6	7	1	1	1	1
Colon Adenocarcinoma								1	1	
Esophageal Adenocarcinoma								1		
Esophageal Squamous Carcinoma		2	1				1	1	3	11
Gastric Cancer	2									
GCB Lymphomas				1		2	4	11	52	4
Hepatocellular Carcinoma			2	3	7	11	15	24	5	1
Kidney Renal Clear Cell Carcinoma	14	26	18	10	4	5		1		2
Kidney Renal Papillary Cell Carcinoma	38	24	13	3						
Lung Squamous Cell Carcinoma							1	2	1	2
Ovarian Serous Cystadenocarcinoma		1	4	11	17	25	15	1	3	1
Pancreatic Cancer					1		1	2	2	7
Pancreatic Cancer Endocrine neoplasms										3
Renal Cancer	1	8	24	20	15	7			1	
Thyroid Cancer				1	3	3	34	31	4	1
Triple Negative Breast Cancer		1	1							1
Uterine Corpus Endometrial Carcinoma				2						

Supplementary Figure 3: Tumor mutational burden (TMB) in chordoma tumors in relation to other tumor types included in The Cancer Genome Atlas (TCGA)

Tumor types were ordered by median TMB values (Log10), from high to low. SKCM: skin cutaneous melanoma; LUSC: lung squamous cell carcinoma; LUAD: lung adenocarcinoma; BLCA: bladder urothelial carcinoma; STAD: stomach adenocarcinoma; COAD: colon adenocarcinoma; HNSC: head and neck squamous cell carcinoma; READ: rectum adenocarcinoma; DLBC: lymphoid neoplasm diffuse large B-cell adenocarcinoma; ESCA: esophageal carcinoma; OV: ovarian serous cystadenocarcinoma; LIHC: liver hepatocellular carcinoma; CESC: cervical squamous cell carcinoma and endocervical adenocarcinoma; UCEC: uterine corpus endometrial carcinoma; KIRP: kidney renal papillary cell carcinoma; GBM: glioblastoma multiforme; KIRC: kidney renal clear cell carcinoma; UCS: uterine carcinosarcoma; BRCA: breast invasive carcinoma; PAAD: pancreatic adenocarcinoma; SARC: sarcoma; LGG: brain lower grade glioma; PRAD: prostate adenocarcinoma; CHOL: cholangiocarcinoma; MESO: mesothelioma; ACC: adrenocortical carcinoma; Chordoma; KICH: kidney chromophobe; UVM: uveal melanoma; TGCT: testicular germ cell tumors; THYM: thymoma; PCPG: pheochromocytoma and paraganglioma; THCA: thyroid carcinoma; LAML: acute myeloid leukemia. The number of samples sequenced for each study is indicated on the top of the figure.



sample.id	%genome with MCN≥2	sample.id	%genome with MCN≥2	sample.id	%genome with MCN≥2	sample.id	%genome with MCN≥2
primary_P01	0.21	primary_P21	0.04	primary_P41	0	primary_P61	0.24
primary_P02	0.24	primary_P22	0.22	primary_P42	0.04	primary_P62	0.26
primary_P03	0.11	primary_P23	0	primary_P43	0.94	primary_P63	0.25
primary_P04	0.01	primary_P24	0.04	primary_P44	0.2	primary_P64	0.18
primary_P05	0.09	primary_P25	0.13	primary_P45	0.09	primary_P65	0
primary_P06	0.23	primary_P26	0	primary_P46	0	primary_P66	0.39
primary_P07	0	primary_P27	0	primary_P47	0	primary_P67	0.15
primary_P08	0.13	primary_P28	0	primary_P48	0	primary_P68	0
primary_P09	0.04	primary_P29	0	primary_P49	0	primary_P69	0
primary_P10	0.01	primary_P30	0	primary_P50	0.07	primary_P70	0
primary_P11	0.07	primary_P31	0.95	primary_P51	0.95	primary_P71	0.25
primary_P12	0.03	primary_P32	0.02	primary_P52	0.07	primary_P72	0
primary_P13	0	primary_P33	0.01	primary_P53	0.17	primary_P73	0
primary_P14	0.05	primary_P34	0.12	primary_P54	0.02	primary_P74	0.01
primary_P15	0.07	primary_P35	0	primary_P55	0	primary_P75	0
primary_P16	0.12	primary_P36	0	primary_P56	0.02	primary_P76	0.11
primary_P17	0	primary_P37	0.13	primary_P57	0.95	primary_P77	0.11
primary_P18	0.02	primary_P38	0.89	primary_P58	0	primary_P78	0.61
primary_P19	0.18	primary_P39	0.86	primary_P59	0.05	primary_P79	0.24
primary_P20	0	primary_P40	0	primary_P60	0.12	primary_P80	0.01

Supplementary Table 4: Percentage of genome showing MCN (major allele copy number) greater than or equal to two.

Supplementary Figure 4: Mutational signatures identified in 80 primary tumors. Bar graphs show the distribution of de novo single-base substitution (SBS) (panel A) and indel (panel B) signatures. Tables below the bar graphs show cosine similarities between de novo identified signatures and COSMIC signatures.

A: Nine de novo single-base substitution (SBS) mutational patterns



De novo extracted	COSMIC SBS Signatures ^a	Similarity ^b
Signature 96-A	Signature SBS1 (1.96%) & Signature SBS5 (83.56%) & Signature SBS34 (14.48%)	0.86
Signature 96-B	Signature SBS1 (4.14%) & Signature SBS2 (23.16%) & Signature SBS5 (45.54%) & Signature SBS13 (27.16%)	0.99
Signature 96-C	Signature SBS1 (12.96%) & Signature SBS5 (11.86%) & Signature SBS12 (31.98%) & Signature SBS21 (13.40%) & Signature SBS44 (29.80%)	0.98
Signature 96-D	Signature SBS1 (1.34%) & Signature SBS5 (57.76%) & Signature SBS43 (17.04%) & Signature SBS47 (16.24%) & Signature SBS52 (7.62%)	0.92
Signature 96-E	Signature SBS1 (4.72%) & Signature SBS5 (75.82%) & Signature SBS57 (19.46%)	0.84
Signature 96-F	Signature SBS1 (35.12%) & Signature SBS5 (64.88%)	0.96
Signature 96-G	Signature SBS1 (6.98%) & Signature SBS9 (20.86%) & Signature SBS18 (18.32%) & Signature SBS40 (53.84%)	0.93
Signature 96-H	Signature SBS1 (0.36%) & Signature SBS5 (65.76%) & Signature SBS39 (33.88%)	0.92
Signature 96-I	Signature SBS1 (8.62%) & Signature SBS5 (34.90%) & Signature SBS8 (48.30%) & Signature SBS19 (8.18%)	0.96

^a: The best linear combination of existing COSMIC signatures that approximates the detected *de novo* SBS in our data. ^b: The cosine similarity between our detected *de novo* mutational signatures and the linear combination of existing COSMIC mutation signatures.

B: Five major *de novo* indel signatures (ID)



De novo extracted	COSMIC Indel Signatures ^a	Similarity ^b
Signature INDEL-A		
Signature INDEL-B	Signature ID1 (66.06%) & Signature ID3 (15.20%) & Signature ID5 (18.74%)	0.99
Signature INDEL-C	Signature ID1 (24.02%) & Signature ID2 (75.98%)	1
Signature INDEL-D	Signature ID5 (28.04%) & Signature ID6 (10.80%) & Signature ID8 (61.16%)	0.98
Signature INDEL-E	Signature ID3 (23.00%) & Signature ID4 (9.42%) & Signature ID5 (40.02%) & Signature ID9 (27.56%)	0.93

^a: The best linear combination of existing COSMIC signatures that approximates the detected *de novo* SBS in our data. ^b: The cosine similarity between our detected *de novo* mutational signatures and the linear combination of existing COSMIC mutation signatures.

Pval #SVs in #CN Pval chr. Pval exponential fragment ID Position breakp. enrich. dist. breakpoints joins Links with other chrs sample segments 11:54996720-131215637;3:33853751-82347706; chrs 0 P09 4:16473677-186968109 96 16 0 0.01 7:80822537-114954467;8:117486242-137195855 9 0 0 0.02 P09 7:80822537-114954467 96 11:54996720-131215637;4:16473677-186968109 6 0 0.01 P09 22:18983600-50619038 96 NA P16 3:1236917-169015428 572 24 0 0 0 10:351295-111311574; 6 0.01 0 0.03 P16 6:56533939-64320531 572 P16 10:351295-111311574 572 18 0 0 0 0 P25 3:4902901-148147061 29 12 0 0.01 P63 6:90080348-170842513 109 76 NA 0 0 5 0 0 0.01 P66 11:15626344-38061210 44 10:43740177-46096635 P79 1:1481021-246175291 170 23 0 0 0 20:14410348-62070724 0 0 P79 19:35069904-54840360 170 17 0.01 5:13050718-40240915 P80 34 6 0.06 0 0 1:1481021-246175291;13:19749310-52735863

Supplementary Table 5: High-confidence chromothripsis events identified by ShatterSeek

Supplementary Figure 5: Comparison of clonal and subclonal mutation signatures. (A): Scatter plots of contribution of signatures of clonal SNVs vs. subclonal SNVs for 10 signatures. Each point represents one tumor. Pearson correlation coefficients are shown on the figure for each signature. (B): Average contribution of mutation signatures based on clonal and subclonal mutations. P values were obtained from Wilcoxon test (two-sided) for the difference of contribution of clonal and subclonal signatures. Figures were plotted based on n=80 skull-base chordoma samples. Differences were not statistically significant after the correction for multiple testing for any of the examined signatures.



Supplementary Table 6: Associations between genomic features and chordoma specific survival (CSS) and recurrence free survival (RFS), based on Cox's proportional hazard model. Models were adjusted for age, sex, pre- and post-surgery radiation therapy. HR=hazard ratio; Lower and Upper=95% confidence interval; SCNA: somatic copy number alteration, reference group is SCNA group 4. *Comparing patients harboring any of these events to those harboring none of these events.

Genomic features	HR	Lower	Upper	Р
PBRM1+	4.79	1.57	14.59	0.0058
CDKN2A/B+	0.88	0.20	3.92	0.86
SCNA_Group1 ^a	2.21	0.39	12.43	0.37
SCNA_Group2 ^a	3.39	0.62	18.48	0.16
SCNA_Group3 ^a	1.90	0.25	14.33	0.53
SCNA_Group5 ^a	1.22	0.11	14.04	0.87
1p deletion	0.94	0.31	2.83	0.92
3p deletion	3.94	1.09	14.29	0.04
3q deletion	2.81	0.89	8.89	0.08
4p deletion	1.06	0.33	3.38	0.93
4q deletion	1.04	0.31	3.42	0.95
9p deletion	2.01	0.56	7.17	0.28
9q deletion	2.46	0.80	7.55	0.12
10p deletion	2.25	0.75	6.80	0.15
10q deletion	2.28	0.75	6.92	0.15
13q deletion	2.79	0.85	9.23	0.09
14q deletion	3.21	0.93	11.06	0.06
18p deletion	3.03	1.08	8.55	0.04
18q deletion	3.08	1.09	8.75	0.03
22q deletion	5.88	1.85	18.68	0.0027
1q amplification	1.51	0.45	5.09	0.50
7p amplification	1.47	0.54	4.03	0.45
7q amplification	1.87	0.63	5.61	0.26
9p21.3	1.44	0.42	4.88	0.56
9p11.2	1.33	0.42	4.16	0.62
9q21.11	1.80	0.61	5.32	0.29
PBRM1+ or 22g deletion [*]	10.55	2.81	39.64	0.001

A. CSS.

B. RFS.

Genomic features	HR	Lower	Upper	Р
PBRM1+	5.72	2.68	12.19	6.4x10 ⁻⁶
CDKN2A/B+	1.68	0.66	4.31	0.28
SCNA_Group1 ^a	2.55	1.06	6.11	0.04
SCNA_Group2 ^a	1.77	0.78	3.99	0.17
SCNA_Group3 ^a	2.22	0.93	5.30	0.07
SCNA_Group5 ^a	1.39	0.48	4.04	0.54
1p deletion	1.41	0.77	2.57	0.26
3p deletion	1.53	0.86	2.73	0.15
3q deletion	1.84	1.05	3.24	0.03
4p deletion	1.66	0.83	3.33	0.15
4q deletion	1.84	0.89	3.81	0.10
9p deletion	3.36	1.74	6.94	0.0003
9q deletion	3.99	2.08	7.65	3.06x10 ⁻⁰⁵
10p deletion	1.71	0.97	3.04	0.07
10q deletion	1.56	0.89	2.71	0.12
13q deletion	1.08	0.62	1.88	0.78
14q deletion	2.13	1.20	3.79	0.01
18p deletion	2.39	1.36	4.20	0.002
18q deletion	2.65	1.51	4.65	0.001
22q deletion	3.74	1.89	7.38	0.0001
1q amplification	1.53	0.81	2.91	0.19
7p amplification	1.68	0.97	2.93	0.07
7q amplification	1.49	0.85	2.59	0.16
9p21.3	2.65	1.42	4.93	0.002
9p11.2	2.54	1.38	4.67	0.003
9q21.11	3.63	1.97	6.69	3.4x10 ⁻⁵
PBRM1+ or 9q21.11 deletion or 22q deletion*	4.22	2.34	7.62	1.77x10 ⁻⁶

C. Sensitivity analyses

CSS	PBRM1			9p21.3				9q	22q		
	HR	p value		HR	p value		HR	p value	HR	p value	
Final modela	4.79	0.01		1.44	0.56		1.80	0.29	5.88	0.003	
Final model + TMBb	5.13	0.01		1.43	0.59		1.85	0.29	6.16	0.003	
Final model + SCNAc	6.02	0.01		1.11	0.89		2.15	0.28	6.31	0.01	
Final model + Ki67	46.05	0.003		10.71	0.02		10.70	0.02	5.04	0.04	
Final model + GRRd	3.70	0.02		1.26	0.69		1.60	0.38	4.51	0.01	
Final modele	21.88	0.001		1.40	0.71		1.42	0.69	15.93	0.01	

RFS	Р	BRM1	9p21.3 9q					22q	
	HR	p value	HR	p value		HR	p value	HR	p value
Final modela	5.72	6.44E-06	2.65	0.002		3.63	3.44E-05	3.74	0.0001
Final model + TMBb	5.69	7.07E-06	2.61	0.003		3.62	5.43E-05	3.72	2.00E-04
Final model + SCNAc	5.17	4.58E-05	2.27	0.03		4.25	4.16E-04	3.26	0.003
Final model + Ki67	6.79	0.001	3.24	0.01		4.36	0.002	3.16	0.02
Final model + GRRd	5.13	3.40E-05	2.55	0.003		3.54	3.70E-05	3.54	3.00E-04
Final modele	7.21	0.004	2.96	0.004		3.01	0.04	5.53	2.00E-04

^{a-} Covariates: age, gender, pre-surgery radiation therapy (yes vs. no), post-surgery radiation therapy (yes vs. no)

^{b-} Tumor mutation burden

^{c-} Somatic copy number alterations

^{d-} Gross resection rate

e Restricting to patients who were diagnosed and operated in our hospital and did not have any treatment prior to surgeries

Supplementary Figure 6: Focal somatic copy number alteration regions (SCNA) identified by GISTIC A: Focal amplification (left) and deletion (right) regions



B: Focal SCNAs correlated with expression levels of key genes in the peak regions; *PBRM1* and *SETD2* expression in tumors without 3p21 deletion (n = 14) and in those with the deletion (n = 13). In B, data distributions are represented as boxplots where the line in the middle is the median, the first and the third quartiles are the box edges, the upper whisker extends from the edge to the largest value equal to $1.5 \times IQR$ from the edge (where IQR is the inter-quartile range) and the lower whisker extends from the edge to the smallest value at most $1.5 \times IQR$ of the edge, while data beyond these whiskers represent the outliers. The p-value of a two-sided Wilcoxon rank sum test is shown in the figure.



Supplementary Figure 7: Chromosomal somatic copy number alteration regions (SCNA) events in tumors without known drivers.



Supplementary Figure 8: Genomic events in relation to expression levels of the most relevant genes. In A– C, data distributions are represented as boxplots where the line in the middle is the median, the first and the third quartiles are the box edges, the upper whisker extends from the edge to the largest value equal to 1.5 × IQR from the edge (where IQR is the inter-quartile range) and the lower whisker extends from the edge to the smallest value at most 1.5 × IQR of the edge, while data beyond these whiskers represent the outliers. The p-value of a two-sided Wilcoxon rank sum test is shown in the figure.

A: *PBRM1* expression in tumors without *PBRM1* alteration (n = 24) and in those with the alteration (n = 3).



B: *CDKN2A* expression in tumors without *CDKN2A/2B* alteration (n = 23) and in those with the alteration (n = 4).



C. *SMARCB1* expression in tumors without arm-level chromosome 22q deletion (n = 18) and in those with the deletion (n = 9).



Supplementary Figure 9: The focal deletion of the *CDKN2A* region in the three paired tumor and metastasis samples. B: Blood (germline), no deletion detected; TM: thoracic metastasis, R: recurrence, LM: lymph node metastasis; *CDKN2A* region deletion was seen in all three paired tumor samples (TM, R, and LM).



Supplementary Figure 10: Images (×200 magnification) of haematoxylin and eosin (H&E) and immunohistochemical (IHC) staining of the two patients with IDH1 mutations (a-c patient P59, d-f patient P26). a and d: H&E staining; b: High expression of BRACHYURY; c and f: Strong positive for CYTOKERATIN; e: Positive EMA staining.

