

Expanded View Figures

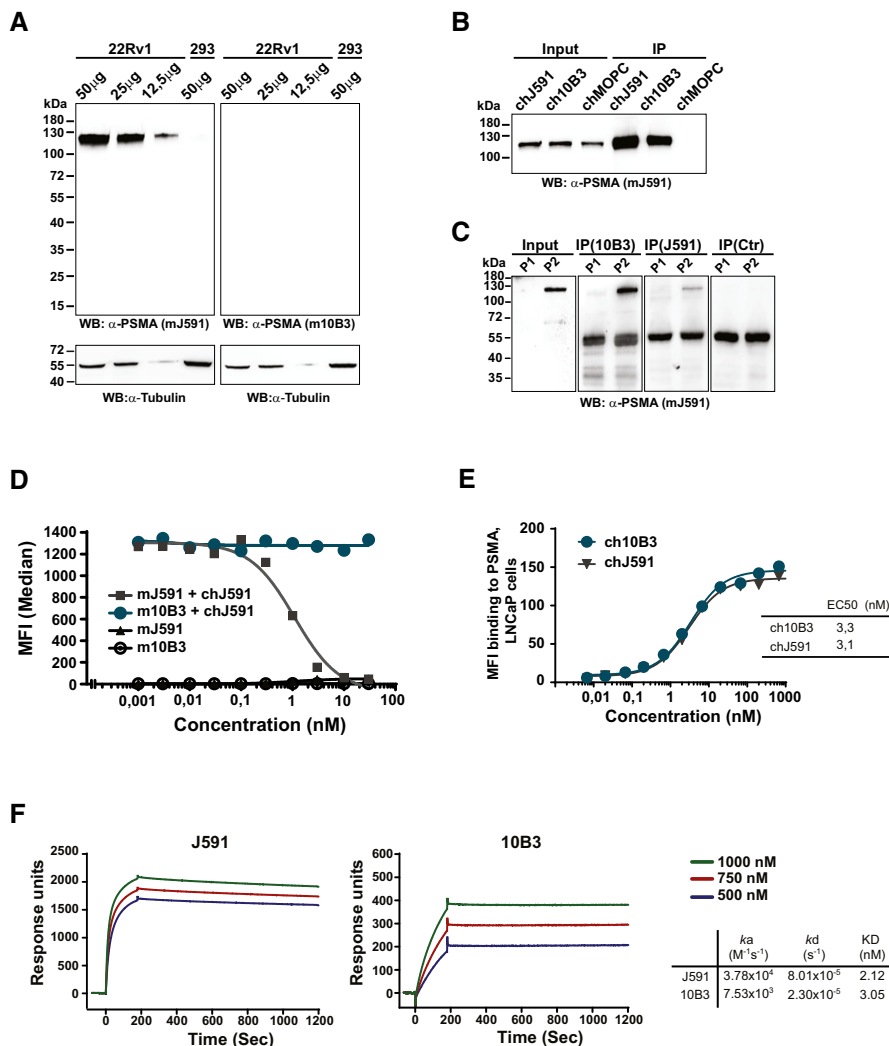


Figure EV1. In vitro characterization of the PSMA antibodies 10B3 vs J591.

- A** Western blot analysis was performed using the indicated amounts of protein extracts prepared from 22Rv1^{high} cells or from PSMA-negative HEK-293 cells. Blots were analyzed using anti-PSMA (mJ591 or m10B3) and anti-tubulin as a loading control.
- B, C** PSMA protein was immunoprecipitated from 22Rv1^{high} cells (B) and lung SCC samples (C) using chimeric J591, 10B3, and MOPC-21 (control) antibodies. The inputs (5%) and immunoprecipitates (30%) were analyzed by Western blot using murine J591 antibody.
- D** Mouse Sp2/0 cells transfected with human PSMA were incubated with the indicated concentrations of primary murine mAbs (mJ591, m10B3), followed by a chimeric J591 antibody (chJ591, 10µg/ml) and finally with fluorescence labeled mouse anti-human antibody for specific detection of chJ591 as indicated.
- E, F** Chimeric versions of both mAbs were bound to LNCaP cells (E) and to a protein A coated sensor chip (F) and analyzed by flow cytometry and SPR, respectively, as described in the Materials and Methods section.

Source data are available online for this figure.

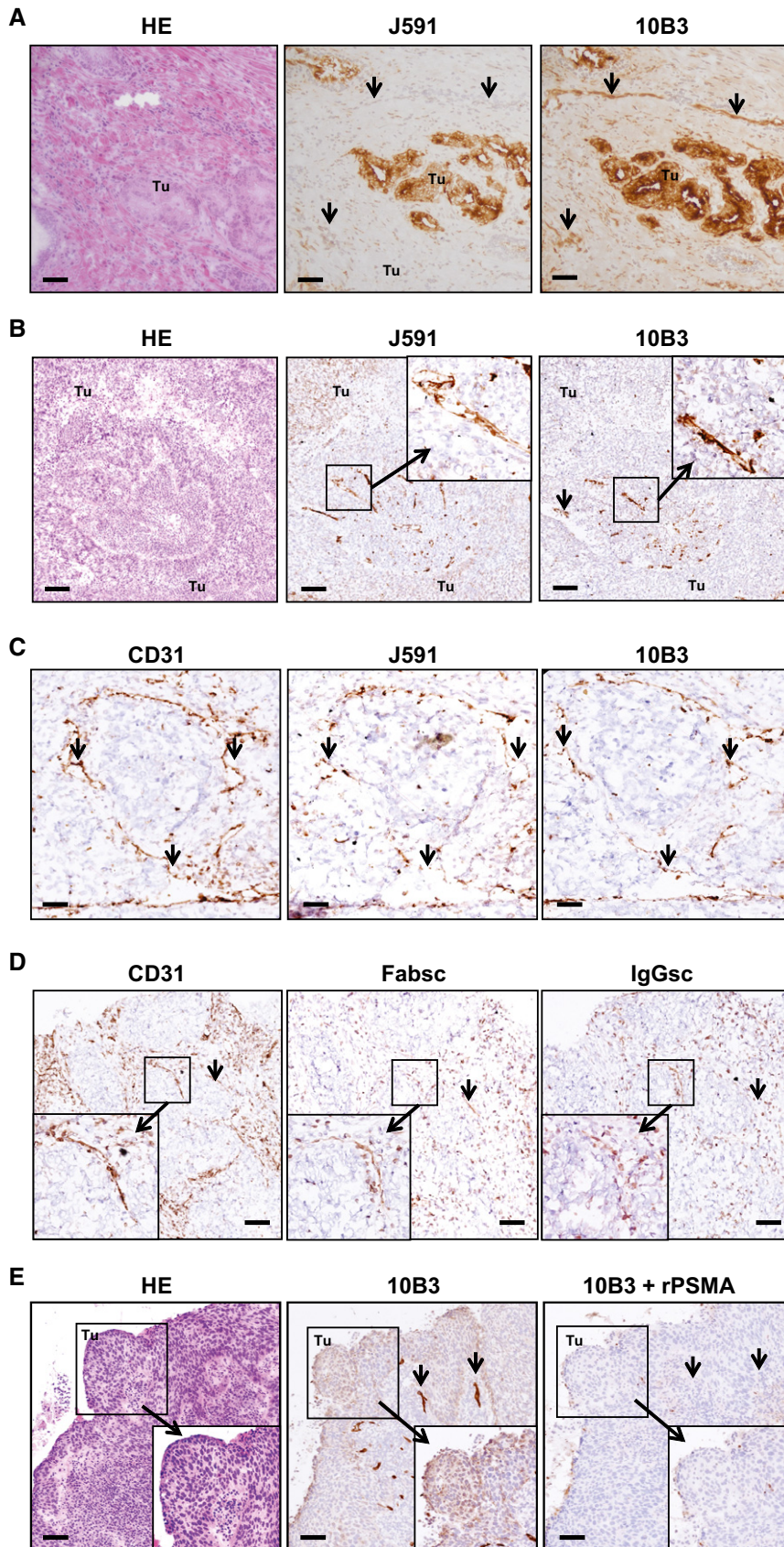


Figure EV2. Staining of exemplary prostate carcinoma and lung SCC samples.

A Directly consecutive 3- μ m sections obtained from prostate carcinoma samples were stained with 10B3 and J591 mAbs. Arrows point to vessels. Tu: tumor. HE: Hematoxylin/Eosin staining. Scale 30 μ m.

B–E Directly consecutive 3- μ m sections (obtained from the same lung SCC sample for each antibody panel) were analyzed by immunohistochemistry as described in the Materials and Methods section. Note that in (C, D) a tumor sample with predominant vascular expression of PSMA (cancer cells PSMA-very low intensity) was chosen to facilitate assessment of vascular staining. (B) Binding of murine 10B3 or J591 mAbs. Arrows point to vessels. Tu: tumor. HE: Hematoxylin/Eosin staining. Scale 30 μ m. (C) Staining with 10B3, J591, or anti-CD31 as marker for vessels. Arrows point to vessels. Scale 10 μ m. (D) Binding of anti-CD31 as well as biotinylated IgGsc and Fabsc molecules. Note that staining intensity with bsAbs is lower compared to CD31 due to the necessity to utilize a different detection protocol, and bsAb may additionally bind to tumor-infiltrating T cells, resulting in differential staining patterns. Arrows point to vessels. Scale 20 μ m. (E) Staining with 10B3 in the absence or presence of 50 μ g recombinant PSMA protein. HE: Hematoxylin/Eosin staining. Scale 40 μ m.

Source data are available online for this figure.

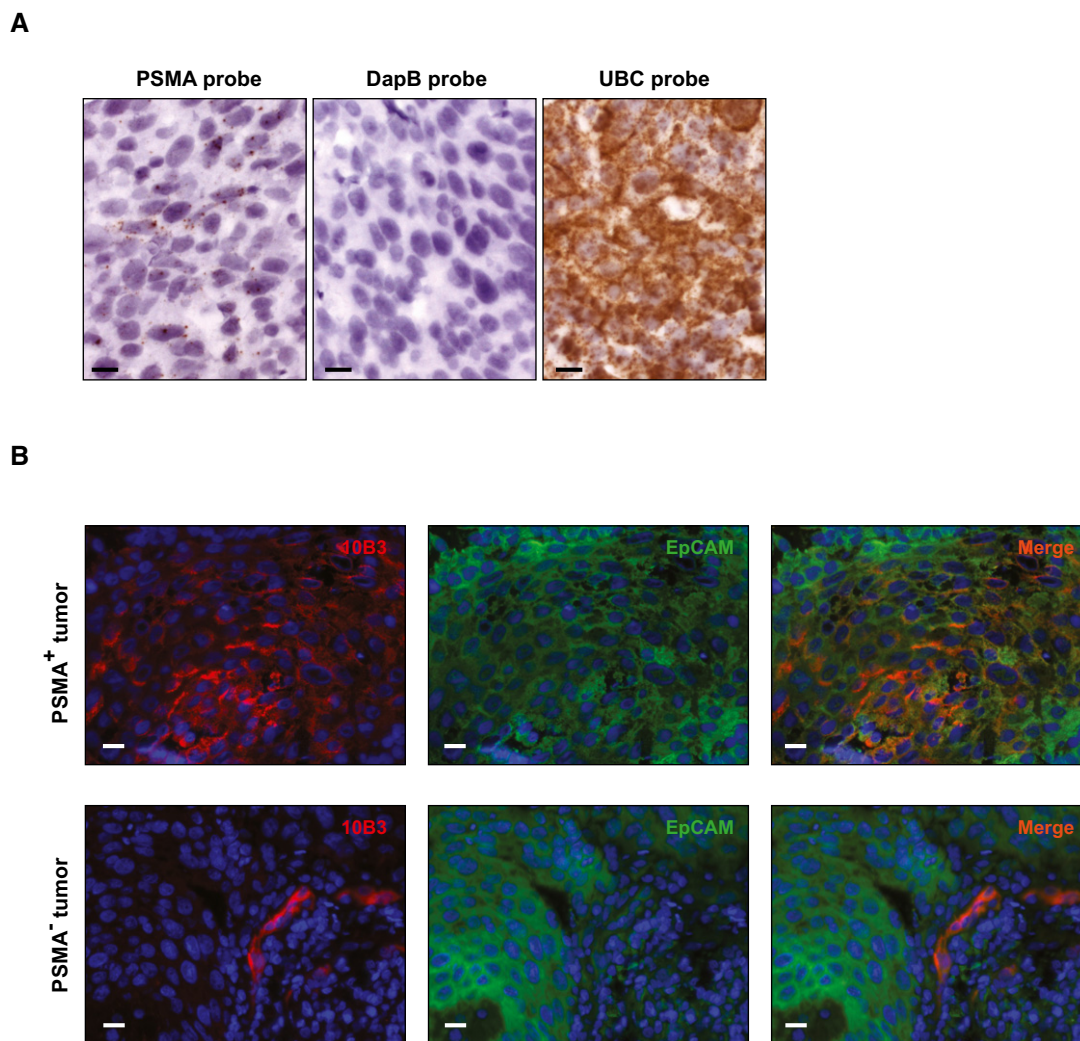


Figure EV3. PSMA RNA expression and membrane staining on lung SCC cryosections.

A RNA *in situ* hybridization was performed using the RNAscope® kit and probes against PSMA, DapB (negative control), and UBC (positive control) as described in the Materials and Methods section. Scale 10 μ m.

B Lung SCC tumor sections, (upper panel with PSMA-positive tumor lower panel with PSMA-negative tumor), were stained with 10B3 and a Cy3-labeled secondary antibody followed by a FITC-labeled antibody to the human EpCAM molecule. Scale 10 μ m.

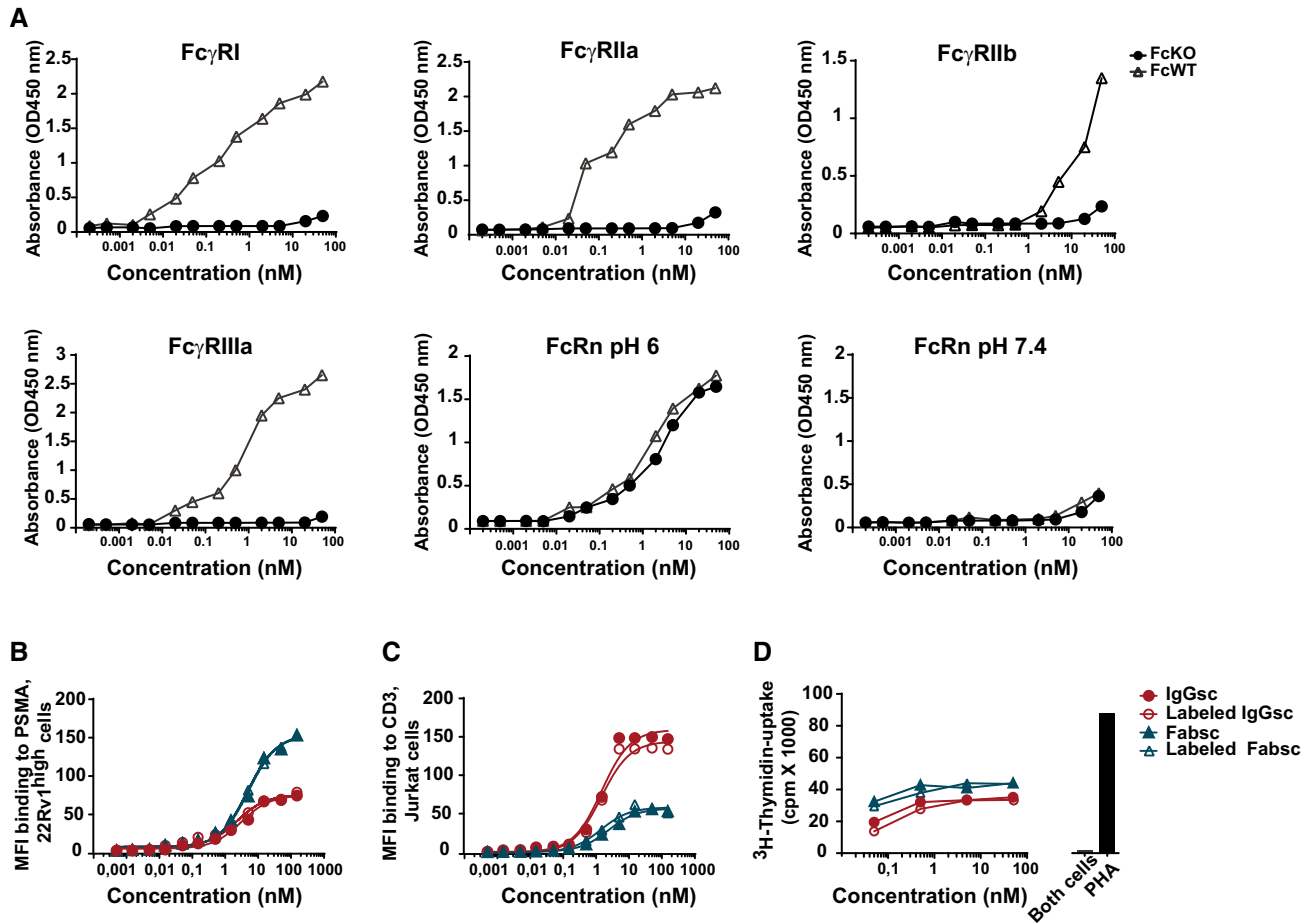


Figure EV4. Consequences of Fc silencing and chelation of CC-1.

- A Binding of CC-1 (FcKO) and a variant containing a wild type Fc part (FcWT) to the indicated his-tagged FcR was determined by ELISA. All experiments were performed at a neutral pH except for FcRn, where binding was additionally evaluated at a pH of 6. Means of duplicate measurements are shown.
- B, C BsAbs in the Fabsc and IgGsc format chelated or not to p-NCS-Bn-NODAGA were incubated with (B) 22Rv1^{high} (PSMA⁺) cells or (C) Jurkat (CD3⁺) cells followed by binding analysis by flow cytometry.
- D LNCaP cells were incubated with PBMC and the indicated concentrations of chelated or native bsAbs, and T-cell activation was assessed using a ³H thymidine uptake assay.

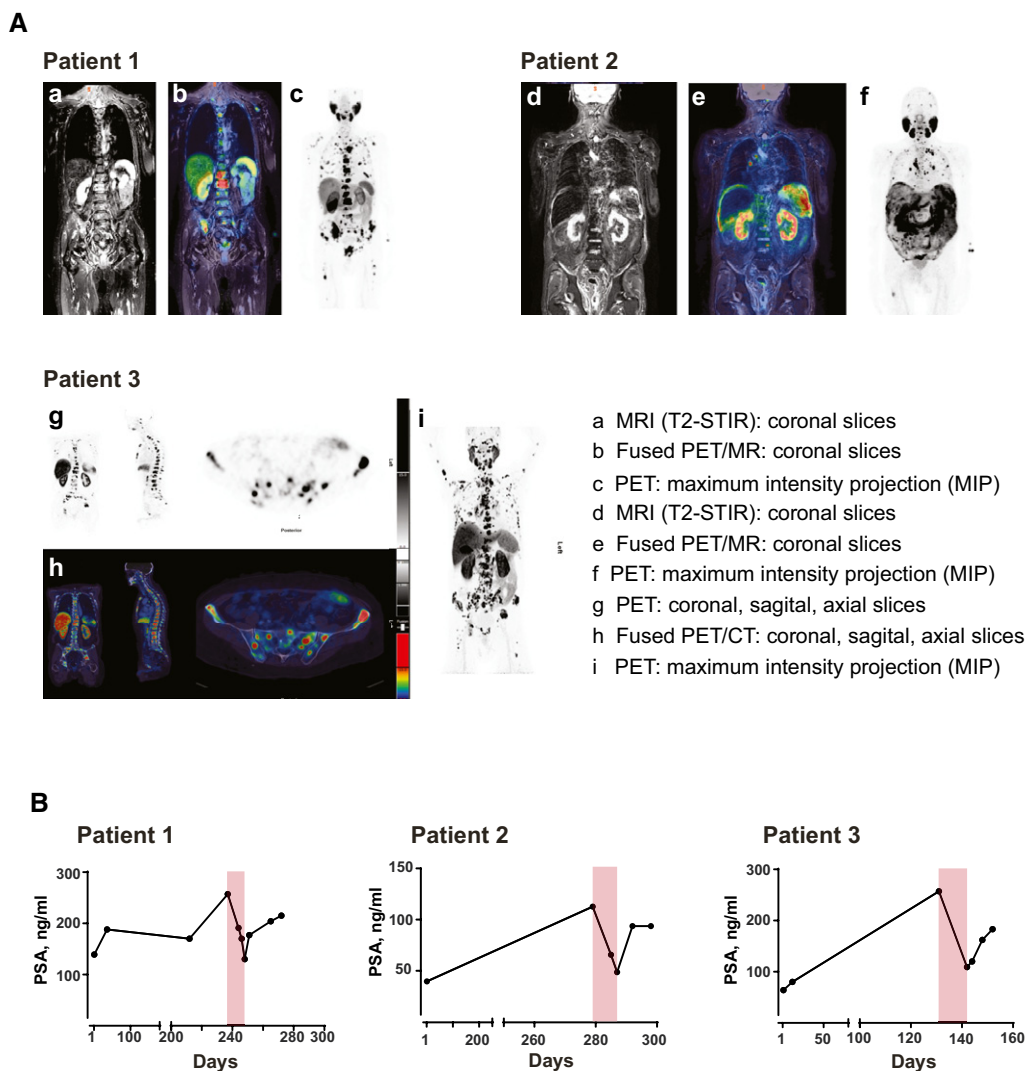


Figure EV5. PET scans and long-term PSA levels of the three treated patients with castrate resistant, metastasized prostate carcinoma.

A Pre-therapeutic PSMA expression of metastatic prostate cancer lesions was visualized by means of contrast enhanced whole body multiparametric PET/MRI (patient 1 and 2) or PET/CT (patient 3). Image data were acquired 60 min after i.v. injection of [^{18}F]-PSMA-1007 (250–325 MBq), a labeled peptide tracer for PET imaging that specifically binds to PSMA. It is used according to §13.2B AMG (German drug law) for PSMA-PET which has become standard clinical care in Germany. Representative images of the patients are shown: patient 1 presented with osseous and lymphonodal metastases with intense PSMA expression (a–c); patient 2 suffered from PSMA-expressing peritoneal carcinomatosis, multiple PSMA-expressing bone and lymph node metastases (d–f); in patient 3, osseous and lymphonodal metastases as well as local recurrence with intense PSMA expression are detected (g–i).

B Long-term PSA values monitored prior, during (highlighted in light red), and after CC-1 therapy. After documented failure of established treatment, patients were free of disease-specific therapy for at least 4 weeks prior to application of CC-1.