

A novel stress-inducible CmtR-ESX3-Zn²⁺ regulatory pathway essential for survival of *Mycobacterium bovis* under oxidative stress

Xiaohui Li¹, Liu Chen¹, Jingjing Liao¹, Jiechen Hui¹, Weihui Li², Zheng-Guo He^{*,1,2}

¹College of Life Science and Technology, Huazhong Agricultural University, Wuhan 430070, China

²State Key Laboratory for Conservation and Utilization of Subtropical Agro-bioresources, College of Life Science and Technology, Guangxi University, Nanning 530004, China.

*To whom correspondence should be addressed: College of Life Science and Technology, Guangxi University, Nanning 530004, China.

Email: hezhennguo2019@163.com

Tel: +86-771-3225146, Fax: +86-771-3225146

Running title: A novel CmtR-triggered antioxidant pathway in mycobacteria

Keywords: *Mycobacterium bovis*; oxidative stress; ESX-3; zinc ion; CmtR

Supplemental Information

Figure S1. Physical map of the *cmtR* gene region and amino acid sequence alignment of CmtR in *M. tuberculosis* H37Rv.

Figure S2. Assays for evaluating the DNA-binding ability of CmtR and the effect of H₂O₂ on the intracellular DNA-binding activity of EthR in *M. bovis* BCG.

Figure S3. Assays for studying the effects of H₂O₂ on the DNA-binding ability of CmtR and its mutant variants.

Figure S4. Assays for studying the effect of CmtR (C24S) on the growth of *M. bovis* BCG under H₂O₂ stress.

Figure S5. The functional categories of the potential target genes of CmtR in *M. tuberculosis* H37Ra.

Figure S6. Assays for studying the effects of H₂O₂ on the expression of *esx-3* operon genes in *M. bovis* BCG strains.

Figure S7. Assays for studying the growth of *M. bovis* BCG strains in 7H9 medium.

Figure S8. Assays for studying the survival of *M. bovis* BCG strains under H₂O₂ stress.

Table S1. Primers used in this study.

Table S2. Strains and plasmids used in this study.

A**B**

Mtu: Rv1994c	MLT CE -----MRESALARLGRALADP TR CRILVALLDGV CYP GGQ LA AHLGLTR	48
Mra: MRA_2010	MLT CE -----MRESALARLGRALADP TR CRILVALLDGV CYP GGQ LA AHLGLTR	48
Mbb: BCG_2011c	MLT CE -----MRESALARLGRALADP TR CRILVALLDGV CYP GGQ LA AHLGLTR	48
Msm: MSMEG_5603	-MQTA-----LHTDALARFGHALSDVTRTRILLSLNESPNYPADLAEQLGVSR	47
Mmi: MMAR_2139	-MPMKIADDAPPTASPREDLAGAVALFHSLSDPTRLAIARRRLADGERRVVDLTRELGLPQ	59
Mtu: Rv1994c	SNVSNHLS CL RG CG LVVATYEGRQVR Y ALADSHLARALGELVQVVLA---VDTDQ PC VAE	105
Mra: MRA_2010	SNVSNHLS CL RG CG LVVATYEGRQVR Y ALADSHLARALGELVQVVLA---VDTDQ PC VAE	105
Mbb: BCG_2011c	SNVSNHLS CL RG CG LVVATYEGRQVR Y ALADSHLARALGELVQVVLA---VDTDQ PC VAE	105
Msm: MSMEG_5603	QTLSNH LA CL RG CG LVVAVPEGR R TRYELADARIGRALDDL M GLVLD---VDPE CR CVGP	104
Mmi: MMAR_2139	STVSS HLA CL R D CG LI AGRPEGRQ V FYALAVPDL LD LFAAA ET VLAATGN AV T LC PNYGT	119
Mtu: Rv1994c	RAASGEAV EM TGS-----	118
Mra: MRA_2010	RAASGEAV EM TGS-----	118
Mbb: BCG_2011c	RAASGEAV EM TGS-----	118
Msm: MSMEG_5603	DGAVCG CG -----	112
Mmi: MMAR_2139	RPTRASASRRRG V KRAAPGVAD W SPGR	146

Figure S1. Physical map of the *cmtR* gene region and amino acid sequence alignment of CmtR in *M. tuberculosis* H37Rv. (A) The *cmtR*(*Rv1994c*) operon genes in the *M. tuberculosis* H37Rv genome are shown. The CmtR-regulated region (*cmtRp*) are indicated by black arrows. **(B)** Amino acid sequence alignment (generated using ClustalW) of CmtR for the analysis of cysteine residue conservation among different mycobacterial species. The sequences shown are Rv1994c from *M. tuberculosis* H37Rv (Mtu), MRA_2010 from *M. tuberculosis* H37Ra (Mra), BCG_2011c from *M. bovis* BCG (Mbb), MSMEG_5603 from *Mycobacterium smegmatis* (Msm), and MMAR_2139 from *Mycobacterium marinum* (Mmi). The terms marked in red represent different cysteine residue sites in the sequence.

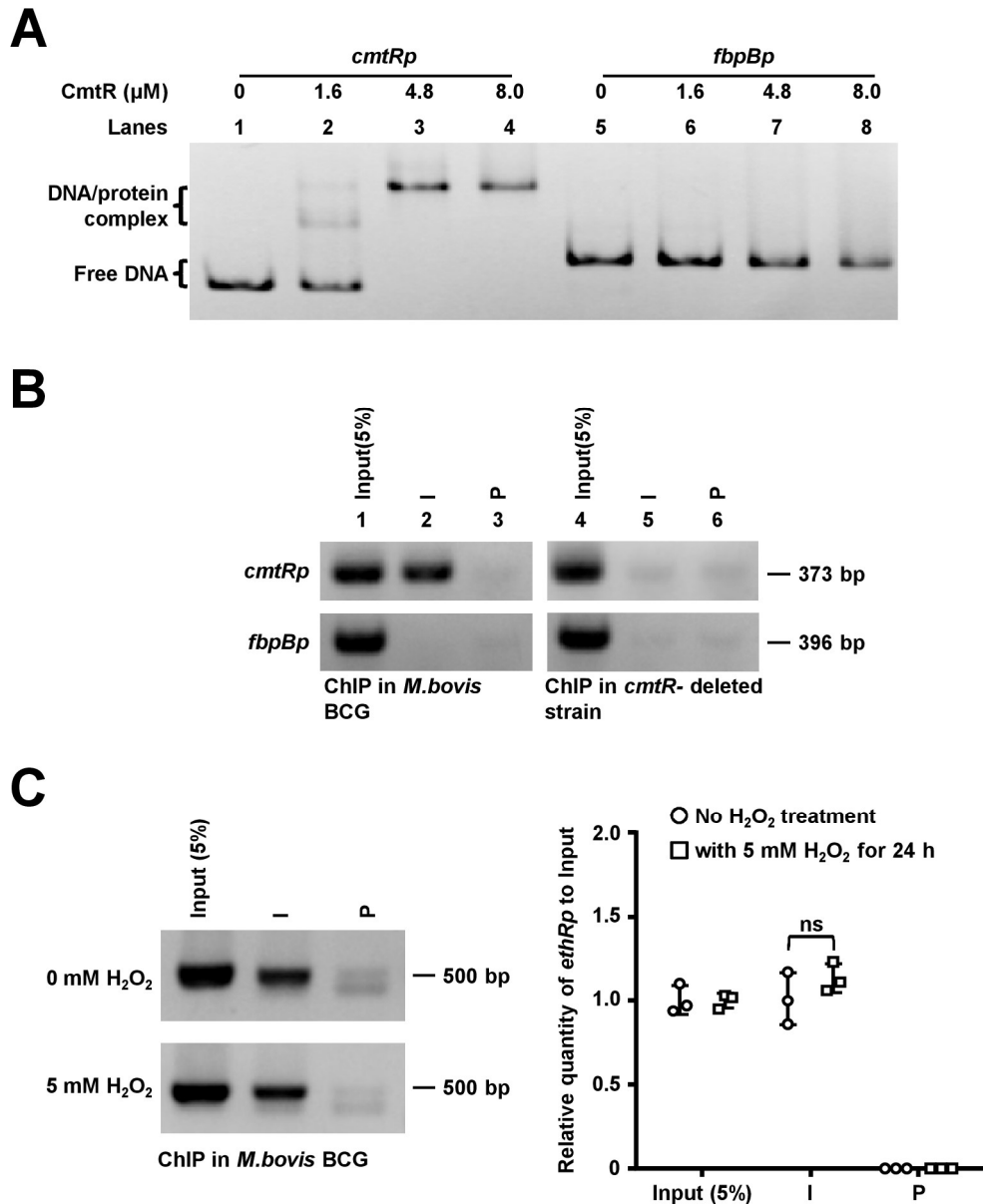
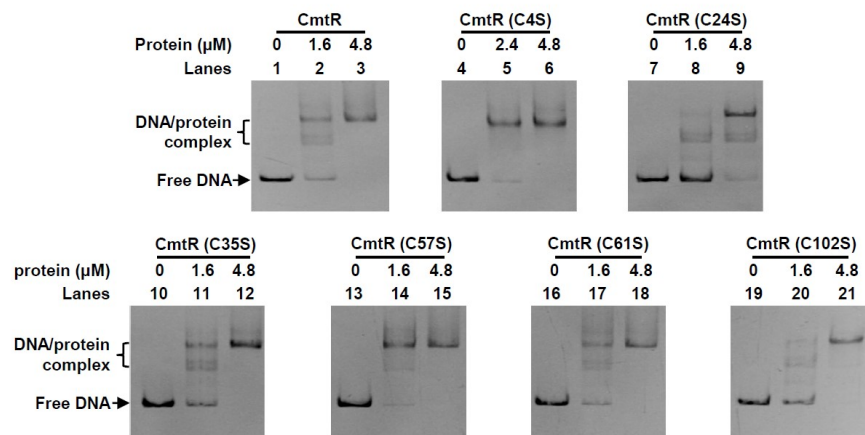


Figure S2. Assays for evaluating the DNA-binding ability of CmtR and the effect of H₂O₂ on the intracellular DNA-binding activity of EthR in *M. bovis* BCG. (A) EMSA. Either *cmtRp* promoter DNA (lanes 1-4) or *fbpBp* (lanes 5-8) were co-incubated with increasing quantities of CmtR, and the DNA-binding activities of the regulator were determined. (B) ChIP assays. ChIP using preimmune (P) or immune sera (I) against *cmtRp*. The mycobacterial promoter *fbpBp* was used as a negative control. The experiments were performed either in *M. bovis* BCG or in the *cmtR*-deleted strains. (C) Quantitative ChIP assays for evaluating the effect of H₂O₂ on the intracellular DNA-binding activity of EthR in *M. bovis* BCG. The Input (5%) indicates that the supernatant of the culture containing disrupted cells was diluted to 5% and used as the template for PCR. ChIP using P or I sera against EthR. Quantification was performed using qPCR (the right panel). Error bars represent the SD from three independent experiments. The *P*-values of the data were calculated by two-tailed Student's *t*-test using GraphPad Prism 7. Asterisks represent significant

difference (ns, not significant, two-tailed Student's *t*-test) between two groups.

A



B

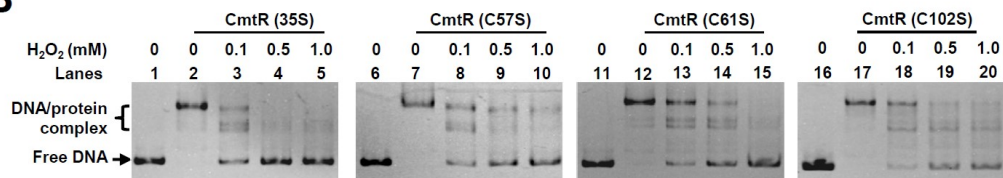


Figure S3. Assays for studying the effects of H₂O₂ on the DNA-binding ability of CmtR and its mutant variants. (A) EMSA for studying the DNA-binding ability of CmtR and its mutant variants. CmtR and its mutant variants were co-incubated with the *cmtRp* DNA substrate; increasing concentrations (1.6-4.8 μM) of the proteins are indicated at the top of the panels. (B) EMSA for studying the effects of H₂O₂ on the DNA-binding activity of CmtR mutant proteins. The protein concentration was 3.2 μM. The concentration (0.1-1 mM) of H₂O₂ is indicated at the top of the panels. The protein/DNA complexes are indicated by arrows on the left side of the panels.

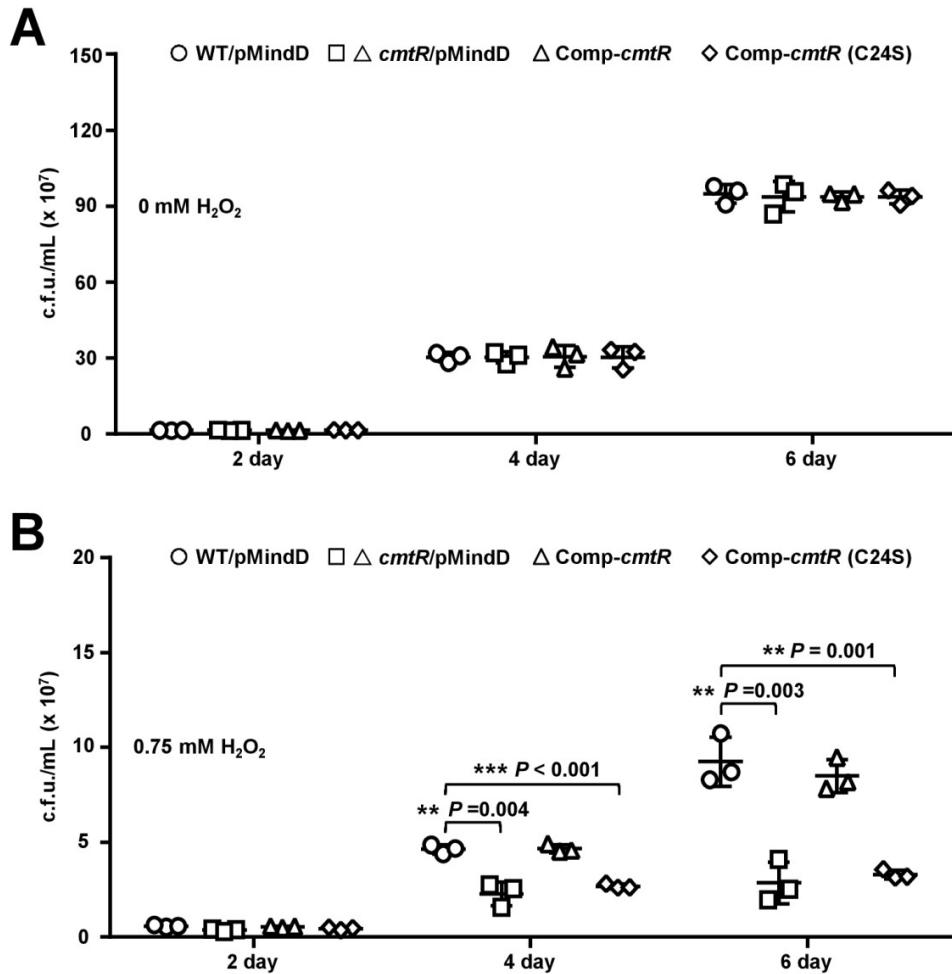


Figure S4. Assays for studying the effect of CmtR (C24S) on the growth of *M. bovis* BCG under H₂O₂ stress. Different mycobacterial strains were cultured in 7H9 medium (A) and the medium supplemented with 0.75 mM H₂O₂ (B). WT/pMindD represents the BCG/pMindD strain; Δ *cmtR*/pMindD represents the BCG *cmtR*::*hyg*/pMindD strain; Comp-*cmtR* represents the BCG *cmtR*::*hyg*/pMindD-*cmtR* strain; Comp-*cmtR* (C24S) represents the BCG *cmtR*::*hyg*/pMindD-*cmtR* (C24S) strain. Error bars represent the SD from three independent experiments. The *P*-values of the data were calculated by unpaired two-tailed Student's *t*-test using GraphPad Prism 7. Asterisks represent significant difference (**, *P*<0.01; ***, *P*<0.001, two-tailed Student's *t*-test) between two groups.

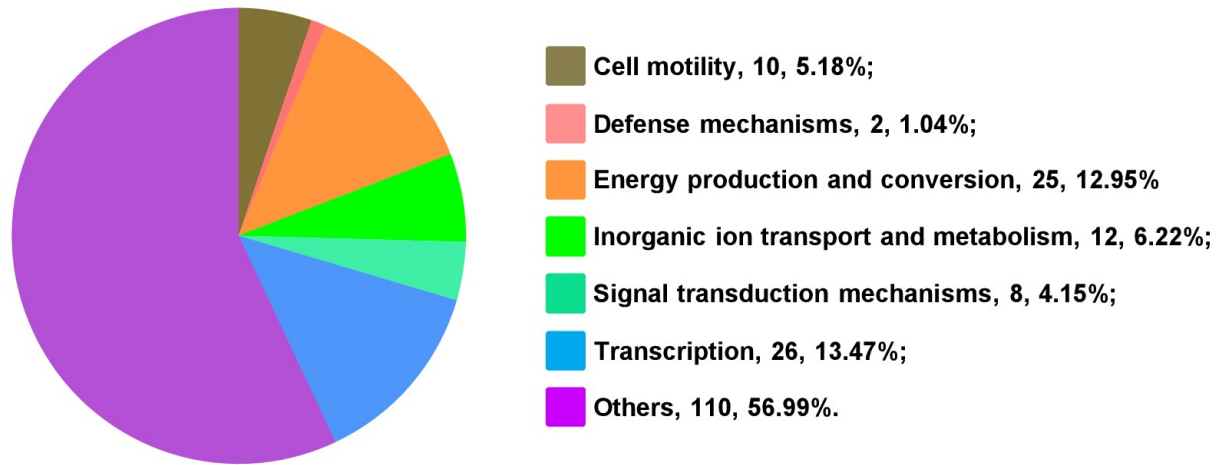


Figure S5. The functional categories of the potential target genes of CmtR in *M. tuberculosis* H37Ra. A functional classification of differentially expressed genes between wild-type and *cmtR*-deleted *M. tuberculosis* H37Ra strains in the context of Cluster of Orthologous Groups categories.

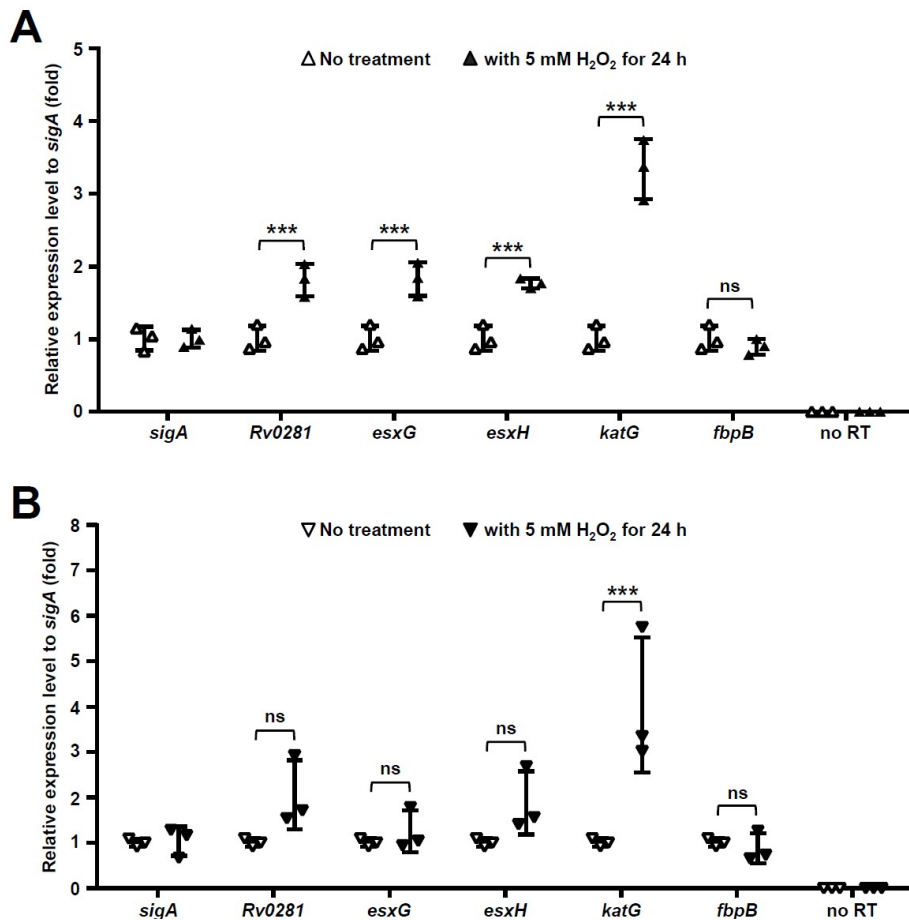


Figure S6. Assays for studying the effects of H₂O₂ on the expression of *esx-3* operon genes in *M. bovis* BCG strains. (A and B) qRT-PCR assays for studying the expression patterns of different genes (*Rv0281*, *esxG*, *esxH*, and *cmtR*) in *M. bovis* BCG WT (A) or *cmtR*-deleted strains (B) under treatment with indicated concentrations (0/5 mM) of H₂O₂ for 24 h. *KatG* and *fbpB* were used as control genes. Relative expression levels of the genes were normalized to those of *sigA*, which was used as an invariant transcript control using the $2^{-\Delta\Delta C_t}$ method. Error bars represent the SD from three independent experiments. The *P*-values of the data were calculated by unpaired two-tailed Student's *t*-test using GraphPad Prism 7. Asterisks represent significant difference (*, $P < 0.001$; ns, not significant, two-tailed Student's *t*-test) between two groups.**

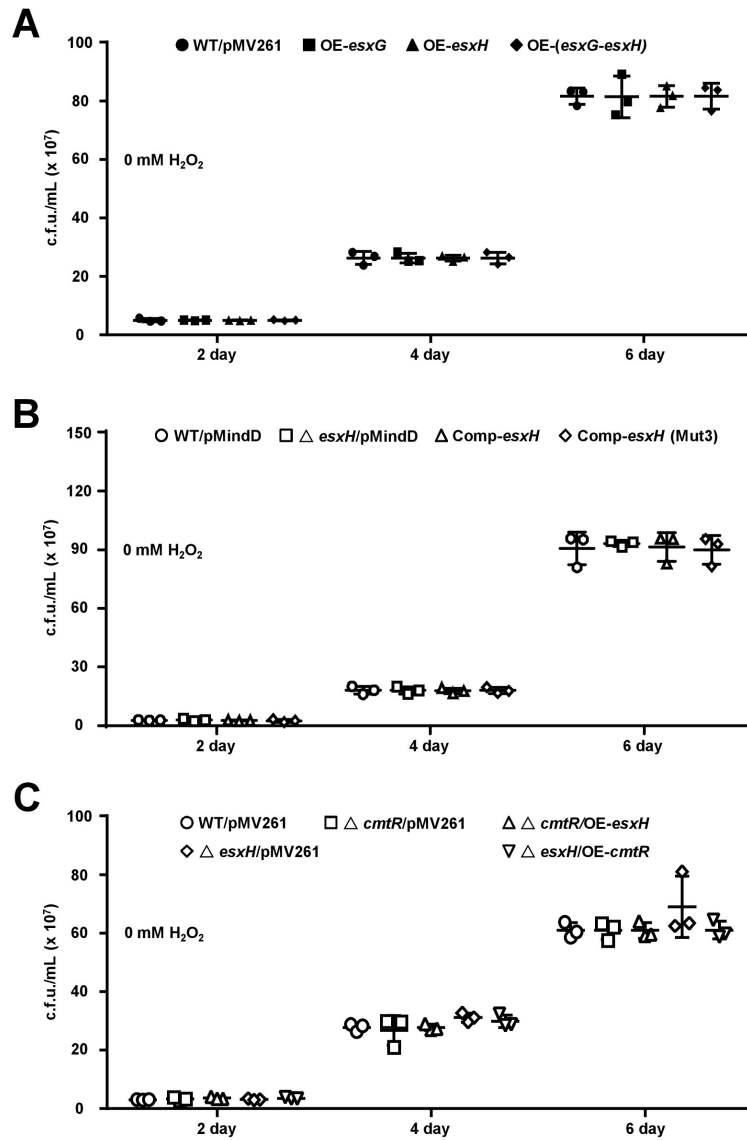


Figure S7. Assays for studying the growth of *M. bovis* BCG strains in 7H9 medium. (A-C) The recombinant mycobacterial strains were cultured in 7H9 medium for 6 days, and the bacterial counts were determined every 2 days. (A) WT/pMV261 represents the BCG/pMV261 strain; OE-*esxG*, OE-*esxH*, and OE-(*esxG-esxH*) represent the BCG/pMV261-*esxG*, BCG/pMV261-*esxH*, and BCG/pMV261-(*esxG-esxH*) strains, respectively; (B) WT/pMindD represents the BCG/pMindD strain; Δ *esxH*/pMindD represents the BCG *esxH*::*hyg*/pMindD strain; Comp-*esxH* represents the BCG *esxH*::*hyg*/pMindD-*esxH* strain; Comp-*esxH*-Mut3 represents the BCG *esxH*::*hyg*/pMindD-*esxH* (Mut3) (Mut3: H14A, H70A, H76A) strain. (C) WT/pMV261 represents the BCG/pMV261 strain; Δ *cmtR*/pMV261 represents the BCG *cmtR*::*hyg*/pMV261 strain; Δ *cmtR*/OE-*esxH* represents the BCG *cmtR*::*hyg*/pMV261-*esxH* strain; Δ *esxH*/pMV261 represents the BCG *esxH*::*hyg*/pMV261 strain; Δ *esxH*/OE-*cmtR* represents the BCG *esxH*::*hyg*/pMV261-*cmtR* strain.

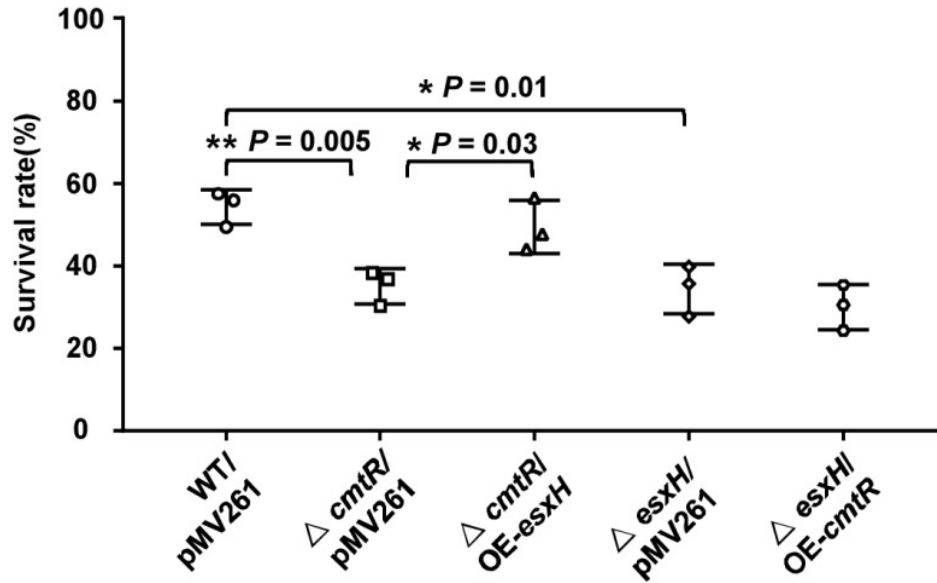


Figure S8. Assays for studying the survival of *M. bovis* BCG strains under H₂O₂ stress. The recombinant strains mentioned in the legend for Figure S7 C were cultured in Sauton's medium supplemented with 3 mM H₂O₂ for 24 h till an OD₆₀₀ value of 0.5 was achieved, and the bacterial counts were determined. The survival rates were calculated in terms of CFU (with H₂O₂)/CFU (without H₂O₂) for each strain. Error bars represent the SD from three independent experiments. The *P*-values of the data were calculated by unpaired two-tailed Student's *t*-test using GraphPad Prism 7. Asterisks represent significant difference (*, *P*<0.05; ***, *P*<0.001, two-tailed Student's *t*-test) between two groups.

Table S1. Primers used in this study

Usage	Construct	Primer name	Sequence (from 5' to 3')
Cloning and expression (pMV261 or pET-28a)	CmtR	Rv1994cf	CCACG <u>GAATTC</u> GTATGCTGACGTGTGAGATGCG
		Rv1994cr	CCGAT <u>CTAGAT</u> TCTCAGCTACCTGTCATCTCGA
Cloning and expression	IdeR	Rv2711f	CCTG <u>GAATTC</u> CCATGAACGAGTTGGTTGATAC
		Rv2711r	TCGTG <u>CTAGAT</u> TCAGACTTTCTCGACCTTGA
Cloning and expression (pET-28a-Sumo)	CmtR	Rv1994cf	TTTGAT <u>GGATCC</u> ATGCTGACCTGCGAGATGCG
		Rv1994cr	GGGGGAA <u>AAAGCTTTT</u> TAGCTGCCGGTCATTTCAACC
Cloning and expression (pET-28a-Sumo)	Zur	Rv2359f	AAAAAT <u>GGATCC</u> ATGAGTGCAGCCGGTGTG
		Rv2359r	CCAA <u>AAAGCTTTT</u> TAGCTCCGGCAGTCTGA
Cloning and expression (pET-28a-Sumo)	MabR	Rv2242f	ATTAG <u>GGATCC</u> GTGAACGACAATCAGTTGGC
		Rv2242r	ATTA <u>AAAGCTTTT</u> CAGTGC GGCGTCGGATAGT
Cloning and expression	EsxG	Rv0287f	CTCC <u>GAATTC</u> GGATGAGCCTTTTGGATGCTCA
		Rv0287r	TACCT <u>CTAGAG</u> GTTAGAACCCGGTATAGGTCG
Cloning and expression	EsxH	Rv0288f	CCCTCC <u>GAATTC</u> GGATGTCGCAAATCATGTAC
		Rv0288r	TAAC <u>CTAGAG</u> GTTAGCCGCCCATTTGGC
site-directed mutagenesis (C4S)	CmtR	C4Sf	TTAT <u>GGATCC</u> ATGCTGACGTCTGAGATGCG
		C4Sr	CGCATCTCAGACGTCAGCAT
site-directed mutagenesis (C24S)	CmtR	C24Sf	CCGACGCGGTCCCGGATTCTG
		C24Sr	CAGAATCCGGGACCGCGTCGG
site-directed mutagenesis (C35S)	CmtR	C35Sf	GATGGCGTTTCTATCCCGGCCAG
		C35Sr	CTGGCCGGGATAGGAAACGCCATC
site-directed mutagenesis (C57S)	CmtR	C57Sf	CATCTGTCGTCTTTGCGGGGCTG
		C57Sr	CAGCCCCGCAAAGACGACAGATG
site-directed mutagenesis (C61S)	CmtR	C61Sf	GTTTGC GGGGCTCCGGGCTGGTAG
		C61Sr	CTACCAGCCCGGAGCCCCGCAAAC
site-directed mutagenesis (C102S)	CmtR	C102Sf	CGACCAACCCTCTGTCGCCGAG
		C102Sr	CTCGGCGACAGAGGGTTGGTGC
site-directed mutagenesis (H14A)	EsxH	<i>EsxH</i> (H14A)f	ATGTTGGGTGCCCGGGGATAT
		<i>EsxH</i> (H14A)r	ATATCCCCGGCGGCACCCAACAT
site-directed mutagenesis (H70A)	EsxH	<i>EsxH</i> (H70A)f	TGCGGGCTATGCTGCGATGTCCA
		<i>EsxH</i> (H70A)r	TGGACATCGCAGCATAGCCCCGA
site-directed mutagenesis (H76A)	EsxH	<i>EsxH</i> (H76A)f	GTCCAGCACCGCTGAAGCCAACAC
		<i>EsxH</i> (H76A)r	GTGTTGGCTTCAGCGGTGCTGGAC
EMSA	Rv0280p	Rv0280pf	ACTGGCTAAATCCGTTGCCG
		Rv0280pr	AGCCGTAAACACCACAGAGG
EMSA	Rv0282p	Rv0282pf	CTACGCTGTTGAACGACTAC
		Rv0282pr	CATGCCAATGTCATCAC
ChIP/EMSA	fbpBp	Rv1886cpf	CATCGCACGCCACAAAC
		Rv1886cpr	GTGCCGATCATCAATCG
ChIP/EMSA	cmtRp	Rv1994cpf	GGGACTTCCTGTTCCGG

		Rv1994cpr	GATTCCCGCATCTCACAC
Clone to pMV261-lacZ	Rv0280p	Rv0280pLf	GACGGAATTCACTGGCTAAATCCGTTGCCG
		Rv0280pLr	GACGTCTAGAGCCGTAAACACCACAGAGG
Clone to pMV261-lacZ	Rv0282p	Rv0282pLf	GAGGGAATTCCTACGCTGTTGAACGACTAC
		Rv0282pLr	GGGTGTCTAGACATGCCAATGTCATCAC
Clone to pMV261-lacZ	fbpBp	Rv1886cpLf	GATGGAATTCATCGCACGCCACAAAC
		Rv1886cpLr	GCACTCTAGAGTGCCGATCATCAATCG
Clone to pMV261-lacZ	cmtRp	Rv1994cpLf	TTTCGAATTCGGGACTTCCTGTTCCGG
		Rv1994cpLr	GGTCTCTAGAGATTCCCGCATCTCACAC
Clone to pMV261-lacZ	LacZ	lacZf	TTCAGCTTATGAGGATGAGGGAAGCAAG
		lacZr	ATGCGCTAGCTTATTTTGACACCAGACCA
knock out of <i>cmtR</i>	CmtR	BCG_2011cUPf	TCACACTTAATTAAGTGTACTACCGACGTTAGCCAG
		BCG_2011cUPr	GGCGACTAGTAGATCAAATAGTACACC
knock out of <i>cmtR</i>	CmtR	BCG_2011cDNf	CCCGAAGCTTGCAGCAGACGACACGACTTGTG
		BCG_2011cDNr	AAACGCTAGCGCGATGGTCATCAGGGTGC
knock out of <i>esxH</i>	EsxH	BCG_0328UPf	CCCGACTTAATTAAGTCGCGGTGGTATCAAGG
		BCG_0328UPr	GGTGACTAGTCATCACAAATCCTCTCG
knock out of <i>esxH</i>	EsxH	BCG_0328DNf	TTAGAAGCTTCTAGCTCGCGCTACATGG
		BCG_0328DNr	TTATTTTCGCTAGCGCAAACGGTGTCCCAGG
qRT-PCR	SigA	RT-sigAf	TCGCGCCTACCTCAAACAG
		RT-sigAr	CGTACAGGCCAGCCTCGAT
qRT-PCR	CmtR	RT-CmtRf	TGCTGGATGGCGTTTGT
		RT-CmtRr	GGCCCTCATAGGTTGCGACTA
qRT-PCR	FbpB	RT-fbpBf	CGCGACATCAAGGTTTCAGTT
		RT-fbpBr	CCGGCATGACTATCGACAGT
qRT-PCR	Rv0281	RT-Rv0281f	GGGCTGCTGATCTATCTCCC
		RT-Rv0281r	TCGCTCGTTGTAGACCAGTT
qRT-PCR	EsxG	RT-esxGf	CGGCTCAGGCGTTTCAC
		RT-esxGr	CCGCCGCCACAAACC
qRT-PCR	EsxH	RT-esxHf	CAGGCCGCGTTGCA
		RT-esxHr	CTGCCACGCTGATACGT
qRT-PCR	KatG	RT-katGf	CCGAGATTGCCAGCCTTAA
		RT-katGr	GTTGACCTCCCACCCGACT

Notes: Restriction enzyme sites are underlined.

Table S2. Strains and plasmids used in this study

Plasmid or Strain	Relevant genotype or feature	Source or reference
Plasmid		
pET28a	Kan ^r , lacZ operon, T7 promotor, His-Tag	Novagen
pET28a- <i>cmtR</i>	<i>cmtR</i> inserted in <i>EcoRI-XbaI</i> of pET28a	This study
pET28a- <i>cmtR</i> (C4S)	<i>cmtR</i> (C4S) inserted in <i>EcoRI-XbaI</i> of pET28a	This study
pET28a- <i>cmtR</i> (C24S)	<i>cmtR</i> (C24S) inserted in <i>EcoRI-XbaI</i> of pET28a	This study
pET28a- <i>cmtR</i> (C35S)	<i>cmtR</i> (C35S) inserted in <i>EcoRI-XbaI</i> of pET28a	This study
pET28a- <i>cmtR</i> (C57S)	<i>cmtR</i> (C57S) inserted in <i>EcoRI-XbaI</i> of pET28a	This study
pET28a- <i>cmtR</i> (C61S)	<i>cmtR</i> (C61S) inserted in <i>EcoRI-XbaI</i> of pET28a	This study
pET28a- <i>cmtR</i> (C102S)	<i>cmtR</i> (C102S) inserted in <i>EcoRI-XbaI</i> of pET28a	This study
pET28a- <i>ideR</i>	<i>ideR</i> inserted in <i>EcoRI-XbaI</i> of pET28a	This study
pET28a-Sumo	Kan ^r , lacZ operon, T7 promotor, His-Tag	Novagen
pET28a-Sumo- <i>cmtR</i>	<i>cmtR</i> inserted in <i>BamHI-HindIII</i> of pET28a-Sumo	This study
pET28a-Sumo- <i>mabR</i>	<i>mabR</i> inserted in <i>BamHI-HindIII</i> of pET28a-Sumo	This study
pET28a-Sumo- <i>zur</i>	<i>zur</i> inserted in <i>BamHI-HindIII</i> of pET28a-Sumo	This study
pMV261	Kan ^r , pAL5000 replicon	This study
pMV261- <i>cmtR</i>	<i>cmtR</i> inserted in <i>EcoRI-XbaI</i> of pMV261	This study
pMV261- <i>esxG</i>	<i>esxG</i> inserted in <i>EcoRI-XbaI</i> of pMV261	This study
pMV261- <i>esxH</i>	<i>esxH</i> inserted in <i>EcoRI-XbaI</i> of pMV261	This study
pMV261- <i>esxG-esxH</i>	<i>esxG-esxH</i> inserted in <i>EcoRI-XbaI</i> of pMV261	This study
pMV261- <i>lacZ</i>	<i>lacZ</i> fused with null-promoter	
pMV261-hsp60- <i>lacZ</i>	hsp60 promoter fused with <i>lacZ</i>	This study
pMV261-Rv0280p- <i>lacZ</i>	Rv0280p promoter fused with <i>lacZ</i>	This study
pMV261-Rv0282p- <i>lacZ</i>	Rv0282p promoter fused with <i>lacZ</i>	This study
pMV261-fbpBp- <i>lacZ</i>	fbpBp promoter fused with <i>lacZ</i>	This study
pMV261-cmtRp- <i>lacZ</i>	cmtRp promoter fused with <i>lacZ</i>	This study
pMindD	Kan ^r , tet ^r , pAL5000 replicon	
pMindD- <i>cmtR</i>	<i>cmtR</i> inserted in <i>EcoRI-XbaI</i> of pMindD	This study
pMindD- <i>cmtR</i> (C24S)	<i>cmtR</i> (C24S) inserted in <i>EcoRI-XbaI</i> of pMindD	This study
pMindD- <i>esxH</i>	<i>esxH</i> inserted in <i>EcoRI-XbaI</i> of pMindD	This study
pMindD- <i>esxH</i> (Mut3)	<i>esxH</i> (H14A, H70A, H76A) inserted in <i>EcoRI-XbaI</i> of pMindD	This study
Strain		
<i>E. coli</i>		
DH5a	Host for plasmid construction	TaKaRa
BL21(λDE3)	Host for protein expression	TaKaRa
<i>M. bovis</i> BCG		
BCG/WT	<i>M. bovis</i> BCG wild-type	ATCC
BCG/ <i>cmtR::hyg</i>	BCG with <i>cmtR</i> replaced by <i>hyg</i>	This study
BCG/ <i>esxH::hyg</i>	BCG with <i>esxH</i> replaced by <i>hyg</i>	This study
BCG/pMV261	BCG with pMV261	This study

BCG <i>cmtR</i> ::hyg/pMV261	<i>cmtR</i> replaced by <i>hyg</i> in BCG, <i>cmtR</i> -deleted strain with pMV261	This study
BCG <i>cmtR</i> ::hyg/pMV261- <i>esxH</i>	<i>cmtR</i> replaced by <i>hyg</i> in BCG, <i>cmtR</i> -deleted strain with pMV261- <i>esxH</i>	This study
BCG <i>esxH</i> ::hyg/pMV261	<i>esxH</i> replaced by <i>hyg</i> in BCG, <i>esxH</i> -deleted strain with pMV261	This study
BCG <i>esxH</i> ::hyg/pMV261- <i>cmtR</i>	<i>esxH</i> replaced by <i>hyg</i> in BCG, <i>esxH</i> -deleted strain with pMV261- <i>cmtR</i>	This study
BCG/pMV261- <i>cmtR</i>	<i>cmtR</i> -overexpression Strain	This study
BCG/pMV261- <i>esxG</i>	<i>esxG</i> -overexpression Strain	This study
BCG/pMV261- <i>esxH</i>	<i>esxH</i> -overexpression Strain	This study
BCG/pMV261- <i>esxG</i> - <i>esxH</i>	<i>esxG</i> - <i>esxH</i> -overexpression Strain	This study
BCG/pMinD	BCG with pMinD	This study
BCG <i>cmtR</i> ::hyg/pMinD	<i>cmtR</i> replaced by <i>hyg</i> in BCG, <i>cmtR</i> -deleted strain with pMinD	This study
BCG <i>cmtR</i> ::hyg/pMinD- <i>cmtR</i>	<i>cmtR</i> replaced by <i>hyg</i> in BCG, <i>cmtR</i> -deleted strain with pMinD- <i>cmtR</i>	This study
BCG <i>cmtR</i> ::hyg/pMinD- <i>cmtR</i> (C24S)	<i>cmtR</i> replaced by <i>hyg</i> in BCG, <i>cmtR</i> -deleted strain with pMinD- <i>cmtR</i> (C24S)	This study
BCG <i>esxH</i> ::hyg/pMinD	<i>esxH</i> replaced by <i>hyg</i> in BCG, <i>esxH</i> -deleted strain with pMinD	This study
BCG <i>esxH</i> ::hyg/pMinD- <i>esxH</i>	<i>esxH</i> replaced by <i>hyg</i> in BCG, <i>esxH</i> -deleted strain with pMinD- <i>esxH</i>	This study
BCG <i>esxH</i> ::hyg/pMinD- <i>esxH</i> (Mut3)	<i>esxH</i> replaced by <i>hyg</i> in BCG, <i>esxH</i> -deleted strain with pMinD- <i>esxH</i> (H14A, H70A, H76A)	This study
BCG/pMVL1	BCG with pMV261- <i>lacZ</i> , negative control of β -galactosidase assays	This study
BCG/pMVL2	BCG with pMV261- <i>hsp60-lacZ</i> , positive control of β -galactosidase assays	This study
BCG/pMVL3	BCG with pMV261-Rv0280p- <i>lacZ</i>	This study
BCG/pMVL4	BCG with pMV261-Rv0282p- <i>lacZ</i>	This study
BCG/pMVL5	BCG with pMV261- <i>fbpBp-lacZ</i>	This study
BCG/pMVL6	BCG with pMV261- <i>cmtRp-lacZ</i>	This study
BCG <i>cmtR</i> ::hyg/pMVL1	<i>cmtR</i> replaced by <i>hyg</i> in BCG, <i>cmtR</i> -deleted strain with pMV261- <i>lacZ</i> , negative control of β -galactosidase assays	This study
BCG <i>cmtR</i> ::hyg/pMVL2	<i>cmtR</i> replaced by <i>hyg</i> in BCG, <i>cmtR</i> -deleted strain with pMV261- <i>hsp60-lacZ</i> , positive control of β -galactosidase assays	This study
BCG <i>cmtR</i> ::hyg/pMVL3	<i>cmtR</i> replaced by <i>hyg</i> in BCG, <i>cmtR</i> -deleted strain with pMV261-Rv0280p- <i>lacZ</i>	This study
BCG <i>cmtR</i> ::hyg/pMVL4	<i>cmtR</i> replaced by <i>hyg</i> in BCG, <i>cmtR</i> -deleted strain with pMV261-Rv0282p- <i>lacZ</i>	This study
BCG <i>cmtR</i> ::hyg/pMVL5	<i>cmtR</i> replaced by <i>hyg</i> in BCG, <i>cmtR</i> -deleted strain with pMV261- <i>fbpBp-lacZ</i>	This study
BCG <i>cmtR</i> ::hyg/pMVL6	<i>cmtR</i> replaced by <i>hyg</i> in BCG, <i>cmtR</i> -deleted strain with pMV261- <i>cmtRp-lacZ</i>	This study
<i>M. tuberculosis</i> H37Ra		
H37Ra/WT	Wild-type <i>M. bovis</i> H37Ra	ATCC
H37Ra/ <i>cmtR</i> ::hyg	H37Ra with <i>cmtR</i> replaced by <i>hyg</i>	This study