

# Figure S1. Area postrema cell types revealed by single-nucleus RNA sequencing, related to Figure 1.

(A) Native Fluoro-Gold fluorescence is visualized in coronal area postrema cryosections after intraperitoneal injection. (B) Workflow for preparation of single-nucleus RNA. (C) UMAP plot of all cell types analyzed by single-nucleus RNA sequencing. (D) Cell type assignments for cell clusters in (C) and associated UMAP plots indicating marker gene expression. (E) UMAP plot based on transcriptome data from all neurons, including seven area postrema cell types (pink, clusters 1-7) and three NTS cell types (green, clusters 8-10). (F) UMAP plots (left) and RNA *in situ* hybridization data from the Allen Brain Atlas (right) showing expression of signature genes in area postrema (pink, clusters 1-7) or NTS (green, clusters 8-10) neurons. (G) UMAP plots indicating expression of some neurotransmitter-related genes in area postrema excitatory neurons and enriched receptor genes in area postrema inhibitory neurons.



#### Figure S2. Validation of genetic tools, related to Figure 1.

(A) In coronal brain cryosections, RNA *in situ* hybridization data from the Allen Brain Atlas for *Slc6a2* (left) or GFRAL immunostaining (right) was compared with native tdTomato fluorescence observed in *Slc6a2-p2a-Cre; Rosa-LSL-tdTomato* (left) or *Gfral-p2a-Cre; Rosa-LSL-tdTomato* (right) mice, scale bars: 100  $\mu$ m. (B) Two color expression analysis in coronal area postrema cryosections of mice indicated, including either RNA *in situ* hybridization for *Glp1r*, *Slc6a2*, or *Calcr* (red), immunochemistry for GFRAL (red), and native GFP fluorescence (green), scale bar: 100  $\mu$ m. The numbers of co-labeled (yellow) or individually labeled (red, green) cells were counted (right).



## Figure S3. Validating chemogenetic control of area postrema neurons, related to Figure 3.

(A) Cre lines indicated were either injected in the area postrema with an AAV encoding Credependent  $G\alpha_s$ -DREADD-mCherry (top) or crossed to mice with a Rosa-LSL-L10GFP reporter allele (bottom). Native GFP and mCherry fluorescence was visualized in coronal area postrema cryosections (scale bar: 100  $\mu$ m), with the number of positive cells per section counted in caudal brainstem regions indicated. (B) Glp1r-ires-Cre mice were either injected in the area postrema with AAV-Flex-G $\alpha_s$ -DREADD (left) or crossed to mice with a Rosa-LSL-L10GFP reporter allele (right). Native GFP and mCherry fluorescence was visualized in wholemounts of vagal ganglia (scale bar: 100  $\mu$ m). (C) The area postrema of *Glp1r-ires-Cre* mice was injected with AAV-Flex- $G\alpha_s$ -DREADD (red) or AAV-Flex- $G\alpha_q$ -DREADD (blue). Area postrema neurons were acutely dissociated and transfected with the cAMP sensor cADDis. Top: Rows indicate responses (F/F, color scale) of individual neurons over time, with CNO (30  $\mu$ M) and forskolin (25  $\mu$ m) applied at times indicated (red bars). Y-axis colored bars (left: red; right: blue) and black bars indicated DREADD-positive and DREADD-negative neurons respectively. Bottom: Representative responses of DREADD-positive (red, blue) and DREADD-negative (black) neurons are shown to CNO (30  $\mu$ M) and forskolin (25  $\mu$ M); x: 100 seconds, y: 0.1 *F/F*. (D) The area postrema of *Glp1rires-Cre* and *Calcr-ires-Cre* mice were injected with AAV-Flex-G $\alpha_s$ -DREADD or AAV-FlextdTomato. CNO was injected intraperitoneally (IP), and two-color expression analysis was subsequently performed in coronal area postrema cryosections for native mCherry fluorescence (red) and Fos immunofluorescence (green), scale bar: 100  $\mu$ m. (E) No preference for grape or cherry flavor was observed in the absence of malaise induction. During conditioning, mice randomly received either cherry or grape stimuli, and all received saline injections (IP), n=14, mean ± sem, circles: individual data points. (F) Glp1r-ires-Cre mice were injected with AAVs encoding Cre-dependent  $G\alpha_q$ -DREADD-mCherry ( $G\alpha_q$ ) or tdTomato (Control), and analyzed for CNO-evoked flavor avoidance, n=5-8, mean  $\pm$  sem, circles: individual data points, p<.05.



# Figure S4. Physiological responses to optogenetic stimulation of area postrema neurons, related to Figure 3.

(A) Glp1r-ires-Cre (left) or Slc6a2-ires-Cre (right) mice were either injected in the area postrema with AAV-Flex-ChR2 or crossed to mice with a Rosa-LSL-L10GFP reporter allele. mCherry immunofluorescence was visualized in coronal area postrema cryosections (scale bar: 100  $\mu$ m), with the number of mCherry (ChR2) and GFP positive cells per section counted in caudal brainstem regions indicated. (B) The area postrema of *Glp1r-ires-Cre* mice was injected with AAV-Flex-ChR2 (AAVAP-ChR2-mCherry mice). Area postrema neurons were acutely harvested and responses to light (blue bar) were measured by whole-cell voltage clamp recordings, x: 2 seconds, y: 20 picoamperes. (C) Two-color expression analysis in area postrema cryosections after area postrema illumination in Glp1r-ires-Cre mice injected with AAV-Flex-ChR2 (top) or AAV-Flex-Yfp (bottom), red: native mCherry fluorescence (top) or pseudocolored native YFP fluorescence (bottom); green: Fos immunofluorescence, scale bar: 100  $\mu$ m. (D) Physiological responses to area postrema illumination (blue bar) in Glp1r-ires-Cre mice previously injected in the area postrema with either AAV-Flex-ChR2 (GLP1R) or AAV-Flex-Yfp (Control), x: 100 seconds, y: 1 cmH<sub>2</sub>O, y': 5 mmHg, y": 1 beat/breath per minute. (E) Quantifying physiological changes to stimulating area postrema neuron subtypes indicated (n=3-14, mean  $\pm$  sem, p<.05, \*\*p<.01, \*\*\*p<.001, statistical comparisons to control).



No-RESCUE Glp1r-RESCUEVAGUS

# Figure S5. Validating and analyzing loss-of-function approaches in area postrema neurons, related to Figures 4, 5.

(A) Two-color expression analysis involving Fos immunohistochemistry (green) and native tdTomato fluorescence (red) in coronal area postrema cryosections after intraperitoneal injection of lithium chloride (LiCl) or lipopolysaccharide (LPS) in Glp1r-ires-Cre; Rosa-LSL-tdTomato or *Calcr-ires-Cre; Rosa-LSL-tdTomato* mice, scale bar: 100  $\mu$ m. The numbers of co-labeled (yellow) or individually labeled (red, green) cells were counted (right). (B) Ad libitum fed Non-ABLATE (black) and *Glp1r*-ABLATE<sup>AP</sup> (red) mice were injected with either exendin-4 (filled bars) or saline (unfilled bars), and cumulative food consumption was measured at 1, 3, 5, and 24 hours after injection, n=3-8, mean ± sem, \*\*p<.01, \*\*\*\*p<.0001. (C) The areas postrema of Gfral-p2a-Cre; Rosa-LSL-DTR and SIc6a2-p2a-Cre; Rosa-LSL-DTR mice were injected with DT (+DT) or saline (-DT). Immunohistochemistry for DTR was performed in coronal area postrema sections, scale bar: 100  $\mu$ m. (D) Counts of DTR-expressing cells in area postrema (AP) and NTS from mice in C, n=3-13 sections from 2-5 mice, circles: individual data points, \*\*p<.01, \*\*\*\*p<.0001. (E) Counts of *Glp1r*-positive cells across sections from *in situ* hybridization experiments described in Figure 5A, n=6-10 sections from 2-4 mice, mean ± sem, \*\*p<.01, \*\*\*\*p<.0001. (F) Fos immunohistochemistry in area postrema sections of mice indicated after intraperitoneal exendin-4 injection, scale bar: 100  $\mu$ m. (G) RNA *in situ* hybridization for *Glp1r* in vagal ganglia cryosections (top, scale bar: 100  $\mu$ m) and flavor avoidance responses conditioned to exendin-4 (bottom, n=7 mice, mean  $\pm$  sem, circles: individual data points) in mice indicated. (H) Counts of *Glp1r*-positive cells from RNA *in situ* hybridization experiments in vagal ganglia of mice indicated, n=4-9 sections from 3-4 mice, mean ± sem, \*\*p<.01.



#### Figure S6. Anatomical mapping of area postrema neuron projections, related to Figure 6.

(A) The PBN and VLM (ventrolateral medulla) of Glp1r-ires-Cre mice were injected with AAVretro-flex-tdTomato and AAVretro-flex-GFP respectively, and tdTomato and GFP immunofluorescence was visualized in coronal brain cryosections containing injection sites (top) or area postrema (bottom), arrows: co-labeled cells, scale bar: 100  $\mu$ m. The numbers of colabeled (yellow) or individually labeled (red, green) cells were counted in a representative section. (B) AAV-Flex-tdTomato was injected into the area postrema of Calcr-ires-Cre; Chat-gfp mice. GFP was visualized by native fluorescence and tdTomato-positive fibers were visualized by immunohistochemistry. (C) AAV-Flex-tdTomato was injected into the area postrema of Gfralp2a-Cre; Calca-gfp (top) and Slc6a2-p2a-Cre; Calca-gfp (bottom) mice. GFP was visualized by native fluorescence (CGRP neurons) and tdTomato-positive fibers were visualized by immunohistochemistry. (D) AAV-Flex-tdTomato was injected into the area postrema of Glp1rires-Cre; Calca-gfp and Calcr-ires-Cre; Calca-gfp mice. GFP was visualized by native fluorescence (CGRP neurons) and tdTomato-positive fibers were visualized by immunohistochemistry in coronal sections of Bregma indicated, AP: area postrema; DMV: dorsal motor nucleus of the vagus; Amb: nucleus ambiguus; RVLM: rostral ventrolateral medulla; PBN: parabrachial nucleus; dl: dorsolateral; el: external lateral; scp: superior cerebellar peduncle; scale bars: 100  $\mu$ m.



# Figure S7. Area postrema GLP1R neurons are functionally connected to PBN neurons, related to Figure 6.

(A) *Glp1r-ires-Cre* mice previously injected in the area postrema with *AAV-flex-ChR2*, which encodes a ChR2-mCherry fusion protein, were treated with (LED stimulation) or without (No stimulation) area postrema optogenetic stimulation (5 Hz, 1 hour, awake mice). Two-color expression analysis was then performed in coronal cryosections of PBN regions involving Fos immunochemistry (green) and native mCherry fluorescence (red), scale bar: 100  $\mu$ m, scp: superior cerebellar peduncle. (B) Fos immunohistochemistry (red) in coronal PBN cryosections 2.5 hours after visceral injection of lithium chloride (LiCl), lipopolysaccharide (LPS) or saline in *Glp1r-ires-Cre*; *Rosa-LSL-DTR* mice previously injected in the area postrema with DT (+DT) or saline (-DT). Left: representative images, scale bar: 100  $\mu$ m, Right: quantification of Fos-positive cells per section, n=6-14, mean ± sem, \*\*\*\*p<.0001.

### Supplementary Table 1

Cluster	Gene numbers	Signature genes
1	1	Olfr78
	2	D130009I18Rik
	3	Gal
	4	Calcr
	5	Ebf1
2	6	Casr
	7	Slc6a2
	8	Rprm
	9	Cyb561
	10	Acsl1
3	11	Agtr1a
	12	Pter
	13	Prlhr
	14	Тох
	15	Tafa1
4	16	Gpr88
	17	Nts
	18	Gfral
	19	Man1a
5	20	Gpr83
	21	Slc5a7
	22	Ptprk
	23	Spon1
	24	Trhde
6	25	Gipr
	26	Chn2
	27	Abcc9
	28	Ngef
	29	Ndnf
7	30	Insrr
	31	F5
	32	Phf24
	33	Etv5
	34	Sst

### Table S1. Signature genes in area postrema neuron subtypes, Related to Figure 1.

A list of the 4-5 most enriched genes for each area postrema neuron cluster, with gene numbers corresponding to rows in Figure 1B.