

Supplementary Information for MobiDetails: Online DNA variants interpretation

David Baux^{1,*}, Charles Van Goethem¹, Olivier Ardouin², Thomas Guignard³, Anne Bergougnoux¹, Michel Koenig¹, Anne-Françoise Roux¹

¹Laboratoire de Génétique Moléculaire, CHU de Montpellier, Université de Montpellier, France

²Plateau de Médecine Moléculaire et Génomique, CHU de Montpellier, Université de Montpellier, France

³Unité de Génétique Chromosomique, CHU de Montpellier, Université de Montpellier, France

*Correspondence : david.baux@inserm.fr

Table of contents :

<i>Supplementary Method 1:</i>	<hr/> 2
<i>Supplementary Table 1: External resources included in MobiDetails</i>	<hr/> 3
<i>Supplementary Figure 1: Global architecture of MobiDetails</i>	<hr/> 5
<i>Supplementary Figure 2: Full view of the variant web page</i>	<hr/> 6
<i>References</i>	<hr/> 9

Supplementary Method 1:

MobiDetails is a python Flask (<https://palletsprojects.com/p/flask/>) application built on top of a PostgreSQL database (Supplementary Figure 1). The database handles gene coordinates, variants and user-specific data. Many external annotations (such as ClinVar¹) are stored as compressed flat files (often in VCF format) and queried via tabix (<https://www.htslib.org/>) by the Flask engine. The engine either renders HTML/CSS/JS web pages using jinja2 templates (<https://jinja.palletsprojects.com/en/2.11.x/>) or returns a JSON object, depending on the caller (web navigation or API call). The API can be accessed via a swagger application for testing purpose (<https://mobidetails.iurc.montp.inserm.fr/MDAPI/>).

Supplementary Table 1: External resources included in MobiDetails

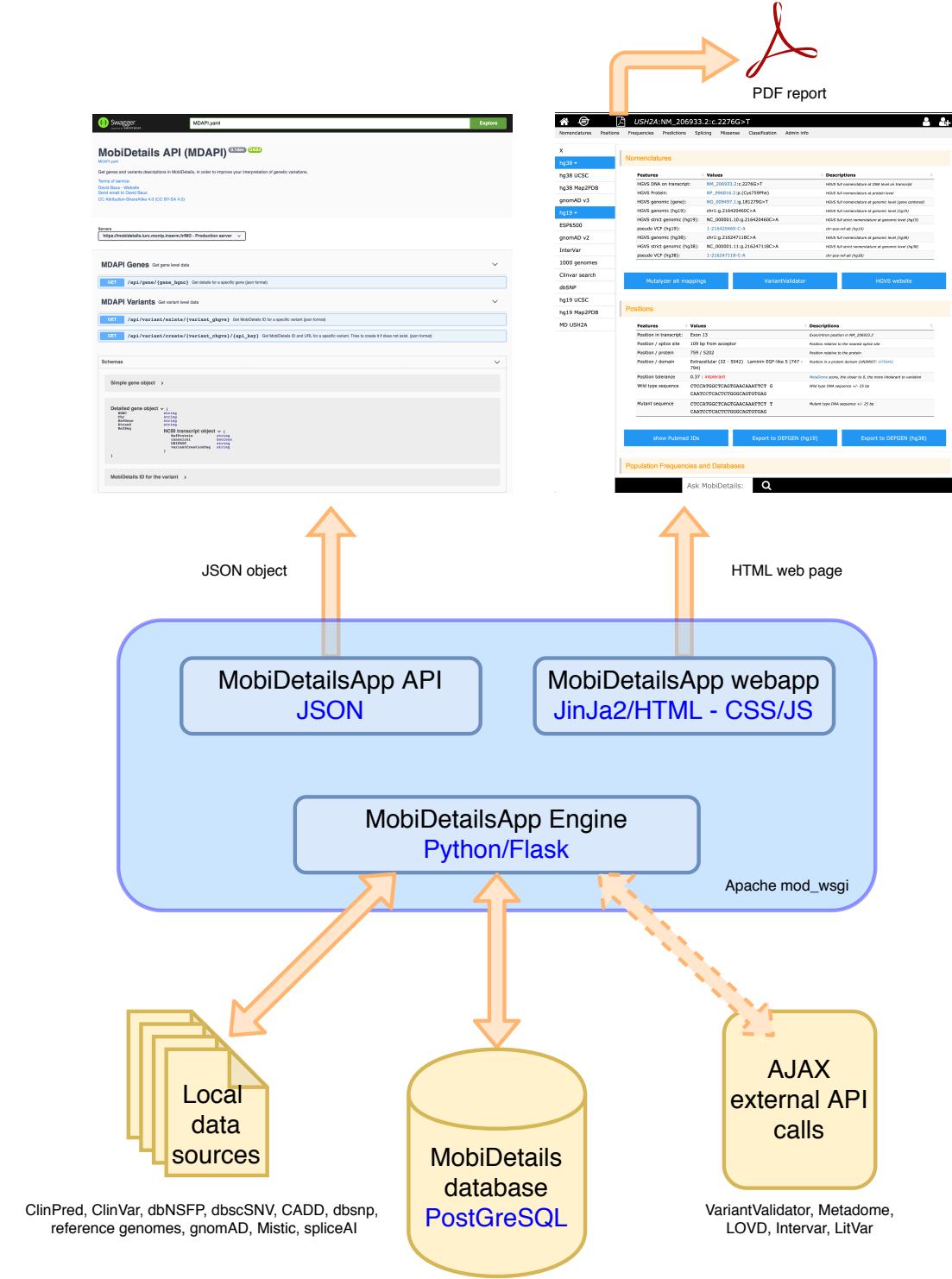
Resource	Version	Access	Reference
CADD	1.5	Local VCF	2
ClinPred	2018_hg19	Local file	3
ClinVar	20200830	Local VCF	1
dbMTS	1.0	Local file	4
dbNSFP	4.0a	Local file	5,6
dbscSNV	1.1	Local file	7
dbSNP	154	Local VCF	8
Eigen	1.1	From dbNSFP or dbMTS	9
FatHMM	2.3	From dbNSFP	10
gnomAD exome/genome	2.0.1	Local VCF	11
gnomAD genome	3	Local VCF	11
InterVar	NA	API	12
LitVar	1	API	13
LOVD	2-3	API	14
MaxEntScan	2004	Perl script	15
MetaDome	1.0.1	API – Local file	16
MetaSVM-LR	NA	From dbNSFP	17
Mistic	1	Local file	18
MPA	1	Internally computed	19
Mutalyzer	2.0.32	API	20
Polyphen-2	2.2.2	From dbNSFP	21
REVEL	NA	From dbNSFP	22
SIFT	Ensembl 66	From dbNSFP	23
spliceAI	1.3	Local VCF	24
TogoWS	NA	API	25
VariantValidator	1.0.4	API	26

NA: not available

Local resources are either automatically updated (ClinVar), i.e. a script periodically checks whether a more recent version of the resource has been released, or must be updated manually, when updates are quite rare (gnomAD, dbNSFP...). The Metadome API returns for each transcript a JSON file containing all the scores for each position of the encoded protein: MobiDetails calls it the first time a gene is visited and keeps a local copy of the JSON file for each gene. Then when a variant page is called, the data is retrieved from the local copy.

VariantValidator API is called when a variant is annotated for the first time. Mutalyzer API is called when a variant is annotated for the first time from its dbSNP identifier (rs id).

Supplementary Figure 1: Global architecture of MobiDetails



The main engine manages the PostGreSQL relational database which stores the genes and variants data. On query, it uses tabix (<https://www.htslib.org/>) to retrieve data from compressed files, and sends AJAX http requests to selected external APIs. Data sources are listed in Suppl. Table 1. SQL queries and data extracted from the files are treated to be rendered as JSON objects (REST API calls) or web pages.

Supplementary Figure 2: Full view of the variant web page

The screenshot shows a detailed view of a variant web page for the USH2A gene. The top navigation bar includes links for 'Navigation: home, about, all genes', 'PDF export', 'Variant' (highlighted with a yellow box), 'Profile', and 'Logout'. Below the navigation is a menu bar with links to 'Nomenclatures', 'Positions', 'Frequencies', 'Predictions', 'Splicing', 'Missense', 'Classification', and 'Admin info'. The main content area is divided into three main sections: 'Nomenclatures', 'Positions', and 'Predictions'.

Nomenclatures Section:

- Left sidebar:** A list of external resources including hg38 UCSC, hg38 Map2PDB, gnomAD v3, hg19 UCSC, hg19 Map2PDB, RegulomeDB, and MD USH2A. A star icon indicates favorite status.
- Content:** A table showing HGVS nomenclature details for the variant NM_206933.2:c.2276G>T across different levels (DNA, protein, genomic).
- Buttons:** 'Mutalyzer alt mappings', 'VariantValidator', and 'HGVS website'.

Positions Section:

- Left sidebar:** Direct links to external resources marked/unmarked as favorites.
- Content:** A table showing the position of the variant in the transcript, relative to splice sites, protein domains, and tolerance scores. It also displays the wild-type and mutant DNA sequences.
- Buttons:** 'show Pubmed IDs' (with a note about displaying links to Pubmed articles citing the variant), 'Export to DEFGEN (hg19)', and 'Export to DEFGEN (hg38)'.

Predictions Section:

- Content:** A table showing predictions from various tools like Splicing, Missense, Classification, and Admin info.
- Buttons:** 'Predictions', 'Splicing', 'Missense', 'Classification', and 'Admin info'.

Annotations and Shortcuts:

- A callout box labeled 'Shortcuts to different categories' points to the 'Profile' and 'Logout' links in the top right.
- A callout box labeled 'Display links to Pubmed articles citing this variant (obtained from Litvar API)' points to the 'show Pubmed IDs' button.
- A callout box labeled 'Direct links to external resources-mark/unmark favorites (star)' points to the star icon in the sidebar.

Population Frequencies and Databases

Features	Values	Descriptions
gnomAD exome:	0.0009	v2.0.1 Exomes global MAF
gnomAD genome:	0.0009	v2.0.1 Genomes global MAF
gnomAD v3:	1.34030e-03	v3 Genomes global MAF
dbSNP rsid:	rs80338902	Identifier for NCBI dbSNP
Clinvar:	Pathogenic	Clinvar interpretation
Intervar:	Pathogenic	Semi-automated ACMG classification - click on the intervar link to adjust
LOVD Matches:	<ul style="list-style-type: none"> Whole genome datasets Global Variome shared LOVD Retinal and hearing Impairment genetic mutation database MSeqDR-LSDB Mitochondrial Disease Locus Specific Database 	LOVD match in public instances
LOVD Effect Reported:	<ul style="list-style-type: none"> unknown: 1 functionProbablyAffected: 16 functionAffected: 178 functionNotAffected: 2 	Effects reported by LOVD submitters
LOVD Effect Concluded:	<ul style="list-style-type: none"> functionAffected: 175 notClassified: 20 functionNotAffected: 2 	Effects concluded by LOVD curators

Direct links to various LOVD instances containing the variant

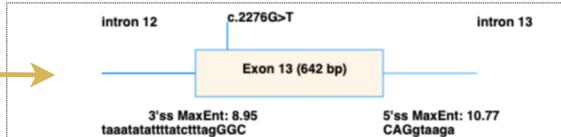
Overall predictions

Features	Values	Descriptions
CADD raw:	3.924570	[-6.41;35.5] The higher the less likely to be observed
CADD phred:	25.4	Phred-like scaling of raw score
Eigen raw:	1.04852691810073	[-3.33;6.84] The higher the less likely to be observed
Eigen phred:	15.52217	Phred-like scaling of raw score
MPA score:	10	Raw score [0;10], 10: high impact
MPA Impact:	Clinvar pathogenic	Impact type

Splicing predictions

The exonic splicing context of the variant including natural splice sites scores is summarized in the graph below:

Representation of the exon, the variant position within the exon and natural 3' and 5' splice sites sequences and MaxEntScan values



MaxEntScan scores are presented in the two following tables. Selected scores have:

- a |variation| > 15% and
- a raw score for mutant or wild-type of at least 3

The upper sequence shows the variation site in red, and the lower sequence the putative splice site considered by MaxEntScan (putative introns are shown as lower case and exons as upper case).

MaxEntScan 5'ss scores			
Wild-type sequence	Score	Mutant sequence	Score
No significant MaxEnt 5'ss scores found.			
MaxEntScan 3'ss scores			
Wild-type sequence	Score	Mutant sequence	Score
No MaxEnt 3'ss score performed (exonic variant far from 3'ss).			

This last table present dbscSNV and spliceAI scores, when available.

Features	Values	Descriptions
dbscSNV ADA:	No match in dbscSNV v1.1	Raw score (0-1), threshold > 0.8 for impact
dbscSNV RF:	No match in dbscSNV v1.1	Raw score (0-1), threshold > 0.8 for impact
spliceAI AG:	0.00 (-20)	Acceptor Gain, 0-1 (distance in bp), thresholds > 0.2 0.5 0.8 for impact
spliceAI AL:	0.00 (12)	Acceptor Loss, 0-1 (distance in bp), thresholds > 0.2 0.5 0.8 for impact
spliceAI DG:	0.01 (-20)	Donor Gain, 0-1 (distance in bp), thresholds > 0.2 0.5 0.8 for impact
spliceAI DL:	0.00 (46)	Donor Loss, 0-1 (distance in bp), thresholds > 0.2 0.5 0.8 for impact

Missense predictions

Click to display a radar chart of splicing or missense predictions

Features	Values	Prediction	Descriptions
SIFT:	0.0	Damaging	Threshold < 0.05 for Damaging
PolypHEN 2 HumDiv:	1.0	Probably Damaging	Thresholds > 0.454 0.957 for Possibly and Probably Damaging
PolypHEN 2 HumVar:	0.999	Probably Damaging	Thresholds > 0.447 0.909 for Possibly and Probably Damaging
REVEL:	0.902	Damaging	Thresholds 0.2 0.5 for Benign, Uncertain, Damaging
ClinPred:	0.88	Damaging	Threshold ≥ 0.5 for Damaging
Fathmm:	-3.4	Damaging	Threshold ≤ -1.5 for Damaging
Meta SVM:	1.0888 (9)	Damaging	Threshold ≥ 0 for Damaging (reliability index: 0-10), 10:high
Meta LR:	0.9471 (9)	Damaging	Threshold ≥ 0.5 for Damaging (reliability index: 0-10), 10:high
Mistic:	0.88	Damaging	Threshold ≥ 0.5 for Damaging

Show radar chart

Classification History

User	Date	ACMG Classification	Comments
No classification defined yet for this variant			

Add your classification

Administrative information

Features	Values	Descriptions
Creation user:	mobidetails	User who created the variant
Creation date:	2019-11-10	Date of creation in MD

Ask MobiDetails:



Full view of a MobiDetails variant page for *USH2A* NM_206933.2:c.2276G>T variant (<https://mobidetails.iirc.montp.inserm.fr/MD/variant/7>). Up to 8 categories can be displayed

on the center panel depending on the nature of the variant (i.e. not all insertions/deletions have splicing predictions, non-missense variants do not present missense predictions, 3'UTR variants present miRNA target sites predictions...). The left panel is dedicated to direct external links and to the button star used to mark/unmark a variant as favorite when logged in. The header is used to present links for navigation (home, about...), for pdf export and administrative tasks (login/logout, register, profile page). Finally, the footer is dedicated to the search engine.

References

1. Landrum MJ, Lee JM, Benson M, Brown G, Chao C, Chitipiralla S, et al. ClinVar: Public archive of interpretations of clinically relevant variants. *Nucleic Acids Research*. 2016;44(D1):D862-8.
2. Rentzsch P, Witten D, Cooper GM, Shendure J, Kircher M. CADD: predicting the deleteriousness of variants throughout the human genome. *Nucleic Acids Research*. 8 janv 2019;47(D1):D886-94.
3. Alirezaie N, Kernohan KD, Hartley T, Majewski J, Hocking TD. ClinPred: Prediction Tool to Identify Disease-Relevant Nonsynonymous Single-Nucleotide Variants. *The American Journal of Human Genetics*. oct 2018;103(4):474-83.
4. Li C, Mou C, Swartz MD, Yu B, Bai Y, Tu Y, et al. dbMTS: A comprehensive database of putative human microRNA target site SNVs and their functional predictions. *Human Mutation*. 2020;41(6):1123-30.
5. Liu X, Jian X, Boerwinkle E. dbNSFP: a lightweight database of human nonsynonymous SNPs and their functional predictions. *Human mutation*. août 2011;32(8):894-9.
6. Liu X, Wu C, Li C, Boerwinkle E. dbNSFP v3.0: A One-Stop Database of Functional Predictions and Annotations for Human Nonsynonymous and Splice-Site SNVs. *Human Mutation*. mars 2016;37(3):235-41.
7. Jian X, Boerwinkle E, Liu X. In silico prediction of splice-altering single nucleotide variants in the human genome. *Nucleic Acids Research*. 16 déc 2014;42(22):13534-44.
8. Sherry ST. dbSNP: the NCBI database of genetic variation. *Nucleic Acids Research*. 2001;29(1):308-11.
9. Ionita-Laza I, McCallum K, Xu B, Buxbaum JD. A spectral approach integrating functional genomic annotations for coding and noncoding variants. *Nat Genet*. févr 2016;48(2):214-20.
10. Shihab HA, Gough J, Cooper DN, Stenson PD, Barker GLA, Edwards KJ, et al. Predicting the Functional, Molecular, and Phenotypic Consequences of Amino Acid Substitutions using Hidden Markov Models. *Human Mutation*. janv 2013;34(1):57-65.
11. Genome Aggregation Database Consortium, Karczewski KJ, Francioli LC, Tiao G, Cummings BB, Alföldi J, et al. The mutational constraint spectrum quantified from variation in 141,456 humans. *Nature*. mai 2020;581(7809):434-43.

12. Li Q, Wang K. InterVar: Clinical Interpretation of Genetic Variants by the 2015 ACMG-AMP Guidelines. *American journal of human genetics*. 2 févr 2017;100(2):267-80.
13. Allot A, Peng Y, Wei C-H, Lee K, Phan L, Lu Z. LitVar: a semantic search engine for linking genomic variant data in PubMed and PMC. *Nucleic Acids Research*. 2 juill 2018;46(W1):W530-6.
14. Fokkema IF a. C, Taschner PEM, Schaafsma GCP, Celli J, Laros JFJ, den Dunnen JT. LOVD v.2.0: the next generation in gene variant databases. *Human Mutation*. 22 mai 2011;32(5):557-63.
15. Yeo G, Burge CB. Maximum Entropy Modeling of Short Sequence Motifs with Applications to RNA Splicing Signals. *Journal of Computational Biology*. 2004;11(2-3):377-94.
16. Wiel L, Baakman C, Gilissen D, Veltman JA, Vriend G, Gilissen C. MetaDome: Pathogenicity analysis of genetic variants through aggregation of homologous human protein domains. *Human Mutation*. 2019;40(8):1030-8.
17. Dong C, Wei P, Jian X, Gibbs R, Boerwinkle E, Wang K, et al. Comparison and integration of deleteriousness prediction methods for nonsynonymous SNVs in whole exome sequencing studies. *Human Molecular Genetics*. 2015;24(8):2125-37.
18. Chennen K, Weber T, Lornage X, Kress A, Böhm J, Thompson J, et al. MISTIC: A prediction tool to reveal disease-relevant deleterious missense variants. Andrade-Navarro MA, éditeur. PLoS ONE. 31 juill 2020;15(7):e0236962.
19. Yauy K, Baux D, Pegeot H, Van Goethem C, Mathieu C, Guignard T, et al. MoBiDiC Prioritization Algorithm, a Free, Accessible, and Efficient Pipeline for Single-Nucleotide Variant Annotation and Prioritization for Next-Generation Sequencing Routine Molecular Diagnosis. *The Journal of Molecular Diagnostics*. juill 2018;20(4):465-73.
20. Wilderman M, van Ophuizen E, den Dunnen JT, Taschner PEM. Improving sequence variant descriptions in mutation databases and literature using the Mutalyzer sequence variation nomenclature checker. *Human mutation*. 2008;29(1):6-13.
21. Adzhubei IA, Schmidt S, Peshkin L, Ramensky VE, Gerasimova A, Bork P, et al. A method and server for predicting damaging missense mutations. *Nature methods*. avr 2010;7(4):248-9.
22. Ioannidis NM, Rothstein JH, Pejaver V, Middha S, McDonnell SK, Baheti S, et al. REVEL: An Ensemble Method for Predicting the Pathogenicity of Rare Missense Variants. *The American Journal of Human Genetics*. oct 2016;99(4):877-85.
23. Kumar P, Henikoff S, Ng PC. Predicting the effects of coding non-synonymous variants on protein function using the SIFT algorithm. *Nature protocols*. janv 2009;4(7):1073-81.
24. Jaganathan K, Kyriazopoulou Panagiotopoulou S, McRae JF, Darbandi SF, Knowles D, Li YI, et al. Predicting Splicing from Primary Sequence with Deep Learning. *Cell*. janv 2019;176(3):535-548.e24.
25. Katayama T, Nakao M, Takagi T. TogoWS: integrated SOAP and REST APIs for interoperable bioinformatics Web services. *Nucleic Acids Research*. 1 juill 2010;38(Web Server):W706-11.

26. Freeman PJ, Hart RK, Gretton LJ, Brookes AJ, Dalgleish R. VariantValidator: Accurate validation, mapping, and formatting of sequence variation descriptions. *Human Mutation*. janv 2018;39(1):61-8.