High resolution cryo EM analysis of HPV16 identifies minor structural protein L2 and describes capsid flexibility

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Supplementary Information



SFig 1. Icosahedral Reconstruction. (A) Representative micrograph 1,164 out of 8,936 collected. (B) Central cross-section of icosahedral structure with the symmetry labeled in the dotted region. (C) Full icosahedral reconstruction colored radially in Angstroms. (D) 2D classes that were used for icosahedral reconstruction. (E) FSC Curve for icosahedral refinement. (F) Representative alpha helix (amino acid residues: 385-394, 396-401) at 4.5Å. (G) Representative beta sheet (amino acid residues: 71-76, 335-325, 153-160, 254-248) at 4.5Å.



SFig 2. Representative density of all side chains in pentavalent and hexavalent capsomers. Representative side chains were chosen from L1 (brown) and compared between the pentavalent (purple density) and hexavalent (green density) capsomer environments.



SFig 3. L2 Density. The surface rendered hexavalent (top) and pentavalent (bottom) capsomer density maps were colored as in Fig. 1A within 2Å of the L1 protein chain with unfilled density (gray) corresponding to L2. The L2 density is stronger in the pentavalent capsomer as can be seen with the changing contour of the map. In the hexavalent capsomers the same internal density can be noted, but the density disappears along with noise. Two distinct areas of gray density can be seen that are correlated to each chain of L1 in the capsomer.



SFig4. Capsomer addresses assigned in ISECC. A) Each capsomer is assigned an address identifying the nearest symmetry vertices. Pentavalent capsomers only have a fivefold designation without any rotational parameter (left, purple). Hexavalent capsomers receive designations for the nearest fivefold (left), threefold (center), and twofold (right) axis, as well as rotational parameters. B) Examples of complete addresses implemented in ISECC are shown as one-part (pentavalent) and three-part (hexavalent) capsomer designations. Addresses are assigned during subparticle generation after normalization of the input vectors to a standard, shared asymmetric unit. This allows refinement parameters for any given subparticle to be correlated with other subparticles from the same parental particle.

Data Collec	ction and Processing	Icosahedral	Pentavalent Capsomer	Hexavalent Capsomer
Magnification		59,000	59,000	59,000
Voltage (kV)		300	300	300
Electron Expo	sure $(e - / \text{\AA}^2)$	60	60	60
Defocus Rang	e (um)	0.5-3.0	0.5-3.0	0.5-3.0
Pixel Size (Å)		1.1	1.1	1.1
Symmetry Imp	posed	I1	C5	C1
Micrographs C	Collected	10,143	-	-
Micrographs F	Rejected (Bad Ice)	1,207	-	-
Micrographs A	Accepted	8,936	-	-
Initial Particle	Number	202,705	-	-
Final Particle	Number	181,299	181,299	181,299
Subparticles per Particle		-	12	60
Final Subparticle Number		-	2,175,588	10,877,940
Map Resolution (Å)		4.46	3.15	3.08
	FSC Threshold	0.143	0.143	0.143
Refinement		Recombined Icosahedral Asymmetric Unit		
Model compos	sition			
_	Non-hydrogen atoms		22492	
	Protein Residues		2864	
B-Factors				
	Protein		-	
R.m.s. Deviations				
	Bond Length (Å)		0.006	
	Bond Angles (°)		1.025	
Validation				
	MolProbity Score		2.65	
	Clash Score		13.17	
	Rotamer Outliers (%)		5.10	
Ramachandran Plot				
	Favored (%)		92.18	
	Outliers (%)		0.46	

Supp. Table 1. Cryo-EM data collection, refinement and validation statistics

HPV Type	Overall Sequence Percent Identity	Sequence Percent Identity of Loop	Sequence
16	-	-	SDAQIFN <u>KPYWLQRAQG</u> HNNGI
31	82.97%	92.3%	SDAQIFN <u>KPYW<mark>M</mark>QRAQG</u> HNNGI
52	76.82%	100%	S <mark>ES</mark> QLFN <u>KPYWLQRAQG</u> HNNGI
58	76.23%	100%	S <mark>ES</mark> QLFN <u>KPYWLQRAQG</u> HNNGI
33	79.60%	100%	S <mark>ES</mark> QLFN <u>KPYWLQRAQG</u> HNNGI
11	68.83%	92.3%	S <mark>E</mark> AQ <mark>L</mark> FN <u>KPYWLQ<mark>K</mark>AQG</u> HNNGI
6	68.59%	92.3%	S <mark>E</mark> AQ <mark>L</mark> FN <u>KPYWLQ<mark>K</mark>AQGHNNGI</u>
45	65.51%	84.6%	SD <mark>S</mark> QLFN <u>KPYWLHKAQG</u> HNNGI
18	65.87%	84.6%	SD <mark>S</mark> QLFN <u>KPYWLHKAQG</u> HNNGV

Supplemental	Table 2. Sequence	Alignment of Ser	306 – Ile328 Loop	o Region

Metadata label	Example	Description
rlnImageOriginalName	000004@ {micrographname}.mrcs	Existing metadata label. Repurposed to carry the identifier for the particle image from which a subparticle was derived.
rlnCustomUID	subparticleUID_ 000000001	Sequential, unique value identifying each subparticle
rlnCustomVertexGroup	Pentavalent: 5f08	Pentavalent capsomers are given a numerical value for the 5f symmetry axis on which they lay. There are 12 unique options
	Hexavalent: 5f08c.3f02b.2f27a	for this metadata label.
		Hexavalent capsomers are designated by the nearest 5f, 3f, and 2f symmetry axis, as well as as a letter (a-e, a-c, a-b) designation the counter-clockwise rotation order about the given symmetry axis. There are 60 unique options for this metadata label.
rlnCustomRelativePose	0.809, +0.309i, -0.500j, +0.000k	Capsomer orientation relative to icosahedrally-refined capsid, in quaternion format
rlnCustomOriginXYZ	-125.7926, 15.1589,	Capsomer origin relative to icosahedrally-
AngstWrtParticleCenter	-102.4911	refined capsid, in Å

Supplemental Table 3: Custom ISECC metadata