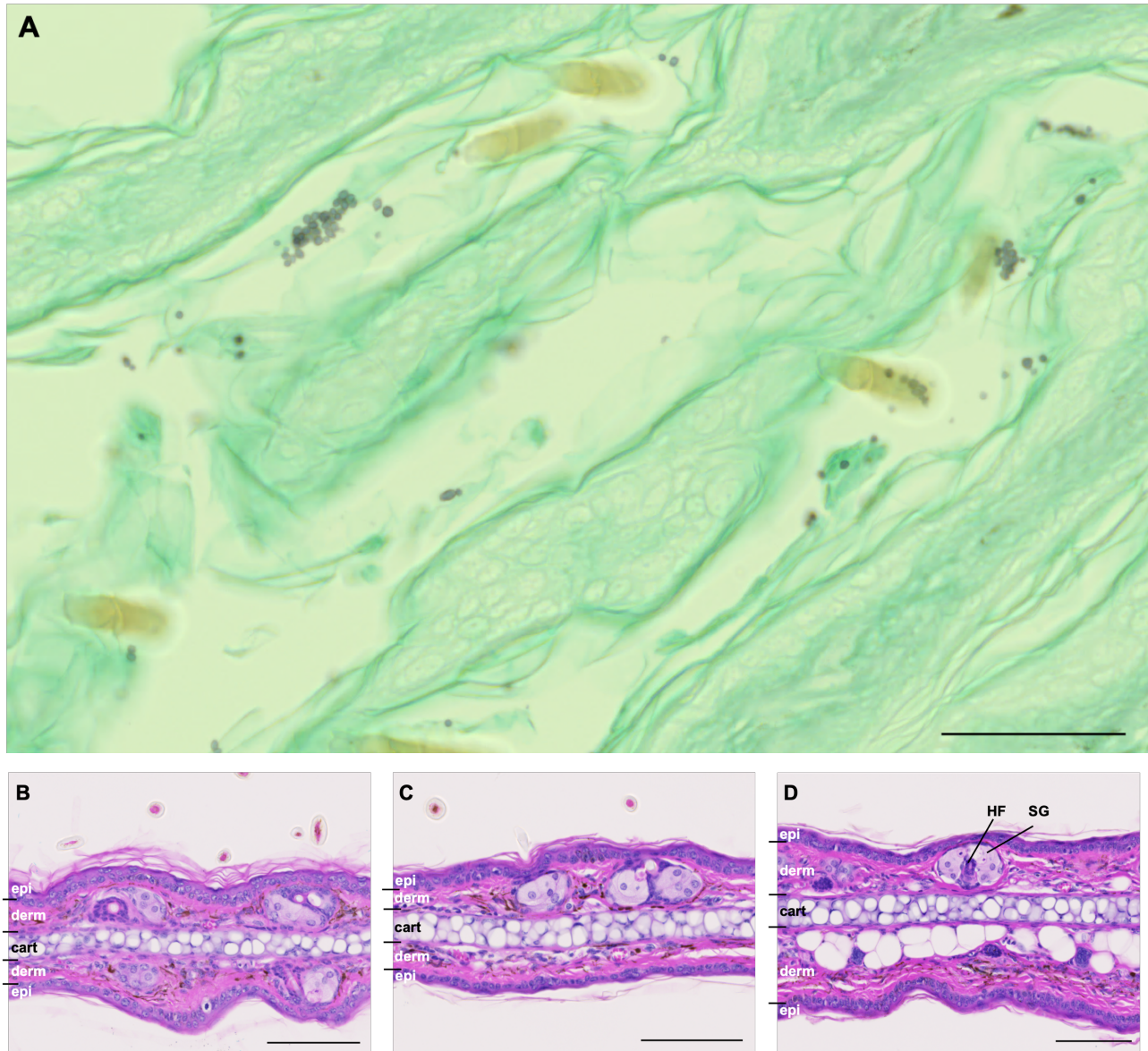
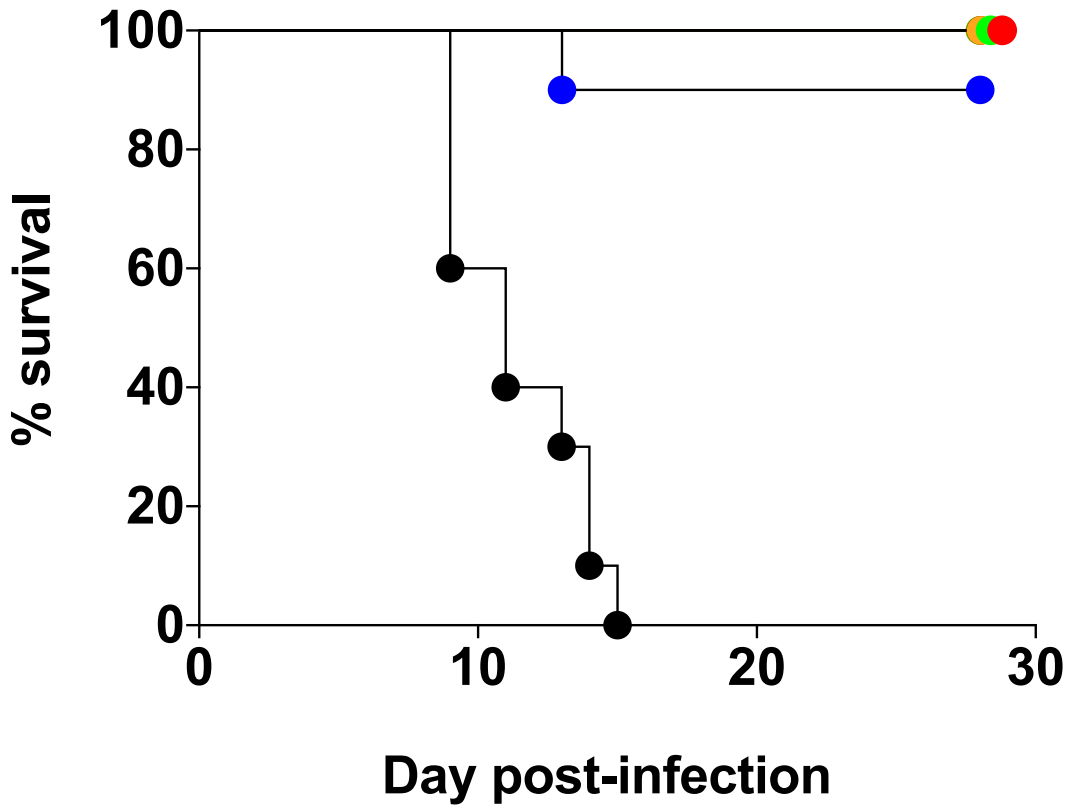


## SUPPLEMENTAL FIGURES AND TABLES



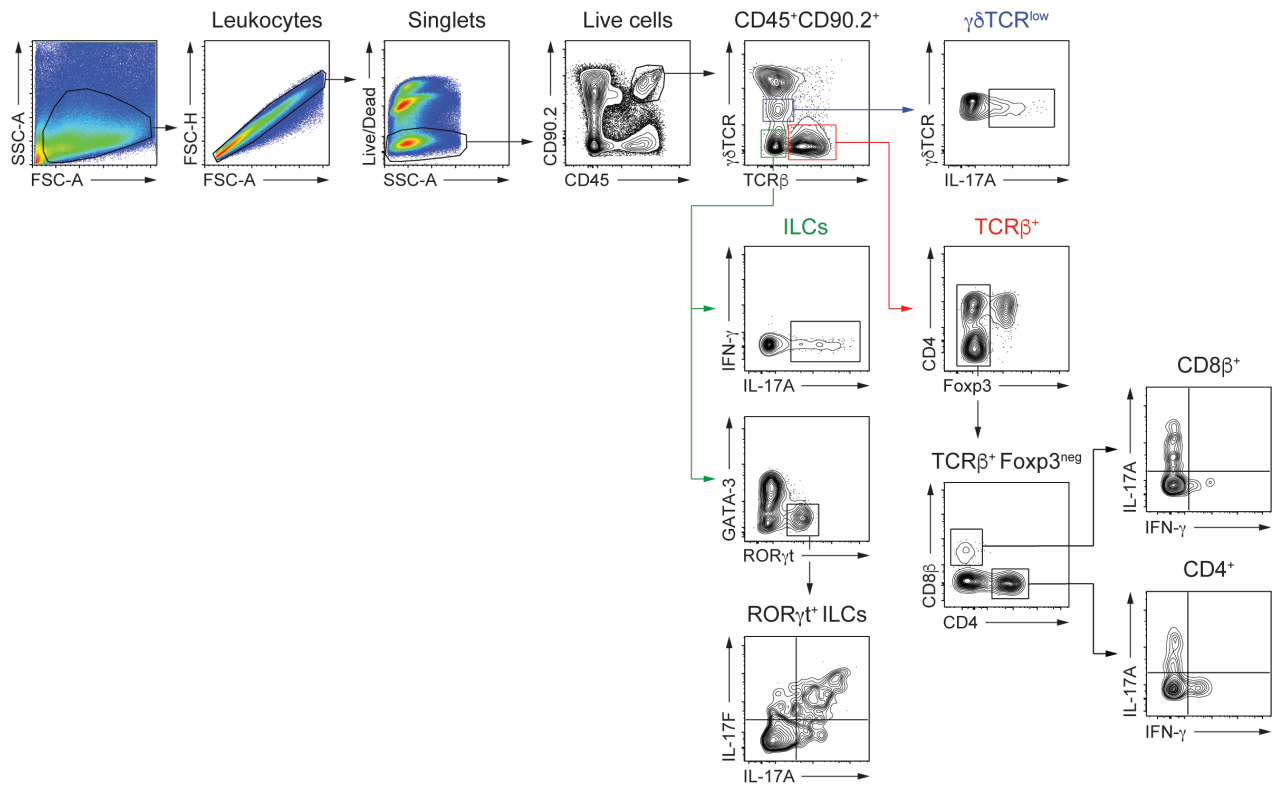
**Supplemental Figure 1. *C. auris* colonization on mouse skin surface and within skin tissue compartment does not elicit overt skin inflammation or skin architecture disruption. Related to Figure 1.** (A) Grocott's methenamine silver stain for fungi on ear pinna longitudinal section of a WT mouse topically associated seven days prior with *C. auris*. Clusters of *C. auris* cells (purple) can be seen on the skin surface (stratum corneum) and located around a hair shaft. 50  $\mu\text{m}$  scale bar is displayed at the bottom right. (B-D) Hematoxylin and eosin staining of pinna skin transverse sections from a naive WT mouse (B), mouse topically associated seven days prior with *C. auris*; and (C), mouse topically associated fourteen days prior with *C. auris* (D). epi: epidermis, derm: dermis, cart: cartilage, HF: hair follicle, SG: sebaceous gland. 100  $\mu\text{m}$  scale bar is displayed at the bottom right of each panel.

- *C. albicans*
- African
- NIH CC strain
- South American
- East Asian
- South Asian

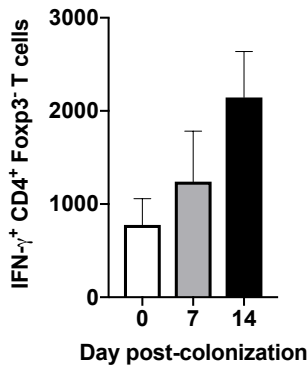
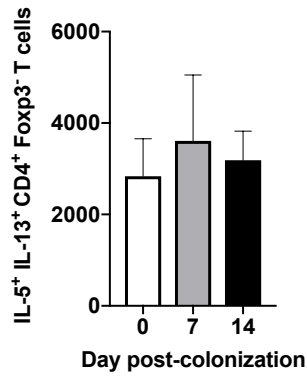
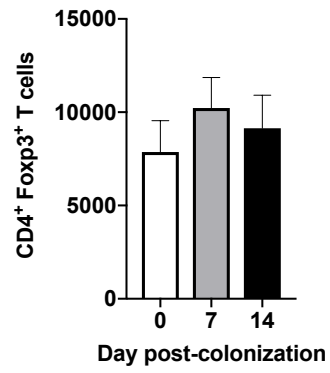
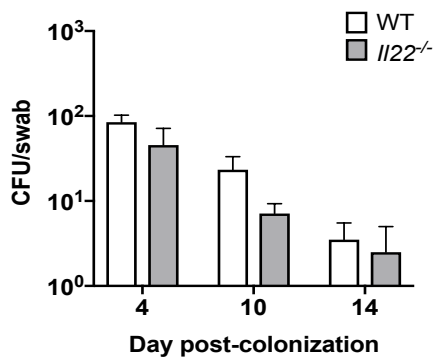
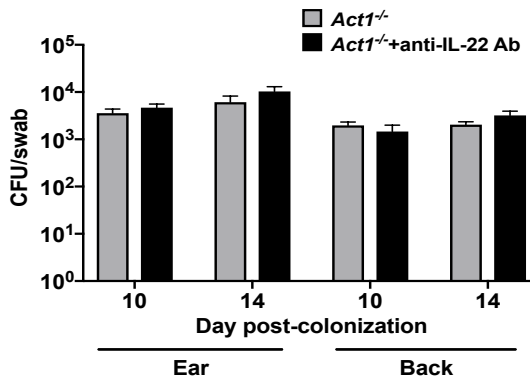
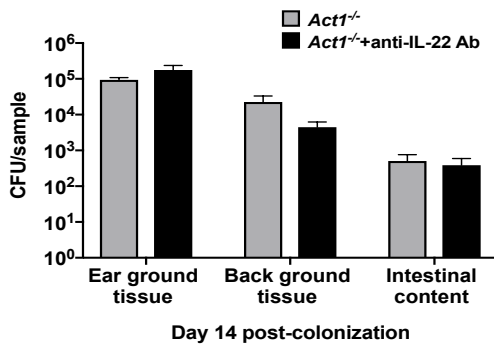
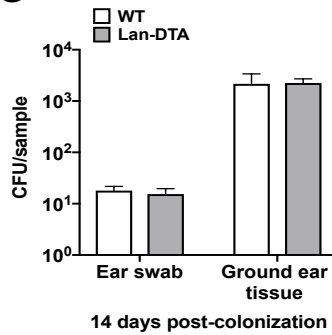
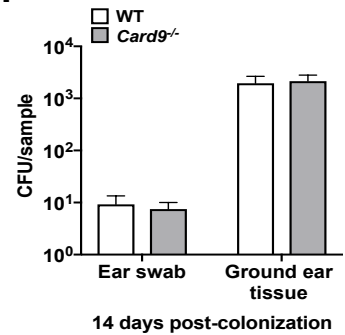


**Supplemental Figure 2. Intravenous injection of *C. albicans*, but not *C. auris*, is lethal to WT mice. Related to Figure 1.**

$1 \times 10^5$  *C. albicans* SC5314, four genetically distinct clades of *C. auris*, and the NIH CC strain (South Asian clade), were injected into the tail vein of WT mice, and survival followed for 30 days. N=10 for each group.



**Supplemental Figure 3. Gating strategy for flow cytometric analysis of IL-17A and IL-17F production by lymphoid cell subsets in the mouse skin. Related to Figure 2.** After initial FSC/SSC gating of immune cells, singlets and live cells were selected (three upper left panels). CD45<sup>+</sup> CD90.2<sup>+</sup> lymphoid cells were delineated as TCRβ<sup>+</sup> TCRγδ<sup>-</sup> αβ T cells, TCRβ<sup>-</sup> TCRγδ<sup>low</sup> γδ T cells, and TCRβ<sup>-</sup> TCRγδ<sup>-</sup> innate lymphoid cells (ILCs). Among TCRβ<sup>+</sup> cells, CD4 T cells and CD8 T cells were delineated as Foxp3<sup>-</sup> CD4<sup>+</sup> CD8β<sup>-</sup> and Foxp3<sup>-</sup> CD4<sup>-</sup> CD8β<sup>+</sup> cells, respectively. Among ILCs, RORγt was used to delineate IL-17A- and IL-17F-producing RORγt<sup>+</sup> ILCs. The total numbers of CD4 T cells, CD8 T cells, γδ T cells and ILCs were quantified and presented in Figure 2A. The IL-17A-producing CD4 T cells (Th17 cells), CD8 T cells (Tc17 cells), γδ T cells and ILCs were quantified and presented in Figure 2B. The IL-17F-producing CD4 T cells, CD8 T cells, γδ T cells and ILCs were quantified and presented in Figure 2C.

**A****B****C****D****E****F****G****H**

**Supplemental Figure 4. Immunological factors not associated with *C. auris* skin colonization. Related to Figures 2 and 3.**

(A-C) *C. auris* skin colonization does not elicit Th1, Th2 or regulatory T cell responses.

Numbers of interferon gamma-producing T helper cells (A), IL-5/IL-13-producing T helper cells (B), and regulatory T cells (C) are shown from naïve WT mice, WT mice colonized by *C. auris* 7 days after first topical association, and 14 days after first topical association.

(D-F) IL-22 is dispensable in regulating *C. auris* colonization.

(D) *C. auris* colonization on ear skin surface of WT and *Il22*<sup>-/-</sup> mice on days 4, 10 and 14 after topical association.

(E) *C. auris* colonization on ear and back skin surface of *Act1*<sup>-/-</sup> mice and *Act1*<sup>-/-</sup> mice injected with IL-22 neutralizing antibody on days 10 and 14 after topical association.

(F) *C. auris* residence in ear and back skin and intestinal compartments of *Act1*<sup>-/-</sup> mice and *Act1*<sup>-/-</sup> mice injected with IL-22 neutralizing antibody.

(G-I) Langerhans cell deficient, *Card9*<sup>-/-</sup> and *Rag2*<sup>-/-</sup> mice have normal *C. auris* skin colonization compared with WT mice.

(G and H) *C. auris* colonization on ear skin surface or disaggregated ear skin tissue of WT and Langerhans cell deficient (G) and *Card9*<sup>-/-</sup> (H) mice.

(I) *C. auris* colonization on back skin surface of WT and *Rag2*<sup>-/-</sup> mice on days 2, 8 and 15 after topical association.

(A-H) N=5 mice for each group. Numbers are shown with mean ± SEM.

**Supplemental Table 1. Within tissue residence by *Candida auris* in WT mice for up to 4 months. Related to Figure 1**

Group	CFU/Pi*
Day58_Pi	40,5,5,0
Day98_Pi	0,/,5,0
Day120_Pi	10,0,5,0
Day153_Pi	0,0,0,0

/: mortality

\* each value represents an individual mouse

WT: wild-type; Pi: pinnae

**Supplemental Table 2. Summary of treatments not contributing to longer-term *Candida auris* colonization or higher fungal load in the skin. Related to Figure 1**

Experimental Treatment	Colonization sites
AMNV <sup>a</sup> pre-exposure for 4 weeks	Back skin
FLU <sup>b</sup> pre-exposure for 4 weeks	Back skin
AMNV+FLU pre-exposure for 4 weeks	Back skin
Diabetic mice (db/db)	Ear and back skin
High-fat (western) diet-fed mice (high body mass index) for 2 weeks	Ear and back skin
TET <sup>c</sup> pre-exposure for 4 weeks	Ear and back skin
TMP-SMX <sup>d</sup> pre-exposure for 4 weeks	Ear and back skin

<sup>a</sup>AMNV: Ampicillin, metronidazole, neomycin and vancomycin

<sup>b</sup>FLU: Fluconazole

<sup>c</sup>TET: Tetracycline

<sup>d</sup>TMP-SMX: Trimethoprim-sulfamethoxazole

**Supplemental Table 3. Chlorhexidine retains *C. auris* genomic DNA on mouse skin.  
Related to Figure 4**

Time	Treatment	Average CFU	Average Ct
Day 3	Chlorhexidine	3540	16.9
	Control	7244	17.3
Day 7	Chlorhexidine	44	22.5
	Control	76	25.2
Day 11	Chlorhexidine	7	27.2
	Control	16	29.6
Day 15	Chlorhexidine	6	29.8
	Control	19	32.0

Two-tail t-test was used to compare differences between groups.

N=5 mice for each group. \*:  $p < 0.05$ ; \*\*:  $p < 0.01$ .