

---

**Supplementary information**

---

**Deep brain optogenetics without  
intracranial surgery**

---

In the format provided by the  
authors and unedited

### **Supplementary Note. Considerations for tissue heating from transcranial optogenetics.**

Tissue heating through high-power illumination can have nonspecific effects on neurophysiology and behavior<sup>1</sup>. For example, delivery of constant 532 nm light at 7 mW into the dorsal striatum has been shown to heat tissue by  $\sim 0.7$  °C, suppress striatal activity, and bias rotational behavior in mice<sup>1</sup>. To address this, we used established models that combine Monte Carlo simulations of photon propagation with Pennes's bio-heat equation to predict light-induced temperature changes in brain tissue based on our stimulation protocols<sup>1,2</sup> (**Supplementary Table 2**). We found that light parameters used to robustly drive behavior were associated with a predicted maximum temperature change from 0.07 °C (40 mW/mm<sup>2</sup>, 5 Hz, 100 ms pulse width) to 0.31 °C (800 mW/mm<sup>2</sup>, 20 Hz, 5 ms pulse width), which were below the temperatures associated with thermal modulation of behavior ( $\sim 0.7$  °C, 7 mW, 222 mW/mm<sup>2</sup>, 10 s)<sup>1</sup>. Based on the estimated temperature profile and histology assessment for neuroinflammation, we recommend using the laser parameters established in this study (635 nm, up to 800 mW/mm<sup>2</sup>, 20 Hz, 5 ms pulse width or up to 400 mW/mm<sup>2</sup>, 1 Hz, 100 ms pulse width) to deliver sufficient photon density for transcranial optogenetics with minimal tissue heating (**Extended Data. 3b and c**). By contrast, previously reported approaches for transcranial optogenetics with the mutated step-function opsin SOUL (473 nm, 400 mW/mm<sup>2</sup>) and near-infrared ChR2 excitation with injection of upconversion nanoparticles (980 nm, 9.5e4 mW/mm<sup>2</sup>) can cause temperature changes exceeding 5 °C<sup>3,4</sup> (**Supplementary Table 2, Extended Data. 3e**).

**Supplementary Table 1. Properties of channelrhodopsins for transcranial optogenetics.**

Opsin <sup>a</sup>	Action spectra <sup>b</sup> (nm)	EPD50 <sup>c</sup> ( $\mu\text{W}/\text{mm}^2$ )	Peak photocurrent (pA)	$\tau_{\text{off}}$ <sup>d</sup> (ms)
ChRmine <sup>5</sup>	390-650 [585]	~30	~4000	40
bReaChES <sup>5</sup>	390-650 [585]	~200	~2000	49
SOUL <sup>3</sup>	350-550 [473]; 525-625 [589]	~10	~400	~1.8*10 <sup>6</sup>

<sup>a</sup> Presented values for ChRmine, bReaChES, and SOUL were obtained from whole-cell patch clamp recordings of murine primary hippocampal neurons from previous publications<sup>3,5</sup>.

<sup>b</sup> Value in square bracket indicate stimulation wavelength for measurements.

<sup>c</sup> Effective power density for 50% activation (EPD50), a measure of opsin photosensitivity independent of expression level.

<sup>d</sup> The rate of channel closure upon light termination ( $\tau_{\text{off}}$ ). Fast off-kinetics are important for precise temporal control.

**Supplementary Table 2. Light parameters used for transcranial optogenetics.**

<i>In vivo</i> Assay <sup>a</sup>	$\lambda$ (nm)	Laser Power (mW)	Irradiance (mW/mm <sup>2</sup> )	$f$ (Hz)	Pulse width (ms)	Duty Cycle (%)	$\Delta T_{\max}^b$ (°C)	$d_{\text{source}}^c$ (mm)	Source
Recordings (mouse)	635	25-200; 0.5-50	200-1600; 4-400	5-40; 1	1-10; 100	1-20; 10	0.32; 0.37	4.5	1c-f, ED1, ED8
Recordings (rat)	635	0.5-50	4-400	1	100	10	0.37	7	ED2
Tissue compatibility	635	100-800	800-6400	20	5 ms	10	2.38	n/a	ED3
RTPP	635	10-100; 5	80-800; 40	20; 5	5; 100	5; 50	0.22; 0.07	4.2	1g, h, ED4, ED5
Lever-press	635	50	400	20	5	10	0.15	4.2	1i, j, ED5
SPP, NOP, OFT	635	100	800	20	5	10	0.31	2.8	2i, j, ED7
Closed-loop EEG	635	5	40	6.7	50	33.5	0.05	1.4	2c-g, ED6
Feeding inhibition (SOUL)	473; 589	50; 25	400; 200	60 s; 100 s		100	6.1	5	Ref. 2
Whisker protraction (ReaChR)	617	100	15	1	100	10	0.2	1	Ref. 3
VTA stimulation (UCNP /ChR2)	980	3000	9.5e4	20	15	30	9.4	4.2	Ref. 4

<sup>a</sup> Listed parameters for ChRmine experiments (unless otherwise noted). Presented values for alternative transcranial optogenetic approaches using ReaChR, SOUL, or ChR2/upconversion nanoparticle (UCNP) stimulation were obtained from previous publications<sup>3,4,6</sup>.

<sup>b</sup> Estimated maximum temperature change in tissue associated with the light parameters used. For irradiance-dependent recordings at 10 Hz and 5 ms pulse width presented in **Fig. 1c-f**, the estimated maximum temperature at 1600 mW/mm<sup>2</sup> was 0.32 °C. The light source dimensions to estimate the temperature changes were based on: a 400- $\mu$ m, 0.39 NA fiber (this study) a 3 mm diameter Luxeon 617 nm LED (ReaChR), a 400- $\mu$ m 0.22 NA fiber (SOUL), and a 200- $\mu$ m 0.37 NA fiber (ChR2 with UCNP).

<sup>c</sup> Proximity of light source to apex of targeted brain structure.

## References

- 1 Owen, S. F., Liu, M. H. & Kreitzer, A. C. Thermal constraints on in vivo optogenetic manipulations. *Nature neuroscience* **22**, 1061-1065 (2019). PMC6592769
- 2 Stujenske, Joseph M., Spellman, T. & Gordon, Joshua A. Modeling the Spatiotemporal Dynamics of Light and Heat Propagation for In Vivo Optogenetics. *Cell Reports* **12**, 525-534 (2015).
- 3 Gong, X. *et al.* An Ultra-Sensitive Step-Function Opsin for Minimally Invasive Optogenetic Stimulation in Mice and Macaques. *Neuron* (2020).
- 4 Chen, S. *et al.* Near-infrared deep brain stimulation via upconversion nanoparticle-mediated optogenetics. *Science* **359**, 679-684 (2018).
- 5 Marshel, J. H. *et al.* Cortical layer-specific critical dynamics triggering perception. *Science* **365**, eaaw5202 (2019).
- 6 Lin, J. Y., Knutsen, P. M., Muller, A., Kleinfeld, D. & Tsien, R. Y. ReaChR: a red-shifted variant of channelrhodopsin enables deep transcranial optogenetic excitation. *Nature neuroscience* **16**, 1499-1508 (2013). PMC3793847