

**Figure S1, related to Figure 1.** PTB granulomas from human and macaque display similar characteristic features.

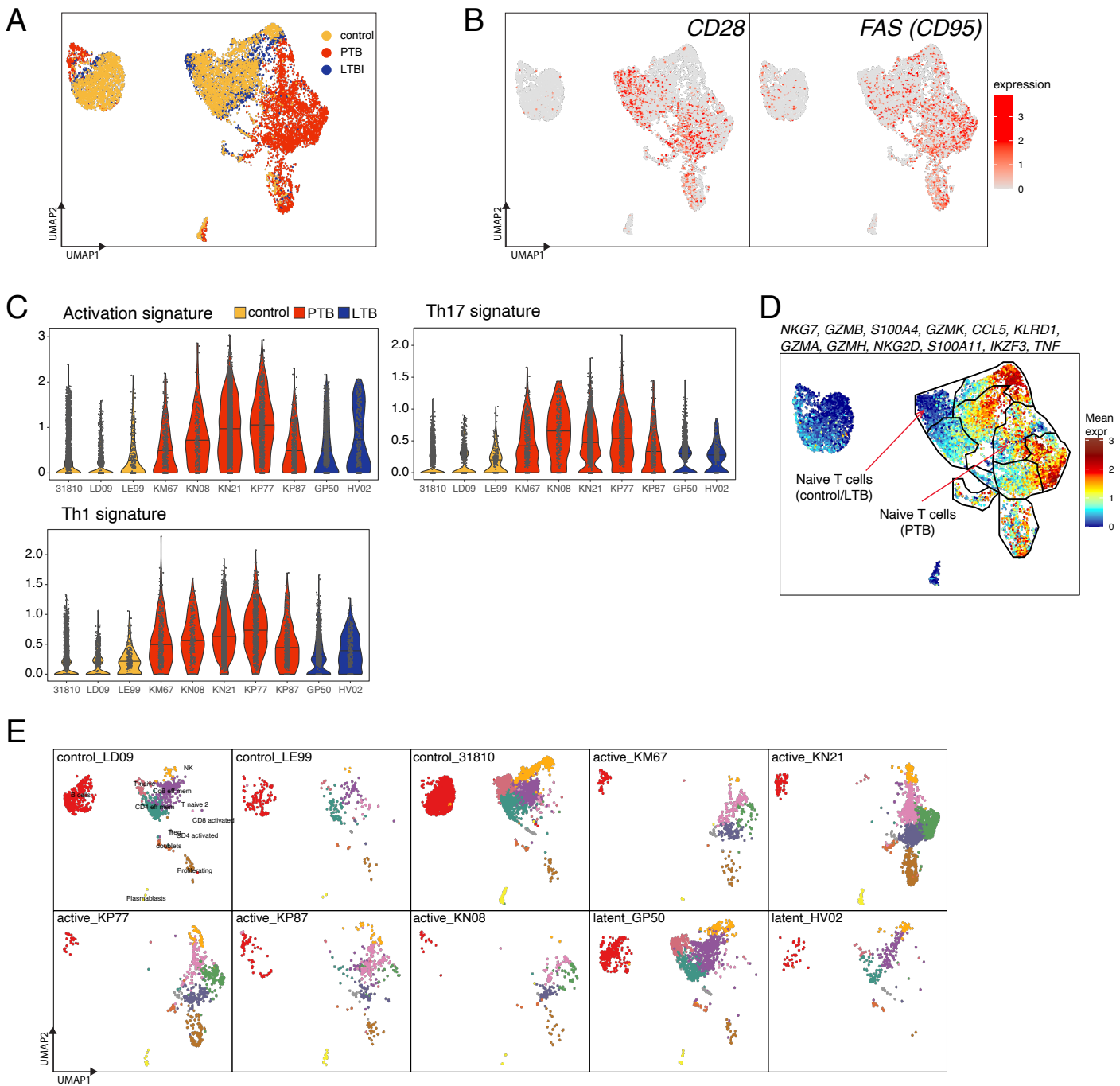
**A** and **B.** Hematoxylin and Eosin staining on formalin-fixed paraffin-embedded sections from lung tissue from macaques with PTB, (n=6) and LTBI (n=4). Right panel display morphometrical quantification of the granuloma size and percentage of inflammation (B) across macaques.

**C.** Immunohistochemistry staining of CD20 and DAPI on formalin-fixed paraffin-embedded serial sections from macaque lung (PTB: n=6 and LTBI; n=4) as used above. Right panel display morphometrical quantification average size of B cell follicle across macaques.

**D.** Immunohistochemistry staining of CD3, CD20, Ror $\gamma$ T, and DAPI on formalin-fixed paraffin-embedded sections from lung tissue from individuals with PTB, (n=6) and macaque lungs (PTB: n=6 and LTBI; n=4).

**E.** UMAP plots of cells from all scRNA-seq samples from macaques, split by condition and colored by the cluster. Control (n=3), LTBI (n=2) and PTB (n=5).

Data represented as mean  $\pm$  SD. \*p < 0.05, \*\*p < 0.01 by Student's T test (A, B, C)



**Figure S2, related to Figure 2.** Disease-dependent changes in lymphoid cells by scRNA-seq for healthy macaques and macaques with PTB and LTBI.

Single cell suspensions from control (n=3), LTBI (n=2) and PTB (n=5) macaques were collected and processed for scRNA-seq as described in Fig 1.

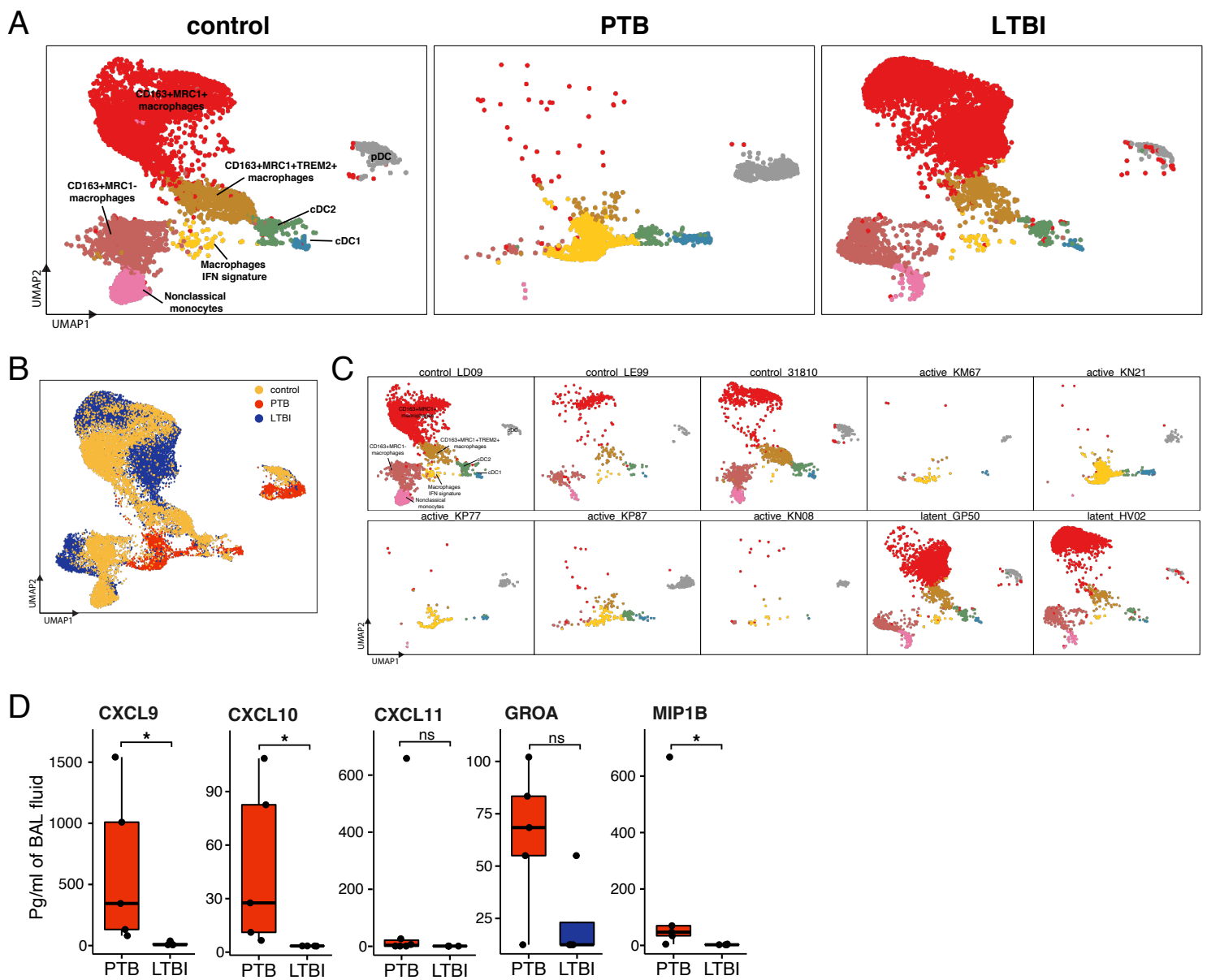
**A.** UMAP plot of lymphoid cells across all samples, colored according to condition.

**B.** UMAP plots of *CD28* and *CD95* expression in lymphoid cells.

**C.** Violin plot of mean gene expression from Th1, Th17, and activation signatures in lymphoid cells, split by subject.

**D.** UMAP plot with the mean expression of genes, associated with T cell activation.

**E.** UMAP plots of lymphoid cells, split by sample and colored according to identified clusters.



**Figure S3, related to Figure 3.** Disease-dependent changes in myeloid cells by scRNA-seq for healthy macaques and macaques with PTB and LTBI.

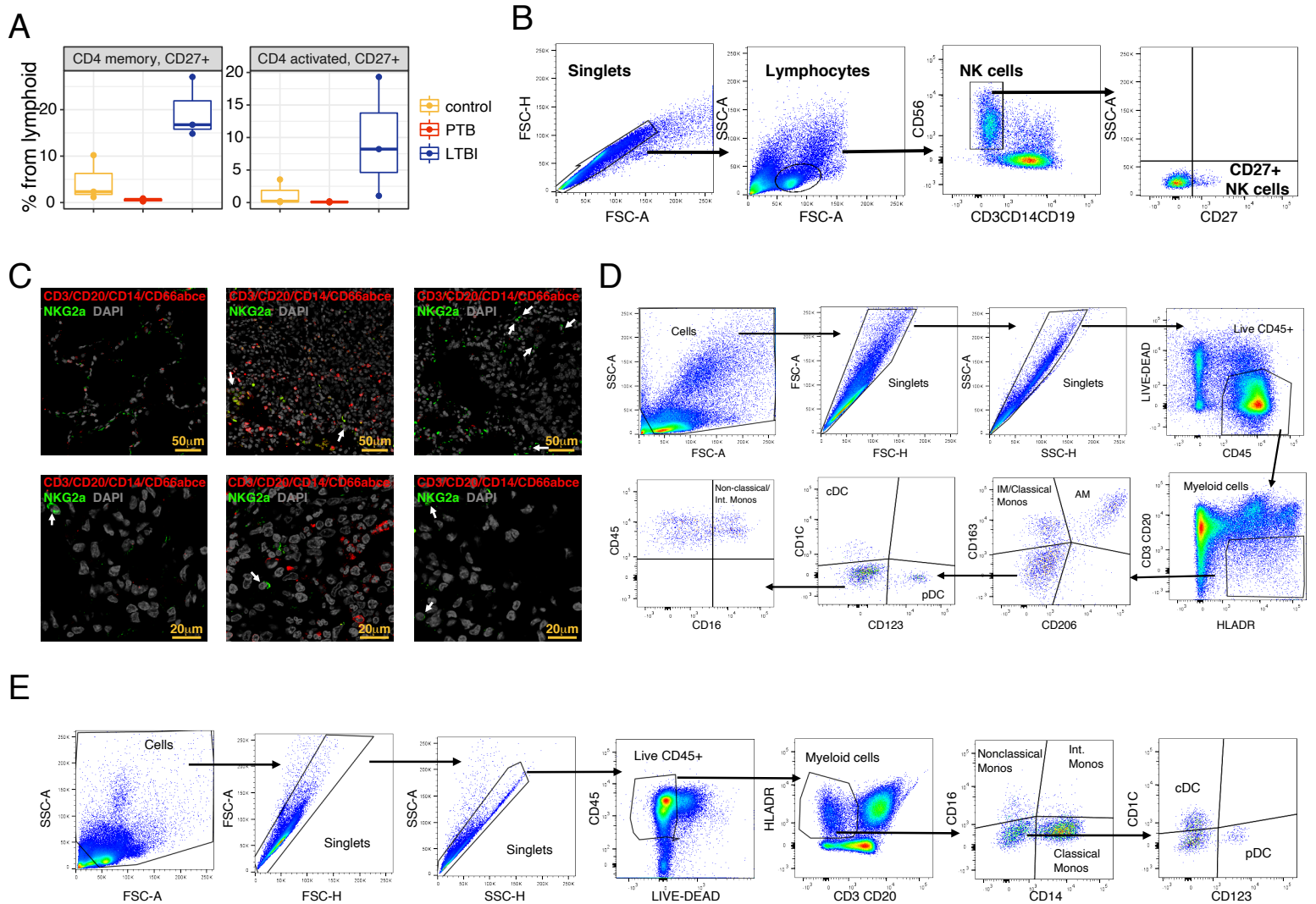
**A.** UMAP plots of myeloid cells split by condition and colored according to identified clusters.

**B.** UMAP plot of myeloid cells across all samples, colored according to condition.

**C.** UMAP plots of myeloid cells, split by sample and colored according to identified clusters.

**D.** Levels of CXCL9, CXCL10, CXCL11, CXCL-1 (GROA), and CCL-4 (MIP-1 ) were measured in the BAL fluid from macaques with PTB (n=5) and LTBI (n=4). Data represented as mean  $\pm$  SD, NS = not significant.

\*p < 0.05 by Student's t-test.



**Figure S4, related to Figure 4.** NK cells are increased in humans and macaques with LTBI. for healthy macaques and macaques with PTB and LTBI.

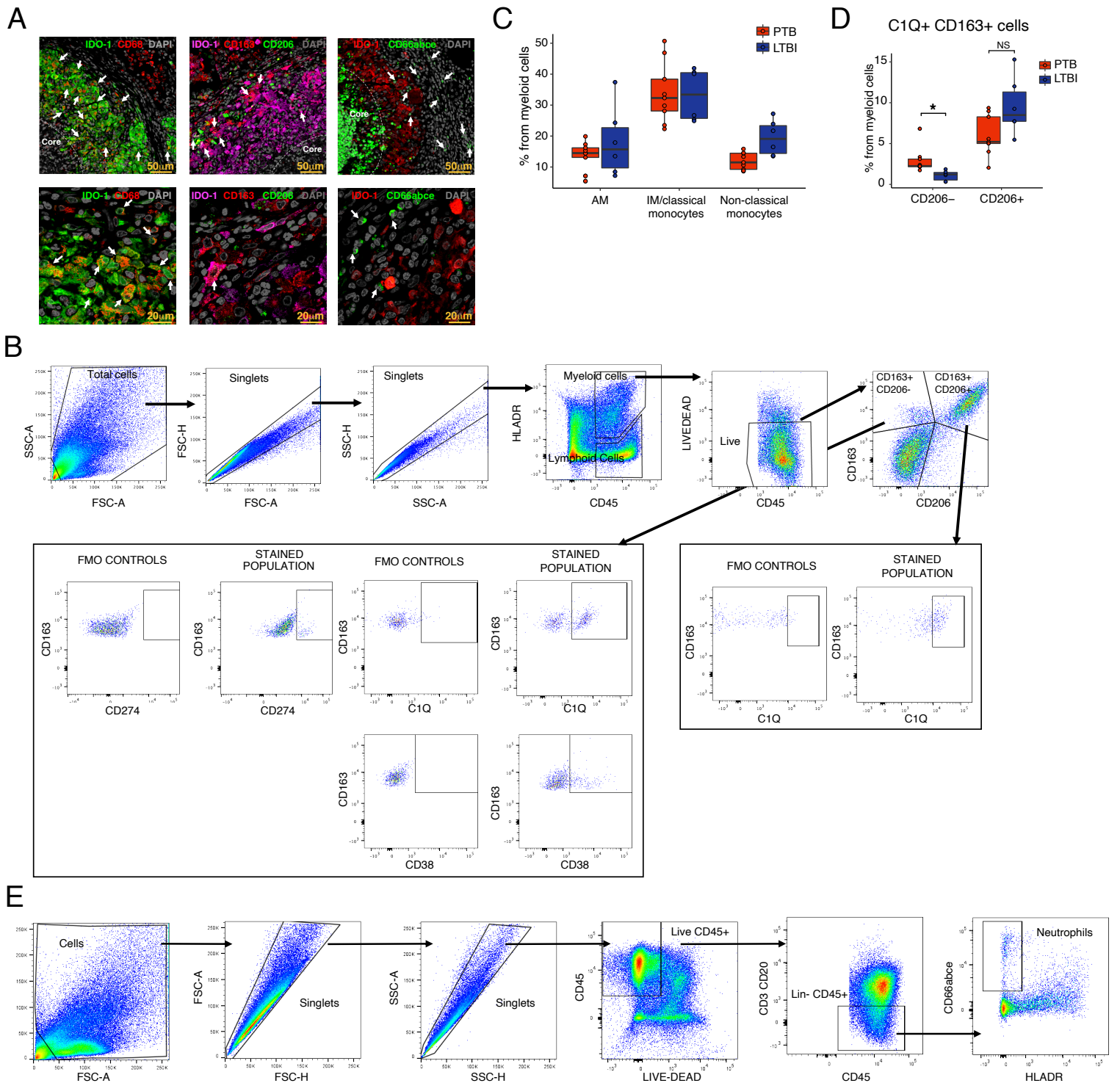
**A.** Proportion of CD4<sup>+</sup> CD27<sup>+</sup> cells in two CD4 clusters from CyTOF.

**B.** PBMCs from healthy controls (HC, n=5) or individuals with PTB (n=25) or LTBI (n=11) were stained with CD3, CD14, CD19, CD56, and CD27 antibodies, and live populations were gated based on the following strategy: NK cells: CD3<sup>+</sup>CD14<sup>+</sup>CD19<sup>+</sup>CD56<sup>+</sup> and CD27<sup>+</sup>.

**C.** Immunohistochemistry staining of CD3, CD20, CD14, Cd66abce, NKG2a, and DAPI on formalin-fixed paraffin-embedded macaque lung (control: n=3, PTB: n=3 and LTBI; n=3). The arrowheads indicate the expected populations.

**D.** Lung single cell suspensions from macaques with PTB (n=9), LTBI (n=6) were stained with CD45, CD3, CD20, HLA-DR, CD206, CD163, CD16, CD1c and CD123 and the live populations were gated based on the following strategy: AMs: CD45<sup>+</sup> CD3/CD20<sup>-</sup> HLA-DR<sup>+</sup> CD163<sup>+</sup> CD206<sup>+</sup>; IMs/Classical monocytes: CD45<sup>+</sup> CD3/CD20<sup>-</sup> HLA-DR<sup>+</sup> CD163<sup>+</sup> CD206<sup>-</sup>; Non-classical monocytes: CD45<sup>+</sup> CD3/CD20<sup>-</sup> HLA-DR<sup>+</sup> CD163<sup>-</sup> CD206<sup>-</sup> CD16<sup>+</sup> and pDC: CD45<sup>+</sup> CD3/CD20<sup>-</sup> HLA-DR<sup>+</sup> CD163<sup>-</sup> CD206<sup>-</sup> CD1c<sup>-</sup> CD123<sup>+</sup>.

**E.** PBMC populations from the blood of individuals and macaques with PTB, LTBI and HCs were stained with CD45, CD3, CD20, HLA-DR, CD16, CD14, CD1c and CD123 and the live populations were gated based on the following strategy: Classical monocytes: CD45<sup>+</sup> CD3/CD20<sup>-</sup> HLA-DR<sup>+</sup> CD14<sup>+</sup> CD16<sup>-</sup>; Non-classical monocytes: CD45<sup>+</sup> CD3/CD20<sup>-</sup> HLA-DR<sup>+</sup> CD16<sup>+</sup> CD14<sup>-</sup> or lo, intermediate monocyte: CD45<sup>+</sup> CD3/CD20<sup>-</sup> HLA-DR<sup>+</sup> CD16<sup>+</sup> CD14<sup>+</sup> and pDC: CD45<sup>+</sup> CD3/CD20<sup>-</sup> HLA-DR<sup>+</sup> CD16<sup>-</sup> CD14<sup>-</sup> CD1c<sup>-</sup> CD123<sup>+</sup>.



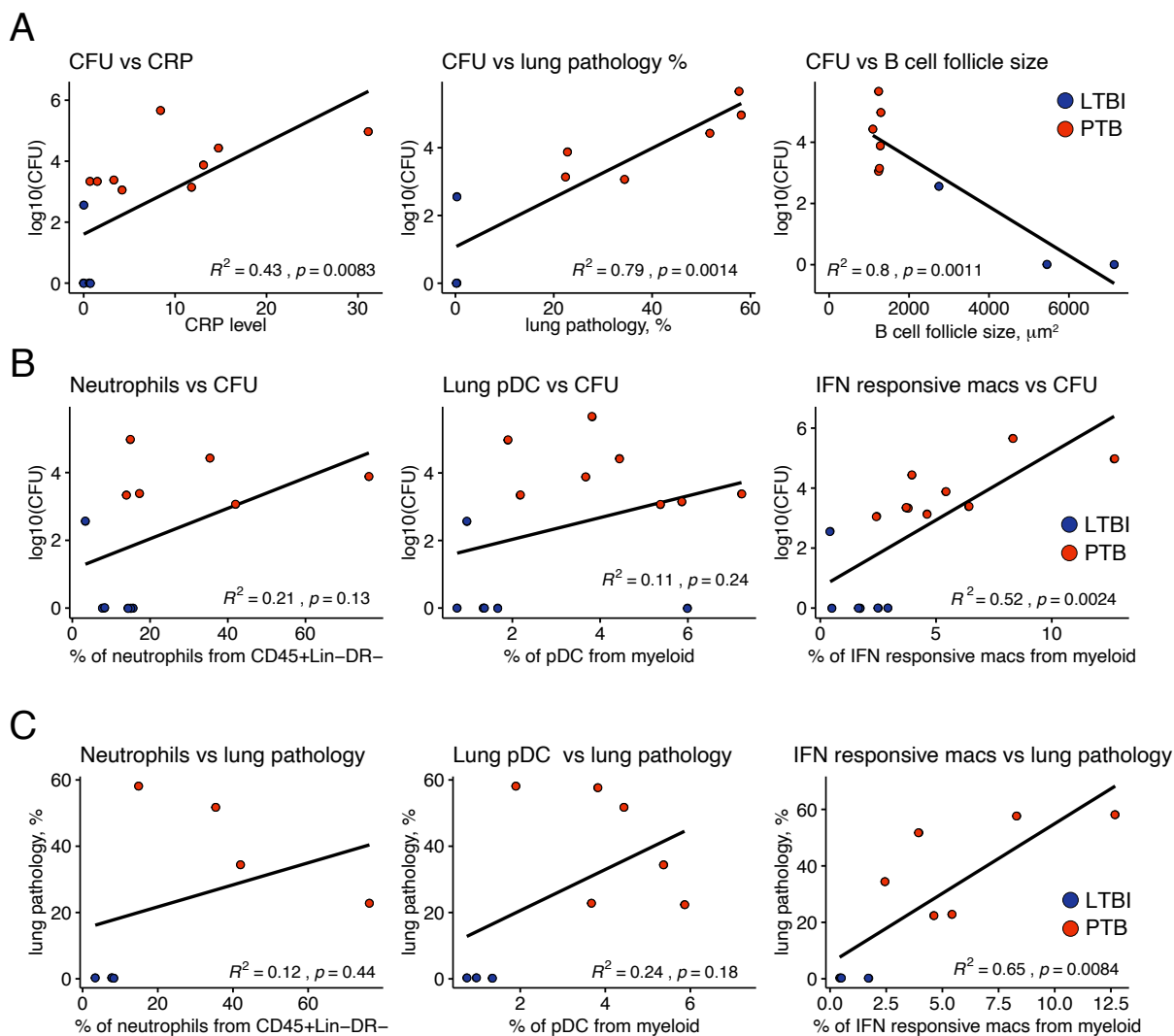
**Figure S5, related to Figure 5.** Lungs of macaques with PTB accumulate IFN-responsive macrophages within the rim of granulomas.

**A.** Immunohistochemistry staining of IDO-1, CD68; IDO-1, CD163, CD206; IDO-1, Cd66abce, and DAPI on formalin-fixed paraffin-embedded macaque lung (PTB: n=3). The arrowheads indicate the expected populations.

**B.** Lung single cell suspensions from macaques with PTB (n=9), LTBI (n=6) were stained with CD45, CD3, CD20, HLA-DR, CD206, CD163, CD38, CD274 and C1q and the live populations were gated based on the following strategy: CD45<sup>+</sup>CD3/CD20<sup>-</sup>HLA-DR<sup>+</sup>CD163<sup>+</sup>CD206<sup>-</sup>CD274<sup>+</sup>; CD45<sup>+</sup> CD3/CD20<sup>-</sup> HLA-DR<sup>+</sup> CD163<sup>+</sup> CD206<sup>-</sup> CD38<sup>+</sup>; CD45<sup>+</sup>CD3/CD20<sup>-</sup>HLA-DR<sup>+</sup>CD163<sup>+</sup>CD206<sup>-</sup>C1Q<sup>+</sup>; and CD45<sup>+</sup> CD3/CD20<sup>-</sup> HLA-DR<sup>+</sup> CD163<sup>+</sup> CD206<sup>+</sup> C1Q<sup>+</sup>. Fluorescence minus one (FMO) controls were used to gate the respective populations.

**C-D.** Percentage of AMs, IMs/classical monocytes, non-classical monocytes, CD163<sup>+</sup> CD206<sup>-</sup> C1Q<sup>+</sup>, CD163<sup>+</sup> CD206<sup>+</sup> C1Q<sup>+</sup> cells were determined by flow cytometry in lung single cell suspension from PTB (n=9) and LTBI (n=6) macaques. Data represented as mean  $\pm$  SD, \*p < 0.01, by Student's t-test with Holms correction.

**E.** Lung single cell suspensions from macaques with PTB (n=6), LTBI (n=6) were stained with CD45, CD3, CD20, HLA-DR, CD66abce and the live populations were gated based on the following strategy: neutrophils: CD45<sup>+</sup>CD3/CD20<sup>-</sup>HLA-DR<sup>-</sup> Cd66abce<sup>+</sup>.



**Figure S6, related to Figure 6.** Accumulation of lung IFN-responsive macrophage population correlates with increased TB disease and *Mtb* burden.

**A.** Linear correlation between log<sub>10</sub> of CFU identified at necropsy, and levels of CRP at necropsy, percent of lung pathology, and average area of B cell follicles measured in the lungs of macaques with PTB (red) and LTBI (blue).

**B.** Linear correlation between log<sub>10</sub> of CFU at necropsy and percentage of neutrophils, pDCs, and IFN-responsive macrophage populations (as a percent of myeloid cells) measured in lungs of macaques with PTB (red) and LTBI (blue).

**C.** Linear correlation between the percentage of lung pathology and percentage of lung neutrophils, pDCs, and IFN-responsive macrophage populations (as a percent of myeloid cells) in the lungs of macaques with PTB (red) and LTBI (blue).

**(A-C)** Pearson's correlation coefficient was used.