

**Supporting Information for “Refining all-atom protein force fields for polar-rich, prion-like, low complexity intrinsically disordered proteins”**

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## Tables

Table S1: Protein sequences used in simulations

Protein	N	Sequence
FUS <sub>0-43</sub> *	44	GMASNDYTQQATQSYGAYPTQPGQGYSSQSSQPYGQQSYSGYSQ
TDP43 <sub>310-350</sub>	41	GMNFGAFSINPAMMAAQAALQSSWGMMGLASQQNQSGPS
RNA Pol II <sub>1927-1970</sub>	44	SPTYSPKSTSPKSTYSPTSPGYSPSTPQYSLTSPAPISPDSSDEEN
FUS <sub>41-84</sub>	44	YSQSTDTSQYGQSSYSSYQSQNTGYGTQSTPQGYGSTGGYGSS
FUS <sub>77-120</sub>	44	STGGYGSSQSSQSSYQSSYQSSYQGYQSPAPSSTSGSYGSSSQSS
FUS <sub>120-163</sub>	44	SSYGQPQSGYSQSPSYGGQQSYGQQQSYNPPQGYGQQNQYNS
FUS <sub>37-97</sub>	61	SYSGYSQSTDTSQYGQSSYSSYQSQNTGYGTQSTPQGYGSTGGYGSSQSSQSSYQSSYQSSY
hnRNPA2 <sub>190-233</sub>	44	GRGGNFGFGDSRGGGNGFPGPGSNFRGGSDGYGSGRFGDGYN
hnRNPA2 <sub>265-308</sub>	44	GNQGGGYYGGYDNYGGGNYGSGNYNDFGNYNQQPSNYGPMKSGN

\*a starting glycine is included in FUS<sub>0-43</sub> as residue 0

Table S2: Summary of simulations performed using ff99SBws, where N is the number of residues in given protein or peptide.

Protein	N	Enhanced Sampling	Total simulation time (ns/replica)	Simulation time analyzed (ns/replica)
TDP43 <sub>310-350</sub>	41	PT-WTE	200	150
FUS <sub>0-43</sub>	44	PT-WTE	200	150
FUS <sub>41-84</sub>	44	PT-WTE	200	150
FUS <sub>77-120</sub>	44	PT-WTE	200	150
FUS <sub>120-163</sub>	44	PT-WTE	200	150
FUS <sub>37-97</sub>	61	PT-WTE	200	150
hnRNPA2 <sub>190-234</sub>	44	PT-WTE	200	150
hnRNPA2 <sub>256-308</sub>	44	PT-WTE	200	150
RNA Pol II <sub>1927-1970</sub>	44	PT-WTE	200	150

Table S3: Summary of simulations performed using ff03ws, where N is the number of residues in given protein or peptide.

Protein	N	Enhanced Sampling	Total simulation time* (ns/replica)	Simulation time analyzed* (ns/replica)
TDP43 <sub>310-350</sub>	41	PT-WTE	200	150
FUS <sub>0-43</sub>	44	PT-WTE	200	150
FUS <sub>41-84</sub>	44	PT-WTE	200	150
FUS <sub>77-120</sub>	44	PT-WTE	200	150
FUS <sub>120-163</sub>	44	PT-WTE	200	150
hnRNPA2 <sub>190-234</sub>	44	PT-WTE	200	150
hnRNPA2 <sub>256-308</sub>	44	PT-WTE	200	150
RNA Pol II <sub>1927-1970</sub>	44	PT-WTE	200	150

Table S4: Summary of simulations performed using modified force fields. All simulations were performed for a total of 200 ns per replica, and analysis conducted on the last 150 ns.

	$k_{\psi}$ (kJ/mol)	$k_{\psi,S}$ (kJ/mol)	$k_{\psi,T}$ (kJ/mol)	$k_{\psi,Q}$ (kJ/mol)	Simulated peptides
ff99SBws	2.00	2.00	2.00	2.00	FUS <sub>0-43</sub> , TDP-43 <sub>310-350</sub> , RNA Pol II <sub>1927-1970</sub>
ff99SBws-S	2.00	1.00	2.00	2.00	FUS <sub>0-43</sub>
ff99SBws-T	2.00	2.00	1.00	2.00	FUS <sub>0-43</sub>
ff99SBws-ST	2.00	1.00	1.00	2.00	FUS <sub>0-43</sub> , TDP-43 <sub>310-350</sub> , RNA Pol II <sub>1927-1970</sub>
ff99SBws-TQ	2.00	2.00	1.00	1.00	FUS <sub>0-43</sub> , TDP-43 <sub>310-350</sub> , RNA Pol II <sub>1927-1970</sub>
ff99SBws-STQ	2.00	1.00	1.00	1.00	FUS <sub>0-43</sub> , TDP-43 <sub>310-350</sub> , RNA Pol II <sub>1927-1970</sub>

Table S5: Root-mean-squared deviation (RMSD) of simulated  $\Delta\delta C_\alpha$ ,  $\Delta\delta C_\beta$ ,  $\Delta\delta C_\alpha - \Delta\delta C_\beta$  (in ppm) from NMR  $^{13}\text{C}$  chemical shifts of three IDPs for ff99SBws, modified ff99SBws variants (S,T,ST,TQ,STQ) (this study), and ff99SB-disp.

$\Delta\delta C_\alpha - \Delta\delta C_\beta$	FUS <sub>0-43</sub>	TDP-43 <sub>310-350</sub>	RNA Pol II <sub>1927-1970</sub>
ff99SBws	0.90 ± 0.11	0.64 ± 0.09	0.68 ± 0.11
ff99SB-disp	0.90 ± 0.11	0.78 ± 0.07	0.74 ± 0.11
ff99SBws-S	0.86 ± 0.10		
ff99SBws-T	0.98 ± 0.13		
ff99SBws-ST	0.64 ± 0.07	0.71 ± 0.08	0.74 ± 0.08
ff99SBws-TQ	0.62 ± 0.08	0.66 ± 0.06	0.71 ± 0.10
ff99SBws-STQ	0.54 ± 0.07	0.51 ± 0.04	0.74 ± 0.08

$\Delta\delta C_\alpha$	FUS <sub>0-43</sub>	TDP-43 <sub>310-350</sub>	RNA Pol II <sub>1927-1970</sub>
ff99SBws	0.58 ± 0.07	0.43 ± 0.06	0.67 ± 0.11
ff99SB-disp	0.59 ± 0.07	0.54 ± 0.05	0.62 ± 0.09
ff99SBws-S	0.51 ± 0.06		
ff99SBws-T	0.61 ± 0.08		
ff99SBws-ST	0.41 ± 0.04	0.41 ± 0.04	0.62 ± 0.08
ff99SBws-TQ	0.36 ± 0.05	0.40 ± 0.04	0.64 ± 0.09
ff99SBws-STQ	0.37 ± 0.04	0.36 ± 0.03	0.62 ± 0.08

$\Delta\delta C_\beta$	FUS <sub>0-43</sub>	TDP-43 <sub>310-350</sub>	RNA Pol II <sub>1927-1970</sub>
ff99SBws	0.43 ± 0.05	0.42 ± 0.05	0.51 ± 0.06
ff99SB-disp	0.39 ± 0.05	0.39 ± 0.06	0.44 ± 0.11
ff99SBws-S	0.45 ± 0.06		
ff99SBws-T	0.44 ± 0.05		
ff99SBws-ST	0.37 ± 0.04	0.44 ± 0.05	0.47 ± 0.10
ff99SBws-TQ	0.40 ± 0.05	0.41 ± 0.06	0.42 ± 0.11
ff99SBws-STQ	0.34 ± 0.04	0.34 ± 0.06	0.47 ± 0.10

## Protein sequences used in NMR experiments

### FUS Low Complexity domain FUS<sub>1-163</sub> (BMRB 26672)<sup>1</sup>

MASNDYTQQATQSYGAYPTQ PGQGYSSQSSQPYGQQSYSG YSQSTDTSGYGQSSYSSYGQ  
SQNTGYGTQSTPQGYGSTGG YGSSQSSQSSYGQQSSYPGY GQQPAPSSTSGSYGSSSQSS  
SYGQPQSGSYSQQPSYGGQQ QSYGQQQSYNPPQGYGQQNQ YNS  
(163 residues)

### TDP-43 Wild Type Low Complexity C-terminal domain TDP43<sub>267-414</sub> (BMRB 26823)<sup>2</sup>

*GHMNRQLERSGRFGGNPGGF* GNQGGFGNSRGGGAGLGNNQ GSNMGGGMNFGAFSINPAMM  
AAAQAALQSSWGMMGLASQ QNQSGPSGNNQNGNMQREP NQAFGSGNNSYSGNSGAAI  
GWGSASNAGSGSGFNNGFGS SMDSKSSGWGM  
(151 residues)

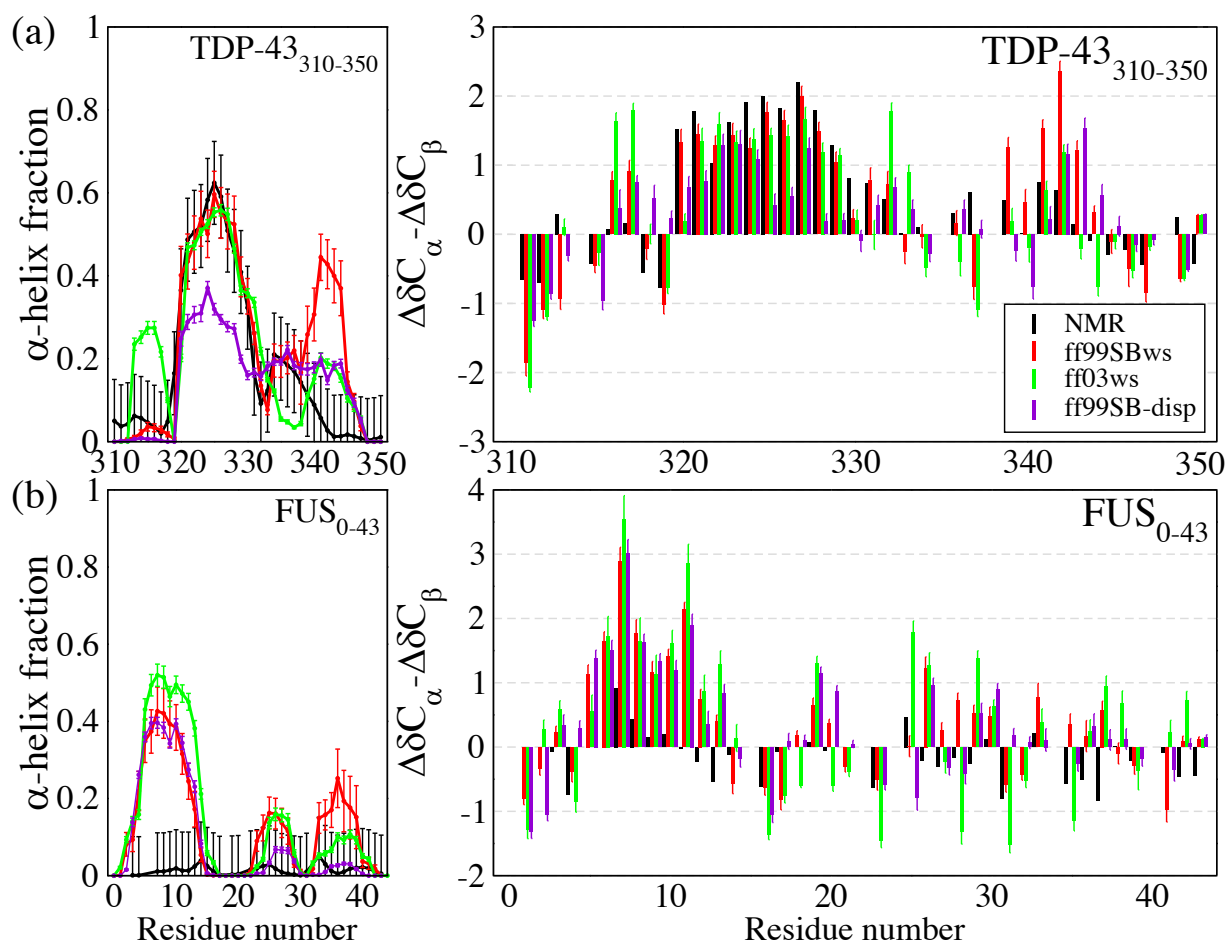
### RNA Pol II C-terminal Domain 27-52 RNA Pol II<sub>1770-1970</sub> (BMRB 27063)<sup>3</sup>

*GHMSPNYTPTSPNYSPTSPS* YSPTSPSYSPTSPSYSPSS RYTPQSPTYTPSSPSYSPSS  
PSYSPASPKYTPTSPSYSPS SPEYTPTSPKYSPTSPKYS TSPKYSPTSPYSPTPKYS  
PTSPTYSPVYTPTSPKY SPTSPTYSPKYSPTSPT YSPTSPKYSTYSPTSPGYSP  
TSPTYSLTSPAISPDDSDEE N  
(201 residues)

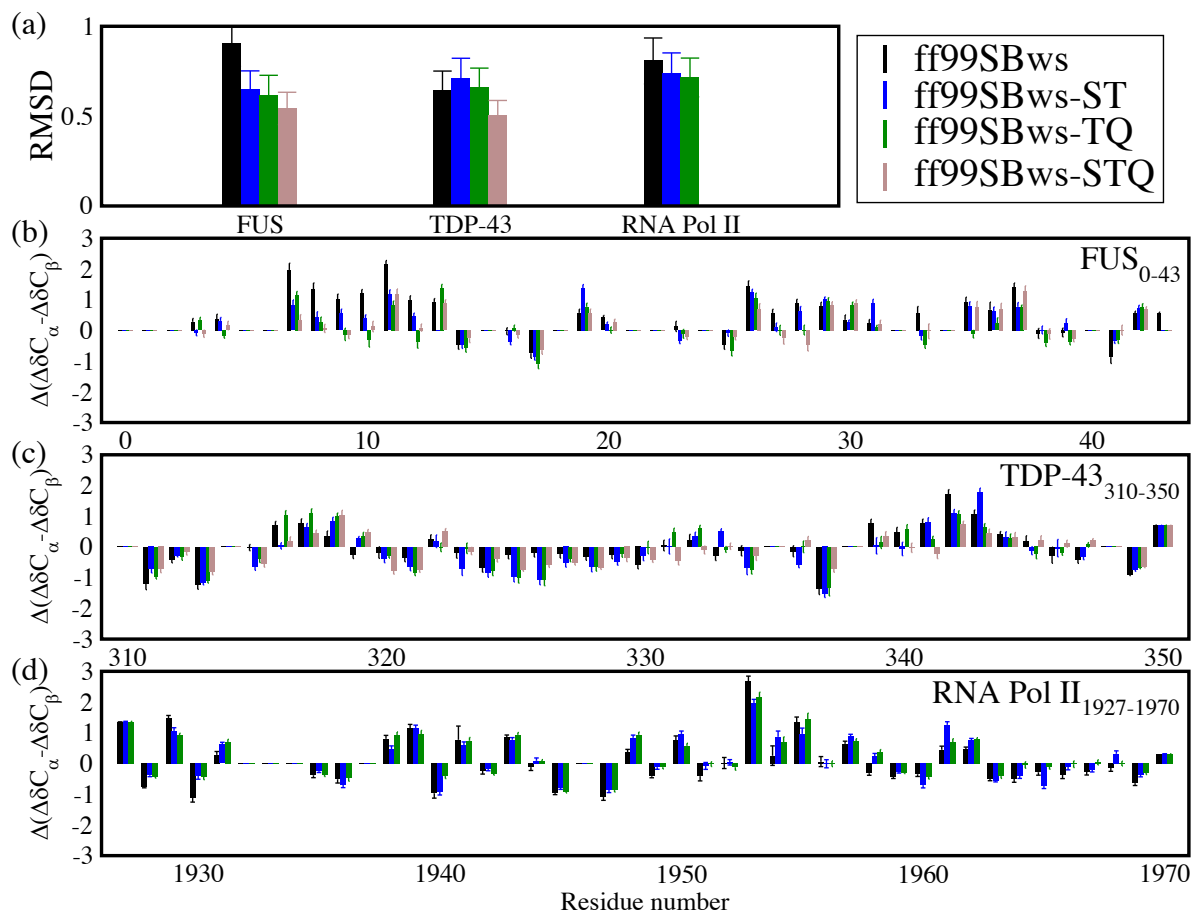
### hnRNPA2 low complexity domain 190-341 hnRNPA2<sub>190-341</sub> (BMRB 27123)<sup>4</sup>

*GHMGRGGNFGFGDSRGGGN* FGPGPSNFRGGSDGYGSGR FFGDGYNGYGGGPGGNFGG  
SPGYGGGRGGYGGGGPGYGN QGGYGGYDNYGGGNYGSG NYNDFGNYNQQPSNYGPMKS  
GNFGGSRNMGGPYGGGNYGP GSGGGSGGYGGRSRY  
(155 residues)

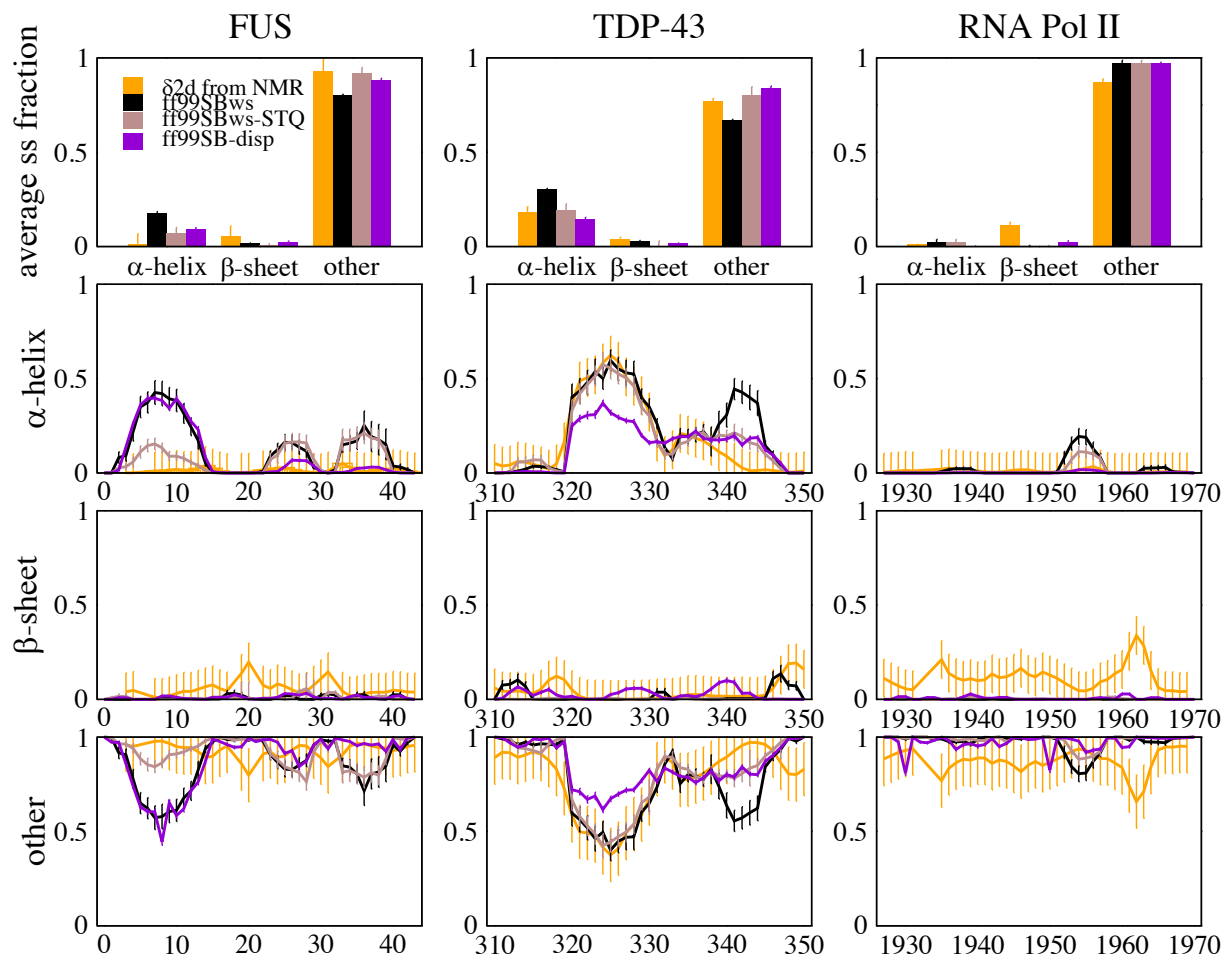
The residues in *italics* correspond to TEV-cleavage and cloning “scar” residues not present in the native sequences.



**Figure S1.** (a)  $\alpha$ -helix propensities and (b) Chemical shifts  $\Delta\delta C_{\alpha} - \Delta\delta C_{\beta}$  of TDP-43<sub>310-350</sub> and FUS<sub>0-43</sub> using all-atom force fields ff99SBws, ff03ws, and ff99SB-disp, compared to experimental results. Chemical shifts are calculated from protein structure using SPARTA+ algorithm. Secondary structures are assigned by DSSP. Experimental values of secondary structure propensities are calculated from NMR chemical shifts using the  $\delta$ 2D program.

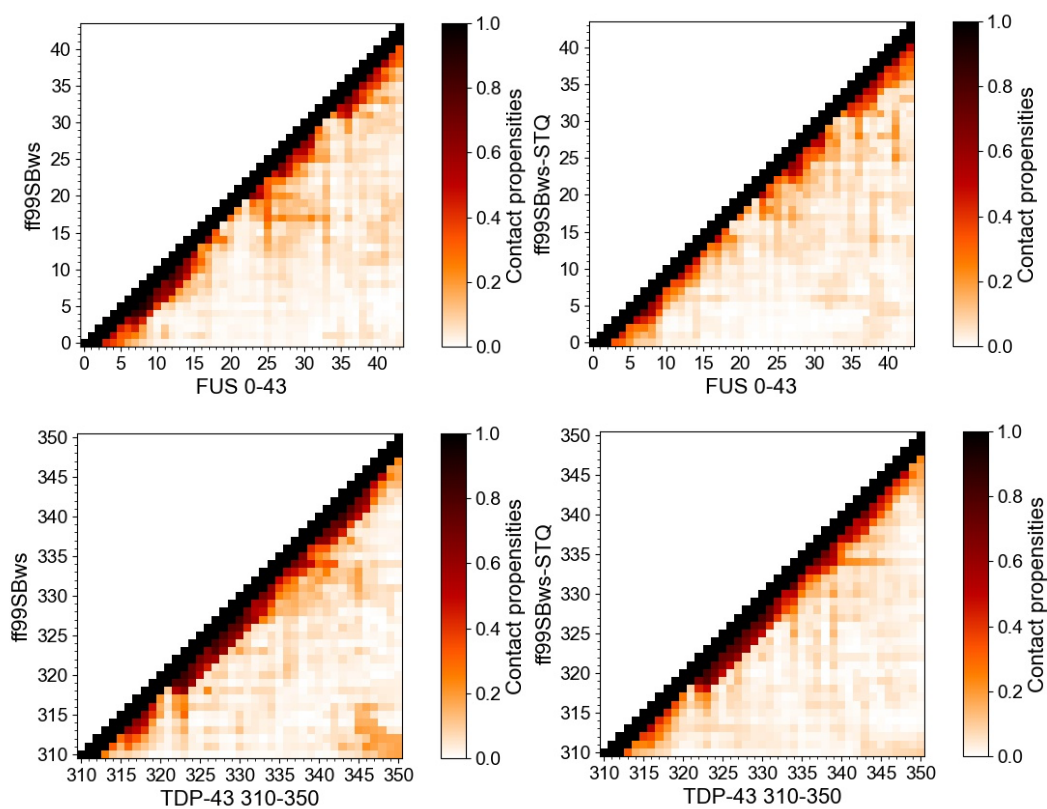


**Figure S2.** (a) Comparison of Root-mean-squared deviation (RMSD) of simulated  $\delta C_{\alpha} - \delta C_{\beta}$  (in ppm) from NMR  $^{13}\text{C}$  chemical shifts (RMSD) of simulated sequences and (b-d) chemical shift deviation from the experiment  $\Delta(\Delta\delta C_{\alpha} - \Delta\delta C_{\beta})$  with ff99SBws and modified force fields (see legend).

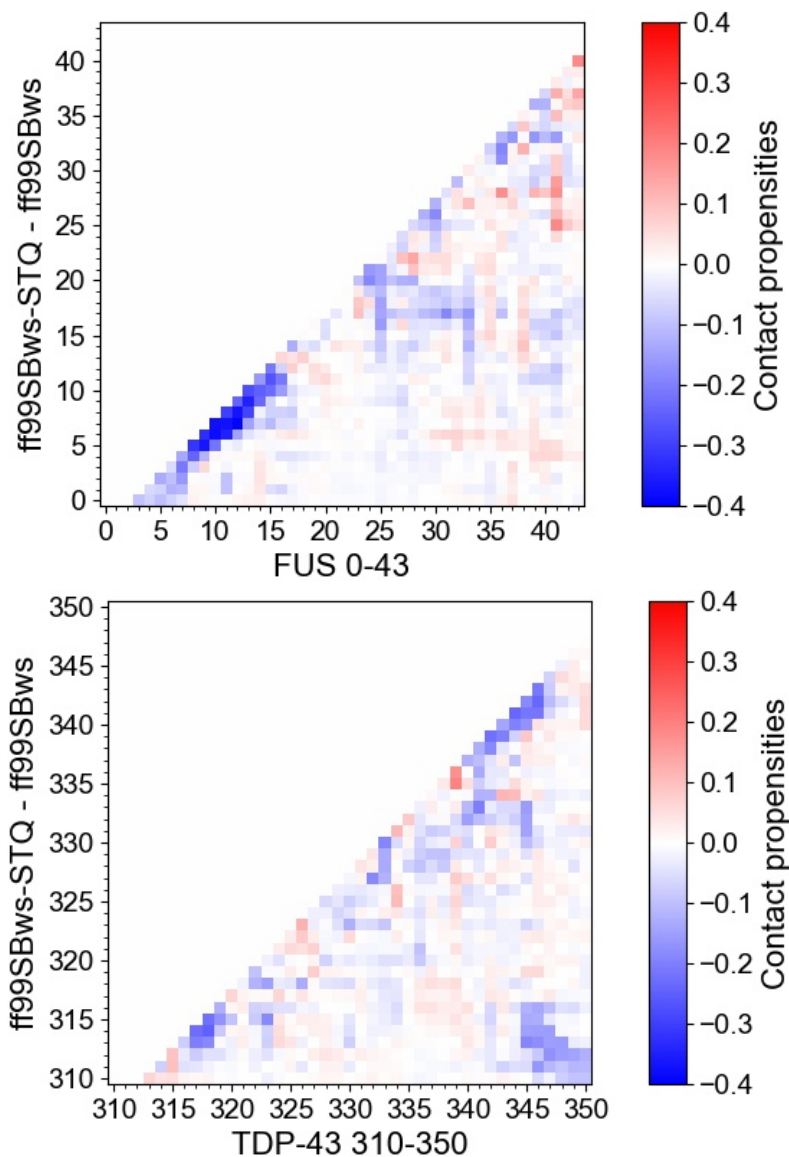


**Figure S3.** Secondary structure propensities of FUS<sub>0-43</sub>, TDP-43<sub>310-350</sub>, and RNA Pol II<sub>1927-1970</sub> with ff99SBws, ff99SBws-STQ (this study) and ff99SB-disp, compared to  $\delta$ 2D secondary structure propensities calculated from experimental NMR chemical shifts.

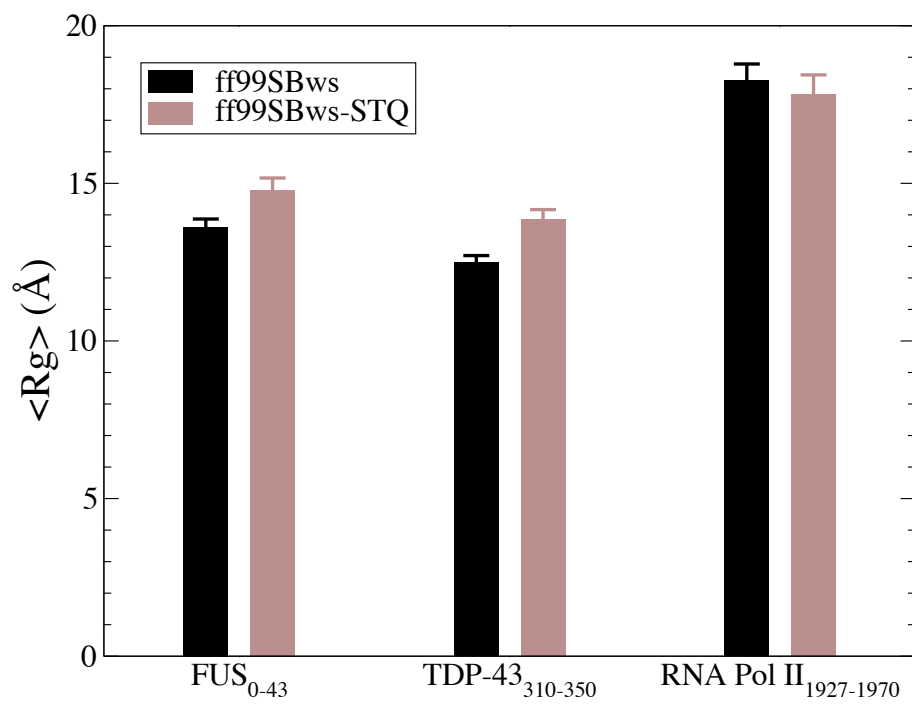




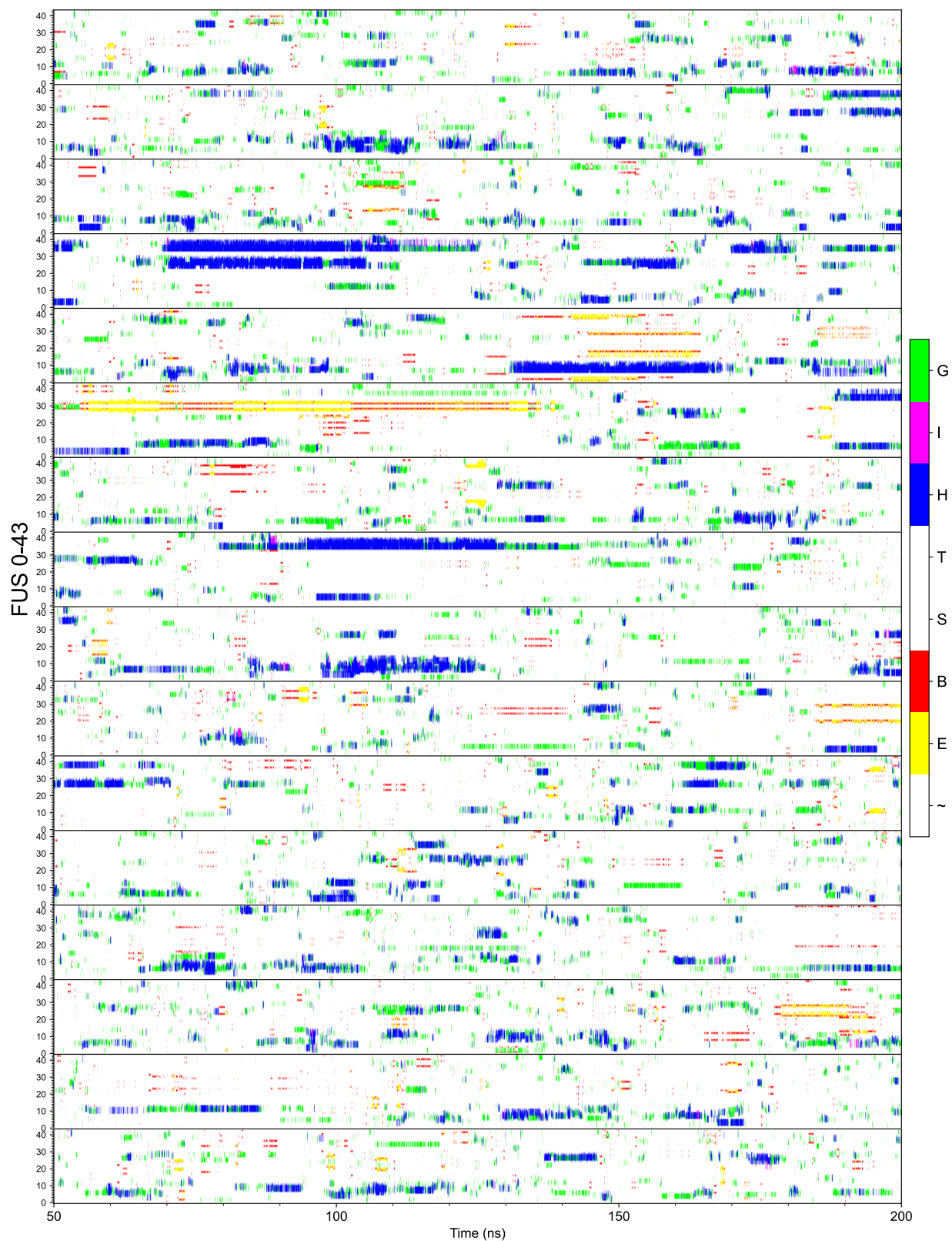
**Figure S4.** Intrachain residue-to-residue contact propensities in FUS<sub>0-43</sub> and TDP-43<sub>310-350</sub> with ff99SBws and ff99SBws-STQ (this study). Two residues are considered in contact when any heavy atoms from each of the residue are within 6 Angstrom in simulation. Contact propensities are the proportion of frames in simulation where two residues are in contact, with contact propensity = 1 being always in contact.



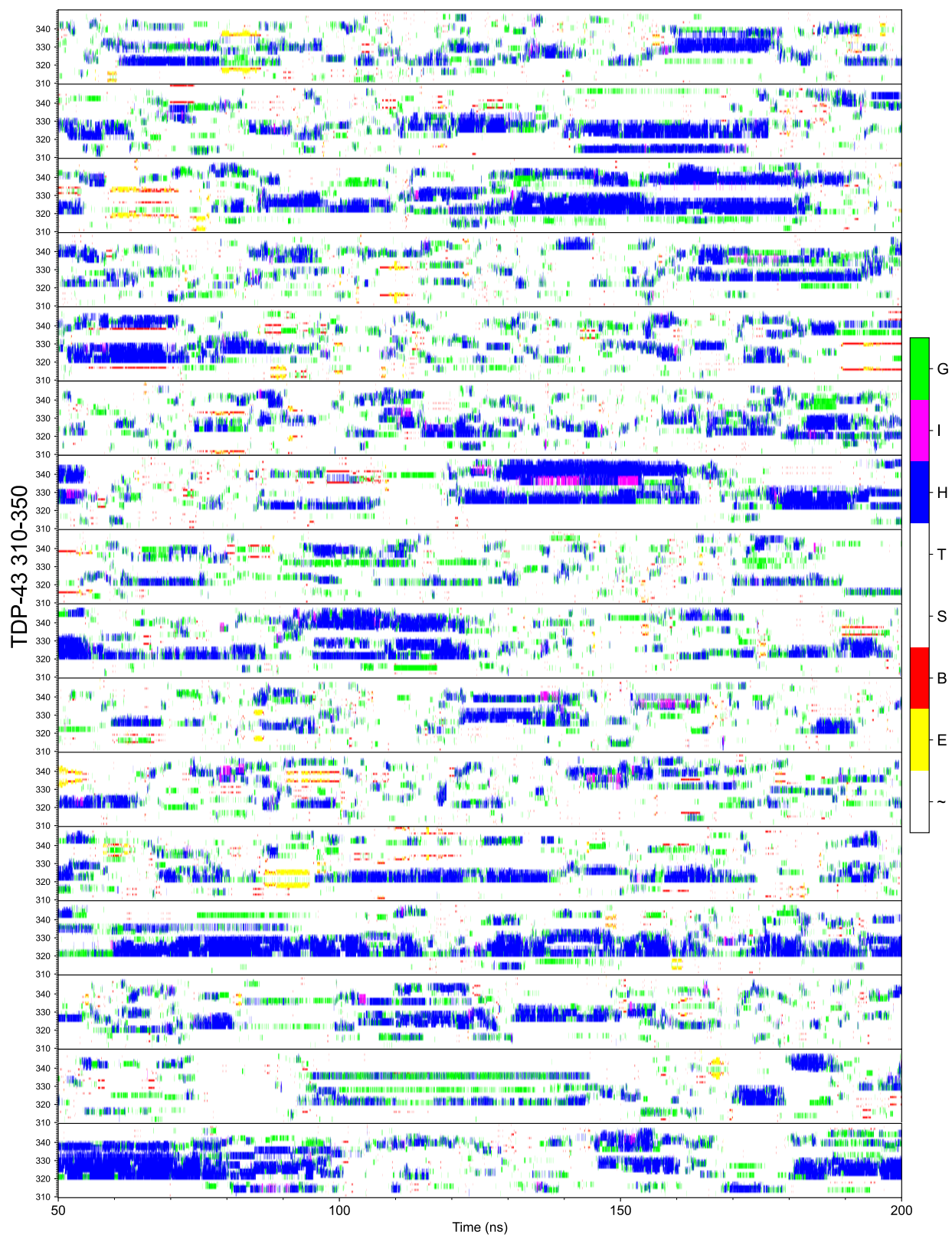
**Figure S5.** Change in Intrachain residue-to-residue contact propensities in FUS<sub>0-43</sub> and TDP-43<sub>310-350</sub> from ff99SBws to ff99SBws-STQ (this study) Two residues are considered in contact when any heavy atoms from each of the residue are within 6 Angstrom in simulation. Contact propensities are the proportion of frames in simulation where two residues are in contact, with contact propensity = 1 being always in contact.



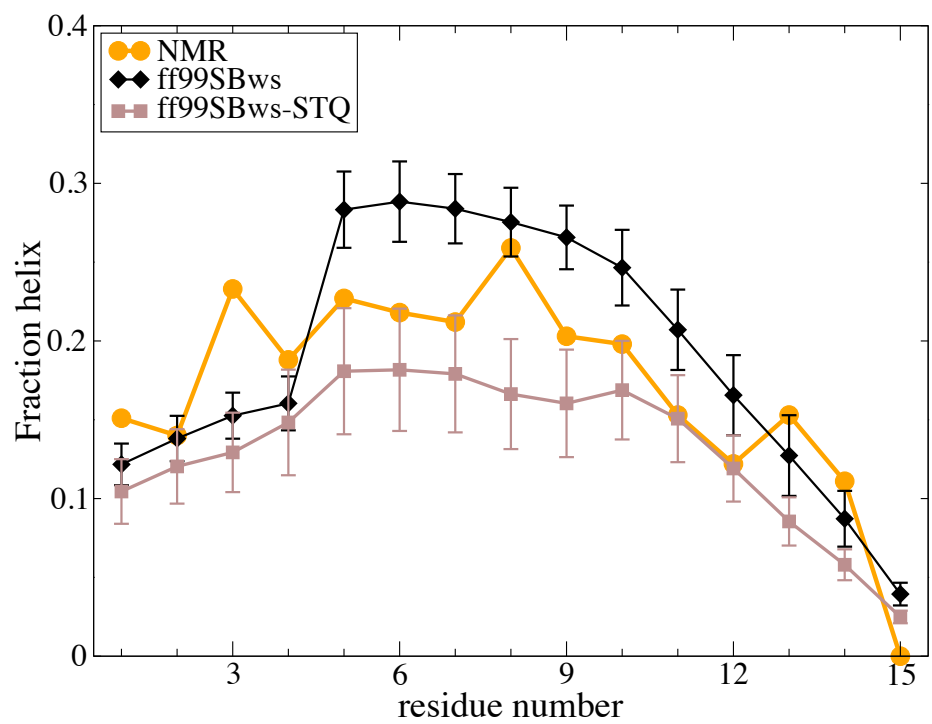
**Figure S6.** Average radius of gyration ( $R_g$ ) in FUS<sub>0-43</sub>, TDP-43<sub>310-350</sub> and RNA Pol II<sub>1927-1970</sub> with ff99SBws and ff99SBws-STQ (this study).



**Figure S7.** DSSP secondary structure as a function of simulation time for each demultiplexed replica moving through the temperature space of FUS<sub>0-43</sub> with ff99SBws-STQ (this study).



**Figure S8.** DSSP secondary structure as a function of simulation time for each demultiplexed replica moving through the temperature space of TDP-43<sub>310-350</sub> with ff99SBws-STQ (this study).



**Figure S9.** Fraction Helix of (AAQAA)<sub>3</sub> with ff99SBws and ff99SBws-STQ (this study) compared to NMR experimental data<sup>5</sup>.

## References

1. Burke, K. A., Janke, A. M., Rhine, C. L. & Fawzi, N. L. Residue-by-Residue View of In Vitro FUS Granules that Bind the C-Terminal Domain of RNA Polymerase II. *Mol. Cell* **60**, 231–241 (2015).
2. Conicella, A. E., Zerze, G. H., Mittal, J. & Fawzi, N. L. ALS Mutations Disrupt Phase Separation Mediated by  $\alpha$ -Helical Structure in the TDP-43 Low-Complexity C-Terminal Domain. *Structure* **24**, 1537–1549 (2016).
3. Janke, A. M. *et al.* Lysines in the RNA Polymerase II C-Terminal Domain Contribute to TAF15 Fibril Recruitment. *Biochemistry* **57**, 2549–2563 (2018).
4. Ryan, V. H. *et al.* Mechanistic View of hnRNPA2 Low-Complexity Domain Structure, Interactions, and Phase Separation Altered by Mutation and Arginine Methylation. *Mol. Cell* **69**, 465–479.e7 (2018).
5. Shalongo, W., Dugad, L. & Stellwagen, E. Distribution of Helicity within the Model Peptide Acetyl(AAQAA)<sub>3</sub>amide. *J. Am. Chem. Soc.* **116**, 8288–8293 (1994).