Pd-catalyzed Formal Mizoroki-Heck Coupling of Unactivated Alkyl Chlorides

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1. General information

Unless otherwise noted, all reactions were performed under inert conditions. Nuclear magnetic resonance (NMR) spectra were recorded in CDCl₃ on a Bruker AVANCE 300 (300 MHz), Bruker AVANCE 400 (400 MHz), or Bruker AVANCE III HD (400 MHz), and the residual solvent signal was used as a reference. High-resolution mass spectrometry (HRMS) was performed at the Organic Chemistry Research Center in Sogang University using the ESI method or the Korea Basic Science Institute (KBSI) using the EI method. Chemical shifts are reported in ppm and coupling constants are given in Hz. Gas chromatography (GC) was carried out using a 7890A or 7890B GC system (Agilent Technologies) equipped with an HP-5 column and a flame ionization detector (FID). Reactions were monitored by thin-layer chromatography (TLC) on EMD Silica Gel 60 F254 plates, and visualized either using UV light (254 nm) or by staining with potassium permanganate and heating. Anhydrous *N*,*N*-dimethylacetamide (DMA) was purchased from Sigma-Aldrich and used after typical degassing procedures. All chemicals were purchased from commercial sources (Sigma-Aldrich, Alfa Aesar, TCI, or Strem) and used without further purification. All photocatalytic reactions were conducted under irradiation by 40 W Blue LED lamps purchased from Kessil (Kessil A160WE) using the maximum light intensity and the shortest wavelength.

2. Substrate preparation

Alkyl chlorides 2d,¹ 2f,² 2g,² 2q,³ 2ai,⁴ and 2aj⁵ were prepared following literature procedures. Other noncommercial alkyl chlorides 2ae, 2af, and 2ag were prepared following the procedure provided below. Olefins $1v^6$ and $1x^7$ were prepared following literature procedures. Olefin 1w was prepared following the procedure provided below. All other substrates were purchased from commercial sources and used directly without further purification and the removal of stabilizers from olefins was not necessary.

Preparation of alkyl chlorides 2ae-2ag



To a flask equipped with a stirrer-bar were added the corresponding carboxylic acid (1.0 mmol, 1.0 equiv), N,N'-dicyclohexylcarbodiimide (DCC, 227 mg, 1.1 mmol, 1.1 equiv), 4-(dimethylamino)pyridine (12.2 mg, 0.10 mmol, 0.10 equiv), 3-chloro-1-propanol (125 μ L, 1.5 mmol, 1.5 equiv), and CH₂Cl₂ (5.0 mL). The resulting mixture was stirred at room temperature for 1 h and filtered to remove the urea by-product. The resulting mixture was concentrated under reduced pressure and purified by flash column chromatography (silica gel, hexanes/EtOAc gradient elution) to afford the desired products **2ae–2ag**.

Preparation of olefin 1w



To a stirred solution of δ -tocopherol (403 mg, 1.0 mmol, 1.0 equiv), triethylamine (418 µL, 3.0 mmol, 3.0 equiv) in CH₂Cl₂ (5.0 mL) was added trifluoromethanesulfonic anhydride (310 mg, 1.1 mmol, 1.1 equiv) dropwise at 0 °C. The resulting mixture was warmed up to room temperature and stirred for 1 h. The resulting mixture was quenched with NaHCO₃ (sat. aq, 10 mL), extracted with CH₂Cl₂ (10 mL × 3), dried (anhydrous Na₂SO₄), filtered, and concentrated under reduced pressure. The resulting mixture was purified by flash column chromatography (silica gel, hexanes/EtOAc gradient elution) to afford the desired product as a yellow oil (530 mg, 99% yield).



To a flask equipped with a stirrer-bar was added δ -tocopherol trifluoromethanesulfonate (428 mg, 0.80 mmol, 1.0 equiv), PdCl₂ (7.1 mg, 0.040 mmol, 0.050 equiv), RuPhos (37.3 mg, 0.080 mmol, 0.10 equiv), potassium vinyltrifluoroborate (214 mg, 2.0 mmol, 2.0 equiv), cesium carbonate (977 mg, 3.0 mmol, 3.0 equiv), THF (2.0 mL) and water (0.30 mL). The resulting mixture was stirred for 16 h at 85 °C and cooled to room temperature before the reaction mixture was quenched with NaHCO₃ (sat. aq, 10 mL), extracted with CH₂Cl₂ (10 mL × 3), dried (anhydrous Na₂SO₄), filtered, and concentrated under reduced pressure. The resulting mixture was purified by flash column chromatography (silica gel, hexanes/EtOAc gradient elution) to afford the desired product **1x** as a viscous colorless oil (248 mg, 75% yield).

3. General procedure for the alkyl chloride Mizoroki-Heck coupling reaction



To a 4 mL vial equipped with a stirrer-bar were added Pd(PPh₃)₄ (5.8 mg, 0.0050 mmol, 0.050 equiv), K₂CO₃ (27.6 mg, 0.20 mmol, 2.0 equiv), the corresponding olefin (0.10 mmol, 1.0 equiv), the corresponding alkyl chloride (0.15 mmol, 1.5 equiv), and *N*,*N*-dimethylacetamide (1.0 mL). The resulting mixture was stirred for 24 h under 40 W blue LED irradiation with fan cooling (\sim 30 °C). The reaction mixture was added brine (10 mL), diluted with EtOAc or Et₂O (10 mL), washed with brine (10 mL), dried (anhydrous Na₂SO₄), filtered, and concentrated under reduced pressure. The resulting residue was purified by flash column chromatography (silica gel, hexanes/EtOAc or hexanes/Et₂O gradient elution) to afford the desired product.

4. Characterization data

New substrates (2ae, 2af, 2ag, 1w) were characterized by ¹H NMR, ¹³C NMR and HRMS. The identity of the reported Heck coupling products (3a–3c, 3f, 3h–3s, 3u–3ab, 4a–7a, 9a–20a, 22a–24a, 3ac, 3ah, 3ai) were confirmed by ¹H NMR and spectral comparison with literature data, while new products (3d, 3e, 3g, 3t, 3y, 8a, 21a, 3ad, 25a, 3ae, 3af, 26a, 3ag, 3aj) were characterized by ¹H NMR, ¹³C NMR and HRMS.

References to characterization data for the reported compounds:

 $(3a, 3b, 3l, 3n, 3o, 3w, 3z, 3aa, 7a, 9a, 12a, 16a, 18a, 20a, 23a, 3ac)^8$; $3c^9$; $(3f, 3p)^{10}$; $(3h, 3i, 3k, 3s, 11a, 17a)^{11}$; $3j^{12}$; $3m^{13}$; $(3q, 15a)^{14}$; $3r^{15}$; $(3u, 3v, 3x)^{16}$; $3ab^{17}$; $(4a, 10a, 13a)^{18}$; $5a^{19}$; $6a^{20}$; $14a^{21}$; $19a^{22}$; $22a^{23}$; $3ah^{24}$; $3ai^{25}$.



3-Chloropropyl 5-(2,5-dimethylphenoxy)-2,2-dimethylpentanoate (2ae)

Colorless oil, 261 mg (0.80 mmol, 80% yield); ¹H NMR (400 MHz, CDCl₃): δ = 7.06 (dd, *J* = 7.5, 1.1 Hz, 1H),

6.71 (d, J = 7.5 Hz, 1H), 6.66 (s, 1H), 4.27 (td, J = 6.1, 0.9 Hz, 2H), 3.98 (tt, J = 4.9, 1.2 Hz, 2H), 3.65 (td, J = 6.4, 0.8 Hz, 2H), 2.36 (s, 3H), 2.24 (s, 3H), 2.18 – 2.08 (m, 2H), 1.82 – 1.76 (m, 4H), 1.28 (d, J = 1.2 Hz, 6H); ¹³C NMR (100 MHz, CDCl₃): $\delta = 177.6$, 156.9, 136.5, 130.3, 123.6, 120.8, 112.0, 67.9, 61.1, 42.2, 41.2, 37.2, 31.6, 25.2, 25.2, 21.4, 15.8; HRMS-ESI (m/z) [M+Na]⁺ calcd for C₁₈H₂₇ClO₃Na, 349.1541; found: 349.1543.



3-Chloropropyl (S)-2-(6-methoxynaphthalen-2-yl)propanoate (2af)

White solid, 278 mg (0.91 mmol, 91% yield); ¹H NMR (600 MHz, CDCl₃): δ = 7.61 (dd, *J* = 9.2, 4.6 Hz, 2H), 7.57 (s, 1H), 7.31 (d, *J* = 8.8 Hz, 1H), 7.06 (d, *J* = 9.0 Hz, 1H), 7.02 (s, 1H), 4.13 (q, *J* = 6.1 Hz, 2H), 3.78 (s, 3H), 3.33 (td, *J* = 6.6, 6.0, 3.3 Hz, 2H), 1.89 (t, *J* = 6.4 Hz, 2H), 1.49 (d, *J* = 7.4 Hz, 2H); ¹³C NMR (75 MHz, CDCl₃): δ = 174.5, 157.7, 135.6, 135.6, 133.7, 129.3, 129.2, 128.9, 127.2, 127.2, 126.2, 126.1, 125.9, 125.9, 119.1, 119.0, 105.6, 105.5, 61.4, 61.3, 55.3, 55.2, 55.2, 45.4, 45.4, 45.4, 41.1, 41.0, 31.5, 31.4, 18.5, 18.4; HRMS-ESI (m/z) [M+Na]⁺ calcd for C₁₇H₁₉ClO₃Na, 329.0915; found: 329.0916.



3-Chloropropyl 5-((3aS,4S,6aR)-2-oxohexahydro-1H-thieno[3,4-d]imidazol-4-yl)pentanoate (2ag)

White solid, 317 mg (0.98 mmol, 98% yield); ¹H NMR (400 MHz, CDCl₃) δ 6.32 (s, 1H), 5.95 (s, 1H), 4.47 (t, J = 6.4 Hz, 1H), 4.33 – 4.22 (m, 1H), 4.18 (t, J = 6.1 Hz, 2H), 3.58 (t, J = 6.4 Hz, 2H), 3.17 – 3.05 (m, 1H), 2.87 (dd, J = 12.7, 4.6 Hz, 1H), 2.70 (d, J = 12.5 Hz, 1H), 2.31 (t, J = 7.5 Hz, 2H), 2.06 (p, J = 6.2 Hz, 2H), 1.74 – 1.53 (m, J = 7.8 Hz, 4H), 1.49 – 1.32 (m, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 173.6, 62.0, 61.1, 60.2, 55.6, 41.4, 40.6, 33.9, 31.5, 28.4, 28.3, 24.8; HRMS-ESI (m/z) [M+Na]⁺ calcd for C₁₃H₂₁ClN₂O₃SNa, 343.0854; found: 343.0856.



(R)-2,8-Dimethyl-2-((4R,8R)-4,8,12-trimethyltridecyl)-6-vinylchromane (1w)

Viscous colorless oil, 272 mg (0.53 mmol, 66% yield); ¹H NMR (300 MHz, CDCl₃): δ = 7.03 (d, *J* = 2.2 Hz, 1H), 6.93 (d, *J* = 2.3 Hz, 1H), 6.57 (dd, *J* = 17.6, 10.9 Hz, 1H), 5.53 (dd, *J* = 17.6, 1.2 Hz, 1H), 5.03 (dd, *J* = 10.8, 1.2 Hz, 1H), 2.72 (t, *J* = 6.7 Hz, 2H), 2.14 (s, 3H), 1.87 – 1.67 (m, 2H), 1.62 – 0.97 (m, 21H), 0.88 – 0.79 (m, 15H); ¹³C NMR (100 MHz, CDCl₃): δ = 152.4, 137.0, 128.7, 126.6, 126.4, 125.3, 120.5, 110.7, 76.5, 40.3, 39.6, 37.7, 37.5, 33.1, 32.9, 31.5, 28.2, 25.1, 24.7, 24.5, 23.0, 22.9, 22.6, 21.2, 20.0, 19.9, 16.4; HRMS-ESI (m/z) [M+H]⁺ calcd for C₂₉H₄₉O, 413.3778; found: 413.3782.



(E)-1-(3,3-Dimethylbut-1-en-1-yl)-4-methoxybenzene (3a)

Colorless oil, 18.2 mg (96% yield); ¹H NMR (300 MHz, CDCl₃): $\delta = 7.32 - 7.20$ (m, 2H), 6.87 - 6.78 (m, 2H), 6.23 (d, J = 16.2 Hz, 1H), 6.10 (d, J = 16.2 Hz, 1H), 3.78 (s, 3H), 1.09 (s, 9H).

1-((*E*)-4-Methoxystyryl)adamantane (3b)

Colorless oil, 24.2 mg (90% yield); ¹H NMR (300 MHz, CDCl₃): δ = 7.28 (d, *J* = 8.7 Hz, 2H), 6.82 (d, *J* = 8.8 Hz, 2H), 6.17 (d, *J* = 16.3 Hz, 1H), 5.95 (d, *J* = 16.3 Hz, 1H), 3.78 (s, 3H), 2.06 - 1.94 (m, 3H), 1.78 - 1.60 (m, 12H).



5-((*E*)-4-Methoxystyryl)adamantan-2-one (3c)

White solid, 28.1 mg (>96% yield); ¹H NMR (300 MHz, CDCl₃): $\delta = 7.30 - 7.24$ (m, 2H), 6.82 (d, J = 8.8 Hz, 2H), 6.23 (d, J = 16.3 Hz, 1H), 5.94 (d, J = 16.3 Hz, 1H), 3.78 (s, 3H), 2.58 (s, 2H), 2.18 (d, J = 3.2 Hz, 1H), 2.05 – 1.89 (m, 10H).

MeO

5-((E)-4-Methoxystyryl)adamantan-2-one (3d)

White solid, 20.6 mg (73% yield); ¹H NMR (300 MHz, CDCl₃): $\delta = 7.32 - 7.26$ (m, 2H), 6.89 - 6.79 (m, 2H), 6.24 (d, J = 16.6 Hz, 1H), 6.12 (d, J = 16.6 Hz, 1H), 3.85 - 3.74 (m, 3H), 2.09 (d, J = 11.7 Hz, 4H), 1.87 - 1.49 (m, 10H), 1.15 (s, 3H); ¹³C NMR (100 MHz, CDCl₃): $\delta = 158.7$, 140.6, 131.5, 127.0, 125.6, 114.1, 114.1, 55.6, 41.4, 39.2, 36.8, 34.3, 33.2, 28.3, 28.1, 27.3; HRMS-EI (m/z) [M]⁺ calcd for C₂₀H₂₆O, 282.1984; found: 282.1982.



(E)-1-(3,3-Dimethylhept-1-en-1-yl)-4-methoxybenzene (3e)

Colorless oil, 22.3 mg (96% yield); ¹H NMR (400 MHz, CDCl₃): δ = 7.34 – 7.26 (m, 2H), 6.83 (d, *J* = 8.8 Hz, 2H), 6.20 (d, *J* = 16.3 Hz, 1H), 6.03 (d, *J* = 16.2 Hz, 1H), 3.79 (s, 3H), 1.36 – 1.30 (m, 2H), 1.30 – 1.15 (m, 4H), 1.06 (s, 6H), 0.87 (t, *J* = 7.0 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ = 158.8, 139.1, 131.2, 127.2, 125.1, 114.1, 55.5, 43.4, 36.3, 27.5, 27.2, 23.7, 14.4; HRMS-EI (m/z) [M]⁺ calcd for C₁₆H₂₄O, 232.1827; found: 232.1827.



(E)-1-(3,3-Dimethyl-5-phenylpent-1-en-1-yl)-4-methoxybenzene (3f)

White solid, 25.3 mg (90% yield); ¹H NMR (300 MHz, CDCl₃): $\delta = 7.36 - 7.21$ (m, 4H), 7.16 (d, J = 7.2 Hz, 3H), 6.94 - 6.76 (m, 2H), 6.28 (d, J = 16.2 Hz, 1H), 6.08 (d, J = 16.2 Hz, 1H), 3.80 (s, 3H), 2.64 - 2.48 (m, 2H), 1.73 - 1.60 (m, 2H), 1.15 (s, 6H).

(E)-1-Methoxy-4-(2-(1-methyl-4-phenylcyclohexyl)vinyl)benzene (3g)

White solid, 29.1 mg (95% yield, 2:1 mixture of diastereomers); ¹H NMR (600 MHz, CDCl₃, mixture of diastereomers): $\delta = 7.38 - 7.31$ (m, 8H), 7.30 - 7.25 (m, 5H), 7.23 - 7.15 (m, 8H), 6.90 - 6.85 (m, 6H), 6.36 (d, J = 16.4 Hz, 2H), 6.32 (d, J = 16.3 Hz, 1H), 6.15 (d, J = 16.1 Hz, 1H), 6.13 (d, J = 16.5 Hz, 2H), 3.82 (s, 6H), 3.82 (s, 3H), 2.55 - 2.47 (m, 3H), 1.97 - 1.91 (m, 4H), 1.84 - 1.46 (m, 20H), 1.21 (s, 3H), 1.11 (s, 6H).; ¹³C NMR (150 MHz, CDCl₃, mixture of diastereomers): $\delta = 158.7$, 158.7, 147.5, 147.5, 141.2, 136.2, 131.0, 130.9, 128.3, 128.3, 127.1, 127.0, 126.9, 126.9, 125.9, 125.8, 123.9, 114.0, 113.9, 55.3, 55.3, 44.4, 44.3, 38.6, 37.5, 36.0, 35.1, 31.9, 30.5, 29.7, 22.2; HRMS-EI (m/z) [M]⁺ calcd for C₂₂H₂₆O, 306.1984; found: 306.1985.



(E)-1-Methoxy-4-(3-methylbut-1-en-1-yl)benzene (3h)

Colorless oil, 16.2 mg (92% yield); ¹H NMR (300 MHz, CDCl₃): $\delta = 7.35 - 7.21$ (m, 2H), 6.87 - 6.77 (m, 2H), 6.26 (d, J = 15.9 Hz, 1H), 6.03 (dd, J = 15.9, 6.8 Hz, 1H), 3.78 (s, 3H), 2.50 - 2.33 (m, 1H), 1.06 (d, J = 6.8 Hz, 6H).



(E)-1-Methoxy-4-(3-methylpent-1-en-1-yl)benzene (3i)

Colorless oil, 19.0 mg (>96% yield); ¹H NMR (300 MHz, CDCl₃): $\delta = 7.29 - 7.25$ (m, 2H), 6.82 (d, J = 8.8 Hz, 2H), 6.27 (d, J = 15.9 Hz, 1H), 5.93 (dd, J = 15.8, 7.8 Hz, 1H), 3.78 (s, 3H), 2.15 (p, J = 6.9 Hz, 1H), 1.44 - 1.32 (m, 2H), 1.04 (d, J = 6.7 Hz, 3H), 0.88 (t, J = 7.4 Hz, 3H).

(E)-1-(2-Cyclobutylvinyl)-4-methoxybenzene (3j)

Colorless oil, 17.0 mg (90% yield); ¹H NMR (300 MHz, CDCl₃): $\delta = 7.32 - 7.23$ (m, 2H), 6.88 - 6.75 (m, 2H), 6.27 - 6.16 (m, 2H), 3.78 (s, 3H), 3.13 - 2.95 (m, 1H), 2.24 - 2.05 (m, 2H), 2.00 - 1.73 (m, 4H).



(E)-1-(2-Cyclopentylvinyl)-4-methoxybenzene (3k)

Colorless oil, 19.6 mg (>96% yield); ¹H NMR (300 MHz, CDCl₃): $\delta = 7.31 - 7.20$ (m, 2H), 6.89 - 6.77 (m, 2H), 6.29 (d, J = 15.8 Hz, 1H), 6.04 (dd, J = 15.8, 7.7 Hz, 1H), 3.78 (s, 4H), 2.62 - 2.48 (m, 1H), 1.90 - 1.76 (m, 2H), 1.70 - 1.51 (m, 4H), 1.45 - 1.27 (m, 2H).

(E)-1-(2-Cyclohexylvinyl)-4-methoxybenzene (3l)

Colorless oil, 20.7 mg (96% yield); ¹H NMR (300 MHz, CDCl₃): δ = 7.26 (dd, *J* = 8.5, 1.8 Hz, 2H), 6.93 – 6.75 (m, 2H), 6.33 – 6.21 (m, 1H), 6.01 (dd, *J* = 16.0, 6.9 Hz, 1H), 3.78 (s, 3H), 2.16 – 1.99 (m, 1H), 1.85 – 1.59 (m, 5H), 1.40 – 1.07 (m, 5H).



(E)-2-(4-Methoxystyryl)-2,3-dihydro-1H-indene (3m)

Bright yellow solid, 24.5 mg (>96% yield); ¹H NMR (300 MHz, CDCl₃): $\delta = 7.38 - 7.13$ (m, 8H), 6.88 - 6.80 (m, 2H), 6.43 (d, J = 15.8 Hz, 1H), 6.22 (dd, J = 15.8, 7.6 Hz, 1H), 3.79 (s, 3H), 3.30 - 3.20 (m, 1H), 3.20 - 3.06 (m, 2H), 2.85 (dd, J = 15.1, 8.0 Hz, 2H).



2-((E)-4-Methoxystyryl)adamantane (3n)

Colorless solid, 17.0 mg (63% yield); ¹H NMR (300 MHz, CDCl₃): $\delta = 7.36 - 7.27$ (m, 2H), 6.87 - 6.80 (m, 2H), 6.41 - 6.31 (m, 2H), 3.78 (s, 3H), 2.52 (s, 1H), 1.99 (d, J = 12.7 Hz, 2H), 1.93 - 1.80 (m, 6H), 1.73 (s, 2H), 1.55 (d, J = 12.5 Hz, 2H).

2-((*E*)-4-Methoxystyryl)bicyclo[2.2.1]heptane (30)

Colorless oil, 21.6 mg (95% yield); ¹H NMR (300 MHz, CDCl₃): $\delta = 7.30 - 7.19$ (m, 2H), 6.86 - 6.77 (m, 2H), 6.22 (d, J = 15.8 Hz, 1H), 5.96 (dd, J = 15.8, 8.1 Hz, 1H), 3.78 (s, 3H), 2.29 - 2.14 (m, 2H), 2.09 (s, 1H), 1.57 - 1.45 (m, 3H), 1.45 - 1.38 (m, 1H), 1.37 - 1.26 (m, 2H), 1.18 - 1.07 (m, 2H).



(E)-4-(4-Methoxystyryl)tetrahydro-2H-pyran (3p)

Yellow oil, 22.1 mg (>96% yield); ¹H NMR (300 MHz, CDCl₃): δ = 7.27 (d, *J* = 8.7 Hz, 2H), 6.82 (d, *J* = 8.7 Hz, 2H), 6.31 (dd, *J* = 15.9, 1.3 Hz, 1H), 6.00 (dd, *J* = 16.0, 6.8 Hz, 1H), 3.98 (ddd, *J* = 11.4, 4.5, 2.0 Hz, 2H), 3.78 (s, 3H), 3.44 (td, *J* = 11.6, 2.3 Hz, 2H), 2.43 – 2.21 (m, 1H), 1.77 – 1.63 (m, 2H), 1.63 – 1.49 (m, 2H).



tert-Butyl (*E*)-4-(4-methoxystyryl)piperidine-1-carboxylate (3q)

Yellow oil, 27.6 mg (87% yield); ¹H NMR (300 MHz, CDCl₃): δ = 7.25 (d, *J* = 8.8 Hz, 2H), 6.82 (d, *J* = 8.7 Hz, 2H), 6.31 (d, *J* = 15.9 Hz, 1H), 5.98 (dd, *J* = 16.0, 6.9 Hz, 1H), 4.10 (s, 2H), 3.78 (s, 3H), 2.75 (t, *J* = 12.6 Hz, 2H), 2.33 - 2.14 (m, 1H), 1.72 (d, *J* = 13.2 Hz, 2H), 1.45 (s, 9H), 1.35 (dt, *J* = 12.5, 6.0 Hz, 2H).



(E)-2-(4-Methoxystyryl)cyclohexan-1-ol (3r)

Yellow oil, 20.1 mg (87% yield, 1:2.7 mixture of diastereomers); ¹H NMR (400 MHz, CDCl₃, reported for the major isomer): $\delta = 7.38 - 7.31$ (m, 2H), 6.91 - 6.84 (m, 2H), 6.50 (d, J = 15.9 Hz, 1H), 5.94 (dd, J = 15.8, 8.9 Hz, 1H), 3.83 (s, 3H), 3.35 (td, J = 10.0, 4.3 Hz, 1H), 2.07 (t, J = 12.7 Hz, 2H), 1.89 - 1.60 (m, 4H), 1.40 - 1.20 (m, 4H).



(*E*)-1-(Hex-1-en-1-yl)-4-methoxybenzene (3s)

Yellow oil, 15.2 mg (80% yield); ¹H NMR (300 MHz, CDCl₃): δ = 7.25 (d, *J* = 8.6 Hz, 2H), 6.81 (d, *J* = 8.7 Hz, 2H), 6.30 (d, *J* = 15.8 Hz, 1H), 6.06 (dt, *J* = 15.7, 6.9 Hz, 1H), 3.78 (s, 3H), 2.24 – 2.11 (m, 2H), 1.48 – 1.31 (m, 4H), 0.90 (t, *J* = 7.0 Hz, 3H).



(E)-(5-(4-Methoxyphenyl)pent-4-en-1-yl)trimethylsilane (3t)

Yellow oil, 18.3 mg (74% yield); ¹H NMR (300 MHz, CDCl₃): δ = 7.27 (d, *J* = 8.8 Hz, 2H), 6.82 (d, *J* = 8.7 Hz, 2H), 6.37 – 6.25 (m, 1H), 6.06 (dt, *J* = 15.8, 6.9 Hz, 1H), 3.78 (s, 3H), 2.18 (qd, *J* = 7.1, 1.4 Hz, 2H), 1.54 – 1.36 (m, 2H), 0.62 – 0.45 (m, 2H), -0.03 (s, 9H); ¹³C NMR (100 MHz, CDCl₃): δ = 158.8, 131.0, 129.5, 129.1, 127.2, 114.1, 55.5, 37.1, 24.3, 16.6, -1.4; HRMS-EI (m/z) [M]⁺ calcd for C₁₅H₂₄OSi, 248.1596; found: 248.1595.

(E)-1-Methoxy-4-(5-phenylpent-1-en-1-yl)benzene (3u)

Yellow oil, 21.0 mg (83% yield); ¹H NMR (300 MHz, CDCl₃): δ = 7.31 – 7.22 (m, 4H), 7.22 – 7.14 (m, 3H), 6.85 – 6.77 (m, 2H), 6.32 (d, *J* = 15.8 Hz, 1H), 6.08 (dt, *J* = 15.6, 6.8 Hz, 1H), 3.78 (s, 3H), 2.72 – 2.59 (m, 2H), 2.26 – 2.17 (m, 2H), 1.88 – 1.71 (m, 2H).

(E)-1-(6-Chlorohex-1-en-1-yl)-4-methoxybenzene (3v)

Yellow oil, 16.7 mg (74% yield); ¹H NMR (300 MHz, CDCl₃) δ 7.29 – 7.22 (m, 2H), 6.85 – 6.78 (m, 2H), 6.32 (d, J = 15.8 Hz, 1H), 6.03 (dt, J = 15.8, 6.9 Hz, 1H), 3.78 (s, 3H), 3.55 (t, J = 6.7 Hz, 2H), 2.25 – 2.15 (m, 2H), 1.86 – 1.74 (m, 2H), 1.66 – 1.56 (m, 3H).



Ethyl (E)-6-(4-methoxyphenyl)hex-5-enoate (3w)

Yellow oil, 16.7 mg (67% yield); ¹H NMR (300 MHz, CDCl₃): $\delta = 7.27 - 7.23$ (m, 2H), 6.81 (d, J = 8.7 Hz, 2H), 6.32 (d, J = 15.8 Hz, 1H), 6.01 (dt, J = 15.7, 6.9 Hz, 1H), 4.10 (q, J = 7.1 Hz, 2H), 3.78 (s, 3H), 2.32 (t, J = 7.5 Hz, 2H), 2.21 (q, J = 6.6 Hz, 2H), 1.78 (p, J = 7.3 Hz, 2H), 1.23 (t, J = 7.1 Hz, 3H).



(E)-6-(4-Methoxyphenyl)hex-5-enenitrile (3x)

Yellow oil, 16.7 mg (67% yield); ¹H NMR (300 MHz, CDCl₃): δ = 7.28 – 7.20 (m, 2H), 6.83 (d, *J* = 8.7 Hz, 2H), 6.38 (d, *J* = 15.8 Hz, 1H), 5.96 (dt, *J* = 15.7, 7.1 Hz, 1H), 3.79 (s, 3H), 2.43 – 2.27 (m, 4H), 1.81 (p, *J* = 7.1 Hz, 2H).



(E)-7-(4-Methoxyphenyl)hept-6-en-2-one (3y)

Yellow oil, 15.3 mg (70% yield); ¹H NMR (400 MHz, CDCl₃): δ = 7.25 (d, *J* = 7.4 Hz, 2H), 6.82 (d, *J* = 8.8 Hz, 2H), 6.31 (d, *J* = 15.8 Hz, 1H), 6.00 (dt, *J* = 15.8, 7.0 Hz, 1H), 3.78 (s, 3H), 2.45 (t, *J* = 7.4 Hz, 2H), 2.18 (qd, *J* = 7.1, 1.4 Hz, 2H), 2.11 (s, 3H), 1.73 (p, *J* = 7.3 Hz, 2H).;¹³C NMR (100 MHz, CDCl₃): δ = 209.2, 159.0, 130.6, 130.2, 127.8, 127.3, 114.1, 55.5, 43.1, 32.5, 30.2, 23.6; HRMS-EI (m/z) [M]⁺ calcd for C₁₄H₁₈O₂, 218.1307; found: 218.1304.

,OH

(E)-8-(4-Methoxyphenyl)oct-7-en-1-ol (3z)

Yellow oil, 16.1 mg (69% yield); ¹H NMR (300 MHz, CDCl₃): $\delta = 7.30 - 7.20$ (m, 2H), 6.81 (d, J = 8.7 Hz, 2H), 6.41 - 6.22 (m, 1H), 6.05 (dt, J = 15.7, 6.9 Hz, 1H), 3.78 (s, 3H), 3.63 (t, J = 6.6 Hz, 2H), 2.17 (qd, J = 7.0, 1.4 Hz, 2H), 1.58 - 1.33 (m, 8H).

(E)-2-(3-(4-Methoxyphenyl)allyl)tetrahydrofuran (3aa)

Yellow oil, 16.7 mg (77% yield); ¹H NMR (300 MHz, CDCl₃): δ = 7.31 – 7.22 (m, 2H), 6.81 (d, *J* = 8.8 Hz, 2H), 6.45 – 6.30 (m, 1H), 6.07 (dt, *J* = 15.7, 7.1 Hz, 1H), 3.99 – 3.84 (m, 2H), 3.78 (d, *J* = 0.7 Hz, 3H), 3.77 – 3.66 (m, 1H), 2.56 – 2.28 (m, 2H), 2.06 – 1.75 (m, 3H), 1.64 – 1.48 (m, 1H).



(E)-1-(Hept-1-en-6-yn-1-yl)-4-methoxybenzene (3ab)

Yellow oil, 18.3 mg (91% yield); ¹H NMR (300 MHz, CDCl₃): δ = 7.25 (d, *J* = 8.9 Hz, 2H), 6.82 (d, *J* = 8.8 Hz, 2H), 6.34 (d, *J* = 15.8 Hz, 1H), 6.03 (dt, *J* = 15.8, 7.0 Hz, 1H), 3.78 (s, 3H), 2.39 – 2.17 (m, 4H), 1.95 (t, *J* = 2.6 Hz, 1H), 1.68 (p, *J* = 7.2 Hz, 2H).



(E)-(3,3-Dimethylbut-1-en-1-yl)benzene (4a)

Yellow oil, 14.8 mg (92% yield); ¹H NMR (300 MHz, CDCl₃): δ = 7.37 – 7.30 (m, 2H), 7.30 – 7.24 (m, 2H), 7.19 – 7.12 (m, 1H), 6.36 – 6.14 (m, 2H), 1.10 (s, 9H).



(E)-1-(3,3-Dimethylbut-1-en-1-yl)-2-methylbenzene (5a)

Yellow oil, 17.4 mg (>96% yield); ¹H NMR (300 MHz, CDCl₃): $\delta = 7.47 - 7.33$ (m, 1H), 7.19 - 7.04 (m, 3H), 6.47 (d, J = 16.0 Hz, 1H), 6.10 (d, J = 16.0 Hz, 1H), 2.32 (s, 3H), 1.15 - 1.09 (m, 9H).



(E)-1-(3,3-Dimethylbut-1-en-1-yl)-4-methylbenzene (6a)

Yellow oil, 14.8 mg (85% yield); ¹H NMR (400 MHz, CDCl₃): $\delta = 7.31 - 7.21$ (m, 2H), 7.08 (d, J = 7.9 Hz,

(E)-1-(tert-Butyl)-4-(3,3-dimethylbut-1-en-1-yl)benzene (7a)

Yellow oil, 21.6 mg (>96% yield); ¹H NMR (300 MHz, CDCl₃): δ = 7.29 (d, *J* = 1.2 Hz, 4H), 6.34 – 6.12 (m, 2H), 1.29 (s, 9H), 1.09 (s, 9H).

(E)-2-(3,3-Dimethylbut-1-en-1-yl)-1,3,5-trimethylbenzene (8a)

Yellow oil, 11.8 mg (58% yield); ¹H NMR (600 MHz, CDCl₃): $\delta = 6.85$ (s, 2H), 6.18 (d, J = 16.6 Hz, 1H), 5.66 (d, J = 16.6 Hz, 1H), 2.26 (s, 3H), 2.24 (s, 6H), 1.13 (s, 9H); ¹³C NMR (150 MHz, CDCl₃): $\delta = 146.4$, 146.4, 135.8, 135.4, 128.3, 121.7, 33.6, 29.6, 20.9, 20.7; HRMS-EI (m/z) [M]⁺ calcd for C₁₅H₂₂, 202.1722; found: 202.1722.

(E)-1-(3,3-Dimethylbut-1-en-1-yl)-4-fluorobenzene (9a)

Yellow oil, 17.4 mg (>96% yield); ¹H NMR (300 MHz, CDCl₃): $\delta = 7.34 - 7.26$ (m, 2H), 7.01 - 6.91 (m, 2H), 6.29 - 6.20 (m, 1H), 6.19 - 6.08 (m, 1H), 1.09 (d, J = 1.3 Hz, 9H).

(E)-1-(3,3-Dimethylbut-1-en-1-yl)-4-chlorobenzene (10a)

Yellow oil, 12.4 mg (64% yield); ¹H NMR (300 MHz, CDCl₃): $\delta = 7.26 - 7.23$ (m, 4H), 6.22 (d, J = 0.6 Hz, 2H), 1.09 (s, 9H).

(E)-5-(3,3-Dimethylbut-1-en-1-yl)benzo[d][1,3]dioxole (11a)

Yellow oil, 20.6 mg (>96% yield); ¹H NMR (300 MHz, CDCl₃): $\delta = 6.90$ (d, J = 1.6 Hz, 1H), 6.81 – 6.67 (m, 2H), 6.19 (d, J = 16.1 Hz, 1H), 6.06 (d, J = 16.2 Hz, 1H), 5.91 (s, 2H), 1.08 (s, 9H).



(*E*)-4-(3,3-Dimethylbut-1-en-1-yl)-1,1'-biphenyl (12a)

Yellow oil, 21.3 mg (90% yield, 1.2:1 ratio of stereoisomers); ¹H NMR (300 MHz, CDCl₃, mixture of isomers, calibrated to the minor isomer): $\delta = 7.64 - 7.21$ (m, 19.8H), 6.41 (d, J = 12.7 Hz, 1H), 6.31 (d, J = 2.1 Hz, 2.4H), 5.62 (d, J = 12.7 Hz, 1H), 1.13 (s, 10.8H), 1.01 (s, 9H).

(E)-(3-(3,3-Dimethylbut-1-en-1-yl)phenyl)methanol (13a)

Yellow oil, 19.2 mg (>96% yield); ¹H NMR (300 MHz, CDCl₃): $\delta = 7.37$ (s, 1H), 7.31 – 7.23 (m, 2H), 7.17 (ddd, J = 5.4, 2.9, 1.7 Hz, 1H), 6.28 (d, J = 0.9 Hz, 2H), 4.67 (s, 2H), 1.10 (s, 9H).



(E)-1-(3,3-Dimethylbut-1-en-1-yl)-3-methoxybenzene (14a)

Yellow oil, 16.3 mg (86% yield); ¹H NMR (300 MHz, CDCl₃): $\delta = 7.19$ (t, J = 7.9 Hz, 1H), 6.99 – 6.89 (m, 1H), 6.92 – 6.85 (m, 1H), 6.78 – 6.68 (m, 1H), 6.24 (d, J = 0.8 Hz, 2H), 3.80 (s, 3H), 1.10 (s, 9H).



(E)-4-(3,3-Dimethylbut-1-en-1-yl)phenyl acetate (15a)

Yellow oil, 21.2 mg (>96% yield); ¹H NMR (300 MHz, CDCl₃): δ = 7.34 (d, *J* = 8.6 Hz, 2H), 6.99 (d, *J* = 8.6 Hz, 2H), 6.31 – 6.12 (m, 2H), 2.27 (s, 3H), 1.09 (s, 9H).



(E)-2-(4-(3,3-Dimethylbut-1-en-1-yl)phenyl)-4,4,5,5-tetramethyl-1,3,2-dioxaborolane (16a)

Yellow oil, 18.3 mg (64% yield); ¹H NMR (300 MHz, CDCl₃): $\delta = 7.77 - 7.64$ (m, 2H), 7.38 - 7.31 (m, 2H), 6.29 (d, J = 1.0 Hz, 2H), 1.32 (s, 12H), 1.10 (s, 9H)

(E)-(4-(3,3-Dimethylbut-1-en-1-yl)phenyl)(methyl)sulfane (17a)

Yellow solid, 20.9 mg (>96% yield); ¹H NMR (300 MHz, CDCl₃): $\delta = 7.31 - 7.22$ (m, 2H), 7.21 - 7.13 (m, 2H), 6.28 - 6.14 (m, 2H), 2.46 (s, 3H), 1.09 (s, 9H).



(E)-2-(3,3-Dimethylbut-1-en-1-yl)naphthalene (18a)

White solid, 13.5 mg (64% yield, 1:2 ratio of stereoisomers); ¹H NMR (300 MHz, CDCl₃, mixture of isomers, calibrated to the minor isomer): $\delta = 7.85 - 7.67$ (m, 10H), 7.66 - 7.57 (m, 3H), 7.54 - 7.37 (m, 6H), 7.31 (dd, J = 8.4, 1.7 Hz, 2H), 6.53 (d, J = 12.6 Hz, 2H), 6.50 - 6.32 (m, 2H), 5.68 (d, J = 12.6 Hz, 2H), 1.15 (s, 9H), 0.99 (s, 18H).



(3,3-dimethylbut-1-ene-1,1-diyl)dibenzene (19a)

Colorless oil, 19.7 mg (83% yield); ¹H NMR (300 MHz, CDCl₃): $\delta = 7.37 - 7.26$ (m, 5H), 7.21 - 7.14 (m, 5H), 6.07 (s, 1H), 0.95 (s, 9H)



(E)-9-(3,3-Dimethylbut-1-en-1-yl)-9H-carbazole (20a)

Yellow solid, 18.8 mg (75% yield); ¹H NMR (300 MHz, CDCl₃): $\delta = 8.12 - 8.02$ (m, 2H), 7.58 - 7.53 (m, 2H), 7.53 - 7.36 (m, 2H), 7.30 - 7.20 (m, 2H), 6.86 (dd, J = 14.5, 1.8 Hz, 1H), 6.19 (dd, J = 14.4, 2.0 Hz, 1H), 1.27 (d, J = 1.9 Hz, 9H).

(E)-4,4-Dimethyl-1-morpholinopent-2-en-1-one (21a)

Yellow solid, 10.6 mg (54% yield); ¹H NMR (600 MHz, CDCl₃, mixture of rotamers): δ = 7.06 (d, *J* = 15.3 Hz, 1H), 6.22 (d, *J* = 15.3 Hz, 1H), 3.90 – 3.66 (m, 8H), 1.23 (s, 9H); ¹³C NMR (150 MHz, CDCl₃, mixture of rotamers): δ = 166.2, 157.1, 114.3, 33.8, 28.8 (The morpholine-derived carbon signals did not appear due to the formation of rotamers); HRMS-ESI (m/z) [M+Na]⁺ calcd for C₁₁H₁₉NO₂Na, 220.1308; found: 220.1310.

(E)-1-(3,3-Dimethylbut-1-en-1-yl)pyrrolidin-2-one (22a)

Colorless oil, 11.3 mg (68% yield); ¹H NMR (300 MHz, CDCl₃): $\delta = 6.83$ (d, J = 14.8 Hz, 1H), 4.97 (d, J = 14.8 Hz, 1H), 3.56 - 3.40 (m, 2H), 2.52 - 2.41 (m, 2H), 2.14 - 1.98 (m, 2H), 1.04 (s, 9H).



1,4-Bis((E)-3,3-dimethylbut-1-en-1-yl)benzene (23a)

Colorless oil, 18.6 mg (77% yield, 2:1 ratio E/Z); ¹H NMR (400 MHz, CDCl₃): $\delta = 7.28 - 7.23$ (m, 4H), 7.14 - 7.07 (m, 2H), 6.36 (d, J = 12.6 Hz, 1H), 6.27 - 6.22 (m, 4H), 5.57 (d, J = 12.6 Hz, 1H), 1.10 (s, 18H), 0.98 (s, 9H).



(8R,9S,13S,14S)-3-((*E*)-3,3-Dimethylbut-1-en-1-yl)-13-methyl-6,7,8,9,11,12,13,14,15,16-decahydro-17H-cyclopenta[a]phenanthren-17-one (24a)

Colorless oil, 23.6 mg (70% yield); ¹H NMR (400 MHz, CDCl₃): δ = 7.30 – 7.22 (m, 1H), 7.22 – 7.16 (m, 1H), 7.13 (s, 1H), 6.34 – 6.18 (m, 2H), 2.93 (dd, *J* = 9.1, 4.2 Hz, 2H), 2.53 (dd, *J* = 18.8, 8.7 Hz, 1H), 2.44 (dd, *J* = 9.9, 4.7 Hz, 1H), 2.32 (dt, *J* = 14.3, 7.3 Hz, 1H), 2.25 – 1.92 (m, 4H), 1.73 – 1.41 (m, 6H), 1.14 (s, 9H), 0.94 (s, 3H); ¹³C NMR (100 MHz, CDCl₃): δ = 220.9, 141.4, 138.4, 136.5, 135.7, 126.6, 125.5, 124.2, 123.5, 50.5, 48.0, 44.4, 38.3, 35.9, 33.3, 31.6, 29.6, 29.4, 26.6, 25.8, 21.6, 13.9; HRMS-ESI (m/z) [M+Na]⁺ calcd for C₂₄H₃₂ONa, 359.2345; found: 359.2348.



(88,98,10R,13R,148,17R)-3-(4-Methoxystyryl)-10,13-dimethyl-17-((R)-6-methylheptan-2-yl)-2,3,4,7,8,9,10,11,12,13,14,15,16,17-tetradecahydro-1H-cyclopenta[a]phenanthrene (3ac)

White solid, 41.5 mg (80% yield, 3:4 ratio of stereoisomers); ¹H NMR (300 MHz, CDCl₃, shown only for the major isomer): $\delta = 7.29 - 7.22$ (m, 2H), 6.81 (d, J = 8.4 Hz, 2H), 6.33 (d, J = 16.0 Hz, 1H), 6.18 (dd, J = 16.0, 7.1 Hz, 1H), 5.32 (t, J = 4.9 Hz, 2H), 3.78 (s, 3H), 2.60 (d, J = 10.9 Hz, 2H), 2.15 – 1.79 (m, 4H), 1.70 – 0.79 (m, 35H), 0.68 (s, 3H).



1-(2-((2S,5R)-2-isopropyl-5-methylcyclohexyl)vinyl)-4-methoxybenzene (3ad)

White solid, 18.4 mg (68% yield, 1:2 ratio of stereoisomers, calibrated to the minor isomer); ¹H NMR (400 MHz, CDCl₃): δ = 7.39 – 7.29 (m, 6H), 6.94 – 6.83 (m, 6H), 6.42 – 6.28 (m, 4H), 5.96 – 5.85 (m, 2H), 3.84 (d, *J* = 1.3 Hz, 9H), 2.81 – 2.70 (m, 1H), 2.16 – 2.02 (m, 2H), 1.99 – 1.89 (m, 2H), 1.87 – 1.61 (m, 10H), 1.52 – 1.23 (m, 6H), 1.18 – 0.86 (m, 30H), 0.81 – 0.76 (m, 6H); ¹³C NMR (100 MHz, CDCl₃, mixture of isomers): δ = 158.7, 158.6, 133.9, 131.1, 130.9, 129.5, 129.4, 128.0, 127.1, 127.0, 113.9, 113.9, 55.3, 47.6, 47.5, 45.2, 43.3, 43.2, 40.7, 35.9, 35.2, 32.5, 30.5, 28.3, 27.0, 25.7, 24.3, 22.9, 22.7, 21.5, 21.1, 20.8, 15.5; HRMS-EI (m/z) [M]⁺ calcd for



(R)-6-((E)-3,3-Dimethylbut-1-en-1-yl)-2,8-dimethyl-2-((4R,8R)-4,8,12-trimethyltridecyl)chromane (25a)

Yellow oil, 30.8 mg (66% yield); ¹H NMR (300 MHz, CDCl₃): $\delta = 6.98$ (d, J = 2.2 Hz, 1H), 6.89 (d, J = 2.3 Hz, 1H), 6.17 (d, J = 16.2 Hz, 1H), 6.12 – 5.99 (m, 1H), 2.71 (t, J = 6.8 Hz, 2H), 2.14 (s, 3H), 1.87 – 1.63 (m, 2H), 1.64 – 1.41 (m, 4H), 1.41 – 0.99 (m, 26H), 0.90 – 0.75 (m, 15H); ¹³C NMR (150 MHz, CDCl₃): $\delta = 151.3$, 138.8, 128.8, 126.3, 125.9, 124.5, 124.2, 120.4, 76.0, 39.9, 39.4, 37.4, 37.3, 33.1, 32.8, 32.7, 31.4, 29.7, 28.0, 24.8, 24.4, 24.2, 22.7, 22.6, 22.3, 21.0, 19.7, 19.6, 16.1; HRMS-ESI (m/z) [M+Na]⁺ calcd for C₃₃H₅₆ONa, 469.4404; found: 469.4408.



(E)-5-(4-Methoxyphenyl)pent-4-en-1-yl 5-(2,5-dimethylphenoxy)-2,2-dimethylpentanoate (3ae)

Yellow oil, 28.2 mg (75% yield); ¹H NMR (600 MHz, CDCl₃): δ = 7.29 – 7.21 (m, 2H), 7.00 (dd, *J* = 7.5, 2.7 Hz, 1H), 6.86 – 6.80 (m, 2H), 6.65 (d, *J* = 7.5 Hz, 1H), 6.61 (d, *J* = 3.1 Hz, 1H), 6.34 (d, *J* = 15.7 Hz, 1H), 6.05 (dt, *J* = 15.8, 7.0 Hz, 1H), 4.11 (t, *J* = 6.5 Hz, 2H), 3.92 (t, *J* = 5.6 Hz, 2H), 3.79 (s, 3H), 2.30 (s, 3H), 2.29 – 2.25 (m, 2H), 2.18 (s, 3H), 1.83 – 1.77 (m, 2H), 1.75 – 1.72 (m, 4H), 1.23 (s, 6H). ¹³C NMR (150 MHz, CDCl₃): δ = 177.8, 158.8, 156.9, 136.4, 130.3, 130.3, 130.1, 127.1, 127.1, 123.6, 120.7, 113.9, 111.9, 67.9, 63.8, 55.3, 42.1, 37.1, 29.4, 28.5, 25.2, 21.4, 15.8; HRMS-ESI (m/z) [M+Na]⁺ calcd for C₂₇H₃₆O₄Na, 447.2506; found: 447.2507.



(E)-5-(4-Methoxyphenyl)pent-4-en-1-yl (S)-2-(6-methoxynaphthalen-2-yl)propanoate (3af)

Yellow oil, 28.2 mg (70% yield, 1:1.3 ratio of stereoisomers); ¹H NMR (600 MHz, CDCl₃, only the major isomer indicated): $\delta = 8.16 - 8.05$ (m, 3H), 7.82 (dd, J = 26.1, 8.4 Hz, 1H), 7.62 - 7.49 (m, 4H), 7.23 (ddd, J = 17.6, 8.7, 1.8 Hz, 2H), 6.56 (d, J = 15.8 Hz, 1H), 6.36 (dt, J = 15.0, 6.9 Hz, 1H), 4.53 (q, J = 6.9 Hz, 2H), 4.32 (s, 3H), 4.28 (q, J = 7.2 Hz, 1H), 4.20 (s, 3H), 2.54 (q, J = 7.3 Hz, 2H), 2.14 (p, J = 7.2 Hz, 2H), 2.00 (d, J = 7.2 Hz, 3H); ¹³C NMR (150 MHz, CDCl₃, mixture of stereoisomers, overlapping peaks not repeated): $\delta = 175.2, 158.1, 134.1, 130.5, 130.3, 129.7, 129.5, 129.4, 127.5, 126.7, 126.4, 119.4, 114.3, 114.0, 106.0, 64.6, 55.7, 46.0, 29.6, 29.3, 28.8, 25.4, 18.9.; HRMS-ESI (m/z) [M+Na]⁺ calcd for C₂₆H₂₈O₄Na, 427.1880; found: 427.1882.$

(E)-4-(3,3-Dimethylbut-1-en-1-yl)-2-methoxyphenyl acetate (26a)

Yellow oil, 23.7 mg (95% yield); ¹H NMR (600 MHz, CDCl₃): $\delta = 6.97 - 6.86$ (m, 3H), 6.26 (d, J = 16.1 Hz, 1H), 6.19 (d, J = 16.1 Hz, 1H), 3.84 (s, 3H), 2.30 (s, 3H), 1.11 (s, 9H); ¹³C NMR (150 MHz, CDCl₃): $\delta = 169.2$, 151.0, 142.2, 138.5, 137.2, 124.0, 122.6, 118.5, 109.7, 55.8, 55.8, 33.4, 29.6, 20.7, 20.7; HRMS-ESI (m/z)



(E)-5-(4-Methoxyphenyl)pent-4-en-1-yl 5-((3aS,4S,6aR)-2-oxohexahydro-1H-thieno[3,4-d]imidazol-4-yl)pentanoate (3ag)

Yellow oil, 34.0 mg (81% yield); ¹H NMR (600 MHz, CDCl₃) δ 7.25 (d, J = 8.5 Hz, 2H), 6.82 (d, J = 8.4 Hz, 2H), 6.33 (d, J = 15.8 Hz, 1H), 6.03 (dt, J = 15.8, 6.9 Hz, 1H), 5.82 (brs, 1H), 5.43 (brs, 1H), 4.47 (t, J = 6.3 Hz, 1H), 4.34 – 4.23 (m, 1H), 4.10 (t, J = 6.6 Hz, 2H), 3.78 (s, 3H), 3.12 (brs, 1H), 2.88 (dd, J = 13.1, 4.8 Hz, 1H), 2.72 (d, J = 12.8 Hz, 1H), 2.32 (t, J = 7.5 Hz, 2H), 2.24 (q, J = 7.4 Hz, 2H), 1.78 (p, J = 6.8 Hz, 2H), 1.69 – 1.59 (m, 4H), 1.46 – 1.38 (m, 2H); ¹³C NMR (150 MHz, CDCl₃) δ 173.9, 164.0, 158.9, 130.5, 130.1, 127.2, 127.2, 114.0, 64.0, 62.1, 60.3, 55.5, 55.4, 40.7, 34.1, 29.5, 28.5, 28.5, 28.4, 24.9. HRMS-ESI (m/z) [M+Na]⁺ calcd for C₂₂H₃₀N₂O₄SNa, 441.1818; found: 441.1820.



1-(Cyclohexylidenemethyl)-4-methoxybenzene (3ah)

Yellow oil, 12.2 mg (60% yield); ¹H NMR (600 MHz, CDCl₃): δ = 7.12 (d, *J* = 8.3 Hz, 2H), 6.84 (d, *J* = 8.4 Hz, 2H), 6.15 (s, 1H), 3.79 (s, 3H), 2.34 (t, *J* = 6.1 Hz, 2H), 2.22 (t, *J* = 5.9 Hz, 2H), 1.63 – 1.52 (m, 6H).



4-(4-Methoxybenzylidene)tetrahydro-2H-pyran (3ai)

Colorless oil, 14.4 mg (70% yield); ¹H NMR (400 MHz, CDCl₃): δ = 7.19 – 7.12 (m, 2H), 6.89 (d, *J* = 8.8 Hz, 2H), 6.29 (s, 1H), 3.83 (s, 3H), 3.82 – 3.78 (m, 2H), 3.68 (t, *J* = 5.6 Hz, 2H), 2.57 – 2.51 (m, 2H), 2.42 – 2.37 (m, 2H); ¹³C NMR (150 MHz, CDCl₃) δ 158.0, 136.4, 130.0, 127.2, 123.3, 113.6, 69.5, 68.6, 55.3, 37.2, 30.6; HRMS-ESI (m/z) [M]⁺ calcd for C₁₃H₁₆O₂, 204.1150; found: 204.1152.



tert-Butyl 4-(4-methoxybenzylidene)piperidine-1-carboxylate (3aj)

Yellow oil, 26.1 mg (86% yield); ¹H NMR (300 MHz, CDCl₃): δ = 7.17 – 7.03 (m, 2H), 6.84 (d, *J* = 8.7 Hz, 2H), 6.27 (s, 1H), 3.78 (s, 3H), 3.47 (t, *J* = 6.0 Hz, 2H), 3.37 (t, *J* = 5.9 Hz, 2H), 2.43 (t, *J* = 5.9 Hz, 2H), 2.29 (t, *J* = 5.9 Hz, 2H), 1.45 (s, 9H).



4-(4-Methoxybenzylidene)-1-phenylpiperidine (3ak)

White solid, 20.9 mg (75% yield); ¹H NMR (300 MHz, CDCl₃): δ = 7.39 – 7.26 (m, 2H), 7.24 – 7.16 (m, 2H), 7.07 – 6.96 (br, 2H), 6.96 – 6.90 (m, 3H), 6.35 (s, 1H), 3.85 (s, 3H), 3.38 (t, *J* = 5.8 Hz, 2H), 3.27 (t, *J* = 5.8 Hz, 2H), 2.70 (br, 2H), 2.55 (br, 2H); ¹³C NMR (150 MHz, CDCl₃, mixture of rotamers) δ 158.0, 151.5, 137.8, 130.2, 130.1, 129.2, 123.3, 120.2, 116.6, 113.6, 55.3, 51.5, 50.7, 35.9, 28.9; HRMS-ESI (m/z) [M]⁺ calcd for C₁₉H₂₁NO, 279.1623; found: 279.1622.



Methyl 4-(4-(4-(4-methoxybenzylidene)piperidin-1-yl)phenyl)butanoate (3al)

Yellow oil, 28.7 mg (76% yield); ¹H NMR (400 MHz, CDCl₃): δ = 7.18 – 7.10 (m, 2H), 7.06 (d, *J* = 8.5 Hz, 2H), 6.86 (d, *J* = 8.7 Hz, 4H), 6.29 (s, 1H), 3.80 (s, 3H), 3.65 (s, 3H), 3.27 (t, *J* = 5.8 Hz, 2H), 3.16 (t, *J* = 5.8 Hz, 2H), 2.63 (t, *J* = 5.7 Hz, 2H), 2.56 (t, *J* = 7.5 Hz, 2H), 2.54 – 2.43 (m, 2H), 2.31 (t, *J* = 7.5 Hz, 2H), 2.00 – 1.82 (m, 2H); ¹³C NMR (150 MHz, CDCl₃, mixture of rotamers, due to severe peak broadening, only tentatively assigned peaks are indicated): δ = 174.1, 158.0, 149.7, 138.2, 132.7, 130.0, 128.6, 123.2, 116.9, 113.6, 112.2, 55.3, 51.9, 51.5, 51.1, 36.0, 34.2, 33.1, 28.9, 26.6; HRMS-ESI (m/z) [M+H]⁺ calcd for C₂₄H₃₀NO₃, 380.2220; found: 380.2223.

5. *E*/*Z* isomerization experiment

An isolated mixture of **18a** (E/Z = 1:2) was subjected to the original reaction condition to probe the origin of the formation of the Z product. The E/Z ratio and product recovery was measured by ¹H NMR using nitrobenzene as an internal standard.



Supplementary Fig. 1. *E*/*Z* isomerization experiment.

By comparing the NMR spectra before and after irradiation, we confirmed that isomerization occurs after product formation. Similar visible-light mediated styrene E/Z isomerization reactions have been reported.²⁶⁻²⁹

6. Radical trapping/clock experiments

TEMPO trapping experiment



To a 4 mL vial equipped with a stirrer-bar were added Pd(PPh₃)₄ (5.8 mg, 0.0050 mmol, 0.050 equiv), K₂CO₃ (27.6 mg, 0.20 mmol, 2.0 equiv), **1a** (13.3 μ L, 0.10 mmol, 1.0 equiv), **2a** (16.5 μ L, 0.15 mmol, 1.5 equiv), (2,2,6,6-tetramethylpiperidin-1-yl)oxyl (TEMPO, 15.6 mg, 0.10 mmol, 1.0 equiv) and *N*,*N*-dimethylacetamide (1.0 mL). The resulting mixture was stirred for 24 h under 40 W blue LED irradiation with fan cooling (~30 °C). The resulting residue was analyzed by gas chromatography after the addition of dodecane (10.0 μ L) as an internal standard and revealed no presence of **3a**.

Radical Clock Experiment



To a 4 mL vial equipped with a stirrer-bar were added Pd(PPh₃)₄ (5.8 mg, 0.0050 mmol, 0.050 equiv), K₂CO₃ (27.6 mg, 0.20 mmol, 2.0 equiv), **1a** (13.3 μ L, 0.10 mmol, 1.0 equiv), **2am** (13.9 μ L, 0.15 mmol, 1.5 equiv) and *N*,*N*-dimethylacetamide (1.0 mL). The resulting mixture was stirred for 24 h under 40 W blue LED irradiation with fan cooling (~30 °C). The resulting residue was analyzed by ¹H NMR after the addition of nitrobenzene (10.0 μ L) as an internal standard. No **3am** was detected and **3an** was generated in 53% yield as the sole product. A crude NMR of the reaction is presented in Supplementary Fig. 2 and all integrated peaks shown below corresponds to the product signals except for the internal standard.



Supplementary Fig. 2. Crude NMR of the radical clock experiment.

7. Stern-Volmer quenching experiments

All emission spectra of the samples were collected under an argon atmosphere. A solution of $Pd(PPh_3)_4$ (1.0 mM) in DMA was prepared in a cuvette. *tert*-Butyl chloride (**2a**) was used as quenchers. The prepared solution was excited at 420 nm and the emission intensity at 650 nm was observed. The Stern-Volmer quenching experiments resulted in positive dependence between I_0/I and the concentration of quenchers. The results indicate that alkyl chlorides quench the excited state of Pd(PPh_3)_4, where it engages in a single-electron transfer (SET) event with the photoexcited Pd(0) complex.



Supplementary Fig. 3. Stern-Volmer quenching experiment.

8. Kinetic isotope effect measurements

Preparation of 3a-β-d₂



To a stirred solution of 4-methoxyphenylacetylene (132 mg, 1.0 mmol, 1.0 equiv) in THF (5.0 mL) at 0 °C was added *n*-BuLi (0.70 mL, 1.6 M in hexanes, 1.1 mmol, 1.1 equiv) dropwise. The reaction mixture was quenched with D_2O (0.20 mL) after 30 minutes and further stirred at room temperature for 5 minutes. The resulting solution was dried (MgSO₄), filtered and concentrated under reduced pressure to yield 4-methoxyphenylacetylene- d_1 (132 mg, 100% yield) as a colorless oil, which was used directly without further purification. The deuterium incorporation was determined to be 98% by ¹H NMR. The NMR spectrum was identical with the starting material with the acetylene proton being absent.



To a stirred solution of 4-methoxyphenylacetylene- d_1 (132 mg, 1.0 mmol, 1.0 equiv) in CH₂Cl₂ (5.0 mL) at 0 °C was added Cp₂ZrHCl (273 mg, 1.1 mmol, 1.1 equiv). The resulting mixture was warmed to room temperature and stirred for 2 h before it was quenched with D₂O (0.20 mL) and stirred for further 30 min. The resulting solution was dried (MgSO₄), filtered and concentrated under reduced pressure. The resulting residue was purified by column chromatography (silica gel, hexanes:EtOAc gradient elution) to yield **1a-\beta-d_2** (130 mg, 99% yield) as a colorless oil. The deuterium incorporation was determined to be 95% by ¹H NMR. The NMR spectrum was identical with the starting material **1a** with the olefinic proton being absent. ¹H NMR (300 MHz, CDCl₃): δ = 7.39 – 7.29 (m, 2H), 6.92 – 6.80 (m, 2H), 6.63 (s, 1H), 3.79 (s, 3H). *Residual H signal: 5.57 (d, *J* = 17.6 Hz, 0.06 H), 5.09 (d, *J* = 10.9 Hz, 0.05 H).

Preparation of 2n-d₁



To a stirred solution of 2-adamantanone (285 mg, 1.9 mmol, 1.0 equiv) in MeOH (10 mL) at room temperature was added NaBD₄ (88 mg, 2.1 mmol, 1.1 equiv). The reaction mixture stirred for 10 min before it was quenched with NH₄Cl (sat. aq., 10 mL). The resulting mixture was extracted with CH₂Cl₂ (10 mL \times 3), dried (anhydrous Na₂SO₄), filtered, and concentrated under reduced pressure. The resulting material was used directly for the next step.



To a stirred solution of thionyl chloride (2.5 mL) was added the crude mixture of 2-adamantanol (291 mg, 1.9 mmol, 1.0 equiv). The reaction mixture was stirred for 3 h at 75 °C before it was quenched with MeOH (5 mL) and NaHCO₃ (sat. aq., 10 mL) in a cooling bath. The resulting mixture was extracted with CH₂Cl₂ (10 mL × 3), dried (anhydrous Na₂SO₄), filtered, and concentrated under reduced pressure. The resulting residue was purified by column chromatography (silica gel, hexanes) to afford the desired product **2n**-*d*₁ (260 mg, 80% over two steps). The deuterium incorporation was determined to be >99% by ¹H NMR. ¹H NMR (600 MHz, CDCl₃) δ = 2.26 (d, J = 12.2 Hz, 2H), 2.07 (brs, 2H), 1.97 – 1.92 (m, 2H), 1.88 – 1.84 (m, 2H), 1.81 – 1.77 (m, 2H), 1.75 (m, 2H), 1.59 – 1.55 (m, 2H).

Preparation of 3a-β-d₁



To a stirred solution of 4-methoxyphenylacetylene (132 mg, 1.0 mmol, 1.0 equiv) in THF (5.0 mL) at 0 °C was added n-BuLi (0.70 mL, 1.6 M in hexanes, 1.1 mmol, 1.1 equiv) dropwise. The reaction mixture was quenched with D_2O (0.20 mL) after 30 minutes and further stirred at room temperature for 5 minutes. The resulting solution was dried (MgSO₄), filtered and concentrated under reduced pressure to yield 4-methoxyphenylacetylene- d_1 (132 mg, 100% yield) as a colorless oil, which was used directly without further purification. The deuterium incorporation was determined to be 98% by ¹H NMR. The NMR spectrum was identical with the starting material with the acetylene proton being absent.



To a stirred solution of 4-methoxyphenylacetylene- d_1 (132 mg, 1.0 mmol, 1.0 equiv) in CH₂Cl₂ (5.0 mL) at 0 °C was added Cp₂ZrHCl (273 mg, 1.1 mmol, 1.1 equiv). The resulting mixture was warmed to room temperature and stirred for 2 h before it was quenched with H₂O (0.20 mL) and stirred for further 30 min. The resulting solution was dried (MgSO₄), filtered and concentrated under reduced pressure. The resulting residue was purified by column chromatography (silica gel, hexanes:EtOAc gradient elution) to yield **1a-β-d**₁ (130 mg, 99% yield) as a colorless oil. The deuterium incorporation was determined to be 98% by ¹H NMR. The NMR spectrum was identical with the starting material **1a** with the olefinic proton being absent. ¹H NMR (300 MHz, CDCl₃): δ = 7.37 – 7.29 (m, 2H), 6.88 – 6.81 (m, 2H), 6.63 (dt, *J* = 10.9, 2.6 Hz, 1H), 5.09 (d, *J* = 10.9 Hz, 1H), 3.79 (s, 4H). *Residual H signal: 5.59 (dd, *J* = 17.7 Hz, 1.1 Hz, 0.02 H).

Preparation of 27-d2



To a stirred solution of 4'-methoxypropiophenone (164 mg, 1.0 mmol, 1.0 equiv) in D₂O (1.5 mL) and dioxane (1.5 mL) at room temperature was added pyrrolidine (7.1 mg, 0.10 mmol, 0.10 equiv). The reaction mixture was heated to 80 °C and stirred for 16 h. The reaction mixture was quenched with water (5 mL), extracted with Et₂O (10 mL), washed with brine (10 mL), dried (anhydrous Na₂SO₄), filtered, and concentrated under reduced pressure to afford 1-(4-methoxyphenyl)propan-1-one-2,2- d_2 (160 mg, 99% yield) as a colorless oil which was used directly for the next step. The deuterium incorporation was determined to be 98.5% by ¹H NMR. ¹H NMR (300 MHz, CDCl₃): $\delta = 7.99 - 7.88$ (m, 2H), 6.97 - 6.81 (m, 2H), 3.84 (d, J = 0.7 Hz, 3H), 1.17 (s, 3H). Residual proton signal: 2.90 (s, 0.03H).



To a stirred solution of 1-(4-methoxyphenyl)propan-1-one-2,2- d_2 (160 mg, 0.99 mmol, 1.0 equiv) in MeOH (5.0 mL) at room temperature was added NaBH₄ (41.6 mg, 1.1 mmol, 1.1 equiv). The reaction mixture stirred for 10 min before it was quenched with NH₄Cl (sat. aq., 10 mL). The resulting mixture was extracted with CH₂Cl₂ (10 mL × 3), dried (anhydrous Na₂SO₄), filtered, and concentrated under reduced pressure. The resulting residue was purified by column chromatography (silica gel hexanes:EtOAc gradient elution) to afford 1-(4-methoxyphenyl)propan-2,2- d_2 -1-ol (145 mg, 86% yield) as a colorless oil. ¹H NMR (300 MHz, CDCl₃): δ = 7.24 (d, *J* = 8.8 Hz, 2H), 6.86 (d, *J* = 8.7 Hz, 2H), 4.51 (s, 1H), 3.78 (s, 3H).



To a stirred solution of 1-(4-methoxyphenyl)propan-1-one-2,2- d_2 (145 mg, 0.86 mmol, 1.0 equiv) in MeOH (3.0 mL) at -78 °C was added SOCl₂ (75 µL, 1.03 mmol, 1.2 equiv) dropwise. The reaction mixture stirred for 15 min before it was quenched with water (10 mL). The resulting mixture was extracted with CH₂Cl₂ (10 mL × 3), dried (anhydrous Na₂SO₄), filtered, and concentrated under reduced pressure. The resulting residue was pure enough and unstable upon silica gel column chromatography and was used directly for the kinetic study. ¹H NMR (300 MHz, CDCl₃): δ = 7.31 – 7.24 (m, 2H), 6.89 – 6.82 (m, 2H), 4.74 (s, 1H), 3.79 (s, 3H).

KIE measurement with $1a-\beta-d_2$



To two 4 mL vial equipped with a stirrer-bar were added Pd(PPh₃)₄ (5.8 mg, 0.0050 mmol, 0.050 equiv), K₂CO₃ (27.6 mg, 0.20 mmol, 2.0 equiv), dodecane (10.0 μ L), **2a** (16.5 μ L, 0.15 mmol, 1.5 equiv), **1a** or **1a-** β -*d*₂ respectively (13.3 μ L, 0.10 mmol, 1.0 equiv) and *N*,*N*-dimethylacetamide (2.0 mL). The resulting mixture was kept stirring under 40 W blue LED irradiation and aliquots of both reaction mixtures were taken every 10 minutes. They were analyzed by gas chromatography to determine the individual reaction rates of **1a** and **1a-** β -*d*₂.





Supplementary Fig. 4. KIE measurement with 1a-β-d₂.

The KIE was determined to be 1.0.

KIE measurement with $3a-\alpha-d_1$



Substrate $1a-a-d_1$ was prepared following literature procedures.³⁰ To two 4 mL vial equipped with a stirrer-bar were added Pd(PPh₃)₄ (5.8 mg, 0.0050 mmol, 0.050 equiv), K₂CO₃ (27.6 mg, 0.20 mmol, 2.0 equiv), dodecane (10.0 µL), **2a** (16.5 µL, 0.15 mmol, 1.5 equiv), **1a** or **1a-a-d_1** respectively (13.3 µL, 0.10 mmol, 1.0 equiv) and *N*,*N*-dimethylacetamide (2.0 mL). The resulting mixture was kept stirring under 40 W blue LED irradiation and aliquots of both reaction mixtures were taken every 10 minutes. They were analyzed by gas chromatography to determine the individual reaction rates of **1a** and **1a-a-d_1**.





Supplementary Fig. 5. KIE measurement with $1a - \alpha - d_1$.

The KIE was determined to be 1.0.

KIE measurement with 2n-d₁



To two 4 mL vial equipped with a stirrer-bar were added Pd(PPh₃)₄ (5.8 mg, 0.0050 mmol, 0.050 equiv), K₂CO₃ (27.6 mg, 0.20 mmol, 2.0 equiv), dodecane (10.0 μ L), **2n** or **2n**-*d*₁, respectively, (25.6 mg, 0.15 mmol, 1.5 equiv), **1a** (13.3 μ L, 0.10 mmol, 1.0 equiv) and *N*,*N*-dimethylacetamide (2.0 mL). The resulting mixture was kept stirring under 40 W blue LED irradiation and aliquots of both reaction mixtures were taken every 10 minutes. They were analyzed by gas chromatography to determine the individual reaction rates of **1a** and **1a**- α -*d*₁.





Supplementary Fig. 6. KIE measurement with 2n-d1.

The KIE was determined to be 1.6.

Intramolecular competition experiment



To a 4 mL vial equipped with a stirrer-bar were added Pd(PPh₃)₄ (5.8 mg, 0.0050 mmol, 0.050 equiv), K₂CO₃ (27.6 mg, 0.20 mmol, 2.0 equiv), **1a-\beta-d₁** (13.3 µL, 0.10 mmol, 1.0 equiv), **2a** (16.5 µL, 0.15 mmol, 1.5 equiv) and *N*,*N*-dimethylacetamide (1.0 mL). The resulting mixture was stirred for 24 h under 40 W blue LED irradiation with fan cooling (~30 °C). The reaction mixture was added brine (10 mL), diluted with EtOAc or Et₂O (10 mL), washed with brine (10 mL), dried (anhydrous Na₂SO₄), filtered, and concentrated under reduced pressure. The resulting residue was analyzed by ¹H NMR to determine the ratio of **3a** and **3a-d₁** to be 0.45:0.55 by integrating the olefinic protons ([P_D]/[P_H] = 1.2).



Supplementary Fig. 7. Intramolecular competition experiment.

Elimination kinetic isotope effect experiment



To a 4 mL vial equipped with a stirrer-bar were added **27** and **27**-*d*₂ (9.3 mg each, 0.10 mmol, 1.0 equiv), K₂CO₃ (276 mg, 2.0 mmol, 20.0 equiv) and *N*,*N*-dimethylacetamide (1.0 mL). The resulting mixture was stirred for 1 h under 40W blue LED irradiation with fan cooling (~30 °C) to ensure identical physical conditions. The reaction mixture was added brine (10 mL), diluted with Et₂O (10 mL), washed with brine (10 mL), dried (anhydrous Na₂SO₄), filtered, and concentrated under reduced pressure. The resulting residue was analyzed by ¹H NMR to determine the ratio of **2am** and **2am**-*d*₁ to be 0.46:0.54 by integrating the olefinic protons ([P_D]/[P_H] = 1.2).



Supplementary Fig. 8. Elimination KIE measurement.

9. Elimination of benzyl chloride intermediate



To a 4 mL vial equipped with a stirrer-bar were added Pd(PPh₃)₄ (5.8 mg, 0.0050 mmol, 0.050 equiv), K₂CO₃ (27.6 mg, 0.20 mmol, 2.0 equiv), **28** (26.7 mg, 0.10 mmol, 1.0 equiv), and *N*,*N*-dimethylacetamide (1.0 mL). The resulting mixture was stirred for 24 h under 40 W blue LED irradiation with fan cooling (~30 °C). The reaction mixture was analyzed by ¹H NMR with nitrobenzene (10 μ L) as an internal standard to determine the yield (92%) of **3ap**. The crude NMR of the reaction mixture is shown in Supplementary Fig. 9.



Supplementary Fig. 9. Crude NMR of the elimination experiment.

10. Control experiment with tert-butyl N-(acyloxy)phthalimide



To a 4 mL vial equipped with a stirrer-bar were added Pd(PPh₃)₄ (5.8 mg, 0.0050 mmol, 0.050 equiv), K₂CO₃ (27.6 mg, 0.20 mmol, 2.0 equiv), **1a** (13.3 μ L, 0.10 mmol, 1.0 equiv), **2al** (37.1 mg, 0.15 mmol, 1.5 equiv), LiCl if appropriate (4.2 mg, 0.10 mmol, 1.0 equiv), and *N*,*N*-dimethylacetamide (1.0 mL). The resulting mixture was stirred for 24 h under 40 W blue LED irradiation with fan cooling (~30 °C). The reaction mixture was analyzed by gas chromatography using dodecane as an internal standard to determine the yield of product **3a**. The results are shown below.

Supplementary Table 1. Control experiment with 2al

Entry	LiCl	Yield
1	_	13%
2	1.0 equiv	22%

11. Computational details

General information

All calculations were carried out using DFT as implemented in the Gaussian 09^{31} program packages. Gas phase geometry optimizations were conducted with the B3LYP³² hybrid functional including Grimme's D3 dispersion correction³³ and the 6-31G** basis set and LanL2DZ^{34,35} basis set for Pd. The energies of the optimized structures were reevaluated by additional single point calculations using B3LYP hybrid functional including Grimme's D3 dispersion correction and the 6-311++G** basis set and the SDD basis set for Pd. The integral equation formalism variant of the Polarizable Continuum Model (IEFPCM) was employed as implemented to account for the solvation effects for DMA ($\varepsilon = 37.781$). Analytical vibrational frequencies within the harmonic approximation were computed with the 6-31G**/LanL2DZ(Pd) basis set to confirm proper convergence to well-defined minima (no imaginary frequency) or saddle points (one and only one imaginary frequency) on the potential energy surface. All thermal corrections from the vibrational frequency calculations were performed at 25 °C (298.15 K).

Time-dependent density functional theory (TD-DFT) calculations were carried out as implemented in the Gaussian 09 program package using the hybrid exchange/correlation CAM-B3LYP³⁶ functional with long-range corrections and the 6-311++G**/SDD(Pd) basis set. The solvation effects for DMA ($\varepsilon = 37.781$) were accounted for using the integral equation formalism variant of the Polarizable Continuum Model (IEFPCM) as implemented.

Computation of redox potentials

The standard reduction potentials, E^{o}_{red} were obtained from the electron attachment energy in the solution phase, $\Delta G^{EA}(sol)$, and subsequent application of the following relationships, where n is the number of electrons, E is the absolute potential and F is the Faraday constant. $\Delta G^{EA}(sol)$ was computed by subtracting the Gibbs free energies of the oxidized species from those of the reduced species.

> $\Delta G^{EA}(sol) = -nFE$ E^o_{red} (V vs. N.H.E.) = -E - E^o (N.H.E.) = -E - 4.43 V

Here, we employed the absolute potential that was measured to be 4.43 V for the normal hydrogen electrode.³⁷ The species are denoted as "**I-OMe-rad**", "**I-OMe-cat**" and their analogues according to the substituent on the aryl group. The reduction potentials of Pd(PPh₃)₃Cl and Pd(PPh₃)₂Cl₂ were also computed accordingly.



Supplementary Fig. 10. Computed reduction potentials of Pd complexes.

Kinetic isotope effect computations

Kinetic isotope effect computations were conducted by using the (Iso=2) keyword on the hydrogen atom(s) of interest and re-performing the frequency calculations. The KIE value was calculated from the following reaction through intramolecular competition, H-TS and D-TS, and applying the Boltzmann distribution.



Supplementary Table 2. KIE computations

G (H-TS) (kcal/mol)	G (D-TS) (kcal/mol)	∆G (kcal/mol)	KIE
-1384623.614	-1384622.662	0.952	5.0

Time-dependent density functional theory computations

Time-dependent density functional theory (TD-DFT) computations were conducted on a model complex of **II** shown below. The molecular orbitals diagrams and TD-DFT results are summarized below.



State	Wavelength (nm)	f	Transitions
1 (4)	423.37	0.0787	H-3 → L (7.8%) H-2 → L (2.7%) H-1 → L (38.2%) H → L (35.8%)
2 (7)	392.54	0.0368	H-13 → L (8.7%) H-2 → L (56.7%) H-1 → L (18.0%) H → L (5.4%)



Supplementary Fig. 11. Time-dependent density functional theory computations.

Energy components of DFT-optimized structures

	E(SCF)/(Hartree)	Thermal Corr. to G (Hartree)	G(sol)/(kcal/mol)
	6-311++G**/SDD	6-31G**/LanL2DZ	
I-OMe-rad	-582.2024828	0.246983	-365182.3137
I-OMe-cat	-582.043615	0.250666	-365080.3116
I-OAc-rad	-695.5966132	0.252701	-436334.563
I-OAc-cat	-695.4228344	0.253819	-436224.8137
I-F-rad	-566.9106881	0.209082	-355610.3581
I-F-cat	-566.7356712	0.211211	-355499.1975
I-Cl-rad	-927.2670096	0.206688	-581738.6953
I-Cl-cat	-927.089249	0.209216	-581625.5626
I-CF ₃ -rad	-804.792516	0.216696	-504878.5682
I-CF ₃ -cat	-804.6095457	0.218972	-504762.3245
I-CHO-rad	-581.0109057	0.225376	-364448.1469
I-CHO-cat	-580.8199019	0.226123	-364327.8216
I-CN-rad	-559.9180952	0.214513	-351219.0351
I-CN-cat	-559.7262494	0.216342	-351097.5025
Pd(PPh ₃) ₃ Cl	-3697.953287	0.737484	-2320036.191
Pd(PPh ₃) ₃ Cl ⁻	-3698.082242	0.734904	-2320118.731
Pd(PPh ₃) ₂ Cl ₂	-3121.647	0.476436	-1958562.619
$Pd(PPh_3)_2Cl_2^-$	-3121.779546	0.476796	-1958645.567
H-TS	-2207.028208	0.48836	-1384623.614
D-TS	-2207.028208	0.489876	-1384622.662

12. ¹H and ¹³C NMR spectra



Supplementary Fig. 12. ¹H NMR spectrum of compound 2ae.



Supplementary Fig. 13. ¹³C NMR spectrum of compound 2ae.



Supplementary Fig. 14. ¹H NMR spectrum of compound 2af.



Supplementary Fig. 15. ¹³C NMR spectrum of compound 2af.


Supplementary Fig. 16. ¹H NMR spectrum of compound 2ag.



Supplementary Fig. 17. ¹³C NMR spectrum of compound 2ag.



Supplementary Fig. 18. ¹H NMR spectrum of compound 1w.



Supplementary Fig. 19. ¹³C NMR spectrum of compound 1w.



Supplementary Fig. 20. ¹H NMR spectrum of compound 3a.



Supplementary Fig. 21. ¹H NMR spectrum of compound 3b.



Supplementary Fig. 22. ¹H NMR spectrum of compound 3c.



Supplementary Fig. 23. ¹H NMR spectrum of compound 3d.



180 170 160 f1 (ppm)





Supplementary Fig. 25. ¹H NMR spectrum of compound e3.



180 170 160 150 140 f1 (ppm)

Supplementary Fig. 26. ¹³C NMR spectrum of compound 3e.



Supplementary Fig. 27. ¹H NMR spectrum of compound 3f.



Supplementary Fig. 28. ¹H NMR spectrum of compound 3g.



Supplementary Fig. 29. ¹³C NMR spectrum of compound 3g.



Supplementary Fig. 30. ¹H NMR spectrum of compound 3h.



Supplementary Fig. 31. ¹H NMR spectrum of compound 3i.



Supplementary Fig. 32. ¹H NMR spectrum of compound 3j.



Supplementary Fig. 33. ¹H NMR spectrum of compound 3k.



Supplementary Fig. 34. ¹H NMR spectrum of compound 3l.



Supplementary Fig. 35. ¹H NMR spectrum of compound 3m.



Supplementary Fig. 36. ¹H NMR spectrum of compound 3n.



Supplementary Fig. 37. ¹H NMR spectrum of compound 30.



Supplementary Fig. 38. ¹H NMR spectrum of compound 3p.



Supplementary Fig. 39. ¹H NMR spectrum of compound 3q.



Supplementary Fig. 40. ¹H NMR spectrum of compound 3r.



Supplementary Fig. 41. ¹H NMR spectrum of compound 3s.





6.0

5.5

5.0

4.5

4.0

f1 (ppm)

3.5

3.0

2.5

2.0

1.5

1.0

0.5

7.5

7.0

6.5



Supplementary Fig. 43. ¹³C NMR spectrum of compound 3t.



Supplementary Fig. 44. ¹H NMR spectrum of compound 3u.



Supplementary Fig. 45. ¹H NMR spectrum of compound 3v.



Supplementary Fig. 46. ¹H NMR spectrum of compound 3w.



Supplementary Fig. 47. ¹H NMR spectrum of compound 3x.



Supplementary Fig. 48. ¹H NMR spectrum of compound 3y.



Supplementary Fig. 49. ¹³C NMR spectrum of compound 3y.



Supplementary Fig. 50. ¹H NMR spectrum of compound 3z.



Supplementary Fig. 51. ¹H NMR spectrum of compound 3aa.



Supplementary Fig. 52. ¹H NMR spectrum of compound 3ab.



Supplementary Fig. 53. ¹H NMR spectrum of compound 4a.



Supplementary Fig. 54. ¹H NMR spectrum of compound 5a.



Supplementary Fig. 55. ¹H NMR spectrum of compound 6a.



Supplementary Fig. 56. ¹H NMR spectrum of compound 7a.



Supplementary Fig. 57. ¹H NMR spectrum of compound 8a.



180 170 160 150 140 130 f1 (ppm)

Supplementary Fig. 58. ¹³C NMR spectrum of compound 8a.



Supplementary Fig. 59. ¹H NMR spectrum of compound 9a.



Supplementary Fig. 60. ¹H NMR spectrum of compound 10a.



Supplementary Fig. 61. ¹H NMR spectrum of compound 11a.



Supplementary Fig. 62. ¹H NMR spectrum of compound 12a.



Supplementary Fig. 63. ¹H NMR spectrum of compound 13a.



Supplementary Fig. 64. ¹H NMR spectrum of compound 14a.



Supplementary Fig. 65. ¹H NMR spectrum of compound 15a.



Supplementary Fig. 66. ¹H NMR spectrum of compound 16a.



Supplementary Fig. 67. ¹H NMR spectrum of compound 17a.



Supplementary Fig. 68. ¹H NMR spectrum of compound 18a.



Supplementary Fig. 69. ¹H NMR spectrum of compound 19a.



Supplementary Fig. 70. ¹H NMR spectrum of compound 20a.



Supplementary Fig. 71. ¹H NMR spectrum of compound 21a.



180 170 160 150 140 130 f1 (ppm)





Supplementary Fig. 73. ¹H NMR spectrum of compound 22a.







Supplementary Fig. 75. ¹H NMR spectrum of compound 24a.



Supplementary Fig. 76. ¹³C NMR spectrum of compound 24a.



Supplementary Fig. 77. ¹H NMR spectrum of compound 3ac.



Supplementary Fig. 78. ¹H NMR spectrum of compound 3ad.



Supplementary Fig. 79. ¹³C NMR spectrum of compound 3ad.



Supplementary Fig. 80. ¹H NMR spectrum of compound 25a.



Supplementary Fig. 81. ¹³C NMR spectrum of compound 25a.



Supplementary Fig. 82. ¹H NMR spectrum of compound 3ae.



Supplementary Fig. 83. ¹³C NMR spectrum of compound 3ae.



Supplementary Fig. 84. ¹H NMR spectrum of compound 3af



Supplementary Fig. 85. ¹³C NMR spectrum of compound 3af.







Supplementary Fig. 87. ¹³C NMR spectrum of compound 26a.


Supplementary Fig. 88. ¹H NMR spectrum of compound 3ag.



Supplementary Fig. 89. ¹³C NMR spectrum of compound 3ag.



Supplementary Fig. 90. ¹H NMR spectrum of compound 3ah.



Supplementary Fig. 91. ¹H NMR spectrum of compound 3ai.



Supplementary Fig. 92. ¹H NMR spectrum of compound 3ai.



Supplementary Fig. 93. ¹H NMR spectrum of compound 3aj.



Supplementary Fig. 94. ¹H NMR spectrum of compound 3ak.



Supplementary Fig. 95. ¹³C NMR spectrum of compound 3ak.



Supplementary Fig. 96. ¹H NMR spectrum of compound 3al.



Supplementary Fig. 97. ¹³C NMR spectrum of compound 3al.

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