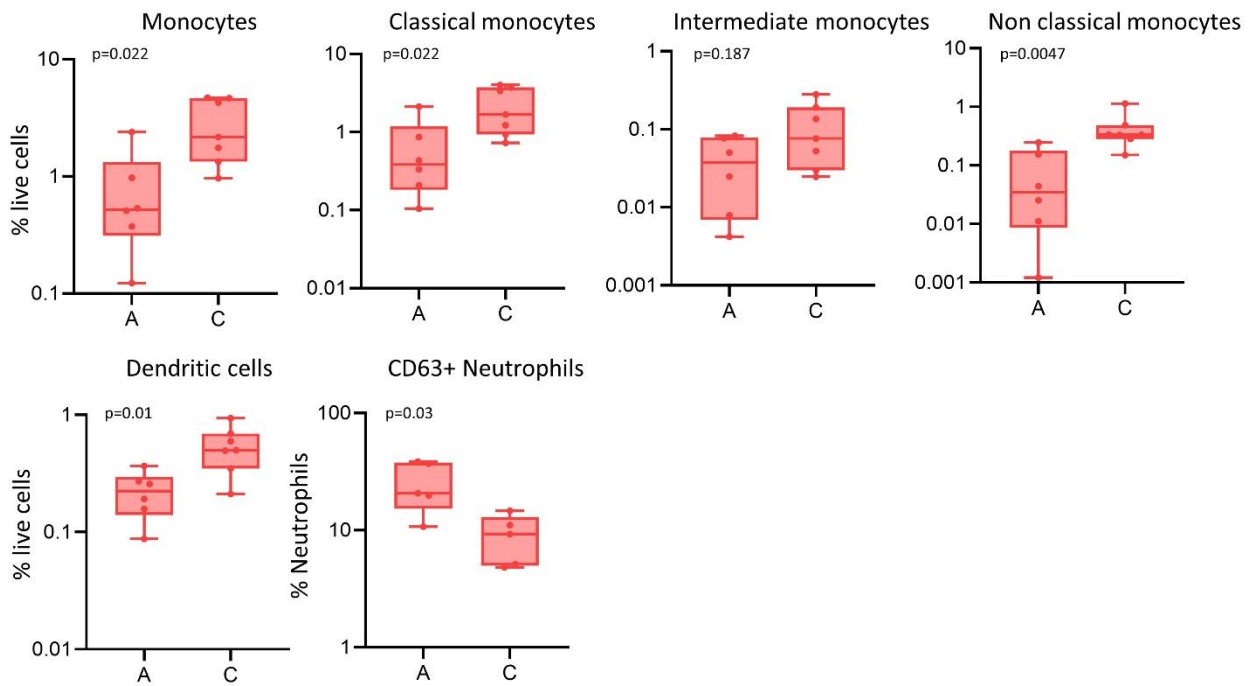
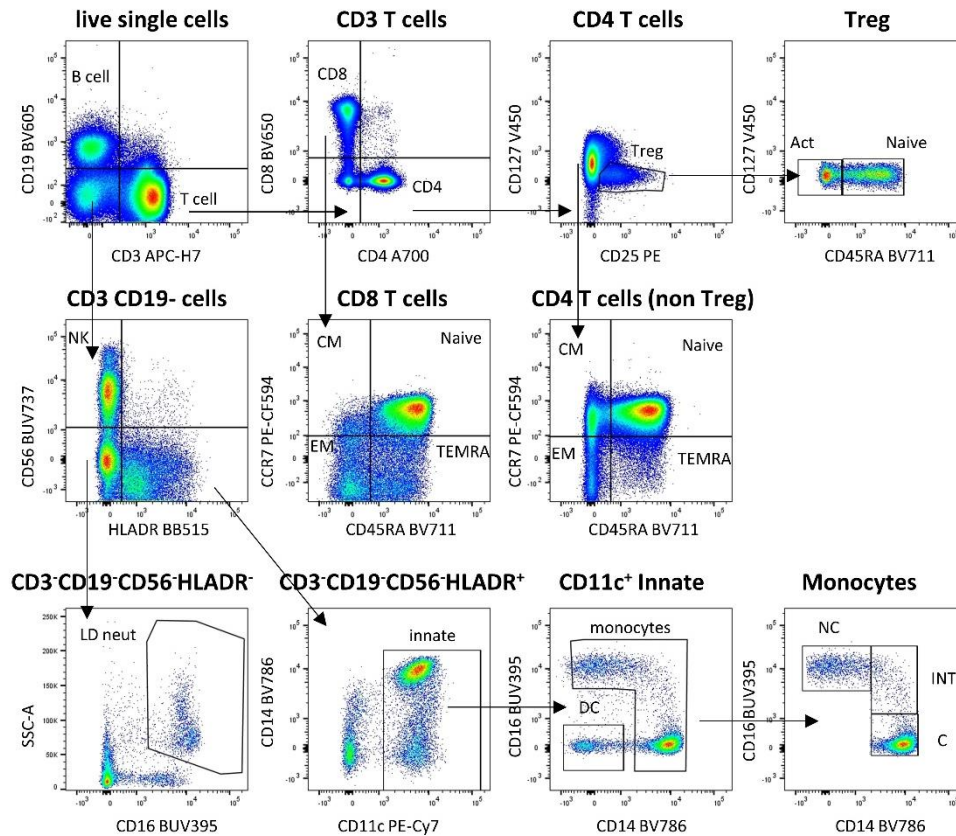


Supplementary Figure 1. Innate cell profiles of adults infected with or exposed to SARS-CoV-2 during the acute and convalescent phase. (a) Frequency of monocytes and subsets (classical, non-classical, intermediate), dendritic cells, NK cells and low-density immature neutrophils in PBMC samples of adults during infection (SARS-CoV-2 positive $n=17$, SARS-CoV-2 exposed $n=21$) and in convalescence (SARS-CoV-2 positive $n=8$, SARS-CoV-2 exposed $n=22$). (b) Frequency of neutrophils and eosinophils in fresh whole blood samples of adults during infection (SARS-CoV-2 positive $n=14$, SARS-CoV-2 exposed $n=20$) and in convalescence (SARS-CoV-2 positive $n=6$, SARS-CoV-2 exposed $n=19$). P values by Kruskal-Wallis rank sum test and Dunn's multiple comparison testing. All statistical tests were performed two-sided. Boxplots show the medians, the 1st and 3rd quartile as well as the smallest and largest values as whiskers.

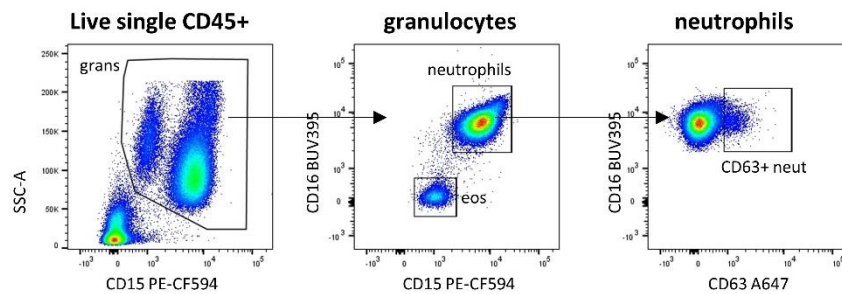


Supplementary Figure 2. Innate cell profiles of children with SARS-CoV-2 during the acute (A) and convalescent (C) phase, excluding children with weak positive PCR results (CT>30). Frequency of monocytes and subsets (classical, non-classical, intermediate), dendritic cells and CD63+ neutrophils in samples from SARS-CoV-2 positive children with initial CT<30 (samples collected during acute phase (n=6) and samples collected during convalescence (n=7)). P values by two-sided Mann-Whitney U test. Boxplots show the medians, the 1st and 3rd quartile as well as the smallest and largest values as whiskers.

A. Representative PBMC gating



B. Representative whole blood gating



Supplementary Figure 3. Representative flow cytometry gating strategies for PBMC and whole blood. (A) Within the PBMC fraction, B cells were selected based on CD19 expression, and the total T cell fraction based on CD3 expression. CD4 and CD8 T cells, and their naïve, effector, memory and regulatory (Treg) subsets were also quantified. CD3⁻CD19⁻ cells were classified into NK cells (CD56⁺) and innate cells (HLA-DR⁺). Within the innate cell fraction, CD14⁺ monocytes and CD11c⁺ DCs were identified. Monocyte subsets were identified based on CD16 expression and classified into classical, intermediate, and non-classical subsets. Low density neutrophils were observed in the PBMC fraction at convalescence, characterised by a high SSC profile and CD16 expression. (B) For whole blood, granulocytes were selected within CD45⁺ leukocytes based on their SSC profile and CD15 expression. Neutrophils were CD15⁺CD16⁺ and eosinophils were CD15⁺CD16⁻. CD63 expression was investigated within the neutrophil population.

Supplementary Table 1. Flow cytometry antibody cocktails

Surface Marker	Fluorophore	Clone	Final Dilution
Whole blood panel			
CD14	BV786	M5E2	1:50
CD11b	BUV805	ICRF44	1:100
CD45	BV711	HI30	1:100
CD56	BUV737	NCAM16.2	1:100
CD11c	PE-Cy7	B-ly6	1:100
CD63	A647	H5C6	1:100
CD4	A700	RPA-T4	1:100
CD3	BB515	UCHTI	1:100
PD1	BB700	EH12.1	1:100
CD15	PE-CF594	W6D3	1:200
HLADR	V500	G46-6	1:200
CD19	BV605	SJ25C1	1:200
CD8	BV650	RPA-T8	1:200
CD16	BUV395	3G8	1:400
Live/dead	N-IR		
PBMC panel			
CD25	PE	M-A251	1:25
CD127	V450	HIL7RM21	1:50
CD3	APCH7	SK7	1:50
CD14	BV786	M5E2	1:50
CD45RA	BV711	HI100	1:100
HLADR	BB515	G46-6	1:100
CD56	BUV737	NCAM16.2	1:100
CD11c	PE-Cy7	B-ly6	1:100
CD4	A700	RPA-T4	1:100
PD1	BB700	EH12.1	1:100
CCR7	PE-CF594	150503	1:200
CD19	BV605	SJ25C1	1:200
CD8	BV650	RPA-T8	1:200
CD16	BUV395	3G8	1:400
Live/dead	BV510		