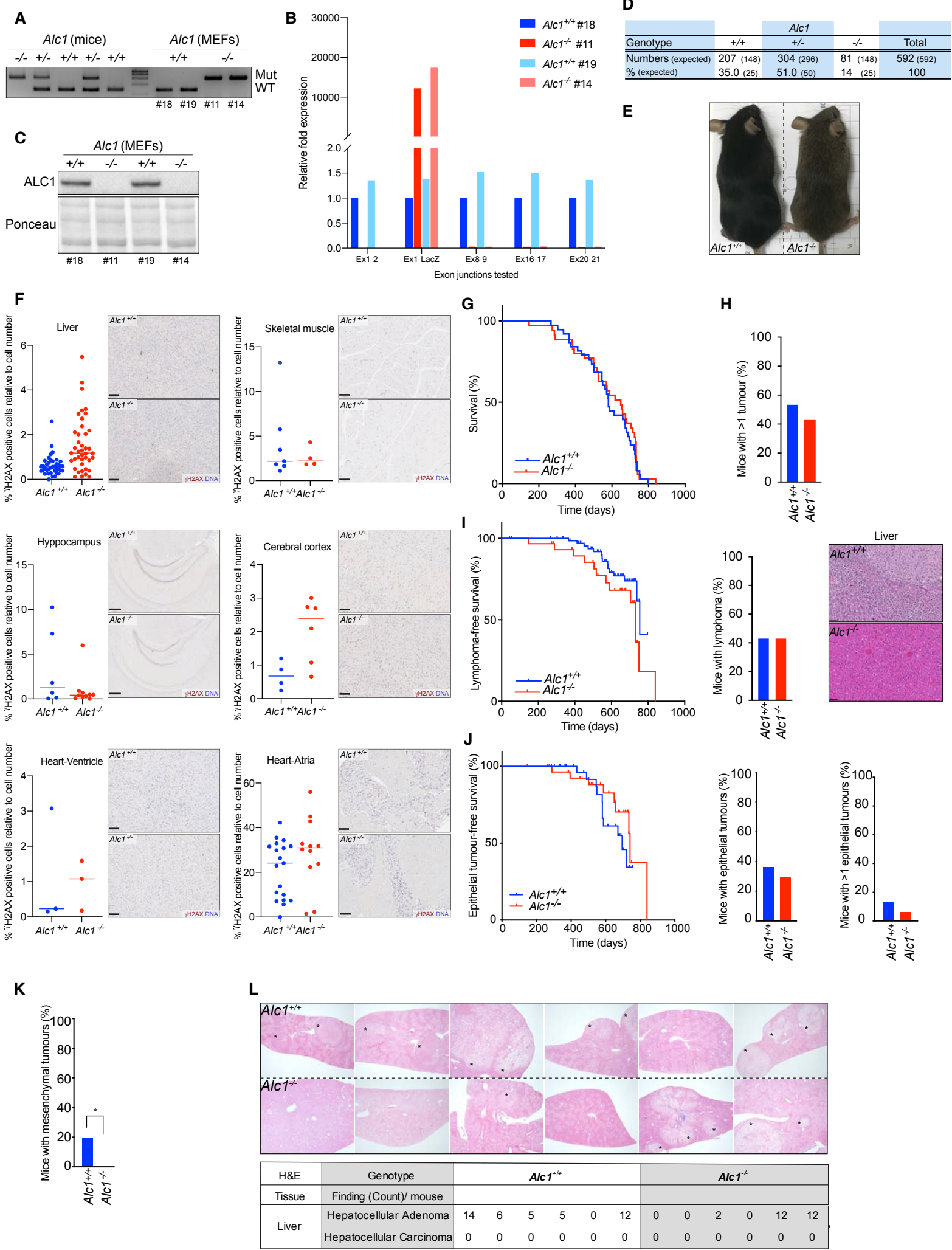


Supplemental Information

**Defective ALC1 nucleosome remodeling confers PARPi
sensitization and synthetic lethality with HRD**

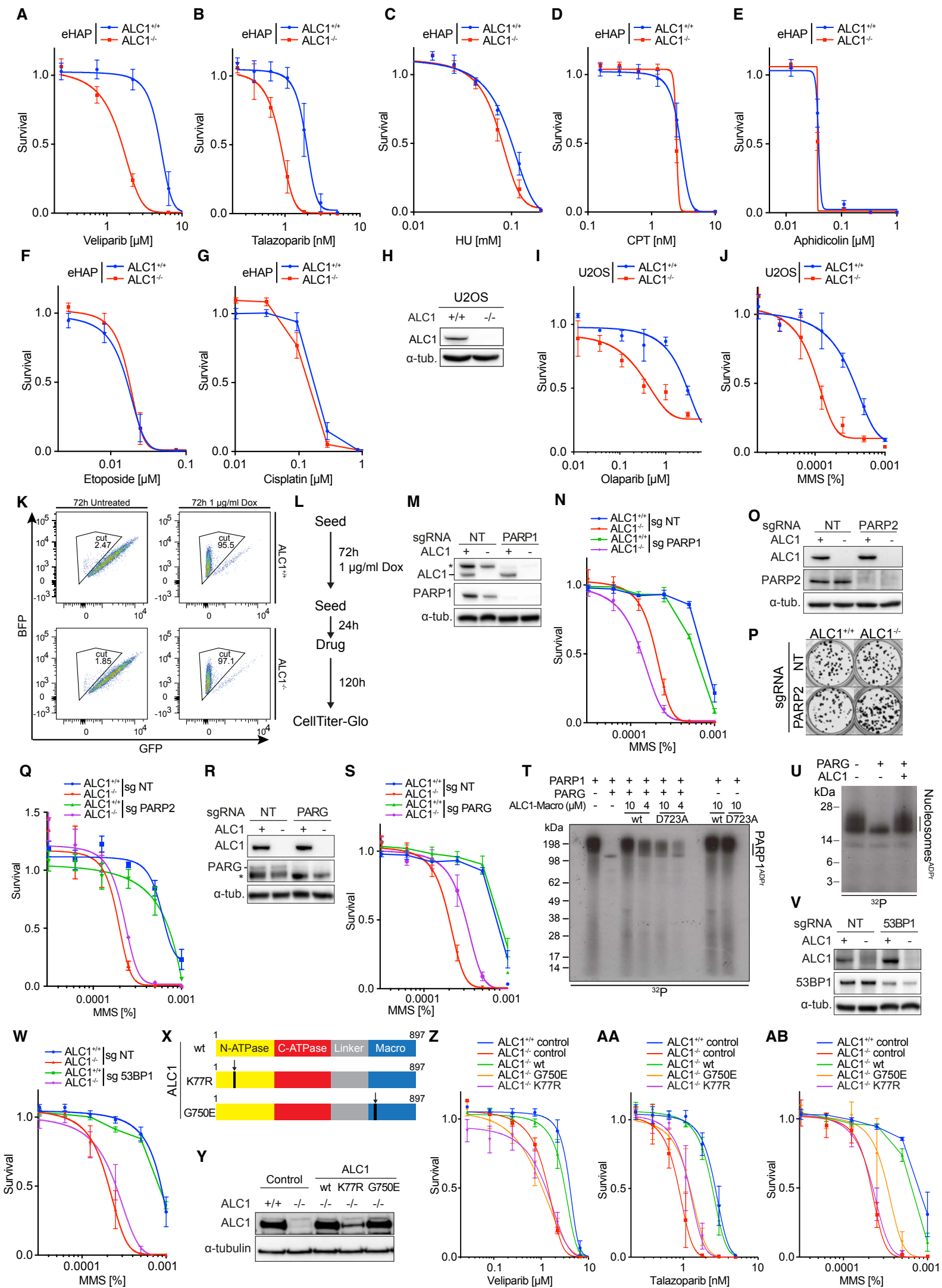
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Supplementary Figure. 1

Figure S1, relating to Figure 1. Tumour prevalence in *Alc1*^{-/-} mice.

(A) *Alc1* mice PCR genotyping strategy. Upper band represents the mutant allele; lower band, the WT allele. (B) Fold reduction of *Alc1* transcript in *Alc1* mice relative to *Alc1*^{+/+} analyzed by qRT-PCR using primers spanning the junction of different exons along the *Alc1* gene. (C) ALC1 protein expression in MEFs. Ponceau was used as loading control. (D) Mendelian ratios of *Alc1* heterozygous mice breeding. Numbers and percent in bracket are expected numbers and ratios. $\chi^2=54.068$, $p<0.0001$. (E) Representative images of 14-week old *Alc1*^{+/+} and *Alc1*^{-/-} littermates showing a slight weight difference. (F) DNA damage analysis in different mitotic and post-mitotic *Alc1*^{+/+} and *Alc1*^{-/-} mice tissues. (G) Overall survival of *Alc1* mice. Significance: Mantel-Cox test, $p=0.6$. $n=30$ *Alc1*^{+/+} and $n=30$ *Alc1*^{-/-}. Mice culled due to nonspecific phenotypes (e.g., dermatitis, overgrown teeth, and fits) were excluded from this study. (H) Frequency of *Alc1* mice which develop more than 1 tumour. Note that there is no difference between both groups. (I) (Left) Lymphoma-free survival of *Alc1* mice. Significance: Mantel-Cox test, $p=0.1$. $n=30$ *Alc1*^{+/+} and $n=30$ *Alc1*^{-/-}. Mice culled due to nonspecific phenotypes (e.g., dermatitis, overgrown teeth, and fits) were excluded from this study. (Middle) Frequency of *Alc1* mice which develop lymphomas. (Right) Representative images of liver lymphoma section stained with hematoxylin and eosin. Note the presence of lymphoma in *Alc1*^{+/+} liver. Scale bars represent 50 μ m. (J) (Left) Epithelial tumour-free survival of *Alc1* mice. Significance: Mantel-Cox test, $p=0.2$. $n=30$ *Alc1*^{+/+} and $n=30$ *Alc1*^{-/-}. Mice culled due to nonspecific phenotypes (e.g., dermatitis, overgrown teeth, and fits) were excluded from this study. Note the longer survival of mice lacking ALC1. (Middle) Frequency of *Alc1* mice which develop epithelial tumours. (Right) Frequency of *Alc1* mice which develop more than 1 epithelial tumour. Fisher's exact test, $p=0.6$. Note the tendency for the *Alc1*^{-/-} mice to develop less epithelial tumours. (K) Frequency of *Alc1* mice which develop mesenchymal tumours. Fisher's exact test, $p=0.02$. *Alc1*^{-/-} mice develop less mesenchymal tumours. (L) (Top) Representative images of haematoxylin and eosin stained liver sections following DEN-induced tumorigenesis. Asterisks identify hepatocellular adenomas. (Bottom) Quantification of the number of hepatocellular adenomas identified in each section by a board-certified veterinary pathologist. Note the tendency of the *Alc1*^{-/-} mice to develop less hepatocellular adenomas.



Supplementary Figure. 2

Figure S2, relating to Figure 2. Response to genotoxins and mechanisms of PARPi resistance in *ALC1*^{-/-} cells.

(A,B) Reduced survival of eHAP *ALC1*^{-/-} cells after treatment with indicated PARPi. Data are mean ± s.e.m normalised to untreated cells (n = 3 independent biological experiments). Solid lines show a nonlinear least-squares fit to a four-parameter dose–response model. **(C-G)** Survival of eHAP *ALC1*^{-/-} cells after treatment with indicated genotoxin. Data are mean ± s.e.m normalised to untreated cells (n = 3 independent biological experiments). Solid lines show a nonlinear least-squares fit to a four-parameter dose–response model. **(H)** CRISPR-mediated inactivation of ALC1 in U2OS. Immunoblot of WCEs in *ALC1*^{+/+} and *ALC1*^{-/-} cells, probed for ALC1. α -tubulin was used as a loading control. **(I)** Reduced survival of U2OS *ALC1*^{-/-} cells after treatment with Olaparib. Data are mean ± s.e.m normalised to untreated cells (n = 3 independent biological experiments). Solid lines show a nonlinear least-squares fit to a four-parameter dose–response model. **(J)** Reduced survival of U2OS *ALC1*^{-/-} cells after treatment with MMS. Data are mean ± s.e.m normalised to untreated cells (n = 3 independent biological experiments). Solid lines show a nonlinear least-squares fit to a four-parameter dose–response model. **(K)** Single cell clones of *ALC1*^{+/+} and *ALC1*^{-/-} were isolated and screened for CAS9 activity using GFP-BFP reporter \pm 72h Dox. BFP⁺ cells were gated and the loss of GFP expression was measured using FACS and displayed as pseudo colour FACS plots. Gates labelled cut represent the percentage of BFP⁺ GFP⁻ cells. **(L)** Schematic representation of survival assays using CellTiter-Glo in inducible CAS9 cells. **(M)** Immunoblot of WCEs from *ALC1*^{+/+} and *ALC1*^{-/-} iCAS9 cells expressing sgRNA against NT or PARP1 following 72h Dox, probed for ALC1 and PARP1. α -tubulin was used as a loading control. **(N)** Increased MMS sensitivity in iCAS9 *ALC1*^{+/+} and *ALC1*^{-/-} eHAP expressing PARP1 sgRNA following 72h Dox. Data are mean ± s.e.m normalised to untreated cells (n = 3 independent biological experiments). Solid lines show a nonlinear least-squares fit to a four-parameter dose–response model. **(O)** Immunoblot of WCEs from *ALC1*^{+/+} and *ALC1*^{-/-} iCAS9 cells expressing sgRNA against NT or PARP2 following 72h Dox, probed for ALC1 and PARP2. α -tubulin was used as a loading control. **(P)** Representative images (n = 3 biologically independent experiments) of clonogenic survival assays in *ALC1*^{+/+} and *ALC1*^{-/-} iCAS9 cells expressing indicated sgRNA following 72h Dox. **(Q)** MMS sensitivity in iCAS9 *ALC1*^{+/+} and *ALC1*^{-/-} eHAP expressing PARP2 sgRNA following 72h Dox. Data are mean ± s.e.m normalised to untreated cells (n = 3 independent biological experiments). Solid lines show a nonlinear least-squares fit to a four-parameter dose–response model. **(R)** Immunoblot of WCEs from *ALC1*^{+/+} and *ALC1*^{-/-} iCAS9 cells expressing sgRNA against NT or PARG following 72h Dox, probed for ALC1 and PARG. α -tubulin was used as a loading control. **(S)** Rescue of MMS sensitivity in iCAS9 *ALC1*^{-/-} eHAP expressing PARG sgRNA following 72h Dox. Data are mean ± s.e.m normalised to untreated cells (n = 3 independent biological experiments). Solid lines show a nonlinear least-squares fit to a four-parameter dose–response model. **(T)** Radioactive ADP-ribosylation assay of PARP1 \pm PARG incubated with macro-ALC1(wt or mutant) at the indicated concentrations. **(U)** Radioactive ADP-ribosylation assay of nucleosomes \pm PARG incubated with macro-ALC1. **(V)** Immunoblot of WCEs from *ALC1*^{+/+} and *ALC1*^{-/-} iCAS9 cells expressing sgRNA against NT or 53BP1 following 72h Dox, probed for ALC1 and 53BP1. α -tubulin was used as a loading control. **(W)** MMS sensitivity in inducible CAS9 *ALC1*^{+/+} and *ALC1*^{-/-} eHAP expressing NT or 53BP1 sgRNA following 72h Dox. Data are mean ± s.e.m normalised to untreated cells (n = 3 independent biological experiments). **(X-AB)** PARPi and MMS sensitivity are associated with defective nucleosome remodelling. **(X)** Schematic of domain structure of indicated ALC1 constructs. **(Y)** Immunoblot of WCEs in *ALC1*^{+/+} and *ALC1*^{-/-} cells transduced with indicated constructs, probed for ALC1. α -tubulin was used as a loading control. **(Z,AA)** PARPi sensitivity in *ALC1*^{+/+} and *ALC1*^{-/-} cells transduced with the indicated constructs. Data are mean ± s.e.m normalised to untreated cells (n = 3 independent biological experiments). Solid lines

show a nonlinear least-squares fit to a four-parameter dose–response model. **(AB)** MMS sensitivity in *ALCI*^{+/+} and *ALCI*^{-/-} cells transduced with the indicated constructs. Data are mean \pm s.e.m normalised to untreated cells (n = 3 independent biological experiments). Solid lines show a nonlinear least-squares fit to a four-parameter dose–response model.

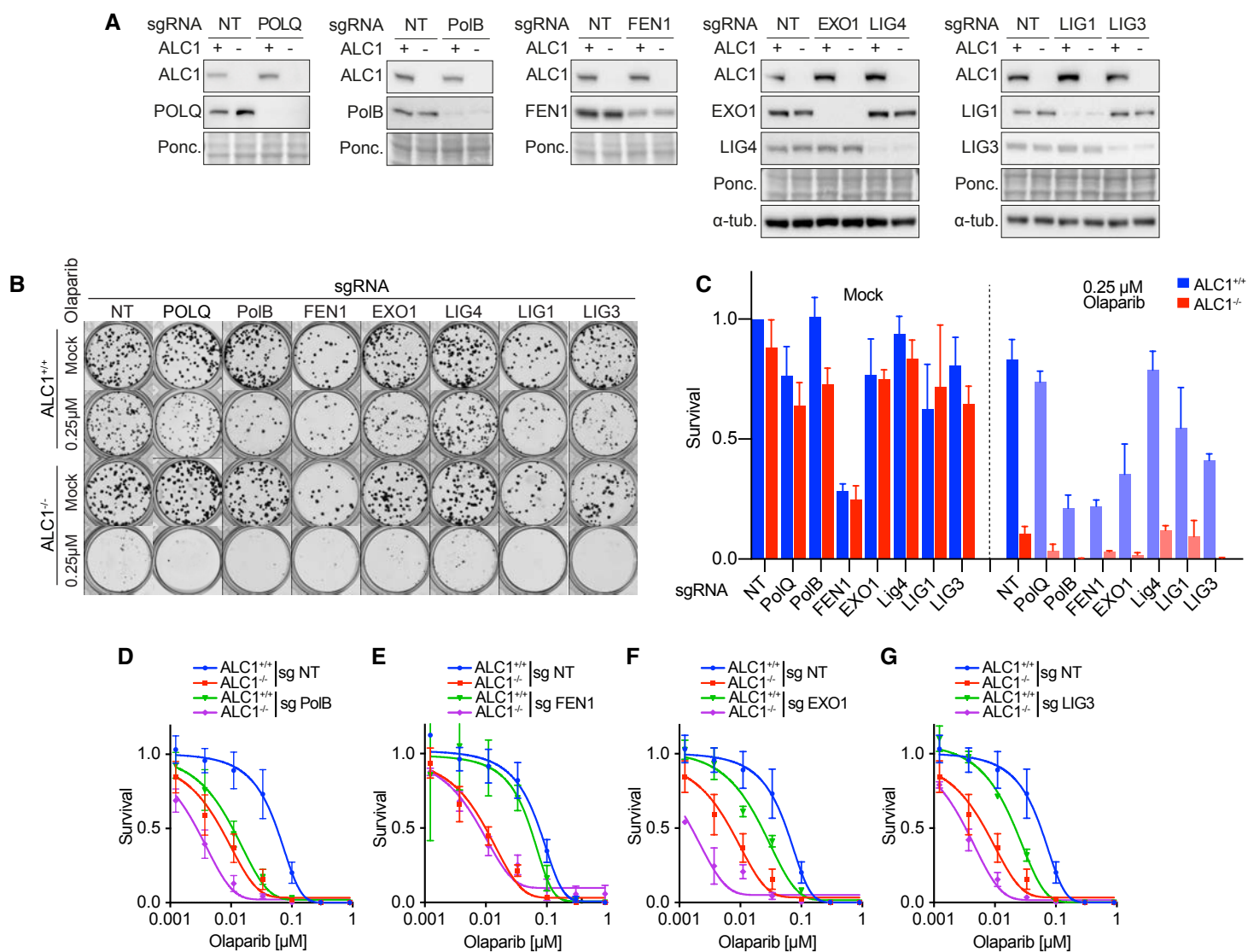


Figure S3, relating to Figure 3. DDR candidate *ALCI*^{-/-} cell panel.

(A) Immunoblots of WCEs in *ALCI*^{+/+} and *ALCI*^{-/-} *iCAS9* cells transduced with indicated sgRNA following 72h Dox. Probed with indicated antibodies. **(B)** Representative images (n = 3 biologically independent experiments) of clonogenic survival assays in *ALCI*^{+/+} and *ALCI*^{-/-} *iCAS9* cells expressing indicated sgRNA following 72h Dox ± 250 nM Olaparib. **(C)** Quantification of clonogenic survival assays in *ALCI*^{+/+} and *ALCI*^{-/-} *iCAS9* cells expressing indicated sgRNA following 72h Dox ± 250 nM Olaparib. Data are mean ± s.e.m normalised to non-treated *ALCI*^{+/+} NT sgRNA (n = 3 biologically independent experiments). **(D)** Olaparib survival of *ALCI*^{+/+} and *ALCI*^{-/-} *iCAS9* cells transduced with NT sgRNA and Polβ sgRNA following 72h Dox. Data are mean ± s.e.m normalised to untreated cells (n = 3 independent biological experiments). Solid lines show a nonlinear least-squares fit to a four-parameter dose–response model. **(E)** Olaparib survival of *ALCI*^{+/+} and *ALCI*^{-/-} *iCAS9* cells transduced with NT sgRNA and FEN1 sgRNA following 72h Dox. Data are mean ± s.e.m normalised to untreated cells (n = 3 independent biological experiments). Solid lines show a nonlinear least-squares fit to a four-parameter dose–response model. **(F)** Olaparib survival of *ALCI*^{+/+} and *ALCI*^{-/-} *iCAS9* cells transduced with NT sgRNA and EXO1 sgRNA following 72h Dox. Data are mean ± s.e.m normalised to untreated cells (n = 3 independent biological experiments). Solid lines show a nonlinear least-squares fit to a four-parameter dose–response model. **(G)** Olaparib survival of *ALCI*^{+/+} and *ALCI*^{-/-} *iCAS9* cells transduced with NT sgRNA and LIG3 sgRNA following 72h Dox. Data are mean ± s.e.m normalised to untreated cells (n = 3 independent biological experiments). Solid lines show a nonlinear least-squares fit to a four-parameter dose–response model.

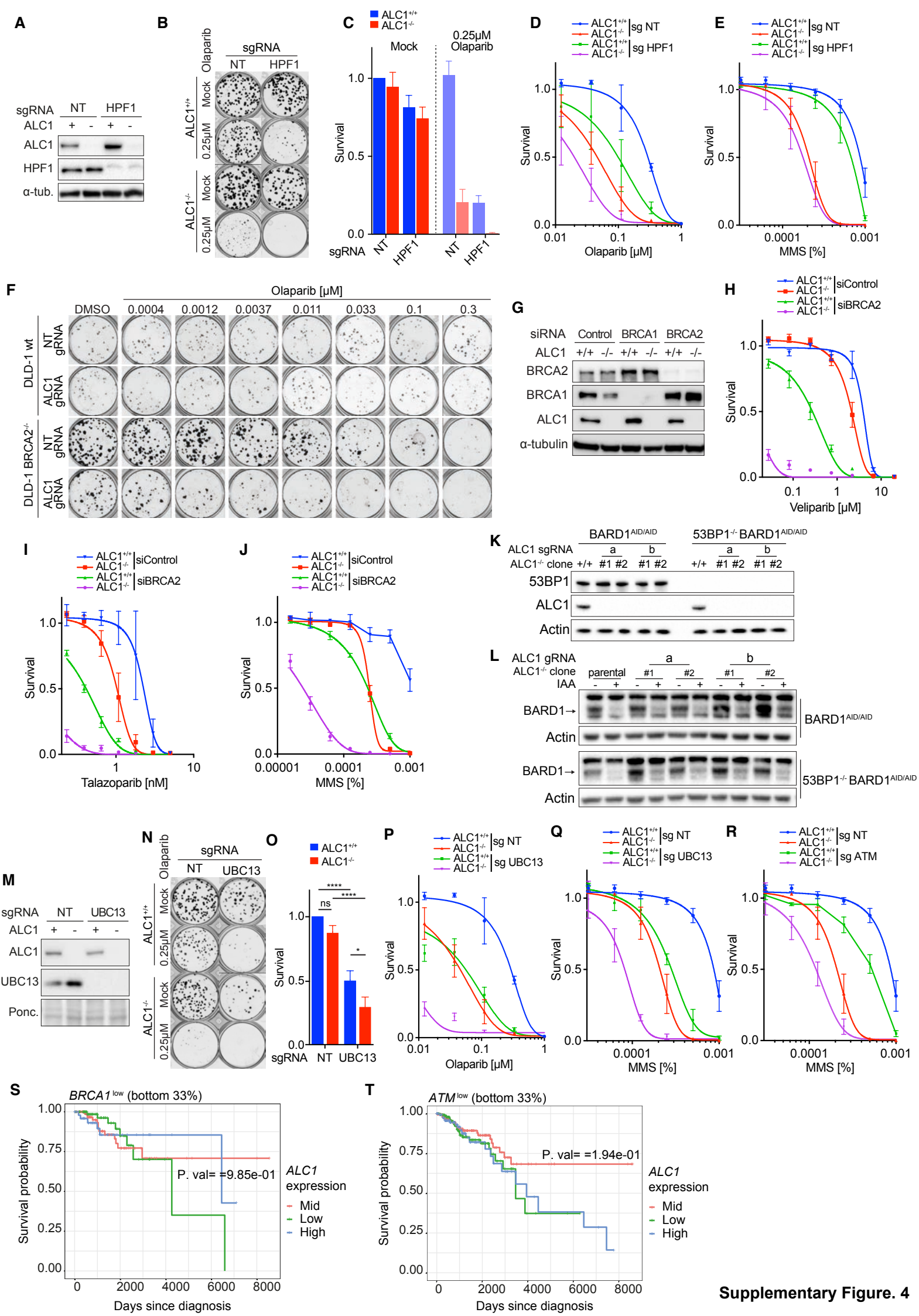
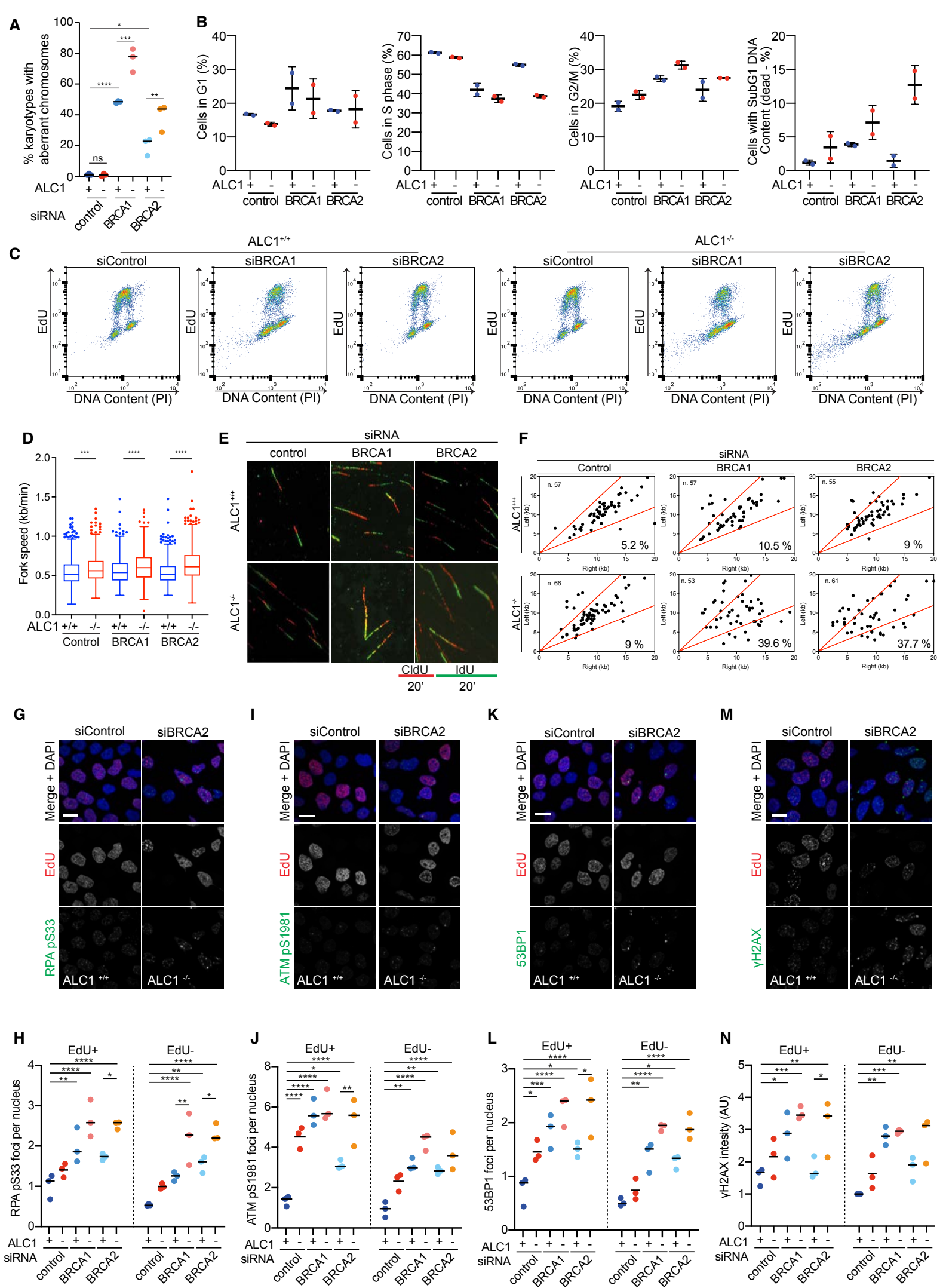


Figure S4, relating to Figure 4. PARPI hypersensitivity in *ALC1*^{-/-} cells in HRD and ATM deficient cells.

(A) Immunoblots of WCEs in *ALC1*^{+/+} and *ALC1*^{-/-} *iCAS9* cells transduced with NT or HPF1 sgRNA following 72h Dox, probed with antibodies against ALC1 and HPF1. α -tubulin is used as a loading control. (B) Representative images (n = 3 biologically independent experiments) of clonogenic survival assays in *ALC1*^{+/+} and *ALC1*^{-/-} *iCAS9* cells expressing NT and HPF1 sgRNA following 72h Dox \pm 250 nM Olaparib. (C) Quantification of clonogenic survival assays in *ALC1*^{+/+} and *ALC1*^{-/-} *iCAS9* cells expressing NT sgRNA and HPF1 sgRNA following 72h Dox \pm 250 nM Olaparib. Data are mean \pm s.e.m normalised to non-treated *ALC1*^{+/+} NT sgRNA (n = 3 biologically independent experiments). (D,E) Olaparib and MMS survival of *ALC1*^{+/+} and *ALC1*^{-/-} *iCAS9* cells transduced with NT sgRNA and HPF1 sgRNA following 72h Dox. Data are mean \pm s.e.m normalised to untreated cells (n = 3 independent biological experiments). Solid lines show a nonlinear least-squares fit to a four-parameter dose-response model. (F) Representative images (n = 3 biologically independent experiments) of clonogenic survival assays in DLD-1 *BRCA2*^{+/+} *ALC1*^{+/+}, *BRCA2*^{+/+} *ALC1*^{-/-}, *BRCA2*^{-/-} *ALC1*^{+/+} and *BRCA2*^{-/-} *ALC1*^{Low expression} (Fig. 4A,B) with the indicated dose of Olaparib. (G) Immunoblot of WCEs in *ALC1*^{+/+} and *ALC1*^{-/-} eHAP cells transfected with non-targeting or BRCA1/2-targeting siRNAs probed for ALC1, BRCA1 and BRCA2. α -tubulin was used as a loading control. (H-J) *ALC1*^{+/+} and *ALC1*^{-/-} eHAP cells were transfected with non-targeting or BRCA2-targeting siRNAs and treated with genotoxin indicated. Data are mean \pm s.e.m normalised to untreated cells (n = 3 biologically independent experiments). Solid lines show a nonlinear least-squares fit to a four-parameter dose-response model. (K) Immunoblots of WCEs in parental (*ALC1*^{+/+}) and *ALC1*^{-/-} *BARD1*^{AID/AID} and parental (*ALC1*^{+/+}) and *ALC1*^{-/-} *53BP1*^{-/-} *BARD1*^{AID/AID} cells, probed with antibodies against ALC1 and 53BP1. Actin is used as a loading control. (L) Immunoblots of WCEs in parental and *ALC1*^{-/-} in *BARD1*^{AID/AID} cells (top) and parental and *ALC1*^{-/-} *53BP1*^{-/-} *BARD1*^{AID/AID} cells (bottom) \pm 2h IAA, probed with antibodies against BARD1. Actin is used as a loading control. (M-O) Loss of UBC13 is synthetic lethal in *ALC1*^{-/-} cells. (M) Immunoblot of WCEs from *ALC1*^{+/+} and *ALC1*^{-/-} *iCAS9* cells expressing sgRNA against NT or UBC13 following 72h Dox, probed for ALC1 and UBC13. Ponceau was used as a loading control. (N) Representative images (n = 3 biologically independent experiments) of clonogenic survival assays in *ALC1*^{+/+} and *ALC1*^{-/-} *iCAS9* cells expressing sgRNA against NT or UBC13 following 72h Dox \pm 250 nM Olaparib. (O) Quantification of non-treated control clonogenic survival assays in *ALC1*^{+/+} and *ALC1*^{-/-} *iCAS9* cells expressing NT or UBC13 sgRNA following 72h Dox. Data are mean \pm s.e.m normalised to *ALC1*^{+/+} NT sgRNA (n = 3 biologically independent experiments). (P,Q) Survival of *ALC1*^{+/+} and *ALC1*^{-/-} eHAP cells expressing NT or UBC13 sgRNA following 72h Dox treated with genotoxin indicated. Data are mean \pm s.e.m normalised to untreated cells (n = 3 biologically independent experiments). Solid lines show a nonlinear least-squares fit to a four-parameter dose-response model. (R) MMS survival of *ALC1*^{+/+} and *ALC1*^{-/-} *iCAS9* cells transduced with NT sgRNA and ATM sgRNA following 72h Dox. Data are mean \pm s.e.m normalised to untreated cells (n = 3 independent biological experiments). Solid lines show a nonlinear least-squares fit to a four-parameter dose-response model. (S) KM survival analysis of BRCA1low breast cancer patients from TGCA according to ALC1 expression. (T) KM survival analysis of ATMlow breast cancer patients from TGCA according to ALC1 expression.



Supplementary Figure. 5

Figure S5, relating to Figure 5. Phenotypic characterisation of cell cycle, replication and DDR following loss of HR in *ALC1*^{-/-} cells.

(A) Loss of *ALC1* exacerbates genomic instability in HR deficient cells. Quantification of % of karyotypes with aberrant metaphase spreads from *ALC1*^{+/+} and *ALC1*^{-/-} eHAP cells transfected with non-targeting, BRCA1-targeting and BRCA2-targeting siRNAs. Data are means from individual experiments; bar represents median (n = 3 independent biological experiments). (B) Quantification of % of cells in different cell-cycle stages in *ALC1*^{+/+} and *ALC1*^{-/-} eHAP cells transfected with non-targeting, BRCA1-targeting or BRCA2-targeting siRNAs. Data are n=2 independent biological experiments. (C) Pseudo colour FACS plots showing EdU incorporation and DNA content in *ALC1*^{+/+} and *ALC1*^{-/-} eHAP cells transfected with non-targeting, BRCA1-targeting or BRCA2-targeting siRNAs. Data are representative of n=2 independent biological experiments. (D) Boxplot showing replication fork speed (measured as IdU track length/min) in *ALC1*^{+/+} and *ALC1*^{-/-} eHAP transfected with the indicated siRNAs. Data are from >500 fibres, mean ± s.d from n = 2 independent biological experiments. (E) Lower: scheme of the nucleotide labelling strategy used in the fiber assay experiments. Upper: Representative DNA fiber immunofluorescence images from *ALC1*^{+/+} and *ALC1*^{-/-} eHAP cells transfected with the indicated siRNAs. (F) Analysis of replication fork symmetry from *ALC1*^{+/+} and *ALC1*^{-/-} eHAP cells transfected with control siRNA or siRNAs targeting BRCA1 or BRCA2. The total number of newly activated replication forks analysed is indicated. (G) Representative micrographs of *ALC1*^{+/+} and *ALC1*^{-/-} eHAP cells transfected with non-targeting and BRCA2-targeting siRNAs stained with RPA pSer33 antibody, EdU click-iT and DAPI. Scale bar = 10 um. (H) Quantification of nuclear RPA pS33 foci in CSK pre-extracted EdU+ and EdU- *ALC1*^{+/+} and *ALC1*^{-/-} eHAP cells transfected with indicated siRNAs 72h following knockdown. Data are means from individual experiments, bar represents median (n = 3 biologically independent experiments). (I) Representative micrographs of *ALC1*^{+/+} and *ALC1*^{-/-} eHAP cells transfected with non-targeting and BRCA2-targeting siRNAs stained with ATM pS1981 antibody, EdU click-iT and DAPI. Scale bar = 10 um. (J) Quantification of nuclear ATM pS1981 foci in CSK pre-extracted EdU+ and EdU- *ALC1*^{+/+} and *ALC1*^{-/-} eHAP cells transfected with indicated siRNAs 72h following knockdown. Data are means from individual experiments, bar represents median (n = 3 biologically independent experiments). (K) Representative micrographs of *ALC1*^{+/+} and *ALC1*^{-/-} eHAP cells transfected with non-targeting, BRCA1-targeting and BRCA2-targeting siRNAs stained with 53BP1 antibody, EdU click-iT and DAPI. Scale bar = 10 um. (L) Quantification of nuclear 53BP1 foci in CSK pre-extracted EdU+ and EdU- *ALC1*^{+/+} and *ALC1*^{-/-} eHAP cells transfected with indicated siRNAs 72h following knockdown. Data are means from individual experiments, bar represents median (n = 3 biologically independent experiments). (M) Representative micrographs of *ALC1*^{+/+} and *ALC1*^{-/-} eHAP cells transfected with non-targeting and BRCA2-targeting siRNAs stained with γ H2AX antibody, EdU click-iT and DAPI. Scale bar = 10 um. (N) Quantification of nuclear γ H2AX intensity in CSK pre-extracted EdU+ and EdU- *ALC1*^{+/+} and *ALC1*^{-/-} eHAP cells transfected with indicated siRNAs 72h following knockdown. Data are means from individual experiments normalised to *ALC1*^{+/+} siControl (EdU-), bar represents median (n = 3 biologically independent experiments).

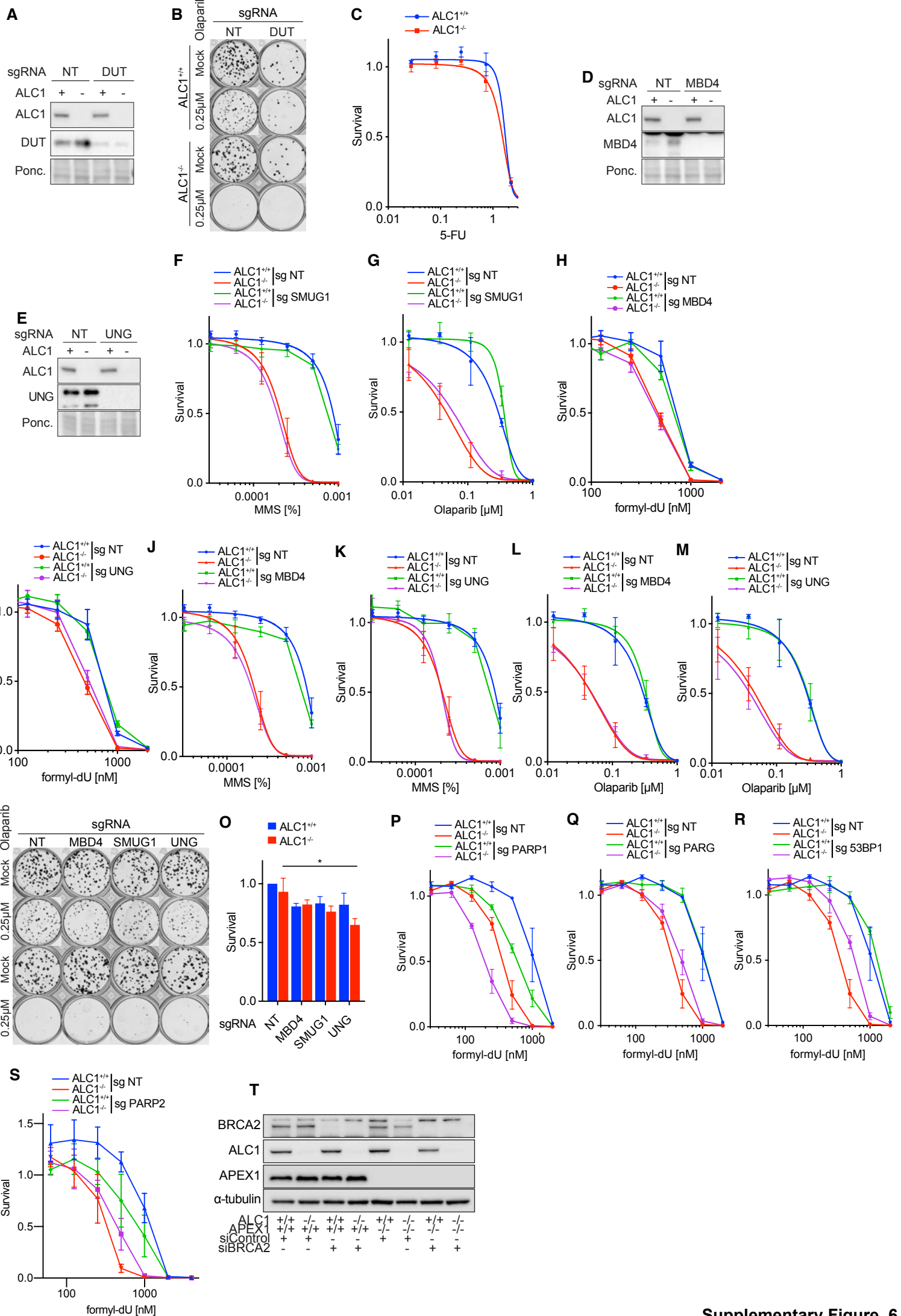


Figure S6, relating to Figure 6. Uracil metabolism in *ALCI*^{-/-} cells.

(A) Immunoblots of WCEs in *ALCI*^{+/+} and *ALCI*^{-/-} *iCAS9* cells transduced with DUT sgRNA following 72h Dox. Probed with antibodies against ALC1 and DUT. Ponceau is used as a loading control. (B) Representative images (n = 3 biologically independent experiments) of clonogenic survival assays in *ALCI*^{+/+} and *ALCI*^{-/-} *iCAS9* cells expressing NT and DUT sgRNA following 72h Dox ± 250 nM Olaparib. (C) 5-FU sensitivity in *ALCI*^{+/+} and *ALCI*^{-/-} eHAP. Data are mean ± s.e.m normalised to untreated cells (n = 3 biologically independent experiments). Solid lines show a nonlinear least-squares fit to a four-parameter dose–response model. (D,E) Immunoblot of WCEs *ALCI*^{+/+} and *ALCI*^{-/-} eHAP *iCAS9* cells expressing indicated sgRNA following 72h Dox. Probed for ALC1, MBD4 and UNG. Ponceau was used as a loading control. (F,G) SMUG1 knockout does not rescue ALC1 dependent sensitivity to Olaparib or MMS. (F) MMS survival of *ALCI*^{+/+} and *ALCI*^{-/-} *iCAS9* cells transduced with NT sgRNA and SMUG1 sgRNA following 72h Dox. Data are mean ± s.e.m normalised to untreated cells (n = 3 independent biological experiments). (G) Olaparib survival of *ALCI*^{+/+} and *ALCI*^{-/-} *iCAS9* cells transduced with NT sgRNA and SMUG1 sgRNA following 72h Dox. Data are mean ± s.e.m normalised to untreated cells (n = 3 independent biological experiments). (H,I) Formyl-dU survival of *ALCI*^{+/+} and *ALCI*^{-/-} *iCAS9* cells transduced with indicated sgRNA following 72h Dox. Data are mean ± s.e.m normalised to untreated cells (n = 3 independent biological experiments). (J,K) MMS survival of *ALCI*^{+/+} and *ALCI*^{-/-} *iCAS9* cells transduced with indicated sgRNA following 72h Dox. Data are mean ± s.d normalised to untreated cells (n = 3 independent biological experiments). (L,M) Olaparib survival of *ALCI*^{+/+} and *ALCI*^{-/-} *iCAS9* cells transduced with indicated sgRNA following 72h Dox. Data are mean ± s.d normalised to untreated cells (n = 3 independent biological experiments). (N) Representative images (n = 3 biologically independent experiments) of clonogenic survival assays in *ALCI*^{+/+} and *ALCI*^{-/-} *iCAS9* cells expressing indicated sgRNA following 72h Dox ± 250 nM Olaparib. (O) Quantification of non-treated control clonogenic survival assays in *ALCI*^{+/+} and *ALCI*^{-/-} *iCAS9* cells expressing indicated sgRNA following 72h Dox. Data are mean ± s.e.m normalised to *ALCI*^{+/+} NT sgRNA (n = 3-5 biologically independent experiments). (P) Increased formyl-dU sensitivity in *iCAS9* *ALCI*^{+/+} and *ALCI*^{-/-} eHAP expressing PARP1 sgRNA following 72h Dox. Data are mean ± s.e.m normalised to untreated cells (n = 3 independent biological experiments). (Q) Formyl-dU sensitivity in *iCAS9* *ALCI*^{-/-} eHAP expressing PARG sgRNA following 72h Dox. Data are mean ± s.e.m normalised to untreated cells (n = 3 biologically independent experiments). (R) Partial rescue of formyl-dU sensitivity in *iCAS9* *ALCI*^{-/-} eHAP expressing 53BP1 sgRNA following 72h Dox. Data are mean ± s.e.m normalised to untreated cells (n = 3 biologically independent experiments). (S) Partial rescue of formyl-dU sensitivity in *iCAS9* *ALCI*^{-/-} eHAP expressing PARP2 sgRNA following 72h Dox. Data are mean ± s.e.m normalised to untreated cells (n = 3 biologically independent experiments). (T) Immunoblot of WCEs in *ALCI*^{+/+} *APEX1*^{+/+}, *ALCI*^{+/+} *APEX1*^{-/-}, *ALCI*^{-/-} *APEX1*^{+/+} and *ALCI*^{-/-} *APEX1*^{-/-} eHAP cells transfected with non-targeting or BRCA2-targeting siRNAs, probed for ALC1, APEX1 and BRCA2. α -tubulin was used as a loading control.

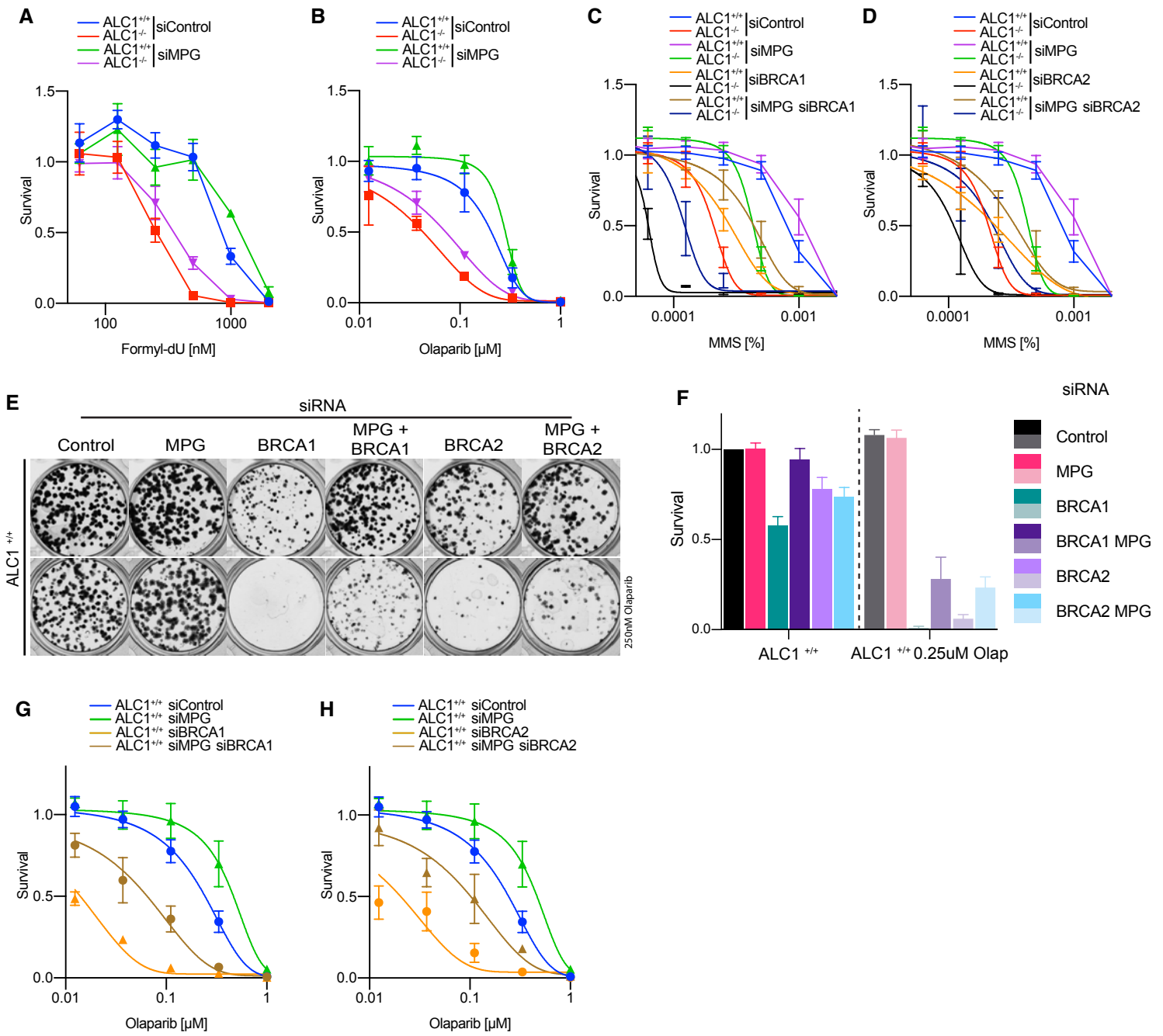


Figure S7, relating to Figure 7. The role of MPG in MMS and PARPi response in $ALC1^{-/-}$ cells.

(A) Formyl-dU survival of $ALC1^{+/+}$ and $ALC1^{-/-}$ eHAP cells transfected with non-targeting or MPG-targeting siRNAs. Data are mean \pm s.e.m normalised to untreated cells ($n = 3$ independent biological experiments). **(B)** Olaparib survival of $ALC1^{+/+}$ and $ALC1^{-/-}$ eHAP cells transfected with non-targeting or MPG-targeting siRNAs. Data are mean \pm s.e.m normalised to untreated cells ($n = 3$ independent biological experiments). Solid lines show a nonlinear least-squares fit to a four-parameter dose–response model. **(C)** MMS survival of $ALC1^{+/+}$ and $ALC1^{-/-}$ eHAP cells transfected with non-targeting, BRAC1 and MPG-targeting siRNAs. Data are mean \pm s.e.m normalised to untreated cells ($n = 3$ independent biological experiments). Solid lines show a nonlinear least-squares fit to a four-parameter dose–response model. **(D)** MMS survival of $ALC1^{+/+}$ and $ALC1^{-/-}$ eHAP cells transfected with non-targeting, BRAC2 and MPG-targeting siRNAs. Data are mean \pm s.e.m normalised to untreated cells ($n = 3$ independent biological experiments). Solid lines show a nonlinear least-squares fit to a four-parameter dose–response model. **(E)** Representative images ($n = 3$ biologically independent experiments) of clonogenic survival assays in $ALC1^{+/+}$ eHAP cells transfected with non-targeting, MPG and BRCA1/2-targeting siRNAs \pm 250 nM Olaparib. **(F)** Quantification of clonogenic survival assays in $ALC1^{+/+}$ eHAP cells transfected with non-targeting, MPG and BRCA1/2-targeting siRNAs \pm 250 nM Olaparib. Data are mean \pm s.e.m normalised to non-treated $ALC1^{+/+}$ non-targeting siRNA ($n = 3$ biologically independent experiments). **(G)** Olaparib survival of $ALC1^{+/+}$ eHAP cells transfected with non-targeting, BRCA1 and MPG-targeting siRNAs. Data are mean \pm s.e.m normalised to untreated cells ($n = 3$ independent biological experiments). Solid lines show a nonlinear least-squares fit to a four-parameter dose–response model. **(H)** Olaparib survival of $ALC1^{+/+}$ eHAP cells transfected with non-targeting, BRCA2 and MPG-targeting siRNAs. Data are mean \pm s.e.m normalised to untreated cells ($n = 3$ independent biological experiments). Solid lines show a nonlinear least-squares fit to a four-parameter dose–response model.