

Figure S1. Analysis of reads from Arabidopsis samples infested with *T. urticae* during 0h (AE7611-13), 30min (AE7614-16), 1h (AE7617-19), 3h (AE7620-22), and 24h (AE7623-25). (A) Principal Component Analysis (PCA) showing the distribution of the samples in the two first dimensions. (B) Percentage of mapped reads per chromosome. (C) Mapping statistics. (D) Number of genes detected with at least one mapping.



Figure S2. Linear graphics comparing the expression pattern of fifteen selected genes by RNA-seq and RT-qPCR techniques.



Figure S3. (A) Scree plot and percentage of variation explained by each PCA dimension. Red circle indicates the present of a point of inflection. (B) Top 15 experiments and (C) top 100 genes with the highest contributions in the eight first PCA dimensions. Red dashed lines indicates the mean value of percentage contribution showed through all experiments or genes.



Figure S4. (A) Representation per cluster of the bootstrapped Confidence Intervals against the log2FC mean from the top 100 clustering-responsible genes. Confidence Intervals significantly different from the log2FC mean for each cluster are coloured in red. (B) Number of genes significantly different to the log2FC mean in each cluster. (C) Venn diagram showing the specific and shared significant genes for each cluster.

Supporting Methods

Real-time RT-qPCR analysis

RT-qPCR assays were performed for validation of the RNA-seq data using a new batch of samples. Rosettes from the ecotype Columbia-0 of *A. thaliana* were sampled and frozen after 0h, 30min, 1h, 3h, and 24h of mite infestation. Total RNA was extracted (Oñate-Sánchez and Vicente-Carbajosa 2008), treated with RQ1 DNase treatment (Promega), and reverse transcribed using Revert AidTM H Minus First Strand cDNA Synthesis Kit (Thermo Fisher). RT-qPCR was performed for three samples coming from three independent experiments using the Light Cycler 480 SYBR Green I Master (Roche). Specific primers were designed through the PRIMER 3 program (Kõressaar et al. 2018). Primer sequences are indicated in **Table S4**. Gene expression was referred to as fold change $(2^{-\Delta\Delta Ct})$, normalizing the expression to ubiquitin (Livak and Schmittgen 2001). The resulting values were log2 transformed (log2 fold change) for comparison with RNA-seq data. The non-parametric Spearman rank correlation test was used. The statistical analysis was performed using GraphPadPrism v6.01.

Analyses of stored microarray and RNA-seq data

Data analysis was developed according to the different nature of the data. For microarray experiments, raw data was obtained as GenePix (GPR) files for two-channel microarray experiments and chip electronic files (CEL) for single-channel microarrays. Bioconductor, affy and limma packages of R v.3.5.2 were used for background correction, normalization and differential expression analyses (Gautier et al. 2004; Gentleman et al. 2004; Ritchie et al. 2015; R Core Team 2018). A gene expression matrix was generated from affy package, using RMA (robust multiarray average), in the case of the single-channel microarrays (Irizarry et al. 2003). For two-channel microarrays, background correction was performed using the 'normexp' approach (Ritchie et al. 2007) and datasets were log-transformed and normalized using the printtip-loess approach (Smyth and Speed 2003). Then, in both cases, statistical analysis and differential expression were performed using linear models and eBayes functions (Phipson et al. 2016). Adjusted p-values were calculated using Benjamini-Hochberg (BH) FDR correction (Benjamini and Hochberg 1995). DEGs were considered those with a log2 ratio (fold change) higher than 1 and an adjusted p-value < 0.05 for all species, except for L. huidobrensis. In this species, to avoid the putative noise inherent to an extremely high number of DEG, we considered a log2 ratio \geq 2 and an adjusted p-value < 0.01.

For the RNA-seq experiments, raw data were obtained as FASTQ files from the NCBI SRA database (Barrett et al. 2012). Reads were mapped against the TAIR10 reference genome of *A*. *thaliana* using STAR aligner (v.2.5.3a and v.2.7.0f) (Dobin et al. 2013). The alignments were sorted with Samtools (Li et al. 2009). HTSeq Phyton library was used for the generation of the

count data files from sorted BAM files through the htseq-count script (Anders et al. 2015). RNAseq read counts were modeled by a generalized linear model considering a specific experimental design for each RNA-seq. To evaluate the DEGs between two different conditions, we performed an analysis with the Bioconductor R package DESeq2 (Love et al. 2014). DEGs were considered those genes with a log2 ratio ≥ 1 and an adjusted p-value < 0.05. These genes were selected for the subsequent comparisons and functional analyses.

Supporting References

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Table S1. Enriched biological processes upon 30min, 1h, 3h, and 24h of *T. urticae* infestation.

30min

GOID	GOTerm	Term PValue	% Associated	Nr. Genes	Associated Genes Found
		Corrected	Genes	Genes	
GO:0009753	response to jasmonic acid	5.53E-33	16.74	37	[5PTASE11, ACS6, AGB1, BT2, BT4, ERF1, GL3, GRX480, JAZ1, JAZ10, JAZ12, JAZ3, JAZ5, JAZ6, JAZ7, JAZ8, MPK4, MPK6, MYB3, MYBR1, MYC2, NINJA, NPR1, NPR3, NPR4, PAP1, PEPR1, PFT1, PHYB, RGA1, RGL1, RGL3, SYP121, SYP122, TIFY10B, TIFY7, TPL]
GO:0009966	regulation of signal transduction	2.34E-28	10.99	40	[ABI1, AT4G16660, EBF2, EXO70B1, GID1B, HAI1, HY5, JAZ1, JAZ10, JAZ12, JAZ3, JAZ5, JAZ6, JAZ7, JAZ8, KEG, MEKK1, MKK2, MKK4, MKK5, MKK6, MKK9, MKP2, MYB73, MYB77, MYB96, MYBR1, NPR1, NPR3, NPR4, PIL6, PP2CA, PTP1, RCAR3, RGA1, RGL1, RGL3, TIFY10B, TIFY7, TMAC2]
GO:0009738	abscisic acid-activated signaling pathway	4.42E-21	12.39	28	[ABF2, ABF3, ABF4, ABI1, BT2, CBL1, CDPK1, CPK32, CPK6, EXO70B1, HAI1, HFR1, HY5, KEG, LTI65, MKK5, MPK3, MPK6, MYB96, NF-YB6, PP2C5, PP2CA, PRN, PYL13, PYL5, PYL7, RCAR3, TMAC2]
GO:0043549	regulation of kinase activity	3.42E-14	12.34	19	[AT2G32710, CKS1, CYCA3;1, CYCD5;1, CYCD6;1, CYCP4;1, HB1, ICK1, ICK5, KRP2, MEKK1, MKK2, MKK4, MKK5, MKK6, MKK9, MKP2, PDK1, PTP1]
GO:0009751	response to salicylic acid	2.92E-13	10.98	19	[AHBP-1B, BAP1, BAP2, BT1, BT2, BT4, BT5, GRX480, MPK4, MYBR1, NPR1, NPR4, OPR1, RGA1, RGL1, RGL3, SYP121, SYP122, WAK1]
GO:0023014	signal transduction by protein phosphorylation	5.92E-12	13.89	15	[MEKK1, MKK2, MKK4, MKK5, MKK6, MKK9, MKP2, MPK11, MPK17, MPK3, MPK4, MPK5, MPK6, PHYB, PTP1]
GO:0006887	exocytosis	6.73E-10	17.46	11	[EXO70B1, EXO70B2, EXO70E2, EXO70H4, EXO70H7, RAB8, SEC15B, SEC1B, SEC8, SYP121, SYP122]
GO:0071482	cellular response to light stimulus	9.74E-10	10.85	14	[CO, HFR1, HY5, NPH4, PAPP2C, PFT1, PHOT1, PHYB, PIF3, PIL5, PIL6, PKS1, PRN, SIB1]
GO:0043405	regulation of MAP kinase activity	4.65E-09	28.57	8	[MEKK1, MKK2, MKK4, MKK5, MKK6, MKK9, MKP2, PTP1]
GO:0004693	cyclin-dependent protein serine/threonine kinase activity	2.01E-08	12.64	11	[AT2G32710, CDC2, CKS1, CYCA3;1, CYCD5;1, CYCD6;1, CYCP4;1, ICK1, ICK5, KRP2, ORG1]
GO:0060548	negative regulation of cell death	6.20E-08	20.51	8	[AT4G16660, BIR1, BON1, BON2, MKP2, RIN4, SYP121, SYP122]

GO:0015036	disulfide oxidoreductase activity	4.97E-07	10.31	10	[AT2G30540, AT4G15660, AT4G15670, AT4G15680, AT4G15700, GRX480, MYB102, NTRC, TH9, TRX5]
GO:0009740	gibberellic acid mediated signaling pathway	2.71E-06	12.12	8	[GID1B, HY5, MYB3, PIF3, PIL5, RGA1, RGL1, RGL3]
GO:0015749	monosaccharide transmembrane transport	4.50E-06	14.00	7	[AT1G08890, ERD6, ESL1, GLT1, MSS1, PMT5, PMT6]
GO:0034620	cellular response to unfolded protein	6.75E-06	12.96	7	[AGB1, BIP3, BZIP28, BZIP60, ERD2, HSP70T-2, mtHsc70-1]
GO:0010182	sugar mediated signaling pathway	7.74E-05	11.11	5	[ABF2, ABI5, BT2, EIN3, PAP1]

1h

GOID	GOTerm	Term	%	Nr.	Associated Genes Found
		PValue	Associated	Genes	
		Corrected	Genes		
GO:0009753	response to jasmonic acid	5.03E-22	11.31	25	[BT2, BT4, ERF1, GL3, GRX480, JAZ1, JAZ10, JAZ12, JAZ3, JAZ5, JAZ6, JAZ7, JAZ8, MPK6, MYB21, MYBR1, MYC2, NINJA, NPR4, PAP1, PFT1, PHYB, TIFY10B, TIFY7, TPL]
GO:0071456	cellular response to hypoxia	3.39E-14	7.63	20	[AGC2-1, AT-HSFB2B, AT1G54050, AT1G59860, AT4G16660, CML37, CML38, HB1, HSFA2, HSP101, HSP17.4, HSP17.6A, HSP17.6II, HSP70, MKK9, PAP1, PBP1, RHL41, STZ, TCH3]
GO:0009966	regulation of signal transduction	1.59E-11	5.49	20	[AT4G16660, CPSF30, HSP70, HY5, JAZ1, JAZ10, JAZ12, JAZ3, JAZ5, JAZ6, JAZ7, JAZ8, MKK4, MKK9, MYB73, MYBR1, NPR4, PTP1, TIFY10B, TIFY7]
GO:0010017	red or far-red light signaling pathway	8.00E-09	14.29	9	[ELF4, HFR1, HY5, HYH, PAP3, PFT1, PHYB, PIF3, PIL5]
GO:0031539	positive regulation of anthocyanin metabolic process	5.81E-08	50.00	5	[GL3, HY5, PAP1, PAP3, PIF3]
GO:0015036	disulfide oxidoreductase activity	3.52E-07	9.28	9	[AT1G06830, AT3G62950, AT4G15660, GRX480, GRXS13, GRXS17, NTRC, ROXY2, TRX5]
GO:0034620	cellular response to unfolded protein	1.15E-06	12.96	7	[AT3G09440, BIP3, ERD2, HSFA2, HSP70, Hsp70-2, MTHSC70-2]
GO:0004707	MAP kinase activity	2.18E-05	11.11	6	[MKK4, MKK9, MPK11, MPK3, MPK6, PTP1]

GO:0009751	response to salicylic acid	3.38E-05	5.20	9	[AHBP-1B, BT1, BT2, BT4, CPSF30, GRX480, MYBR1, NPR4, WRKY62]
GO:0023014	signal transduction by protein phosphorylation	7.59E-05	6.48	7	[MKK4, MKK9, MPK11, MPK3, MPK6, PHYB, PTP1]
GO:0043549	regulation of kinase activity	7.79E-05	5.19	8	[AKINBETA1, CYCA2;3, CYCA3;1, HB1, MKK4, MKK9, PTP1, cycp3;1]
GO:0009739	response to gibberellin	1.40E-04	5.65	7	[BT4, GAMMA-TIP, HY5, MYB21, PAP3, PIF3, PIL5]
GO:0015144	carbohydrate transmembrane transporter activity	5.68E-04	5.10	5	[AT1G08890, AT1G08900, ESL1, GLT1, STP14]
GO:0004693	cyclin-dependent protein serine/threonine kinase activity	6.56E-04	5.75	5	[CDC2, CYCA2;3, CYCA3;1, ORG1, cycp3;1]
GO:0006475	internal protein amino acid acetylation	7.05E-04	6.17	5	[BT1, BT2, BT4, HAF01, HD1]

3h

GOID	GOTerm	Term	%	Nr.	Associated Genes Found
		PValue Corrected	Associated Genes	Genes	
GO:0009753	response to jasmonic acid	5.94E-26	12.22	27	[BT2, BT4, ERF1, GL3, GRX480, JAZ1, JAZ10, JAZ12, JAZ3, JAZ5, JAZ6, JAZ7, JAZ8, MPK4, MPK6, MYB21, MYBR1, MYC2, NINJA, PAP1, PEPR1, PFT1, TIFY10B, TIFY7, TPL, TT4, TT8]
GO:0071456	cellular response to hypoxia	1.41E-10	6.11	16	[AGC2-1, AT4G16660, CML37, CML38, HB2, HSP101, HSP17.4, HSP17.6A, HSP17.6II, HSP70, PAP1, PBP1, RHL41, S6K2, STZ, TCH3]
GO:0009962	regulation of flavonoid biosynthetic process	8.35E-07	17.65	6	[GL3, MYB7, MYC2, PAP1, TT4, TT8]
GO:0009718	anthocyanin-containing compound biosynthetic process	1.82E-06	15.38	6	[DFR, GL3, MYC4, PAP1, TT4, TT8]
GO:0015036	disulfide oxidoreductase activity	1.97E-06	8.25	8	[AT1G06830, AT1G77370, AT2G30540, AT4G15660, ERO1, GRX480, NTRC, TRX5]
GO:0009751	response to salicylic acid	1.38E-05	5.20	9	[BT1, BT2, BT4, BT5, GRX480, KTI1, MPK4, MYBR1, WRKY62]
GO:0004693	cyclin-dependent protein serine/threonine kinase activity	1.28E-04	6.90	6	[CDC2, CKS2, CYCA2;3, CYCA3;1, CYCP4;1, ORG1]

GO:0016573	histone acetylation	7.94E-04	6.17	5	[BT1, BT2, BT4, BT5, HD1]
GO:0006626	protein targeting to mitochondrion	1.33E-03	6.35	4	[AR192, MPPalpha, MYB90, TIM17-1]
GO:0034620	cellular response to unfolded protein	1.48E-03	7.41	4	[BIP3, CNX1, HSP70, HSP70T-2]
GO:0004707	MAP kinase activity	1.48E-03	7.41	4	[MPK11, MPK3, MPK4, MPK6]

24h

GOID	GOTerm	Term PValue	% Associated Genes	Nr. Genes	Associated Genes Found
GO:0009867	jasmonic acid mediated signaling pathway	6.20E-27	20.00	23	[ERF1, GL3, GRX480, JAZ1, JAZ10, JAZ12, JAZ3, JAZ5, JAZ6, JAZ7, JAZ8, MPK6, MYB21, MYBR1, MYC2, NINJA, PAP1, PFT1, SGT1B, TIFY10B, TIFY7, TPL, TT8]
GO:0009753	response to jasmonic acid	7.04E-23	11.31	25	[BT2, BT4, ERF1, GL3, GRX480, JAZ1, JAZ10, JAZ12, JAZ3, JAZ5, JAZ6, JAZ7, JAZ8, MPK6, MYB21, MYBR1, MYC2, NINJA, PAP1, PFT1, SGT1B, TIFY10B, TIFY7, TPL, TT8]
GO:0015036	disulfide oxidoreductase activity	5.24E-10	11.34	11	[AT2G30540, AT2G47870, AT3G21460, AT4G15660, GRX480, GRXC2, NTRC, ROXY2, TH8, TRX3, TRX5]
GO:0009751	response to salicylic acid	1.87E-08	6.94	12	[BT1, BT2, BT4, BT5, GRX480, KTI1, MKK7, MYBR1, WRKY18, WRKY38, WRKY40, WRKY62]
GO:0015144	carbohydrate transmembrane transporter activity	1.72E-07	9.18	9	[AT1G08890, AT5G61520, ESL1, GLT1, INT2, MSS1, PMT5, PMT6, SFP2]
GO:0071456	cellular response to hypoxia	1.48E-06	4.58	12	[AGC2-1, CML37, CML38, HSP101, HSP70, MKK9, PAP1, PBP1, RHL41, S6K2, STZ, TCH3]
GO:0004707	MAP kinase activity	1.39E-05	11.11	6	[MKK7, MKK9, MPK11, MPK3, MPK6, PTP1]
GO:0009738	abscisic acid-activated signaling pathway	2.01E-05	4.42	10	[BT2, CEN2, HAI1, HFR1, KEG, MPK3, MPK6, NF-YB6, PP2CA, PRN]
GO:0009962	regulation of flavonoid biosynthetic process	2.26E-05	14.71	5	[GL3, MYB7, MYC2, PAP1, TT8]
GO:0023014	signal transduction by protein phosphorylation	4.57E-05	6.48	7	[МКК7, МКК9, МРК11, МРК3, МРК6, РНҮА, РТР1]

GO:0016573	histone acetylation	8.85E-05	7.41	6	[BT1, BT2, BT4, BT5, HAF01, HD1]
GO:0009863	salicylic acid mediated signaling pathway	3.82E-04	7.25	5	[GRX480, MKK7, MYBR1, WRKY38, WRKY62]
GO:0010017	red or far-red light signaling pathway	2.05E-03	6.35	4	[ELF4, HFR1, PFT1, PHYA]
GO:0009734	auxin-activated signaling pathway	2.67E-03	4.40	4	[BT2, MYB73, MYBR1, SGT1B]
GO:1902531	regulation of intracellular signal transduction	2.90E-03	5.19	4	[HSP70, MKK7, MKK9, PTP1]

Table S2. Selected transcriptomic experiments for the analysis of the response of plants to herbivore attack. Accession number of the experiment is provided as Gene Expression Datasets Series (GSE) from the Gene Expression Omnibus (GEO) platform, as E-MTAB accession number from ArrayExpress database, as SRP accession number from the Sequence Read Archive (SRA) or as NASCarray experiments from the Nottingham Arabidopsis Stock Centre's microarray database. The presence of an asterisk (*) indicates that a re-analysis of the data was performed.

Accession	Year	Herbivore	Type of herbivore	Type of experiment	Infestation time	Plant tissue	Plant age	Ref.
GSE38281*	2014	Liriomyza huidobrensis	Leafminer (Dipteran)	Microarray	4h	Leaves	4 weeks	(Zhang et al. 2012)
GSE39245*	2013	Brevicoryne brassicae	Aphid	Microarray	72h	Rosettes	4.5 weeks	(Barah et al. 2013)
E-MTAB- 3223	2015	Myzus persicae Myzus cerasi Rhopalosiphum pisum	Aphids	Microarray	3, 6, 24h	Rosettes	4 weeks	(Jaouannet et al. 2015)
NASC-330*	2005	Frankiniella occidentalis	Thrip	Microarray	24, 48h	Leaves	5 weeks	(De Vos et al. 2005)
GSE6516	2006	Bemisia tabaci	Hemiptera	Microarray	21d	Leaves	4 weeks	(Kempema et al. 2007)
GSE114211	2019	Pieris brassicae Mamestra brassicae	Lepidopterans	Microarray	48h	Leaves	8 weeks	(Oberländer et al. 2019)
E-MTAB- 1418	2013	Spodoptera littoralis	Lepidopteran	Microarray	8d	Leaves	4 weeks	(Schweizer et al. 2013)
SRP073212	2016	Pieris rapae	Lepidopteran	RNA-seq	3, 6, 12, 24h	Leaves	5 weeks	(Coolen et al. 2016)
SRP144249	2018	Brevipalpus yothersi	Mite	RNA-seq	6h, 2d, 6d	Leaves	4 weeks	(Arena et al. 2018)
-	2020	Tetranychus urticae	Mite	RNA-seq	30 min 1, 3, 24h	Rosettes	4 weeks	-

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Table S3. Individual genes significantly deregulated in clusters 3 and 4.

Gene ID	Cluster 1	Cluster 2	Cluster 3	Cluster 4	Gene name	Description
AT5G42380	-1.99	-3.28	-0.43	2.72	CML37	Calcium-binding protein CML37 (Q9FIH9)
AT3G01830	-1.89	-2.38	-0.94	2.38	CML40	Probable calcium-binding protein CML40 (Q9SGI8)
AT1G72520	-2.36	-1.11	#N/A	3.24	LOX4	Lipoxygenase 4, chloroplastic (Q9FNX8)
AT3G48520	-2.81	-1.75	#N/A	5.67	CYP94B3	CYP94B3 (A0A178V8H3)
AT4G30280	-2.73	-2.21	#N/A	2.70	XTH18	Xyloglucan endotransglucosylase/hydrolase 18 (Q9M0D2)
AT2G22880	-2.38	-1.46	#N/A	2.47	-	At2g22880 (O81005)
AT2G38240	-2.82	#N/A	-2.04	4.08	ANS	2-oxoglutarate-dependent dioxygenase ANS (O80449)
AT5G05600	-2.73	#N/A	-0.36	2.95	-	2-oxoglutarate-dependent dioxygenase At5g05600 (Q9FFF6)
AT2G27690	-2.81	#N/A	-1.08	2.05	CYP94C1	Cytochrome P450 94C1 (Q9ZUX1)
AT2G24850	-2.04	#N/A	-1.18	2.67	TAT3	Probable aminotransferase TAT3 (Q9SK47)
AT3G44860	-2.64	#N/A	#N/A	2.38	FAMT	Farnesoic acid carboxyl-O-methyltransferase (Q9FYC4)
AT5G13220	-2.59	#N/A	#N/A	2.40	JAZ10	jasmonate-zim-domain protein 10 (cc:AT5G13220)
AT5G63450	-2.86	#N/A	#N/A	2.77	CYP94B1	cytochrome P450, fam 94, subfam B, polypep 1 (AT5G63450)
AT3G25780	-2.06	#N/A	#N/A	3.47	AOC3	Allene oxide cyclase 3, chloroplastic (Q9LS01)
AT5G12340	-2.60	#N/A	#N/A	2.66	-	DUF4228 domain protein (Q1PDX4)
AT1G76470	-2.63	#N/A	#N/A	2.66	-	NAD-binding Rossmann-fold superfamily protein (F4I2E5)
AT2G34600	-2.29	#N/A	#N/A	4.52	TIFY 5B	Protein TIFY 5B (O64687)
AT3G25180	-2.52	#N/A	#N/A	4.34	CYP82G1	Cytochrome P450 82G1 (Q9LSF8)
AT1G61120	-1.89	#N/A	#N/A	3.55	GES	(E,E)-geranyllinalool synthase (Q93YV0)
AT1G28480	-1.81	#N/A	#N/A	2.33	GRXC9	Glutaredoxin-C9 (Q9SGP6)
AT1G66280	-2.60	#N/A	#N/A	2.83	BGLU22	Beta-glucosidase 22 (Q9C8Y9)
Gene ID	Cluster 1	Cluster 2	Cluster 3	Cluster 4	Gene name	Description
AT5G21960	-1.30	-1.63	3.68	#N/A	ERF016	Ethylene-responsive transcription factor ERF016 (Q9C591)
A14G37990	-2.60	-1.41	2.98	#N/A	CAD8	Cinnamyl alcohol dehydrogenase 8 (Q02972)
AT4G15200	-2.73	-2.30	3.20	#N/A	AFH3	formin 3 (cc:AT4G15200)
AT1G33760	-2.10	-2.01	1.11	#N/A	ERF022	Ethylene-responsive transcription factor ERF022 (Q9LQ28)
AT1G/0130	-2.04	-2.34	1.01	#N/A	LECRK52	L-type lectin-domain containing receptor kinase V.2 (004533)
AT4G13395	-1.78	-2.07	2.30	#N/A	DVLIU	
AT3G11480	-2.36	#N/A	1.52	#N/A	BSIMIT	Salicylate/benzoate carboxyl methyltransferase (Q6XMI3)
AT4G11320	-2.69	#N/A	3.11	#N/A	RDL5	Probable cystelle protease RDL5 (Q95059)
AT1G52030	-2.52	#N/A #N/A	1.11	#N/A #N/A		Myrosinase-binding protein 2 (Q9SAV1)
AT4G21830	-1.01	#IN/A #NI/A	1.02	#IN/A #NI/A		Peptide methionine suifoxide reductase B7 (Q8V186)
AT1G54010	-2.45	#IN/A #NI/A	1.70	#IN/A #NI/A	GLL23	Inactive GDSL esterase/Ilpase-like protein 23 (Q8W4H8)
AT5G24770	-2.73	#IN/A #NI/A	2.04	#IN/A #NI/A	VSP2	vegetative storage protein 2 (082122)
A14G24340	-2.30	#IN/A #NI/A	1.74	#IN/A #NI/A	-	Al4g24340 (Q8GW53)
ATTC24420	-2.13	#IN/A #NI/A	0.74	#IN/A #NI/A		Runitz family trypsin (Q9FX28)
AT5G24420	-2.00	#IN/A #NI/A	0.70	#N/A #N/A	PGL5	Alpha (heta Uudrolassa superfemily protein (OE01C7)
AT4G17470	-2.04	#IN/A #NI/A	2.11	#IN/A #NI/A	-	Alpha/beta-Hydrolases superfamily protein (Q501G7)
AT2G24210	-2.04	#IN/A #NI/A	3.80	#IN/A #NI/A	15210	Construction of the system of
AT4G35160	-2.05	#N/A	2.09	#N/A #N/A		O-methyliransierase family protein (Q91003)
AT4G57410	-2.71	#IN/A #NI/A	2.15	#N/A #N/A		Cytochionie P430 81F4 (Q33201)
AT5G07010	-5.10	#IN/A #NI/A	0.40	#N/A #N/A	30115	Sunotransierase (ADA1780G05)
AT4G11911	-3.10	#IN/A #NI/A	2.45	#N/A #N/A	-	DINC (11 hav domain containing protain (ASMS72)
AT2G44578	-2.50	#IN/A #NI/A	1.92	#N/A #N/A	-	Ring/0-box domain-containing protein (A8MS73)
AT1614120	-5.29	#IN/A #NI/A	2.37	#N/A #N/A	-	KITIIC-TIKE CUPINS SUPERTAMILY PROTEIN (F4JLIVIZ)
AT2055070	-2.92	#IN/A #NI/A	1.42	#IN/A #NI/A	- IBC21	
ATAC15210	-2.00	#IN/A #NI/A	2.42	#N/A #N/A		JUZI (AUAI/0V/04)
ATEC44420	-3.23 #NI/A	#IN/A #NI/A	_1.42	#N/A #N/A		Deta-annyldse 5 (F25055) Defensin like protein 16 (005122)
ATEC44420	#IN/A #NI/A	#IN/A #NI/A	-1.49	#IN/A #NI/A		
AT2C2C020	#IN/A #NI/A	#IN/A #NI/A	-2.08	#N/A		ruri.zu (AUA178UK4Z)
AT2020020	#IN/A #NI/A	#IN/A #NI/A	-2.73	#IN/A #NI/A		FUF1.20 (AUA170VQC3)
ATZG26010	#IN/A	#N/A	-2.73	#IN/A	40F1.3	rufi.3 (AUA1/8V330)

Gene ID	Cluster 1	Cluster 2	Cluster 3	Cluster 4	Gene name	Description
AT1G43160	-2.88	-1.96	1.21	5.38	RAP2-6	Ethylene-responsive transcription factor RAP2-6 (Q7G1L2)
AT4G27654	-1.93	-1.88	2.41	4.91	-	At4g27654 (Q3E9V5)
AT1G76650	-2.45	-1.79	1.90	2.12	CML38	CML38 (A0A178WMC5)
AT1G80840	-1.77	-1.89	1.09	2.44	WRKY40	Probable WRKY transcription factor 40 (Q9SAH7)
AT3G02840	-1.71	-3.15	0.50	2.56	-	ARM repeat superfamily protein (Q9M8S5)
AT2G02990	-2.60	-1.89	0.95	4.50	RNS1	Ribonuclease 1 (P42813)
AT1G52890	-2.73	-1.75	1.02	2.63	NAC019	NAC domain-containing protein 19 (Q9C932)
AT1G19180	-2.53	-1.32	0.71	2.61	TIFY10A	TIFY10A (A0A178W7G1)
AT1G28370	-2.15	-1.31	1.24	2.65	ERF11	ERF domain protein 11 (cc:AT1G28370)
AT3G23250	-1.57	#N/A	0.81	3.46	MYB15	Transcription factor MYB15 (Q9LTC4)
AT3G43250	-2.30	#N/A	2.05	3.61	-	Coiled-coil protein (DUF572) (Q9LXK9)
AT5G38120	-3.02	#N/A	2.20	1.83	4CLL8	4-coumarateCoA ligase-like 8 (Q84P26)
AT5G19110	-2.84	#N/A	1.91	1.53	-	Eukaryotic aspartyl protease family protein (cc:AT5G19110)
AT3G53600	-2.64	#N/A	1.71	2.24	-	At3g53600 (Q9LFG0)
AT1G51780	-2.60	#N/A	2.98	4.12	ILL5	IAA-amino acid hydrolase ILR1-like 5 (Q9SWX9)
AT1G76640	-2.14	#N/A	2.58	5.15	CML39	Calcium-binding protein CML39 (Q9SRE7)
AT2G22760	-2.38	#N/A	1.51	4.10	BHLH19	Putative uncharacterized protein At2g22760 (C0SV56)
AT1G76790	-2.73	#N/A	0.89	1.76	IGMT5	Indole glucosinolate O-methyltransferase 5 (Q9SRD4)
AT1G17380	-2.32	#N/A	0.81	2.94	TIFY11A	Protein TIFY 11A (Q9LDU5)
AT1G66370	-2.52	#N/A	2.23	2.00	MYB113	Transcription factor MYB113 (Q9FNV9)
AT5G52400	-2.73	#N/A	1.09	4.16	CYP715A1	Cytochrome P450, fam 715, subfam A, polypep 1 (F4KG63)
AT2G44840	-1.62	#N/A	2.90	2.72	ERF13	Ethylene-responsive transcription factor 13 (Q8L9K1)
AT2G14290	-2.73	#N/A	1.47	1.65	-	F-box protein At2g14290 (Q9ZQ60)
AT2G30830	-2.73	#N/A	2.74	4.62	-	2-oxoglutarate and Fe(II)-dependent oxygenase (O80850)
AT4G34410	-2.13	#N/A	1.60	5.73	ERF109	Ethylene-responsive transcription factor ERF109 (Q9SZ06)
AT5G38130	-2.76	#N/A	2.82	2.21	-	At5g38130 (Q6DBD5)
AT4G02360	-2.56	#N/A	1.12	1.83	-	Putative uncharacterized protein AT4g02360 (O81297)

Table S4. Oligonucleotide sequences for H	RT-qPCR analysis. F, forward; R, reverse.
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ID	Nombre	Secuencia	
At2g47190	ATMYB2	CCTCAAACTCCATTCTCTTTGG	F
		TCTTAGGTGTTTGGCTTGCTTT	R
At4g09570	CDPK4	TCAAGAAAATGGCCTTACGAGT	F
		TTCAAAGGTGATTGTTCCACTG	R
At2g06530	VPS2.1	AATGAACAGGCAGATGAACCTT	F
		CTTCTTCCTCATCTCCTTCCAA	R
At4g08500	MEKK1	GGAAGAGATGGGAGCTAGGTTT	F
		CCATCTGGATAGATAGGCGAAG	R
At5g47220	ERF2	GGGTTAATTCCGGTGAACCT	F
		CACGTGTCTCATCACCAACC	R
At1g51800	IOS1	GCGCTACCACGAAAAAGAAG	F
		AGGTTCAAAACCGTGGTGAG	R
At4g23180	CRK10	TAACGGGCGACCATTAGAAC	F
		GGCACGGGTAAAGTCACAGT	R
At3g48360	BT2	AAGAAAAGAAGGCGACGACA	F
		AACGCCTTACAAGGCTCTGA	R
At1g68690	PERK9	CTCCGGTTTCAAATTCTCCA	F
		TGGTGGTGGTGAGGTAGTCA	R
At1g32640	MYC2	TCCGAGTCCGGTTCATTCT	F
		TCTCGGGAGAAAGTGTTATTGAA	R
At5g24770	VSP2	ATGCCAAAGGACTTGCCCTA	F
		CGGGTCGGTCTTCTCTGTTC	R
At5g44420	PDF1.2A	GTTCTCTTTGCTGCTTTCGAC	F
		GCAAACCCCTGACCATGT	R
At2g26020	PDF1.2B	GTTCTCTTTGCAGCTTTTG	F
		TCCACCATTGTTGGTACTT	R
At2g14610	PR1	TCAGTGAGACTCGGATGTGC	F
		CGTTCACATAATTCCCACGA	R
A+1g72260	THI2.1	CTGCCCTTCCAACCAAGCTA	F
ALIG/2200		TGGCATCCACTCACTTGCAT	R