

## **SUPPLEMENTAL MATERIALS**

Jugular-Vein Injection of High-Titer Lentiviral Vectors Does Not Transduce the Aorta

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## Major Resources Table

In order to allow validation and replication of experiments, all essential research materials listed in the Methods should be included in the Major Resources Table below. Authors are encouraged to use public repositories for protocols, data, code, and other materials and provide persistent identifiers and/or links to repositories when available. Authors may add or delete rows as needed.

### Animals (in vivo studies)

Species	Vendor or Source	Background Strain	Sex	Persistent ID / URL
Mus musculus	The Jackson Laboratory, Bar Harbor, ME	C57BL/6J	Male	<a href="https://www.jax.org/strain/000664">https://www.jax.org/strain/000664</a>

### Genetically Modified Animals

	Species	Vendor or Source	Background Strain	Other Information	Persistent ID / URL
Parent - Male					
Parent - Female					

### Antibodies

Target antigen	Vendor or Source	Catalog #	Working concentration	Lot # (preferred but not required)	Persistent ID / URL
GFP	Invitrogen, Carlsbad, CA	A11122	1:1000	2015993	<a href="https://www.thermofisher.com/antibody/product/GFP-Antibody-Polyclonal/A-11122">https://www.thermofisher.com/antibody/product/GFP-Antibody-Polyclonal/A-11122</a>
Rabbit IgG	Vector Labs, Burlingame, CA	PK-4001	1:1000	ZE0717	<a href="https://vectorlabs.com/vectastain-abc-kit-rabbit-igg.html">https://vectorlabs.com/vectastain-abc-kit-rabbit-igg.html</a>

### DNA/cDNA Clones

Clone Name	Sequence	Source / Repository	Persistent ID / URL

### Cultured Cells

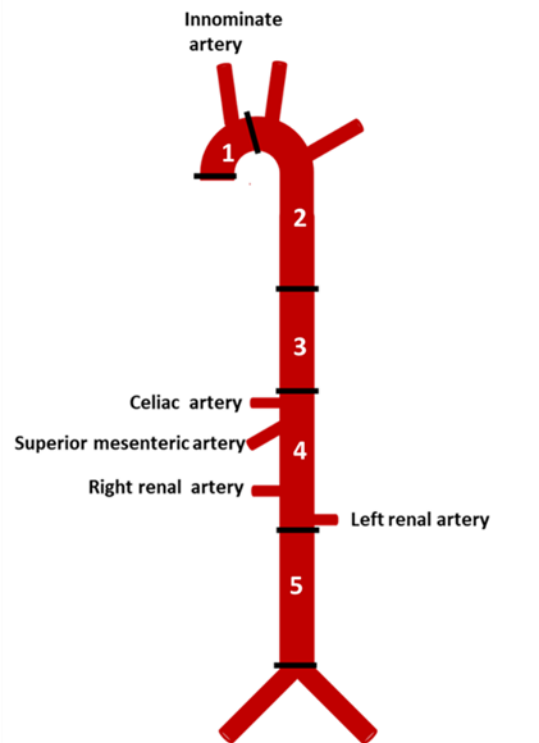
Name	Vendor or Source	Sex (F, M, or unknown)	Persistent ID / URL

### Data & Code Availability

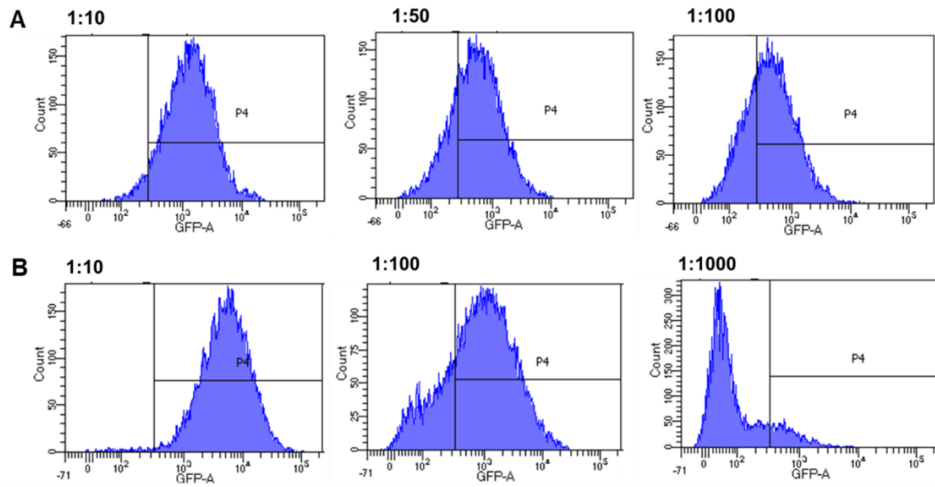
Description	Source / Repository	Persistent ID / URL

**Other**

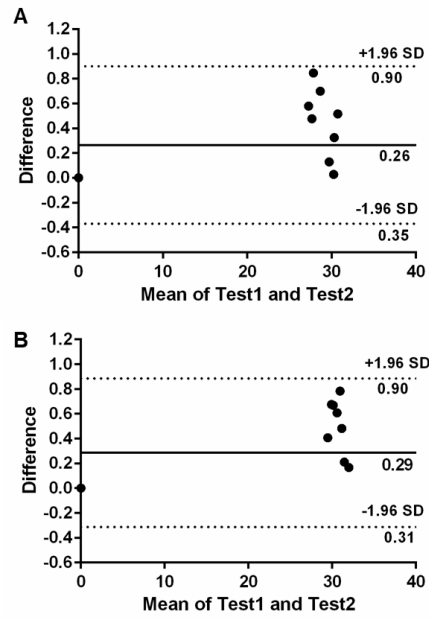
Description	Source / Repository	Persistent ID / URL



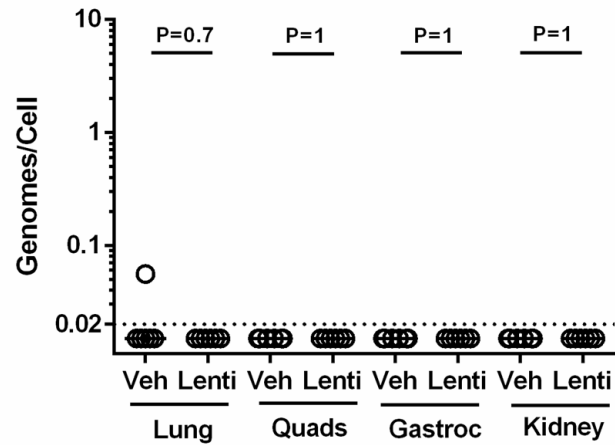
**Figure I. Sectioning protocol for mouse aortas.** Aortas were divided at 6 locations (black bars). Segments 2 and 4 were processed for DNA extraction. Segments 1, 3, and 5 were processed for immunostaining.



**Figure II. Titration of green fluorescent protein (GFP)-expressing lentivirus (LV-GFP) by flow cytometry.** Live Jurkat cells were analyzed for GFP fluorescence after exposure to LV-GFP either obtained from Dr. Juan Miguel Redondo (A) or prepared by the Fred Hutch viral vector core (B). The dilutions of LV-GFP used to generate each histogram are indicated.



**Figure III. Bland-Altman plots comparing repeat measurements of vector DNA.** Extracts of (A) spleen and (B) liver (n=8 each), removed 4 days after injection of a lentiviral vector expressing GFP, were analyzed by quantitative PCR that detects the GFP sequence. The plots compare quantification cycle (Cq) values of duplicate reactions performed independently.



**Figure IV. No transduction of lung, muscle, or kidney after in vivo lentiviral vector (LV) injection.** LV genomes per diploid cell were measured by quantitative PCR for the GFP sequence, performed on DNA extracted from indicated tissues, with reference to a standard curve (see Figure 1A in the main manuscript). Data are from left-sided lung, muscles and kidney; for right-sided results, see Figure 1 in the main manuscript. Dotted line=limit of detection; data points are from individual mice; P values are from Mann-Whitney rank-sum test. Veh=vehicle-injected (n=6 for all tissues); Lenti=lentivirus-GFP injected (n=8 for all tissues); Gastroc=gastrocnemius; Quads=quadriceps.