

Supplementary information

Specific residues in the cytoplasmic domain modulate photocurrent kinetics of channelrhodopsin from *Klebsormidium nitens*

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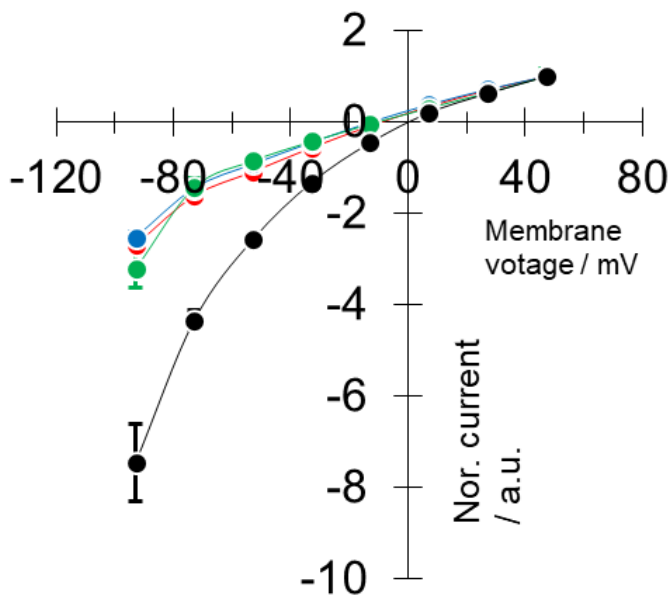
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CrChR2	1	--MDYGGALSAVGRELLFVFNPPVVVNG-SVLVPEDCQYCAGWIESRGT--NGAQTASN	54
PsChR	1	-----MGFQLNPEYLNE-TILLDD---CTPIYLVNVP--LWEQKVARG	37
KnChR	1	MTVDAHSTVDA-----HSTVTDH-SAGNGTESCVAFLGMHHSHEGALYSVYKS	51
GtCCR4	1	-----MTTSAPSLSDPNWQYGMGGWNNPRLPNFNL-HDPTVIG	37
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BR	8	PEWIWALGALMGLGLTYFLVFKMGVSDPAKIFYAITTLVP-----AIAFTMYLSM	60
CrChR2	55	LOWLAAG-----PSILLMFYAYOTWKTSCGWEEIYVCAIEMV-----KVILEFFFEF	102
PsChR	38	TCWFGVI-----LSLAFLIYYIQQYKATCWEELYVCTIEFC-----KIVILELYFEF	85
KnChR	52	LEWGCFL-----ISIGLFVFLYLVITRKTAGWEVVIYIAFIESE-----KYIFEIFWPH	99
GtCCR4	38	VWNLGFL--CLLGLASLALMYKLMSPFKGPD-DQEFFVGYREERCLSIYVNLIAAITWGR	94
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BR	61	LLGYGLTMVPPFGGEQNPIYWARVADWLFITLLELLDALLVDADQG---TIALVGDGI	117
CrChR2	103	---KNPSMLYL-ATGHRVQWLRVYAEWLLTCEVILIHNSNLTGLSNDSRRTMGLLVS DIG	158
PsChR	86	---SPAMIYQ-TNGEVPWLRVYAEWLLTCEVILIHNSNITGLNDDYSGRTMSLITSDLG	146
KnChR	100	---NNPAQLNIYGVNKSVPVWVRYVMEWMITCEVILMAISNISGEEGYTHRSMLLATDQG	151
GtCCR4	95	IC---AHFNNDMGLSLSVNYPKLYLIEFTCEIITLDELWLSNLPYKITYSLFV---GL	146
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BR	118	MGTGLVGLTKVYSYRFVWNAI STAAMLYILYVLFPGFTSKAESM--RP--EVASTFKV	173
CrChR2	159	TIWVGATSAM---ATGYVKVIFPCLGCGANTFFHAARAYIEGYHTVPKGRCRQVVTG	214
PsChR	142	GLCMVVTSALE---SKGWLKWLFEVIGCCYGASTFYHAALIYIESYTTPHGVCKNMVLA	197
KnChR	157	AALCAITAAA---SEGALSAVFYAIGVCYGITFYFCLOIYIEAYFTLE-TCHSAVKW	211
GtCCR4	147	TEACGVFCNAF-EPPARYLWFMGECFIFAFETWISILRLVYARFQOPLNEDAKKIRAPLKL	205
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BR	174	LRNVTVLMSAYPVVWVLIIGSEGAGIVPLNIE TLTFMVLVSAVVGFLILLRRAIFGEA	233
CrChR2	215	MAWLFVVSNGMFEILFPLGPEGFVLSVYGSTVGHTEIDLMSNCGWLLGHYLRVLIHEH	274
PsChR	198	MAAVFETSWMFEPGLFLAGPEGTNALS WAGSTIGHTVADLLSNANGMIGHFLRLIHKH	257
KnChR	212	MAVIFYAGWLCYCFEFLAGSEGWGNLSYEGSAIGHCTADLLSNANGVMHWIRCOLEEY	271
GtCCR4	206	SLTLYFSINCGYPAWLLTE--FGALSQLAHVMTVIMVVAASVYGFALLKPOLGVDKR	263
<hr/>			
BR	234	EAP-----EP---SAGDGAATS D-----	249
CrChR2	275	ILIHG-----D-----	280
PsChR	258	ILIHG-----DVRRIPI---TVNT-----LGREVTVSCEFVDKE-EED	289
KnChR	272	KETHN-----GQLPHY---SLETRAKMRAEAGHIIAGSLGSLVHVAGOH-----	313
GtCCR4	264	DVWLDLDELKSVRYRDPVVPQIRPSKTRRESR-MEYSEDGDFMRPSKGRRAEGDYMNPRWDHHD	322
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BR	250	-----	249
CrChR2	281	-----	280
PsChR	290	EDERISTRKYANRASFMKMRNDMEQRGIQTRKS-LE---MLAPPPALNDGSIV-LA---	340
KnChR	314	-----HHFHFDEDSMGOGAQIQGQPLAQANERANLKASLLNGNAVNAYNQAV	359
GtCCR4	323	DGRRLPDS-----REMDEQVHEKDKQEIS-STMKQIADLNKQLSAMQESBAV-----	367
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BR	250	-----	249
CrChR2	281	-----	280
PsChR	341	VADPMTLTFPTQQLSQ-LDATIRATPAMGQGLEQVLEKGGFDGVLVSP EYIQVGLVQ-	398
KnChR	360	QAQPQAFQIQSGEDGYVISMNQGESASGSRRTTQNTIEGP--RKDPVASAFSKALGSMKS	417
GtCCR4	368	-----	367
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BR	250	-----	249
CrChR2	281	-----	280
PsChR	399	RLKDRKYHMPVYAFGWGKSSPWRSVIEGSGVDCWLEGPYFGSTFDTDALSDAIAEMQRIKT	458
KnChR	418	KVTEHFTKDLPPS-ARPDSPSRGRSARGGADAPTAQPQSMKIDLRD---LANNPQLQQ	472
GtCCR4	368	-----	367
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BR	250	-----	249
CrChR2	281	-----	280
PsChR	459	SYSMVVNGVGMNAGAMN---GVGMN---GMGMDGVGMNAGAMNAGMNGMGGYGSVG	509
KnChR	473	QLSMMLKGQMGAGGYALKTDLKDLANNPQLQQLSMMLKGQMGAGNSNTN-----QGSNN	526
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BR	250	-----	249
CrChR2	281	-----	280
PsChR	510	NNMHSPMMSQ---QAVM-MPQSA---PQMMGMSQTQPAMMGGMQGASPHYVGNLQNM	563
KnChR	527	GSPRARPVSELAQPARLQKAQSA RATSNLDDIEEQAPAQMSRRPSA---RVPRAT--	580
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BR	250	-----	249
CrChR2	281	-----	280
PsChR	564	EQQQAVGSPQVLASSWOQSALHGGMQQQQYGVQVQMPMMVGMQTPAS---PGGVQTPPH	620
KnChR	581	---AAADEEQAPARTLSRKRISRGDGGEPVSEERLQKRPSYRAAKAAAEAADEDMTDAPAP	637
GtCCR4	368	-----	367
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BR	250	-----	249
CrChR2	281	-----	280
PsChR	621	TMAGQPOMSPQOLOQLYFMQOLOQROLOQ---	650
KnChR	638	ALARRPSARAPAADADE-ETAPVRTASLRRAGSKSPGNSLPSSPSSRRVSRANSLRAPVSA	696
GtCCR4	368	-----	367
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BR	250	-----	249
CrChR2	281	-----	280
PsChR	651	---QQYGGTGQR---	660
KnChR	697	SVLLCDADGDMGDFFLDQFAAVPNSKCSVKVATPDELDALATGRAYDFVMVPKGVITS	756
GtCCR4	368	-----	367
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BR	250	-----	249
CrChR2	281	-----	280
PsChR	661	-----	660
KnChR	757	DPEISEEIRAHQATPLVAFGPALGMAVAEYKAHMFELVDDYIGLTKRGEYFVEDLEA	816
GtCCR4	368	-----	367
<hr/>			
BR	250	-----	249
CrChR2	281	-----	280
PsChR	661	-----	660
KnChR	817	LWVKYKKVADDSNKD	831
GtCCR4	368	-----	367

Supplementary Figure 1.
Full amino acid alignments of bacteriorhodopsin (BR), CrChR2, KnChR and GtCCR4
 The transmembrane region was shown based on the membrane topology of CrChR2. Putative FimV domain in KnChR was indicated.

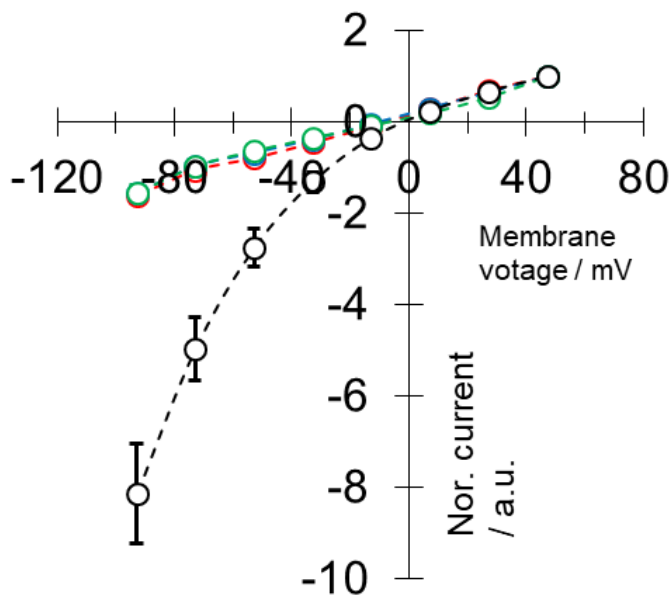
FimV	1	MVRLRLTVRAIAAASVLTSGMAHGLGLGEITLKSALNQPLDAEIEELLEVRDLGSGEVIPS	60
KnChR-FimV	1	-----KALG-----	4
		: **	
FimV	61	LASPEEFKAGVDRLYYLTDLKFTPVVVKPNGKSVIRVTSSKPVQEPYLNFLVQVLWPNGR	120
KnChR-FimV	5	-----SMKSK-----	9
		: : :	
FimV	121	LLREYTVLLDEPLYSPQAAAAPQAEVVSAPRATGAPRAQCAPAPVRTTAPAGSDTYRTVS	180
KnChR-FimV	10	VTEHFTKDLPE-S-----ARPDSESRRGSRGGADAPTA-----Q	43
		: . . : * * * : * : * : * * . * * * .	
FimV	181	NTLWEIAQRNRTDRVSPQAMLAFOELNPGAFVDGNINRLKSGOVLRIETPEOQMLERSP	240
KnChR-FimV	44	PQOSMKIDLRDLA---NNPQLOOQLSMLKQOMAGGYALKTDLKDLANNPQLOOQLSMM-	99
		: : * * : : . ** : : * . . * : . : : * ** *	
FimV	241	REALSQVQAQNSWRGSRNPAAGSAGARQLDATQRNAAGSAPSKVDATDNLRLVSGEGKA	300
KnChR-FimV	100	-----LKG-----GOMGA-----GNSMT	111
		: * * * : * * * : * : : *	
FimV	301	SKGADKGGKGDASKAIADTLAVTKESL-DSTRRENEELQSRMODL-QSQLD-----KIQQL	353
KnChR-FimV	112	NOGSNNGSP-RARPVSEELAQPARLQKAQASARATSNLDEIEEQAPAQMSRRPSARVPA	170
		: * : : * . : : : * * . : * * * : : : * : : : * : : : * : : :	
FimV	354	IQLK---DAQLAKLQGLGAEQOGAA-QPNAALPD---ASQPNAQAQAPAPQPTPAAAP	406
KnChR-FimV	171	TAAADEEQAPARTLSRKIS-RGDGCEPVSEERLQKRPSYRAAKAAAEAA---DEDMTDAP	226
		: * . * . : : . * : * . : * * : * * : * * : * * : * * :	
FimV	407	TPA---PAGEAPAAPAQPPVAPP---APAAEKPPAPAVPAPAPVQAAEQPAPSFIDE	458
KnChR-FimV	227	APALARRPSARAPAADADEETAPVRTASLRRRGSKSPGNSLPSSPSRRV-----	275
		: * * : * * * * * : * * : * * : * * : * * : * * : * * : * * :	
		← Transmembrane region →	
FimV	459	LLANPLWLAVIGGSALLALLVLLMILSRRNAQKEKEEAQAFADTGEEQEDALDLGKDG	518
KnChR-FimV	276	SRANSL-----	281
		* * *	
FimV	519	DDLTLDEPEPQVAAPQVEKTTAQTSDALGEADIYIAYGRFNQAAELLQNAIYDEPORT	578
KnChR-FimV	282	-----	281
FimV	579	DLRLKLMVEVYAEEMGDREGFARQENELREIGGAQPQVEQLKSRYPAMVAVAAVAGLAGAKL	638
KnChR-FimV	282	-----	281
FimV	639	AQDELDSFSLDLSLDDSGHAAKPDAAAGQDLDDAFDLSLDDLGGDDVQADLKS DSGALDD	698
KnChR-FimV	282	-----	281
FimV	699	LTLSDLDLAASTPADKPVDDLDFGLDFAELAETPSQPKHDDLGDGFSLDDLAPEDKLSDD	758
KnChR-FimV	282	-----	281
FimV	759	DFLLSLNDEVPAAPADNEFTLDEAAEPPALSLPDDFDLSLADEPTEPAAPEKGEDSFA	818
KnChR-FimV	282	-----	281
FimV	819	AQLDEVSAQLDELASNLDPEPKSATPSFSAEDA AVASALDGDADDDFD FSPVPTKRRPSWI	878
KnChR-FimV	282	-----	281
FimV	879	WLAPTSTWAIAKARAISSSTKSWPKVMTASRRKPA SCWSAWPDPEMKRPS	927
KnChR-FimV	282	-----	281

Supplementary Figure 2. Full amino acid alignments of FimV from *Pseudomonas aeruginosa* and KnChR. Transmembrane region is indicated.

a Peak

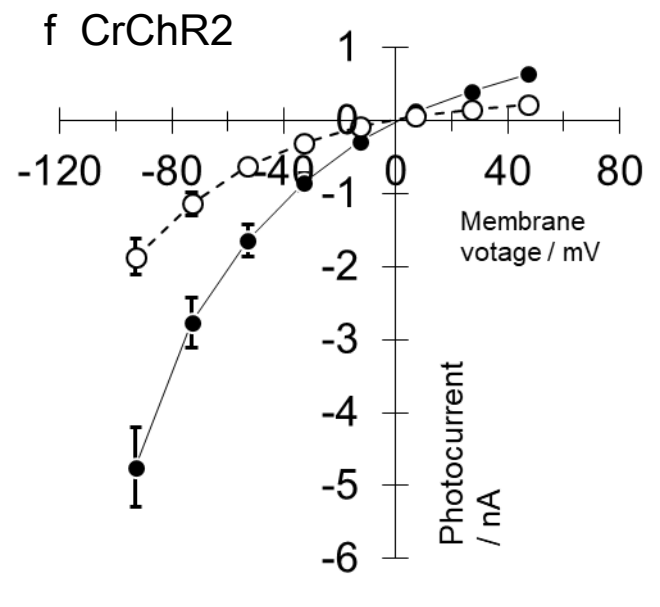
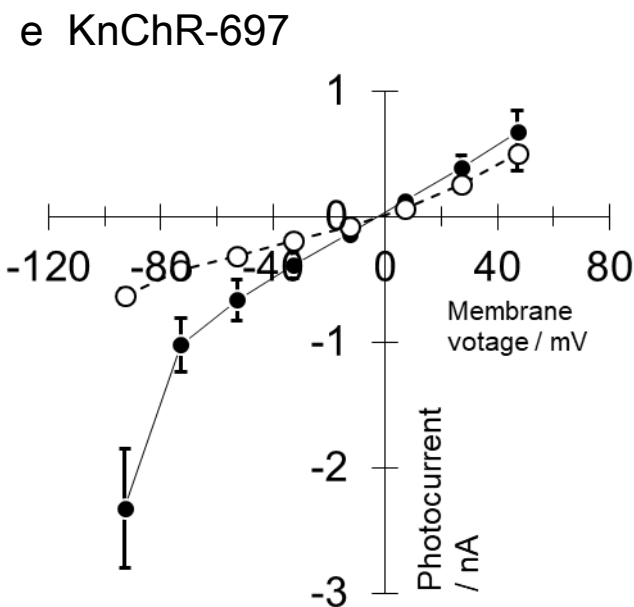
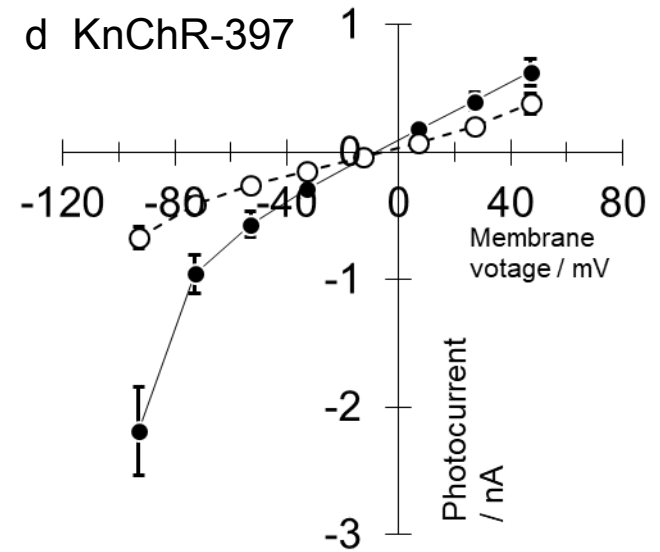
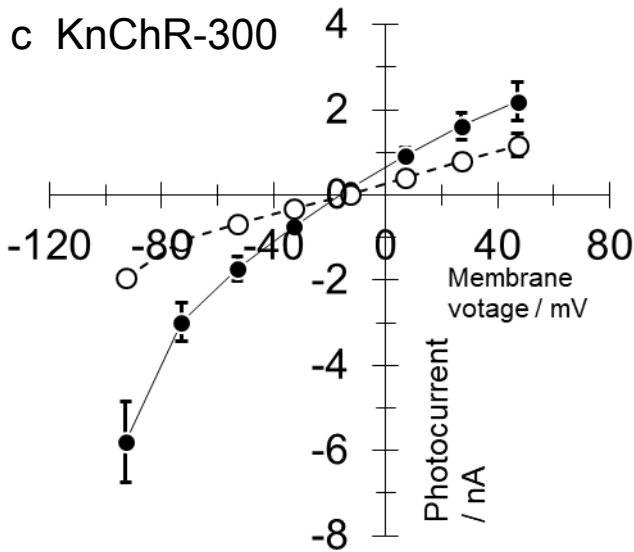
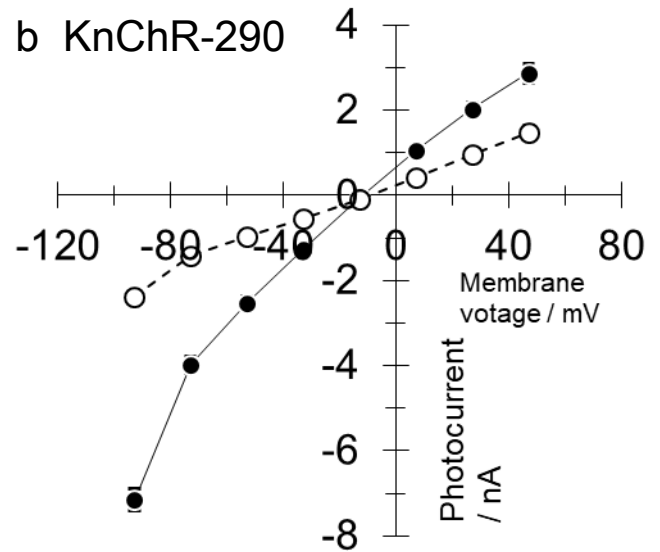
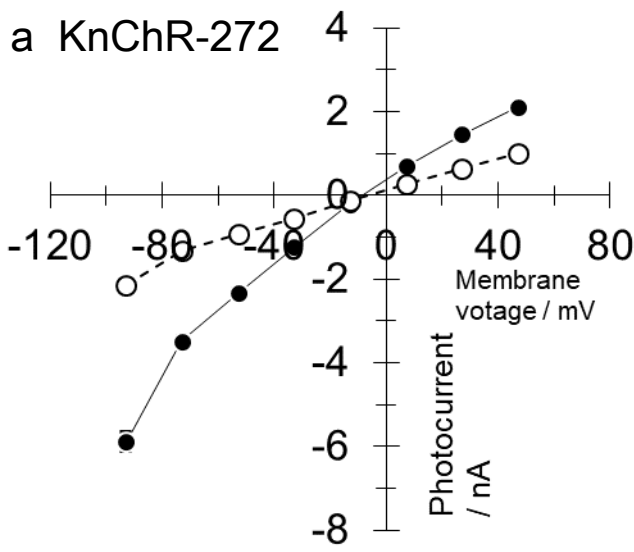


b Steady state



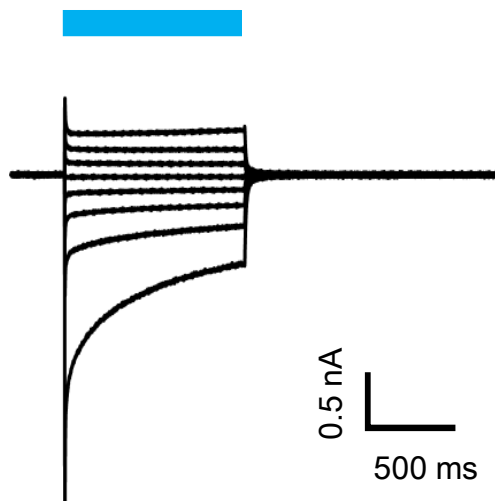
Supplementary Figure 3. I-V relationship of KnChR and CrChR2.

Photocurrent amplitude from each variant and CrChR2 is normalized (at +50 mV as 1.0). Filled (a) and empty (b) symbols indicate peak (I_p) and steady-current (I_s), respectively. Red: KnChR-272 ($N=7$), Blue: KnChR-290 ($N=7$), Green: KnChR-397 ($N=9$), Black: CrChR2 ($N=6$). Standard solutions were used (See *Materials and Methods*). Data were presented as the mean \pm SEM.

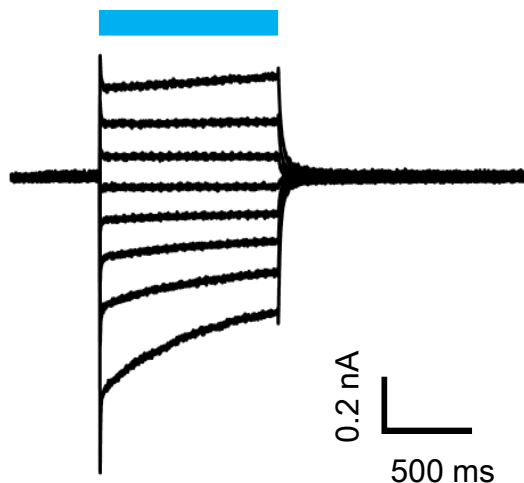


Supplementary Figure 4. I-V relationship of C-terminus length variants of KnChR and CrChR2. Amino acid length of KnChR variants is indicated on each panel (a-e). f. CrChR2. Filled circle, a peak component of the photocurrent (I_p). Empty circle, a steady-state component of the photocurrent (I_s). Standard solutions were used (See *Materials and Methods*). Data were presented as the mean \pm SEM with $N = 6$ (panel c and f), 7 (panels a, b and e), 9 (panel d).

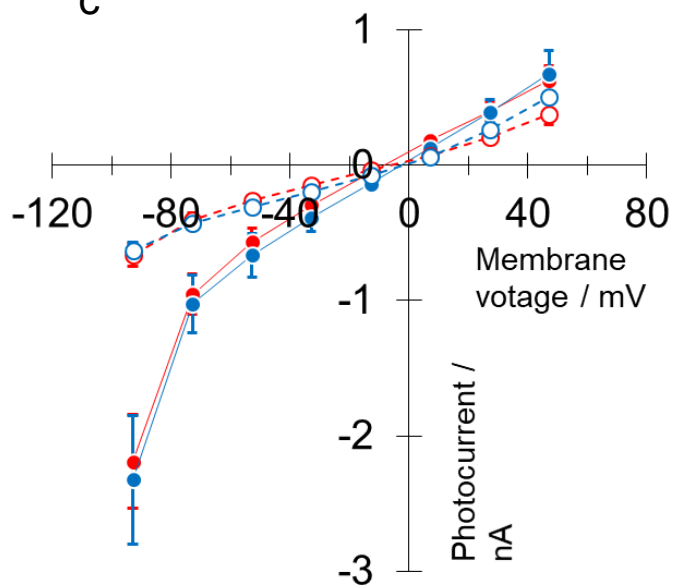
a KnChR-397



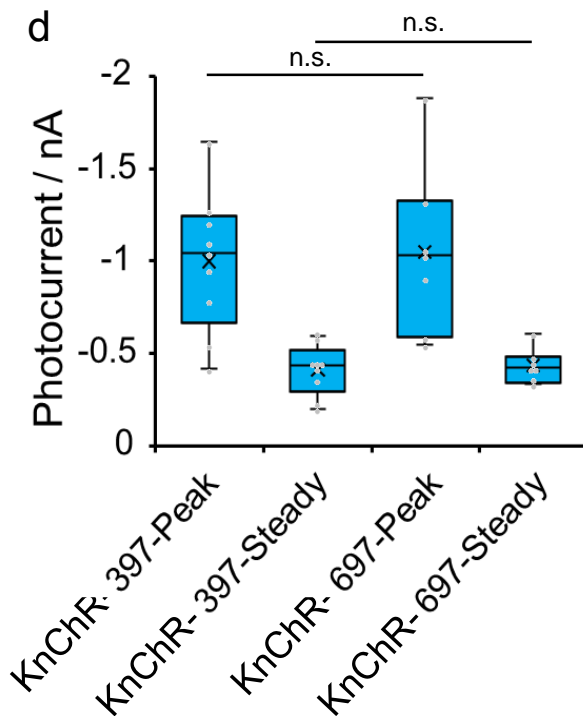
b KnChR-697



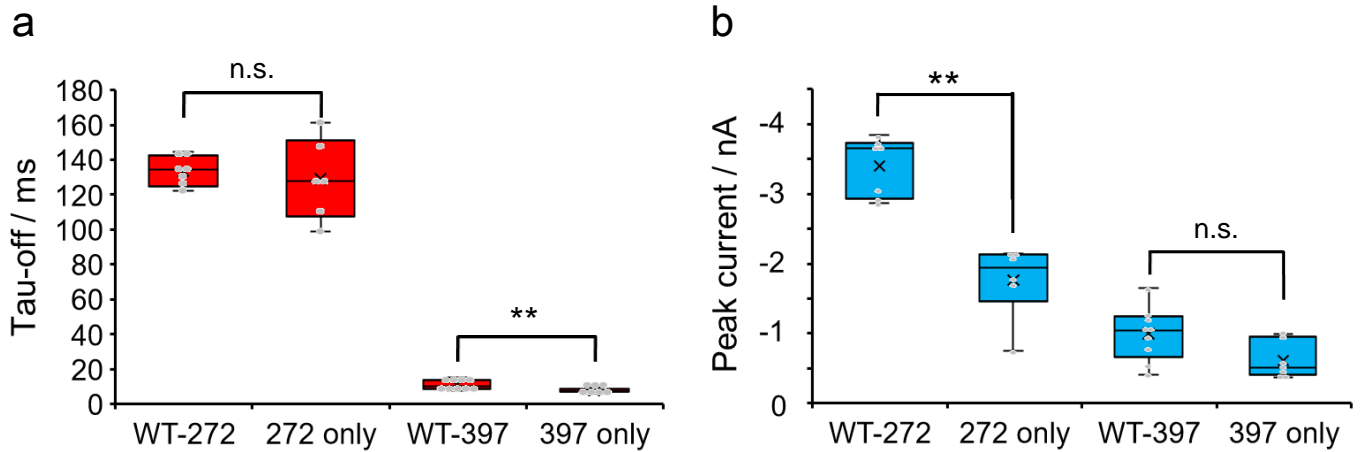
c



d

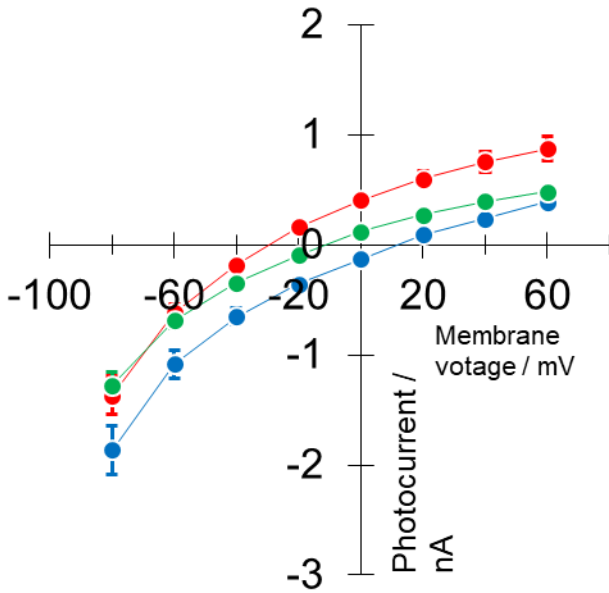


Supplementary Figure 5. Comparison of the length variants KnChR-397 and KnChR-697. Typical photocurrent traces of KnChR-397 amino acids (a) and KnChR-697 amino acids (b). Blue bars indicate light application (480 nm, 12.3 mW/mm²). Membrane voltage was clamped from at -90 to +50 mV by 20 mV steps. Standard solutions were used (See *Materials and Methods*). c, I-V relationship of KnChR-397 (red) and KnChR-697 (blue). Peak component (filled circle) and steady-state component (empty circle) of the photocurrent were depicted. d, Photocurrent amplitudes at -70 mV were shown. Data were presented as the mean \pm SEM with $N=9$ (KnChR-397), and 7 (KnChR-697).

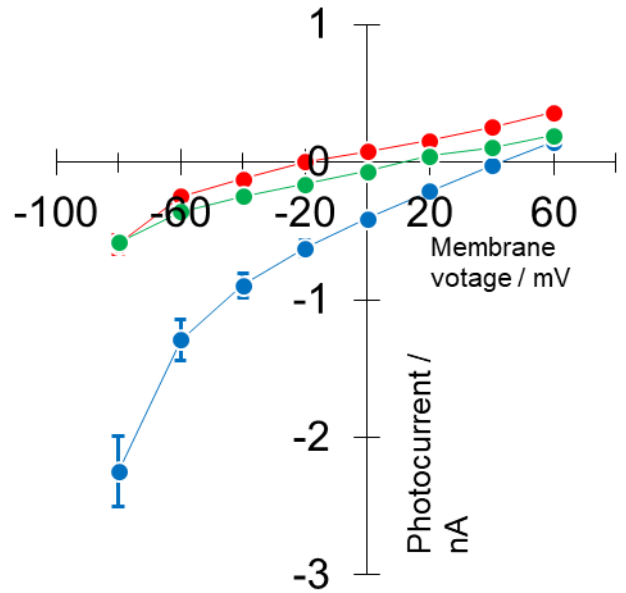


Supplementary Figure 6. eYFP does not alter the channel kinetics of KnChR. Photocurrent amplitudes at -70 mV were analyzed. Time constants (a) and peak current amplitude (b) of KnChR-272 amino acids and -397 amino acids variants with or without eYFP. Standard solutions were used. Data were presented as the mean \pm SEM. $N=7$ (WT-272), 6 (272 only), 9 (WT-397), and 7 (397 only).

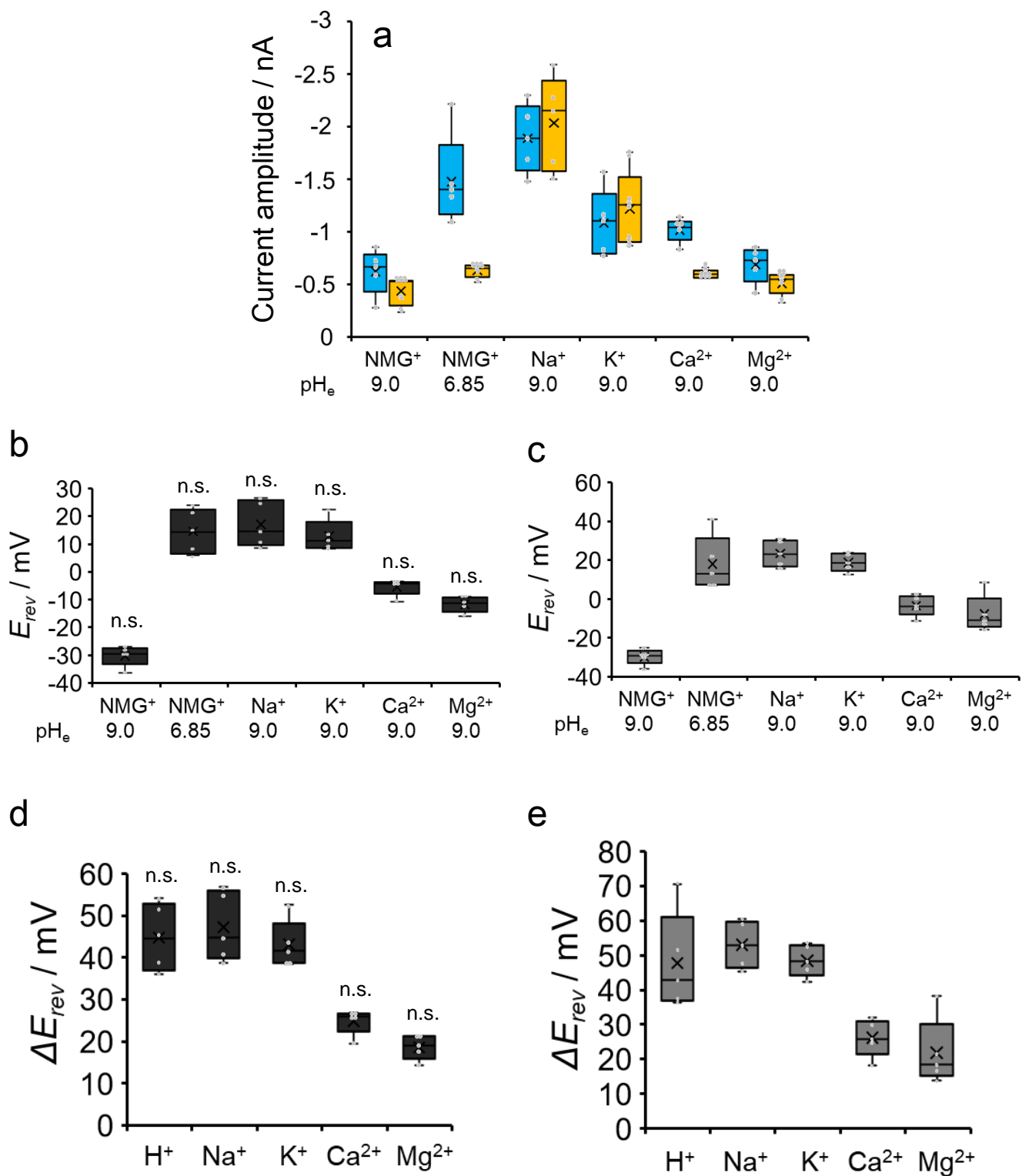
a KnChR



b CrChR2

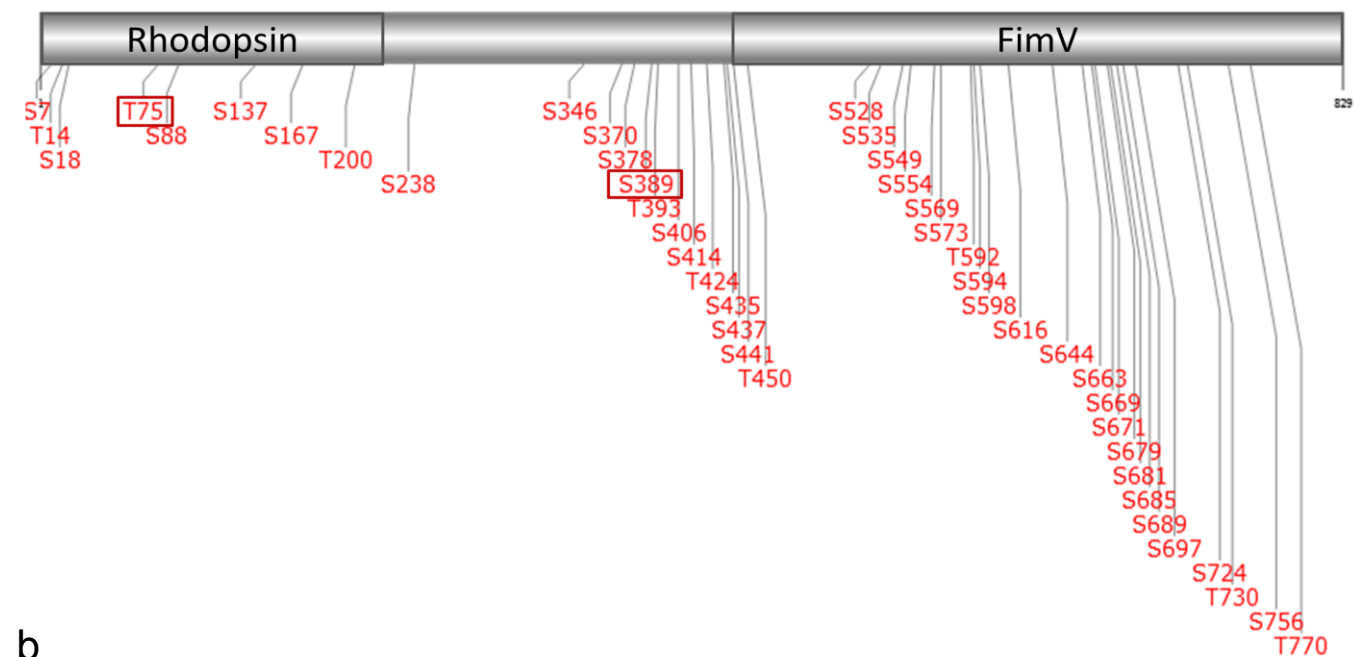


Supplementary Figure 7. I-V relationship of KnChR (a) and CrChR2 (b) in various bath solutions. Red: NMG pH9.0, Blue: K⁺ pH9.0, Green: Mg²⁺ pH9.0. See Supplementary Table 4 for more details of solutions. Data were presented as the mean \pm SEM. $N=5$ (all the conditions for KnChR). $N=8$ (CrChR2 in NMG pH 9.0), 5 (CrChR2 in K⁺ pH 9.0), and 6 (CrChR2 in Mg²⁺ pH 9.0).

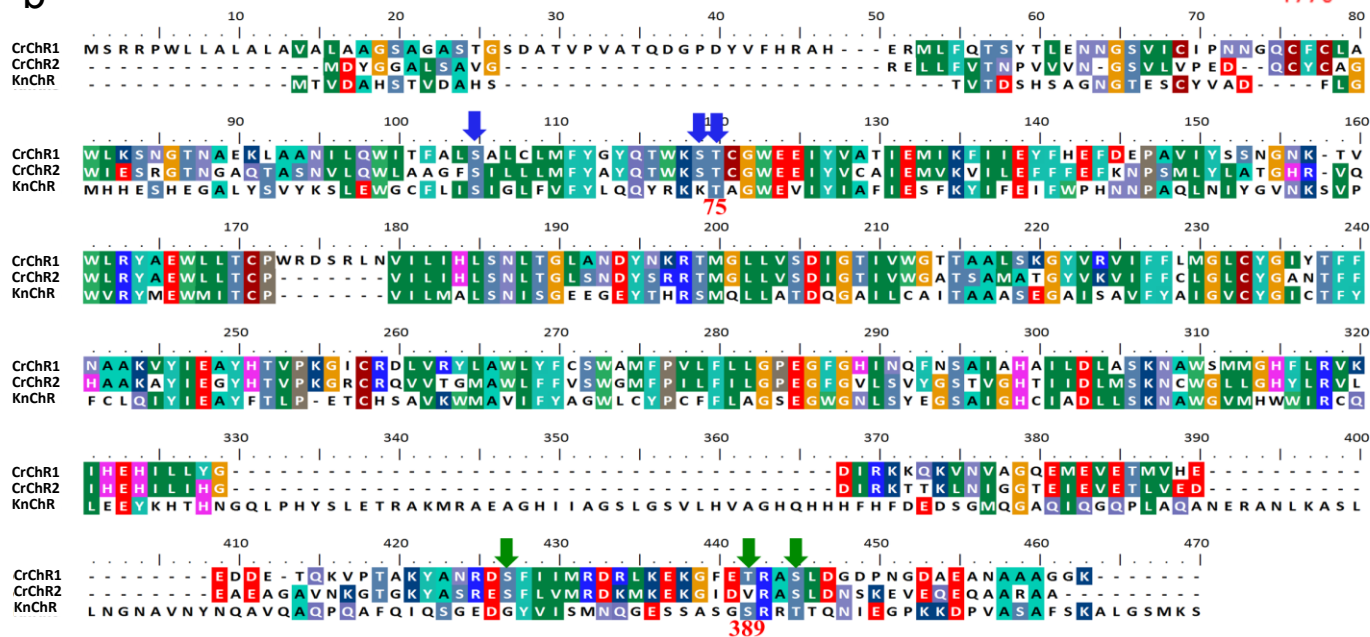


Supplementary Figure 8. Ion selectivity of KnChR. Peak and steady-state component of photocurrents were analyzed. **a**, Photocurrent amplitude at -60 mV under various ionic conditions. blue, peak component. orange, steady-state component. Reversal potential (E_{rev}) of peak component (**b**) and steady-state component (**c**). Shift of E_{rev} , ΔE_{rev} analyzed from peak component (**d**) and from steady-state component (**e**). The Mann-Whitney U test was performed between **b** and **c**, and **d** and **e**. No significant difference was observed between **b** and **c**, and **d** and **e** respectively. Data were presented as the mean \pm SEM with $N=5$.

a Predicted Site:



b



Supplementary Figure 9. Prediction of phosphorylation site in KnChR (a) Prediction of kinase-specific phosphorylation site by GPS 5.0 database. T75 and S389 highlighted in red box correspond to Ser102 and Thr373 in CrChR1 phosphorylation site proposed by Böhm et al. (2019) and Wagner et al. (2008), respectively. (b) Proposed phosphorylation sites in CrChR1; Böhm et al. (2019) suggested multiple phosphorylation sites, one of them (Ser102) being in the first transmembrane domain (TM) and two (Ser 116 and Thr117) in the intracellular vestibule between (TM) domains marked by a blue arrow. One of them (Thr117) was conserved in KnChR (T75 marked by a red number below the sequence). Wagner et al. (2008) proposed three phosphorylation sites (Ser358, Thr373 and Ser376) in CrChR1 and one in CrChR2 (Ser358) marked by a green arrow. Out of three phosphorylation sites, one is common among CrChR1 and CrChR2 (RD/ESpF) but replaced by Gly in KnChR. Ser389 corresponding to Thr373 is marked by a red number (389) below the sequence.

Supplementary Table 1 Solutions for the ion selectivity measurement

		NaCl	KCl	CaCl ₂	MgCl ₂	NMG	EGTA	CHES	MES
Pipette	NMG pH9.0	1	1	2	2	140	10	10	-
Bath	NMG pH9.0	1	1	2	2	140	-	10	-
	NMG pH6.85	1	1	2	2	140	-	-	10
	Na ⁺ pH9.0	140	1	2	2	-	-	10	-
	K ⁺ pH9.0	1	140	2	2	-	-	10	-
	Ca ²⁺ pH9.0	1	1	70	2	-	-	10	-
	Mg ²⁺ pH9.0	1	1	2	70	-	-	10	-

Supplementary Table 2 List of primers used for insertions

Gene	Sense primers	Anti-sense primers
KnChR_cMyc	TCAGAAGAGGATCTGGGGGA TCCAAAGAGC	GATGAGTTTTTGTTCGCTGGC GCTCACAGG
KnChR_272_stop	GTTGGAAGAATACAAGTAAGG GGATCCAAAGAGC	GCTCTTTGGATCCCCTTACTT GTATTCTTCCAAC
KnChR_397_stop	CAACACAGAACATCGAGTAAG GGATCCAAAGAGC	GCTCTTTGGATCCCCTTACTC GATGTTCTGTGTTG

Supplementary Table 3 List of primers used for subcloning into an eYFP-3.0 vector

Gene	Sense primers	Anti-sense primers
KnChR_272 aa	TTCGAATTCGCCACCGCTAGC ATTATGACAGTGGATG	GCTCTTTGGATCCCCCTTGTA TTCTTCCAAGTGGC
KnChR_280 aa	TTCGAATTCGCCACCGCTAGC ATTATGACAGTGGATG	GCTCTTTGGATCCCCCTGGCAA CTGTCCGTTATGAG
KnChR_290 aa	TTCGAATTCGCCACCGCTAGC ATTATGACAGTGGATG	GCTCTTTGGATCCCCCATTTT GGCTCTTGTCTCAAG
KnChR_300 aa	TTCGAATTCGCCACCGCTAGC ATTATGACAGTGGATG	GCTCTTTGGATCCCCGCCAGC AATAATATGCCAG
KnChR_310 aa	TTCGAATTCGCCACCGCTAGC ATTATGACAGTGGATG	GCTCTTTGGATCCCCGCCAGC CACGTGCAAGACGC
KnChR_317 aa	TTCGAATTCGCCACCGCTAGC ATTATGACAGTGGATG	GCTCTTTGGATCCCCGTGAAA ATGATGATGCTGATG
KnChR_321 aa	TTCGAATTCGCCACCGCTAGC ATTATGACAGTGGATG	GCTCTTTGGATCCCCATCCTC ATCAAAGTGAAAATG
KnChR_397 aa	TTCGAATTCGCCACCGCTAGC ATTATGACAGTGGATG	GCTCTTTGGATCCCCCTCGAT GTTCTGTGTTGTCC
KnChR_697 aa	TTCGAATTCGCCACCGCTAGC ATTATGACAGTGGATG	GCTCTTTGGATCCCCGCTGGC GCTCACAGGGGCC
eYFP 3.0 vector	GGGGATCCAAAGAGCAGGAT	GGTGGCGAATTCGAAGCTTG

Supplementary Table 4 List of primers used for site-directed mutagenesis

Mutations	Sense primers	Anti-sense primers
KnChR_397_E285A	GCCACACTATAGCCTTGCGA CAAGAGCCAAAATGA	TCATTTTGGCTCTTGTGCGA AGGCTATAGTGTGGC
KnChR_397_R287A	GTTGCCACACTATAGCCTTG AGACAGCAGCCAAAATGAG G	CCTCATTTTGGCTGCTGTCT CAAGGCTATAGTGTGGCAAC
KnChR_397_K289A	GCCTTGAGACAAGAGCCGC AATGAGGGCTGAAGCTG	CAGCTTCAGCCCTCATTGCG GCTCTTGTCTCAAGGC
KnChR_397_R291A	GAGACAAGAGCCAAAATGG CGGCTGAAGCTGGGCATA	TATGCCCAGCTTCAGCCGCC ATTTTGGCTCTTGTCTC
KnChR_397_E293A	AGCCAAAATGAGGGCTGCA GCTGGGCATATTATTG	CAATAATATGCCCAGCTGCA GCCCTCATTTTGGCT