

## Supplementary Information for

### Characterization of mitochondrial health from human peripheral blood mononuclear cells to cerebral organoids derived from induced pluripotent stem cells

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#### **This PDF file includes:**

Tables S1 to S11

Figures S1 to S2

Clinical and Demographic Characteristics	
Sample Identification	MS005-2014
Sample Type	Whole Blood
Informed consent	Yes
Age	33
Sex	Female
BMI	24.6
Medication	None reported
Ethnicity	Caucasian
BP	120/92

**Table S1. Clinical and demographic characteristics of the donor.** BMI, body mass index; BP, blood pressure.

	Antigen	Host	Dilution	Source	Identifier
iPSC	OCT4	Rabbit	1:400	Invitrogen	A24867
	SOX2	Rat	1:100	Invitrogen	A24759
	SSEA4	Mouse	1:100	Invitrogen	A24866
	TRA-1-60	Mouse	1:100	Invitrogen	A24868
CO	SOX2	Goat	3:200	R&D	AF2018
	NeuN	Rabbit	1:500	Cell Signaling	12943S
Mitochondria	TOMM-20	Rabbit	1:100	Abcam	ab186734
	NDUFS3	Mouse	1:100	Abcam	ab110246
	SDHA	Mouse	1:200	Abcam	ab14715
	UQCRC1	Mouse	1:50	Abcam	ab110246
	COXIV	Mouse	1:200	Abcam	ab33985
	ATP Synthase $\beta$	Mouse	1:500	Invitrogen	A-21351

**Table S2. List of primary antibodies used for fluorescent immunohistochemistry**

**experiments.** OCT4, octamer-binding transcription factor 4; SOX-2, sex determining region Y-box 2; SSEA4, stage-specific embryonic antigen 4; TRA-1-60, podocalyxin; NeuN, neuronal nuclei; TOMM-20, translocase of outer mitochondrial membrane 20; NDUFS3, NADH:ubiquinone oxidoreductase core subunit S3; SDHA, succinate dehydrogenase complex flavoprotein subunit A; UQCRC1, ubiquinol-cytochrome C reductase core protein I; COXIV, cytochrome c oxidase subunit IV; ATP Synthase  $\beta$ , ATP synthase beta subunit; iPSC, induced pluripotent stem cells; CO, cerebral organoid.

	iPSC CO	H9 hESC CO
Electrophysiology	4	4
Cellular IF	2	2
Mitochondria	8	8
TOTAL	14	14

**Table S3. Total number of whole cerebral organoids produced and allocated for each assessment.** iPSC CO induced pluripotent stem cells-derived cerebral organoids; H9 hESC CO, H9 human embryonic stem cells- derived cerebral organoids; IF, immunofluorescence.

Assessment	Sample type	Specific Endpoints	Biological Replicates	Technical Replicates
Characterization	iPSC	Karyotype	2	10
		Alkaline phosphatase	3	3
		Pluripotency validation	3	3
	H9 hESC CO and iPSC CO	Cellular characterization	2	2
Neuronal Activity	H9 hESC CO and iPSC CO	Action potential	4	7-12
		Spontaneous activity	4	4-8
		Na <sup>+</sup> and K <sup>+</sup> currents	4	3-9
Mitochondrial Function	PBMC	MitoTracker	1	3
		MMP	1	3
		OXPPOS	1	3
		ATP	1	3
		mtDNA genetics	1	2
		TEM	1	10
	H9 hESCs and iPSCs	MitoTracker	3	3
		MMP	3	3
		OXPPOS	3	5
		ATP	3	2
		mtDNA genetics	3	1-3
		TEM	2	10
	H9 hESC CO and iPSC CO	MitoTracker	3	3
		MMP	4	3
		OXPPOS	3	5
		ATP	3	3
		mtDNA genetics	3	1-3
		TEM	1	10

**Table S4. Summary of biological and technical replicates used for each experiment.** PBMC, peripheral blood mononuclear cells; iPSC, induced pluripotent stem cells; H9 hESC, H9 human embryonic stem cells; CO, cerebral organoids; MMP, mitochondrial membrane potential; OXPPOS; oxidative phosphorylation; ATP, adenosine triphosphate; mtDNA, mitochondrial DNA; TEM, transmission electron microscopy; Na<sup>+</sup>, sodium; K<sup>+</sup>, potassium.

PCR Primers for Sex Determination		Size
SRY F	CATGAACGCATTCATCGTGTGGTC	254 bp
SRY R	CTGCGGGAAGCAAAGCAATTCTT	
PARP F385	AAGCTTTCCAGGAGATCCTAAC	498 bp
PARP R882	CCTTCACTGTAGGTCCCAAAT	

**Table S5. PCR primers used for sex determination as a quality control.** SRY, sex determining region of Y-chromosome; PARP, poly(ADP-ribose) polymerase; F, forward; R, reverse; bp, base pair.

Sample	PARP	SRY	Sex
PBMC	+	-	F
iPSC	+	-	F
iPSC CO	+	-	F
Controls			
PC – SH-SY5Y	+	-	F
WG1669	+	+	M

**Table S6. Sex determination as a quality control across all stages of cerebral organoid generation.** There is preservation of the female karyotype across PBMC, iPSC and iPSC CO. SH-SY5Y human female neuroblastoma cell line was used as a female control and WG1669 male patient was used as a male control. PBMC, peripheral blood mononuclear cells; iPSC, induced pluripotent stem cells; H9 hESC, H9 human embryonic stem cells; CO, cerebral organoids; PC, positive control; SH-SY5Y, human neuroblastoma cell line; SRY, sex determining region of Y-chromosome; PARP, poly(ADP-ribose) polymerase; F, female; M, male.

Primers for mtDNA amplification for Illumina sequencing	
MT-COIII F	TCACAATTCTAATTCTACTGA
MT-COIII R	CGGATGAAGCAGATAGTGAGG
MT16425 F	CCGCACAAGAGTGCTACTCTCCTC
MT16426 R	GATATTGATTCACGGAGGATGGTG

**Table S7. Primers used for mtDNA amplification and sequencing.** MT, mitochondrial;

F, forward; R, reverse.



	PBMC	iPSC	iPSC-derived CO	NC - SH SY5Y
A73G	+	+	+	-
A153G	+	+	+	-
T195C	+	+	+	-
G225A	+	+	+	-
A263G	+	+	+	+
A750G	+	+	+	+
G769A	+	+	+	-
A1438G	+	+	+	+
G1719A	+	+	+	-
A2706G	+	+	+	-
A4769G	+	+	+	+
T6221C	+	+	+	-
C6371T	+	+	+	-
C7028T	+	+	+	-
A8860G	+	+	+	+
T9722C	+	+	+	-
G11719A	+	+	+	-
C12705T	+	+	+	-
G13368A	+	+	+	-
A13966G	+	+	+	-
T14470C	+	+	+	-
C14766T	+	+	+	-
A15326G	+	+	+	+
G15650A	+	+	+	-
A15924G	+	+	+	-
T16189C	+	+	+	-
C16223T	+	+	+	-
C16278T	+	+	+	-
T16519C	+	+	+	+

**Table S8.** Mitochondrial DNA (mtDNA) variants for the X2g haplogroup across PBMCs, iPSCs and iPSC-derived COs validated by whole mtDNA sequencing and short mtDNA fragment sequencing. SH-SY5Y human neuroblastoma cell line was used as a negative control (NC), validated by only short fragment mtDNA sequencing. (+) denotes positive or presence; (-) denotes negative or absence. PBMC, peripheral blood mononuclear cells; iPSC, induced pluripotent stem cells; CO, cerebral organoids.

PCR Primers		Sequencing primers		SNPs sequenced		
F13464	CAGCCTAGCATTAGCAGGAATA	F14368	CAATCCTACCTCCATCGCTAA C	T14470C	C14766T	
R15211	GTATGGGATGGCGGATAGTAAG					
F458	CCTCCACTCCCATACTACTAA	R1927	CTTAGGTAGCTCGTCTGGTTTC	A1438G	G1719A	
R1997	GGTTTGTCGCCTCTACCTATAAA					
F4708	TCTCCGGACAATGAACCATAAC	R5216	GAGGAGGGTGGATGGAATTA AG	A4769G	C5111T	
R6882	GTGTGGCGAGTCAGCTAAATA	F5993	CACAGCTCTAAGCCTCCTTATT	T6221C	C6371T	
F14562	CCACACCGCTAACAAATCAATAC	F15264	CCACCCTCACACGATTCTTTAC	A15326G	G15650A	A15924G
R16511	GTAGGAACCAGATGTCGGATAC					
F8474	CCCTCACCAAAGCCATAAA	F8804	CACCAACCACCAACTATCT CACTCCATAACGCTCCTCATA C	A8860G		
R11031	CGTGATAGTGGTTCCTGATAAG	F9318		T9722C		
F11644	CCTCGTAGTAACAGCCATTCTC	R11775	CTGTGAGTGCCTCGTAGTT	G11719A		
R14130	GGTTAGGATGAGTGGGAAGAAG	F13595	GCGCCTATAGCACTCGAATAA GAGGTTGAAGTGAGAGGTATG G	A13966G		
		R13451		C12705T	G13368A	
F15323	GCAAACTCCACCTCCTATTC	F458	CCTCCACTCCCATACTACTA A	A750G	G769A	
R943	CTCTTTACGCCGGCTTCTATT	R16548	GGGAACGTGTGGGCTATTTA CCCACACGTTCCCTTAAATA A	T16189C	C16223T	C16278T
		F16536	CGTGAGTGGTTAATAGGGTGA TAGAC	A73G	A153G	T195C
		R34		T16519C	G225A	A263G
F5317	CCACCATCACCTCCTT	F6357	CTAGCAGGTGTCTCCTCTATCT	C7028T		
R7608	CCTACTTGCGCTGCATGTGCC					
F1562	GTAACATGGTAAGTGTACTG	F2209	GCTCAACACCCACTACCTAAA	A2706G		
R3717	GGCTACTGCTCGCAGTG					

**Table S9. PCR primers and sequencing primers used for the determination of the mitochondrial DNA haplogroup, X2g. F, forward; R, reverse; SNPs, single nucleotide polymorphisms.**

PCR Primers for mtDNA Copy Number		
mtDNA	MT-ND1 F	ATGGCCAACCTCCTACTCCT
	MT-ND1 R	CTACAACGTTGGGGCCTTT
Nuclear DNA	$\beta$ 2M F	TGCTGTCTCCATGTTTGATGTATCT
	$\beta$ 2M R	TCTCTGCTCCCCACCTCTAAGT

**Table S10. PCR primers used for mtDNA copy number determination.** MT-ND1, mitochondrially encoded NADH:ubiquinone oxidoreductase core subunit 1;  $\beta$ 2M, Beta 2 Microglobulin; F, forward; R, reverse.

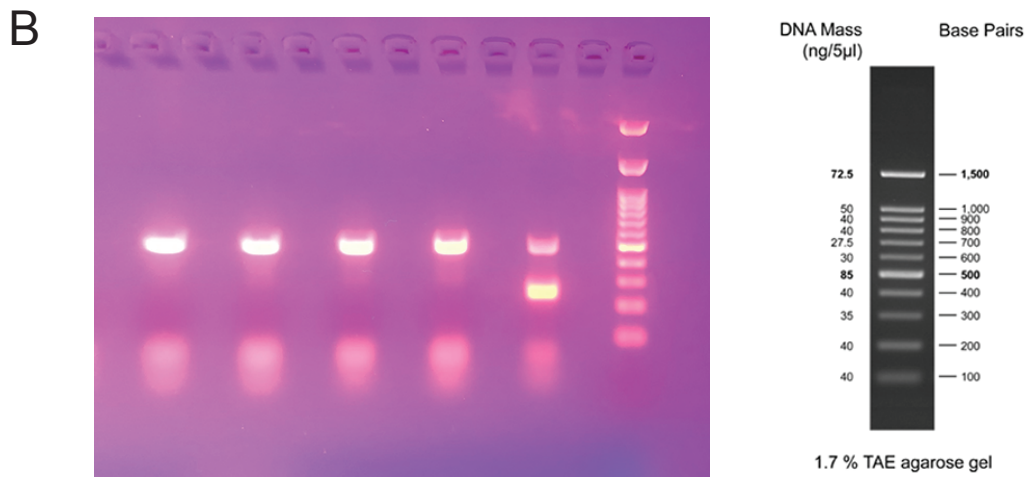
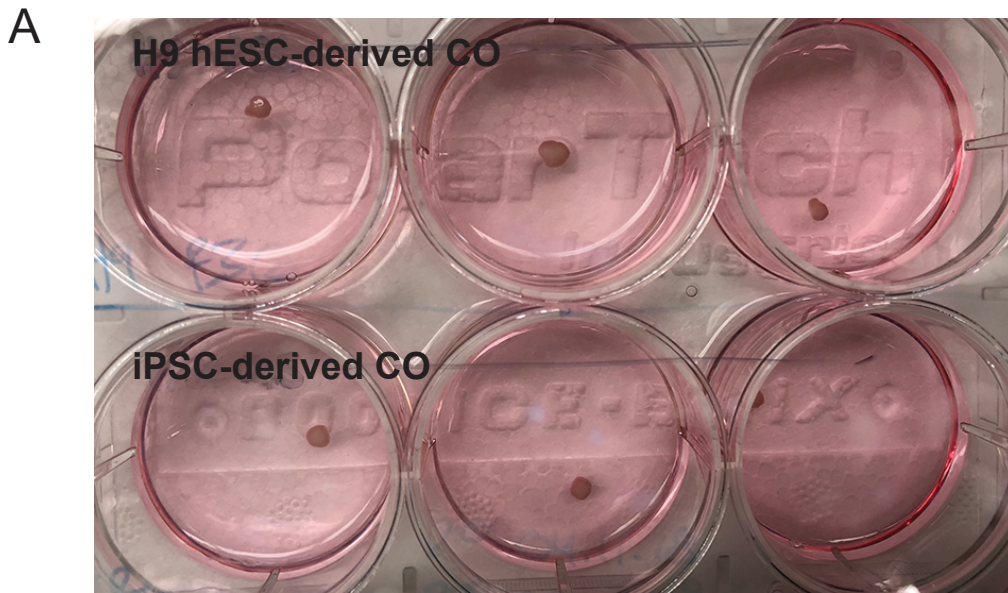
	Resting membrane potential (mV)			Membrane capacitance (pF)			Membrane resistance (M $\Omega$ )		
	Type 1	Type 2	Type 3	Type 1	Type 2	Type 3	Type 1	Type 2	Type 3
iPSC CO	42 $\pm$ 2	42 $\pm$ 3	47 $\pm$ 3	20 $\pm$ 3	29 $\pm$ 3	39 $\pm$ 3	1137 $\pm$ 214	847 $\pm$ 114	558 $\pm$ 48
H9 hESC CO	41 $\pm$ 4	47 $\pm$ 2	56 $\pm$ 3	22 $\pm$ 3	29 $\pm$ 3	32 $\pm$ 2	834 $\pm$ 175	577 $\pm$ 74	574 $\pm$ 102

**Table S11. Electrophysiological properties of neurons from iPSC-derived CO and H9**

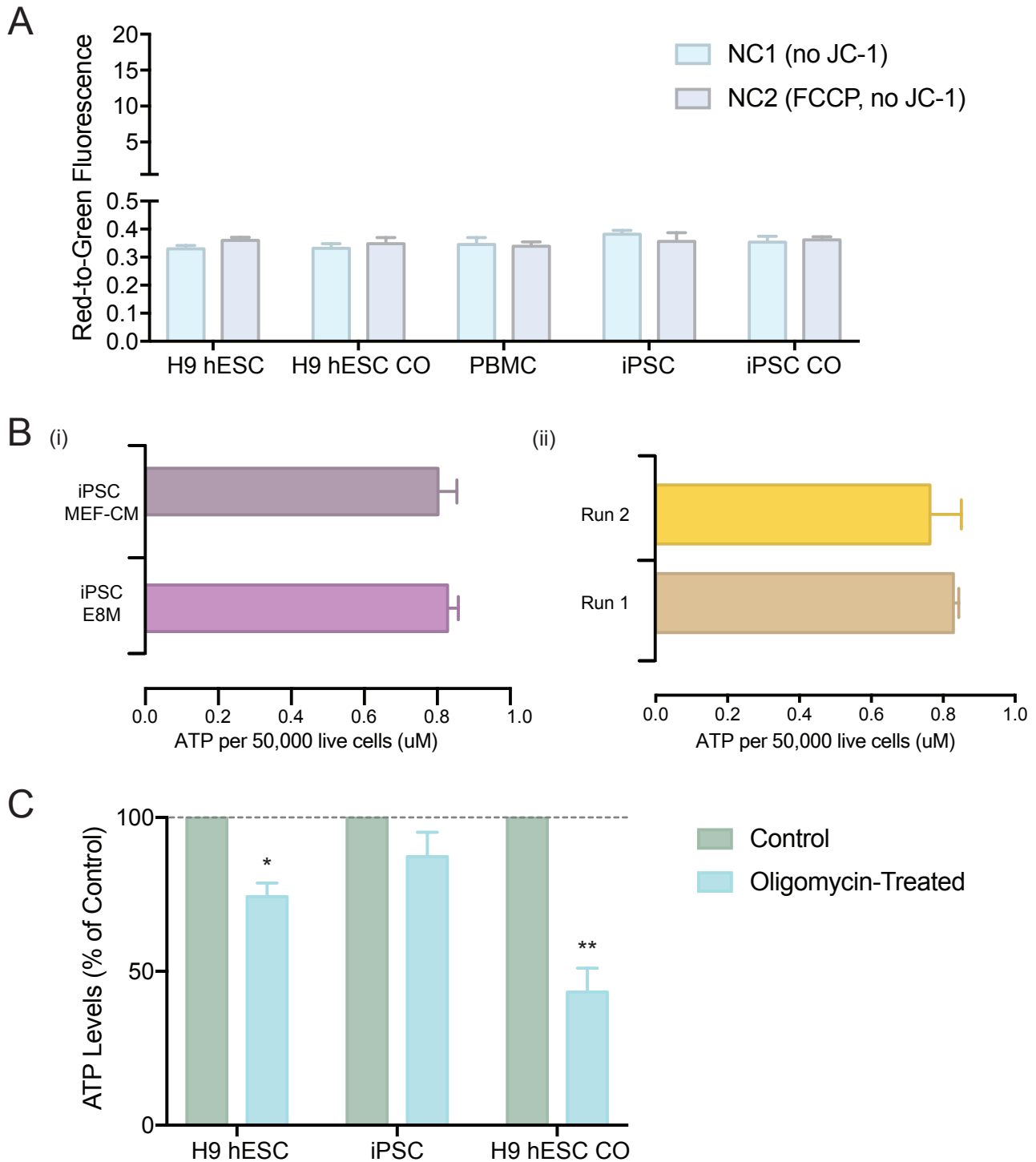
**hESC-derived CO.** Type 1, immature neurons; type 2, developing neurons; type 3, mature

neurons; iPSC, induced pluripotent stem cells; H9 hESC, H9 human embryonic stem cells; CO,

cerebral organoids.



**Fig. S1. Sex determination as a quality control.** (A) Representative cerebral organoid culture in a 6-well plate, H9 hESC-derived CO (top row) and iPSC-derived CO (bottom row). (B) A representative gel image of SRY and PARP PCR products. Left to right: PBMC, iPSC, iPSC-derived CO, SH-SY5Y neuroblastoma cell line, a male control and 100bp ladder.



**Fig. S2. Mitochondrial function experimental quality control and proof-of-concept.** (A) Bar graph summarizing mitochondrial membrane potential as red-to-green fluorescence ratio across CO generation in samples treated without JC-1 (negative control 1, NC-1) and those treated with FCCP and without JC-1 (negative control 2, NC-2), bars, mean  $\pm$  SD. (B) Bar graph showing intracellular ATP levels in iPSCs cultured in Mouse Embryonic Fibroblast (MEF)-Conditioned Media and Essential 8 Media. bars, mean  $\pm$  SD (i). Bar graph showing the intracellular ATP levels in iPSCs in two independent experimental runs, bars, mean  $\pm$  SD (ii). (C) Bar graph showing ATP levels expressed as percent of control in H9 hESCs, iPSCs and H9 hESC COs treated with oligomycin, bars, mean  $\pm$  SD. \* $p < 0.01$ ; \*\* $p < 0.001$ . *P*-values were determined using the independent samples *t*-test.