

Supplementary materials

Supplementary figures

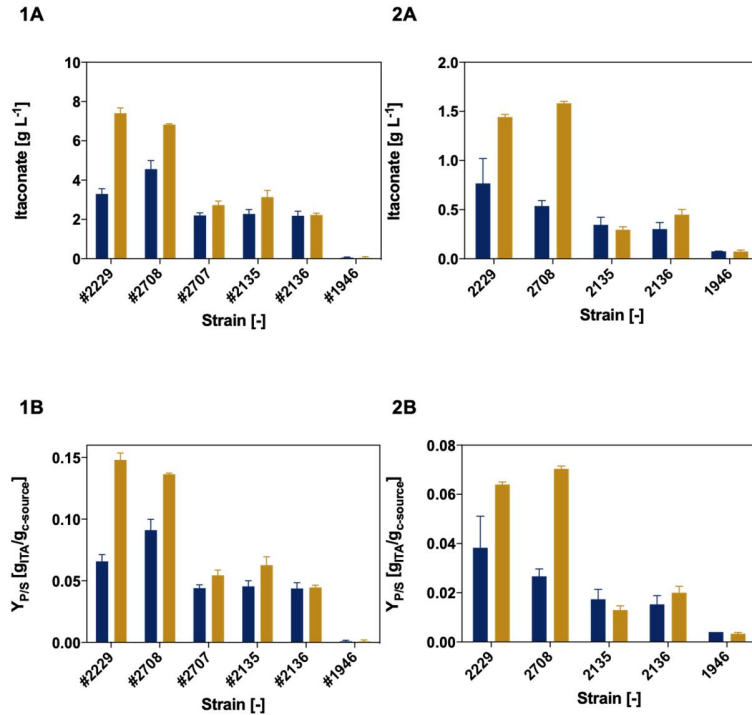


Figure S1 Itaconic acid production of most promising Ustilaginaceae strains using different carbon source concentrations. (A) Maximum itaconic acid production [g L⁻¹] and (B) Y_{P/S} [g_{ITA}/g_{C-source}] during System Duetz® 24-deep well plates cultivation experiments with 1.5 ml MTM medium and 0.8 g L⁻¹ NH₄Cl. Cultivations were performed with two different carbon-source concentrations (1) 50 g L⁻¹ glucose + 6.25 g L⁻¹ acetate and (2) 20 g L⁻¹ glucose + 2.5 g L⁻¹ acetate. Ustilaginaceae candidates using acetate as a co-substrate are shown in orange, respective glucose references are shown in blue. Error bars indicate the deviation from the mean for n=2.

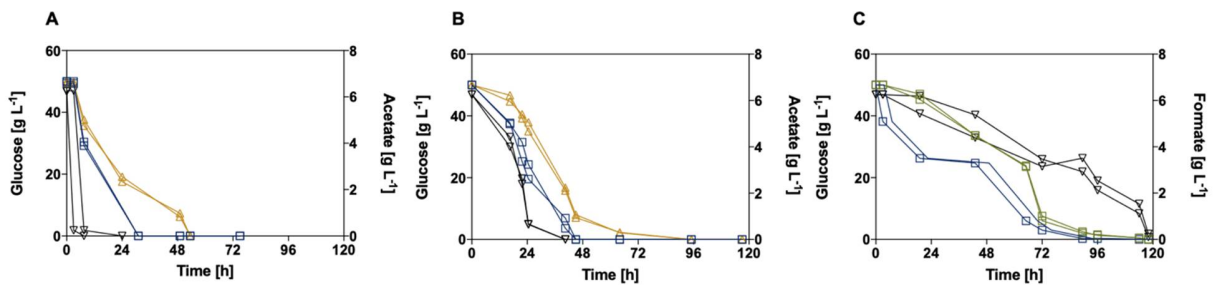


Figure S2 C-source consumption during controlled batch fermentations of selected Ustilaginaceae candidates. (A) *U. maydis* #2229, (B) *U. rabenhorstiana* #2708 and (C) *U. cynodontis* #2705 fermentation in a bioreactor containing MTM medium (0.8 g L⁻¹ NH₄Cl, 30°C, 80% DOT, at pH 6.5). Glucose consumptions using acetate as a co-substrate are shown in orange, formate co-substrate cultivations are shown in green. Respective glucose references (50 g L⁻¹) are shown in blue. Black indicates co-substrate consumption (6.25 g L⁻¹).

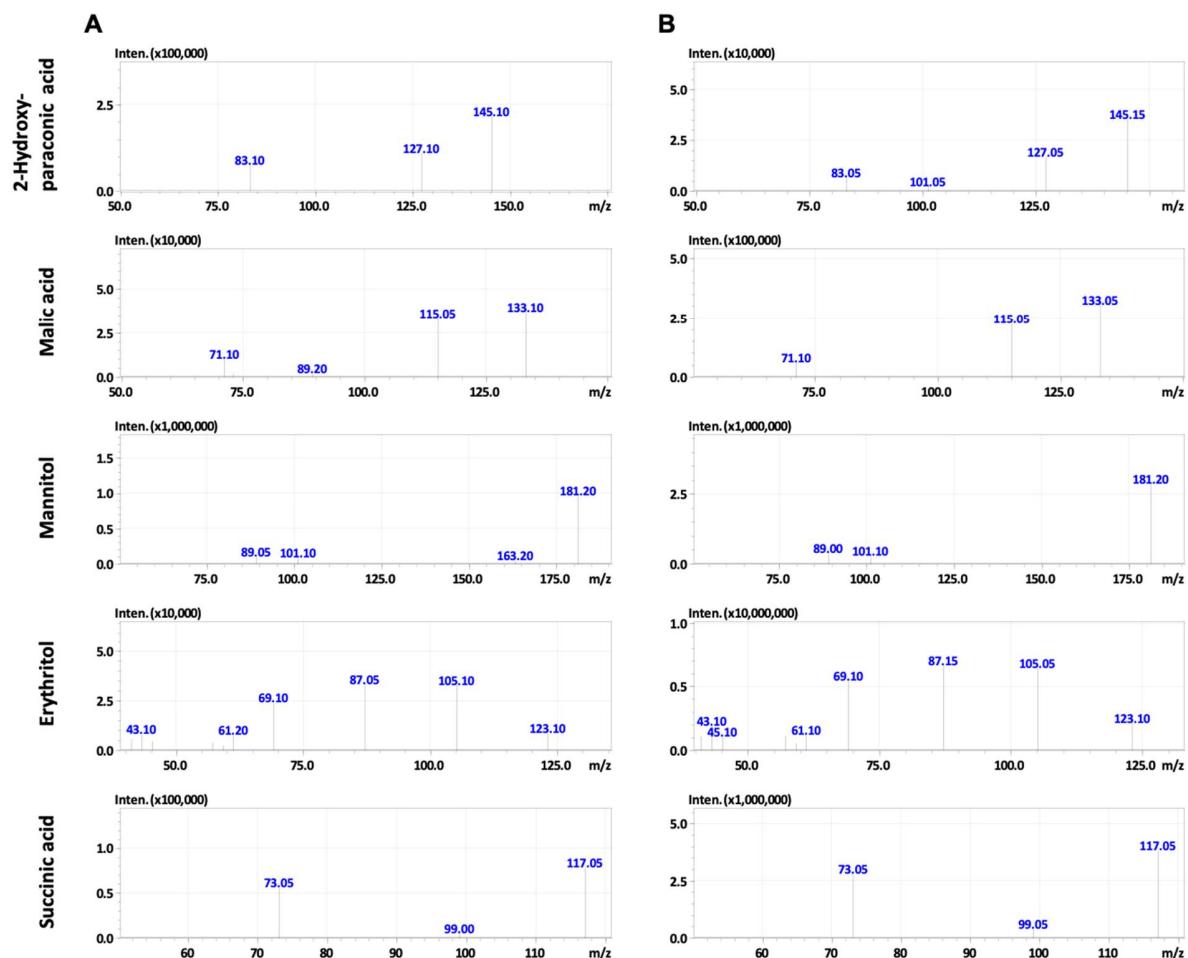


Figure S3 Extracellular metabolite identification via LC-UV/RI-MS/MS. (A) Product ion scan for extracellular metabolite samples and (B) their corresponding standards. Extracellular metabolites in the supernatant formed by *U. maydis* #2229, *U. rabeihorstiana* #2708, and *U. cynodontis* #2705. Samples were taken after 75, 64, and 114 h during fermentation in a bioreactor containing MTM medium (0.8 g L⁻¹ NH₄Cl, 19.5 g L⁻¹ MES, 30°C, 80% DOT, at pH 6.5).

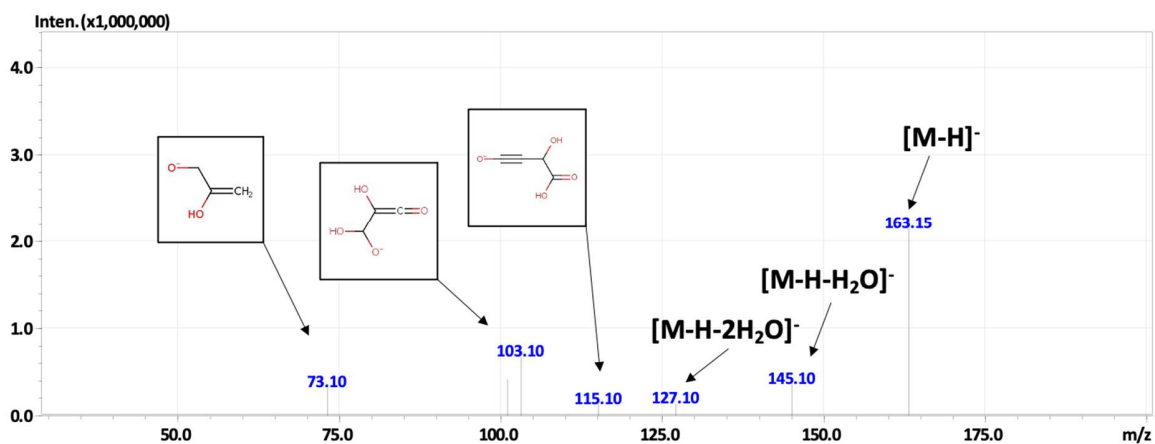


Figure S4 MS/MS spectrum of ITT. Structure prediction of itatartarate (ITT) was performed via CFM-ID 3.0 [1].

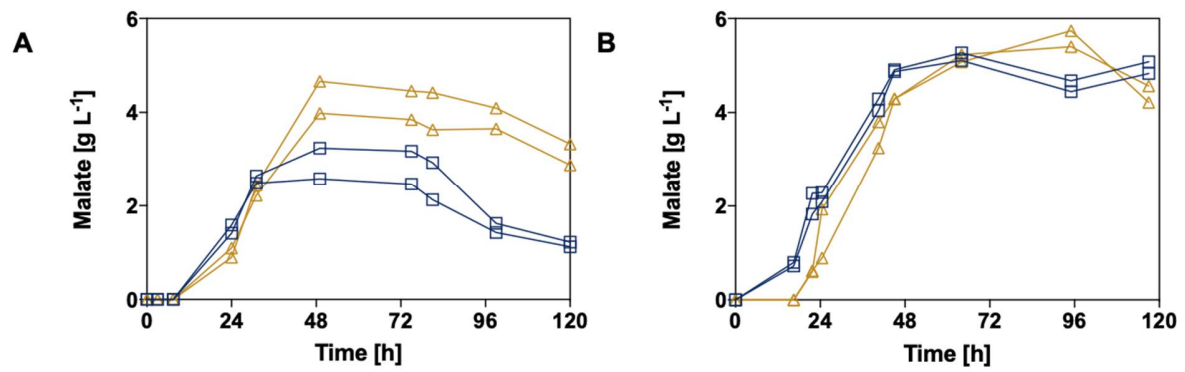


Figure S5 Malate production during controlled batch fermentations of selected Ustilaginaceae candidates. (A) *U. maydis* #2229 and (B) *U. rabenhorstiana* #2708 fermentation in a bioreactor containing MTM medium ($0.8 \text{ g L}^{-1} \text{ NH}_4\text{Cl}$, 30°C , 80% DOT, at pH 6.5). Glucose consumptions using acetate as a co-substrate are shown in orange. Respective glucose references (50 g L^{-1}) are shown in blue. Black indicates co-substrate consumption (6.25 g L^{-1}).

Supplementary tables

Table S1 Ustilaginaceae strains screened in this study.

No.	Strain	Origin
#1946	<i>Pseudozyma antartica</i>	NRRL Y – 7808
#1947	<i>Pseudozyma antartica</i>	NRRL Y - 8295
#1949	<i>Ustilago maydis</i>	DSM 3121
#1951	<i>Ustilago maydis</i>	DSM 14603
#2134	<i>Ustilago maydis</i>	Nr. 196
#2135	<i>Ustilago maydis</i>	Nr. 197
#2136	<i>Ustilago maydis</i>	Nr. 198
#2144	<i>Ustilago maydis</i>	Nr. 206
#2152	<i>Ustilago maydis</i>	Nr. 215
#2158	<i>Ustilago maydis</i>	Nr. 469
#2162	<i>Ustilago maydis</i>	Nr. 474
#2167	<i>Ustilago maydis</i>	Nr. 480
#2169	<i>Ustilago maydis</i>	Nr. 482
#2172	<i>Ustilago maydis</i>	Nr. 485
#2177	<i>Ustilago maydis</i>	Nr. 491
#2178	<i>Ustilago maydis</i>	Nr. 492
#2179	<i>Ustilago maydis</i>	Nr. 495
#2196	<i>Ustilago maydis</i>	FB1 Mating type a1b1
#2197	<i>Ustilago maydis</i>	FB2 Mating type a2b2
#2199	<i>Ustilago maydis</i>	RK 123
#2205	<i>Ustilago maydis</i>	RK 212
#2208	<i>Ustilago maydis</i>	RK215
#2209	<i>Macalpinomyces eriachnes</i>	RK 028
#2210	<i>Sporisorium consanguineum</i>	RK 133
#2211	<i>Sporisorium cruentum</i>	UMa920, Mating type MAT1
#2212	<i>Sporisorium exsertum</i>	RK 033
#2213	<i>Sporisorium scitamineum</i>	UMa698, Sscl4, JS109, RK 109, Mating type MAT1

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CBS 131454

CBS 131454

CBS 131456

CBS 133249

CBS 131457

#2214	<i>Sporisorium walkeri</i>	RK 031	CBS 131462
#2215	<i>Ustanciosporium gigantosporum</i>	UMa706	CBS 131464
#2218	<i>Ustilago filiformis</i>	UMa701	CBS 131469
#2219	<i>Ustilago trichophora</i>	RK 089	CBS 131473
#2220	<i>Ustilago vetiveriae</i>	RK 075	CBS 131474
#2221	<i>Ustilago xerochloae</i>	UMa702	CBS 131476
#2229	<i>Ustilago maydis</i>	MB215	DSM 17144
#2696	<i>Pseudozyma hubeiensis</i>		NBRC 105053
#2697	<i>Pseudozyma hubeiensis</i>		NBRC 105054
#2698	<i>Pseudozyma hubeiensis</i>		NBRC 105055
#2699	<i>Ustilago trichophora</i>		NBRC 100155
#2700	<i>Ustilago trichophora</i>		NBRC 100156
#2701	<i>Ustilago trichophora</i>		NBRC 100157
#2702	<i>Ustilago trichophora</i>		NBRC 100158
#2703	<i>Ustilago trichophora</i>		NBRC 100159
#2704	<i>Ustilago trichophora</i>		NBRC 100160
#2705	<i>Ustilago cynodontis</i>		NBRC 7530
#2706	<i>Ustilago cynodontis</i>		NBRC 9727
#2707	<i>Ustilago cynodontis</i>		NBRC 9758
#2708	<i>Ustilago rabenhorstiana</i>		NBRC 8995
#2710	<i>Pseudozyma tsukubaensis</i>		NBRC 1940
#2813	<i>Sporisorium lanigeri</i>	BRIP 27609 a	
#2814	<i>Macalpinomyces mackinlayi</i>	BRIP 52549 a	
#2815	<i>Sporisorium cenchri-elymoidis</i>	BRIP 26491 a	
#2816	<i>Macalpinomyces ordensis</i>	BRIP 26904 a	
#2817	<i>Ustilago schmidiae</i>	BRIP 26906 a	
#2818	<i>Sporisorium bothriochloae</i>	BRIP 26908 a	
#2819	<i>Sporisorium themedae</i>	BRIP 26917 a	
#2820	<i>Sporisorium tumiforme</i>	BRIP 26919 a	
#2821	<i>Ustilago curta</i>	BRIP 26929 a	
#2822	<i>Ustilago triodiae</i>	BRIP 26907 a	

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#2823	<i>Sporisorium setariae</i>	BRIP 26910 a
#2824	<i>Ustilago cynodontis</i>	BRIP 28040 a
#2825	<i>Sporisorium caledonicum</i>	BRIP 28043 a
#2826	<i>Ustilago lituana</i>	BRIP 46795 a
#2832	<i>Sporisorium iseilematis-ciliati</i>	BRIP 60429 a
#2833	<i>Macalpinomyces spermophorus</i>	BRIP 60430 a
#2834	<i>Macalpinomyces tubiformis</i>	BRIP 60434 a
#2835	<i>Macalpinomyces spermophorus</i>	BRIP 60448 a
#2836	<i>Ustilago xerochloae</i>	BRIP 60876 a
#2838	<i>Sporisorium iseilematis-ciliati</i>	BRIP 60887 a
#2841	<i>Anthracoystis heteropogonicola</i>	BRIP 60896 a
#2842	<i>Anthracoystis bothriochloae</i>	BRIP 60901 a
#2850	<i>Ustilago egenula</i>	BRIP 60884 a
#3600	<i>Ustilago maydis</i>	



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Table S5 Production parameters of biodiversity screening for acetate co-utilization. Growth results of the five most promising strains obtained during experiments using the Growth Profiler device by EnzyScreen with different acetate concentrations as co-substrates in combination with 20 g L⁻¹ glucose (n=2). *U. maydis* #2135 and #2136 obtained best results using 10 g L⁻¹ acetate. Remaining strains showed highest growth using 2.5 g L⁻¹ acetate.

Strain	C-source	Max. OD _{600nm}	Y _{OD/S}	Y _{OD/S}
		[-]	[max. OD _{600nm} /g _{C-source}]	[max. OD _{600nm} /C-mol _{C-source}]
<i>U. maydis</i>	Glucose	46.8	2.34	69.9
		48.3	2.42	72.1
#1946	Glucose + Acetate	55.9	2.48	77.6
		56.5	2.51	78.5
<i>U. cynodontis</i>	Glucose	40.7	2.03	60.7
		41.3	2.07	61.7
#2707	Glucose + Acetate	53.7	2.38	74.5
		54.0	2.40	75.0
<i>U. maydis</i>	Glucose	29.9	1.40	44.6
		19.9	1.00	29.8
#2136	Glucose + Acetate*	51.4	1.71	56.5
		52.8	1.76	58.1
<i>U. maydis</i>	Glucose	19.7	0.99	29.4
		22.5	1.13	33.6
#2135	Glucose + Acetate*	49.5	1.65	54.4
		49.8	1.66	54.8
<i>U. rabenhorstiana</i>	Glucose	38.0	1.90	56.8
		38.2	1.91	57.0
#2708	Glucose + Acetate	48.3	2.14	67.0
		50.0	2.22	69.5

Table S6 Production parameters of biodiversity screening for formate co-utilization. Growth results of the five most promising strains obtained during experiments using the Growth Profiler device by Enzyscreen with 2.5 g L⁻¹ formate as co-substrates in combination with 20 g L⁻¹ glucose (n=2).

Strain	C-source	Max.	Y _{OD/S}	Y _{OD/S}
		OD _{600nm} [-]	[max. OD _{600nm} /g _{C-source}]	[max. OD _{600nm} /C-mol _{C-source}]
<i>U. rabenhorstiana</i> #2708	Glucose	38.0	1.90	56.8
		38.2	1.91	57.0
	Glucose + Formate	56.6	2.52	80.9
		56.7	2.52	81.0
<i>U. cynodontis</i> #2707	Glucose	31.6	1.58	47.2
		33.0	1.65	49.3
	Glucose + Formate	50.0	2.22	71.5
		50.5	2.25	72.2
<i>U. maydis</i> #2177	Glucose	29.0	1.45	43.3
		30.2	1.51	45.0
	Glucose + Formate	42.0	1.86	59.9
		43.1	1.91	61.5
<i>U. cynodontis</i> #2706	Glucose	27.5	1.37	41.0
		27.7	1.38	41.3
	Glucose + Formate	40.4	1.70	57.6
		44.2	1.93	63.1
<i>U. maydis</i> #2196	Glucose	36.3	1.82	54.2
		37.8	1.89	56.4
	Glucose + Formate	42.3	1.88	60.4
		43.4	1.93	62.0

Table S7 Production parameters of System Duetz cultivation for acetate co-utilization. System Duetz® 24-deep well plates cultivation experiments were performed with 1.5 ml MTM medium and 50 g L⁻¹ glucose. 6.25 g L⁻¹ acetate were added during co-culture cultivations. Error bars indicate the deviation from the mean for n=2.

Strain	C-source	Titer _{max}	Y _{P/S}	Y _{P/S}	Max. OD
		[g L ⁻¹]	[g _{ITA} /g _{c-source}]	[C-mol _{ITA} /C-mol _{c-source}]	[-]
<i>U. maydis</i> #2229	Glucose	2.99	0.060	0.069	39
		3.38	0.068	0.078	33
	Glucose + Acetate	7.20	0.144	0.152	49
		7.60	0.152	0.161	48
<i>U. rabenhorstiana</i> #2708	Glucose	4.30	0.086	0.099	39
		4.31	0.101	0.099	40
	Glucose + Acetate	6.88	0.136	0.144	41
		6.80	0.138	0.145	45
<i>U. cynodontis</i> #2707	Glucose	2.24	0.045	0.052	53
		2.05	0.041	0.047	52
	Glucose + Acetate	2.88	0.056	0.060	50
		2.88	0.058	0.061	53
<i>U. maydis</i> #2135	Glucose	2.40	0.048	0.055	46
		2.42	0.048	0.056	47
	Glucose + Acetate	2.97	0.071	0.075	54
		2.35	0.059	0.063	58
<i>U. maydis</i> #2136	Glucose	2.36	0.047	0.054	50
		2.28	0.046	0.053	51
	Glucose + Acetate	2.32	0.046	0.049	49
		2.22	0.044	0.047	50
<i>U. maydis</i> #1946	Glucose	0.09	0.002	0.002	71
		0.06	0.001	0.001	73
	Glucose + Acetate	0.01	0.000	0.00	72
		0.01	0.000	0.00	66

Table S8 Production parameters of System Duetz cultivation for formate co-utilization. System Duetz® 24-deep well plates cultivation experiments were performed with 1.5 ml MTM medium and 50 g L⁻¹ glucose. 6.25 g L⁻¹ formate were added during co-culture cultivations. Error bars indicate the deviation from the mean for n=2.

Strain	C-source	Titer _{max.}	Y _{P/S}	Y _{P/S}	Max. OD
		[g L ⁻¹]	[g _{ITA} /g _{c-source}]	[C-mol _{ITA} /C-mol _{c-source}]	[-]
<i>U. cynodontis</i> #2706	Glucose	6.64	0.133	0.153	51
		6.90	0.138	0.159	50
<i>U. cynodontis</i> #2705	Glucose + Formate	6.90	0.123	0.151	38
		5.81	0.103	0.127	36
<i>U. cynodontis</i> #2705	Glucose	6.18	0.124	0.143	52
		6.32	0.126	0.146	57
<i>U. rabenhorstiana</i> #2708	Glucose + Formate	9.04	0.161	0.198	50
		8.20	0.146	0.179	54
<i>U. rabenhorstiana</i> #2708	Glucose	4.16	0.083	0.096	48
		4.35	0.087	0.100	50
<i>U. maydis</i> #2229	Glucose + Formate	3.98	0.071	0.087	35
		2.64	0.047	0.058	37
<i>U. maydis</i> #2177	Glucose	2.61	0.052	0.060	50
		2.87	0.057	0.066	54
<i>U. maydis</i> #2177	Glucose + Formate	0.30	0.005	0.007	37
		0.31	0.004	0.007	36
<i>U. maydis</i> #2196	Glucose	2.78	0.056	0.064	66
		2.60	0.052	0.060	65
<i>U. maydis</i> #2196	Glucose + Formate	0.43	0.008	0.009	65
		0.49	0.009	0.011	66
<i>U. maydis</i> #2196	Glucose	0.66	0.013	0.015	79
		0.55	0.011	0.013	80
<i>U. maydis</i> #2196	Glucose + Formate	0.04	0.001	0.001	60
		0.04	0.001	0.001	57

Table S9 pH values during System Duetz cultivation for acetate co-utilization. System Duetz[®] 24-deep well plates cultivation experiments were performed with 1.5 ml MTM medium and 50 g L⁻¹ glucose. 6.25 g L⁻¹ acetate were added during co-culture cultivations. Error bars indicate the deviation from the mean for n=2.

Strain	C-source	Min. pH [-]	pH End [-]
<i>U. maydis</i> #2229	Glucose	4.98	5.76
		4.96	5.80
	Glucose + Acetate	5.49	5.78
		5.48	5.80
<i>U. rabenhorstiana</i> #2708	Glucose	2.51	2.86
		2.47	2.85
	Glucose + Acetate	3.11	3.36
		3.13	3.37
<i>U. cynodontis</i> #2707	Glucose	3.37	3.44
		3.44	3.45
	Glucose + Acetate	3.79	3.79
		3.78	3.78
<i>U. maydis</i> #2135	Glucose	4.64	5.68
		4.70	5.71
	Glucose + Acetate	5.07	5.85
		5.09	5.80
<i>U. maydis</i> #2136	Glucose	5.02	5.72
		5.03	5.74
	Glucose + Acetate	5.36	5.78
		5.38	5.79
<i>U. maydis</i> #1946	Glucose	5.61	5.63
		5.64	5.70
	Glucose + Acetate	6.34	6.43
		6.32	6.42

Table S10 pH values during System Duetz cultivation for formate co-utilization. System Duetz® 24-deep well plates cultivation experiments were performed with 1.5 ml MTM medium and 50 g L⁻¹ glucose. 6.25 g L⁻¹ formate were added during co-culture cultivations. Error bars indicate the deviation from the mean for n=2.

Strain	C-source	Min. pH [-]	pH End [-]
<i>U. cynodontis</i> #2706	Glucose	3.04	3.07
		3.03	3.01
	Glucose + Formate	4.96	4.96
<i>U. cynodontis</i> #2705	Glucose	4.94	4.94
		2.86	2.86
	Glucose + Formate	2.83	2.83
<i>U. rabenhorstiana</i> #2708	Glucose	4.26	4.26
		4.34	4.34
	Glucose + Formate	3.80	3.95
<i>U. maydis</i> #2229	Glucose	3.76	3.84
		5.08	5.08
	Glucose + Formate	5.11	5.11
<i>U. maydis</i> #2177	Glucose	4.85	5.87
		4.87	5.88
	Glucose + Formate	5.54	6.05
<i>U. maydis</i> #2196	Glucose	5.56	6.06
		5.11	5.60
	Glucose + Formate	5.10	5.56
<i>U. maydis</i> #2196	Glucose	5.64	6.57
		5.65	6.56
	Glucose + Formate	6.71	7.80
<i>U. maydis</i> #2196	Glucose + Formate	6.75	7.09
		6.66	7.69
		6.70	7.79

Table S11 Welch t-test results obtained for *U. maydis* #2229, *U. rabenhorstiana* #2708, and *U. cynodontis* #2705 during small-scale production and bioreactor experiments.

t-Test: Two-Sample Assuming Unequal Variances			t-Test: Two-Sample Assuming Unequal Variances			t-Test: Two-Sample Assuming Unequal Variances		
#2229 Maximum titer	small-scale		#2708 Maximum titer	small-scale		#2705 Maximum titer	small-scale	
	GLC	co substrate		GLC	co substrate		GLC	co substrate
Mean	3.185	7.4	Mean	4.305	6.84	Mean	6.25	8.62
Variance	0.07605	0.08	Variance	5E-05	0.0032	Variance	0.0098	0.3528
Observations	2	2	Observations	2	2	Observations	2	2
Hypothesized Mean Difference	0		Hypothesized Mean Difference	0		Hypothesized Mean Difference	0	
df	2		df	1		df	1	
t Stat	-15.089703		t Stat	-62.88561		t Stat	-5.56608	
P(T<=t) one-tail	0.00218152		P(T<=t) one-tail	0.0050413		P(T<=t) one-tail	0.0565838	
t Critical one-tail	2.91998558		t Critical one-tail	6.31375151		t Critical one-tail	6.31375151	
P(T<=t) two-tail	0.00436304		P(T<=t) two-tail	0.0101226		P(T<=t) two-tail	0.1131676	
t Critical two-tail	4.30265273		t Critical two-tail	12.7062047		t Critical two-tail	12.7062047	

t-Test: Two-Sample Assuming Unequal Variances			t-Test: Two-Sample Assuming Unequal Variances			t-Test: Two-Sample Assuming Unequal Variances		
#2229 Yield	small-scale		#2708 Yield	small-scale		#2705 Yield	small-scale	
	GLC	co substrate		GLC	co substrate		GLC	co substrate
Mean	0.064	0.148	Mean	0.0935	0.137	Mean	0.125	0.1535
Variance	3.2E-05	3.2E-05	Variance	0.0001125	0.000002	Variance	0.000002	0.0001125
Observations	2	2	Observations	2	2	Observations	2	2
Hypothesized Mean Difference	0		Hypothesized Mean Difference	0		Hypothesized Mean Difference	0	
df	2		df	1		df	1	
t Stat	-14.849242		t Stat	-5.7491218		t Stat	-3.766666	
P(T<=t) one-tail	0.00225226		P(T<=t) one-tail	0.05481825		P(T<=t) one-tail	0.08260158	
t Critical one-tail	2.91998558		t Critical one-tail	6.31375151		t Critical one-tail	6.31375151	
P(T<=t) two-tail	0.00450453		P(T<=t) two-tail	0.10963649		P(T<=t) two-tail	0.16520315	
t Critical two-tail	4.30265273		t Critical two-tail	12.7062047		t Critical two-tail	12.7062047	

t-Test: Two-Sample Assuming Unequal Variances			t-Test: Two-Sample Assuming Unequal Variances			t-Test: Two-Sample Assuming Unequal Variances		
#2229 Maximum titer	bioreactor		#2708 Maximum titer	bioreactor		#2705 Maximum titer	bioreactor	
	GLC	co substrate		GLC	co substrate		GLC	co substrate
Mean	3.25	4.75	Mean	2.1	2.9	Mean	1.65	2.9
Variance	0.005	0.045	Variance	0	0.08	Variance	0.005	0
Observations	2	2	Observations	2	2	Observations	2	2
Hypothesized Mean Difference	0		Hypothesized Mean Difference	0		Hypothesized Mean Difference	0	
df	1		df	1		df	1	
t Stat	-9.486833		t Stat	-4		t Stat	-25	
P(T<=t) one-tail	0.03342936		P(T<=t) one-tail	0.07797913		P(T<=t) one-tail	0.01272561	
t Critical one-tail	6.31375151		t Critical one-tail	6.31375151		t Critical one-tail	6.31375151	
P(T<=t) two-tail	0.06685872		P(T<=t) two-tail	0.15595826		P(T<=t) two-tail	0.02545122	
t Critical two-tail	12.7062047		t Critical two-tail	12.7062047		t Critical two-tail	12.7062047	

t-Test: Two-Sample Assuming Unequal Variances			t-Test: Two-Sample Assuming Unequal Variances			t-Test: Two-Sample Assuming Unequal Variances		
#2229 Yield	bioreactor		#2708 Yield	bioreactor		#2705 Yield	bioreactor	
	GLC	co substrate		GLC	co substrate		GLC	co substrate
Mean	0.07	0.085	Mean	0.04	0.055	Mean	0.033	0.052
Variance	0	0.00005	Variance	0	5E-05	Variance	0.000002	3.9506E-07
Observations	2	2	Observations	2	2	Observations	2	2
Hypothesized Mean Difference	0		Hypothesized Mean Difference	0		Hypothesized Mean Difference	0	
df	1		df	1		df	1	
t Stat	-3		t Stat	-3		t Stat	-17.362419	
P(T<=t) one-tail	0.10241638		P(T<=t) one-tail	0.10241638		P(T<=t) one-tail	0.01831304	
t Critical one-tail	6.31375151		t Critical one-tail	6.31375151		t Critical one-tail	6.31375151	
P(T<=t) two-tail	0.20483276		P(T<=t) two-tail	0.20483276		P(T<=t) two-tail	0.03662607	
t Critical two-tail	12.7062047		t Critical two-tail	12.7062047		t Critical two-tail	12.7062047	

Table S12 Welch t-test results obtained during small-scale production experiments. System Duetz[®] 24-deep well plates cultivation experiments were performed with 1.5 ml MTM medium and 50 g L⁻¹ glucose. 6.25 g L⁻¹ co-substrate were added during co-culture cultivations. Error bars indicate the deviation from the mean for n=2. Experiments performed with acetate are highlighted in yellow whereas formate cultivations are highlighted in grey.

t-Test: Two-Sample Assuming Unequal Variances			t-Test: Two-Sample Assuming Unequal Variances			t-Test: Two-Sample Assuming Unequal Variances			t-Test: Two-Sample Assuming Unequal Variances		
#2707 Maximum titer			#2135 Maximum titer			#2706 Maximum titer			#2229 Maximum titer		
	GLC	co substrate		GLC	co substrate		GLC	co substrate		GLC	co substrate
Mean	2.145	2.8845	Mean	2.41	2.66	Mean	6.77	6.355	Mean	2.74	0.305
Variance	0.01805	4.05E-05	Variance	0.0002	0.1922	Variance	0.0338	0.59405	Variance	0.0338	5E-05
Observations	2	2	Observations	2	2	Observations	2	2	Observations	2	2
Hypothesized Mean Difference	0		Hypothesized Mean Difference	0		Hypothesized Mean Difference	0		Hypothesized Mean Difference	0	
df	1		df	1		df	1		df	1	
t Stat	-7.7754922		t Stat	-0.8060324		t Stat	0.74068772		t Stat	18.7169305	
P(T<=t) one-tail	0.04071409		P(T<=t) one-tail	0.2840559		P(T<=t) one-tail	0.29707281		P(T<=t) one-tail	0.01699037	
t Critical one-tail	6.31375151		t Critical one-tail	6.31375151		t Critical one-tail	6.31375151		t Critical one-tail	6.31375151	
P(T<=t) two-tail	0.08142819		P(T<=t) two-tail	0.5681118		P(T<=t) two-tail	0.59414562		P(T<=t) two-tail	0.03398074	
t Critical two-tail	12.7062047		t Critical two-tail	12.7062047		t Critical two-tail	12.7062047		t Critical two-tail	12.7062047	
t-Test: Two-Sample Assuming Unequal Variances			t-Test: Two-Sample Assuming Unequal Variances			t-Test: Two-Sample Assuming Unequal Variances			t-Test: Two-Sample Assuming Unequal Variances		
#2707 Yield			#2135 Yield			#2706 Yield			#2229 Yield		
	GLC	co substrate		GLC	co substrate		GLC	co substrate		GLC	co substrate
Mean	0.043	0.057	Mean	0.0485	0.065	Mean	0.1305	0.113	Mean	0.0545	0.0045
Variance	8E-06	0.000002	Variance	5E-07	0.000072	Variance	0.0000125	0.0002	Variance	0.0000125	0.000005
Observations	2	2	Observations	2	2	Observations	2	2	Observations	2	2
Hypothesized Mean Difference	0		Hypothesized Mean Difference	0		Hypothesized Mean Difference	0		Hypothesized Mean Difference	0	
df	1		df	1		df	1		df	1	
t Stat	-6.2609903		t Stat	-2.7405008		t Stat	1.69774938		t Stat	19.6116135	
P(T<=t) one-tail	0.05041437		P(T<=t) one-tail	0.11137147		P(T<=t) one-tail	0.16943737		P(T<=t) one-tail	0.01621664	
t Critical one-tail	6.31375151		t Critical one-tail	6.31375151		t Critical one-tail	6.31375151		t Critical one-tail	6.31375151	
P(T<=t) two-tail	0.10082873		P(T<=t) two-tail	0.22274294		P(T<=t) two-tail	0.33897474		P(T<=t) two-tail	0.03243328	
t Critical two-tail	12.7062047		t Critical two-tail	12.7062047		t Critical two-tail	12.7062047		t Critical two-tail	12.7062047	
t-Test: Two-Sample Assuming Unequal Variances			t-Test: Two-Sample Assuming Unequal Variances			t-Test: Two-Sample Assuming Unequal Variances			t-Test: Two-Sample Assuming Unequal Variances		
#2136 Maximum titer			#1946 Maximum titer			#2177 Maximum Titer			#2196 Maximum Titer		
	GLC	co substrate		GLC	co substrate		GLC	co substrate		GLC	co substrate
Mean	2.32	2.27	Mean	0.075	0.011	Mean	2.69	0.46	Mean	0.605	0.0405
Variance	0.0032	0.005	Variance	0.00045	0.000002	Variance	0.0162	0.0018	Variance	0.00605	5E-07
Observations	2	2	Observations	2	2	Observations	2	2	Observations	2	2
Hypothesized Mean Difference	0		Hypothesized Mean Difference	0		Hypothesized Mean Difference	0		Hypothesized Mean Difference	0	
df	2		df	1		df	1		df	1	
t Stat	0.78086881		t Stat	4.25721667		t Stat	23.5062639		t Stat	10.2632123	
P(T<=t) one-tail	0.25831588		P(T<=t) one-tail	0.07343813		P(T<=t) one-tail	0.01353333		P(T<=t) one-tail	0.03091705	
t Critical one-tail	2.91998558		t Critical one-tail	6.31375151		t Critical one-tail	6.31375151		t Critical one-tail	6.31375151	
P(T<=t) two-tail	0.51663176		P(T<=t) two-tail	0.14687626		P(T<=t) two-tail	0.02706666		P(T<=t) two-tail	0.06183411	
t Critical two-tail	4.30265273		t Critical two-tail	12.7062047		t Critical two-tail	12.7062047		t Critical two-tail	12.7062047	
t-Test: Two-Sample Assuming Unequal Variances			t-Test: Two-Sample Assuming Unequal Variances			t-Test: Two-Sample Assuming Unequal Variances			t-Test: Two-Sample Assuming Unequal Variances		
#2136 Yield			#21946 Yield			#2177 Yield			#2196 Yield		
	GLC	co substrate		GLC	co substrate		GLC	co substrate		GLC	co substrate
Mean	0.0465	0.045	Mean	0.0015	0	Mean	0.054	0.0085	Mean	0.012	0.001
Variance	5E-07	0.000002	Variance	0.0000005	0	Variance	8E-06	5E-07	Variance	0.000002	0
Observations	2	2	Observations	2	2	Observations	2	2	Observations	2	2
Hypothesized Mean Difference	0		Hypothesized Mean Difference	0		Hypothesized Mean Difference	0		Hypothesized Mean Difference	0	
df	1		df	1		df	1		df	1	
t Stat	1.34164079		t Stat	3		t Stat	22.0707419		t Stat	11	
P(T<=t) one-tail	0.20388458		P(T<=t) one-tail	0.10241638		P(T<=t) one-tail	0.0144124		P(T<=t) one-tail	0.02885794	
t Critical one-tail	6.31375151		t Critical one-tail	6.31375151		t Critical one-tail	6.31375151		t Critical one-tail	6.31375151	
P(T<=t) two-tail	0.40776917		P(T<=t) two-tail	0.20483276		P(T<=t) two-tail	0.0288248		P(T<=t) two-tail	0.05771588	
t Critical two-tail	12.7062047		t Critical two-tail	12.7062047		t Critical two-tail	12.7062047		t Critical two-tail	12.7062047	

Supplementary references

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