## Supplemental material Table of Contents

Supplement 1.

Periodic acid–Schiff (PAS) staining of NPHS-Cre-Rpt3 flox/flox and NPHS-Cre+Rpt3 flox/WT

Supplement 2. The expression of Rpt3 was decreased in the glomeruli of Rpt3<sup>pdKO</sup> mice at 4 weeks of age.

Supplement 3. The number of WT1 was decreased in the glomeruli of Rpt3<sup>pdKO</sup> kidneys compared with Rpt3<sup>Control</sup> kidneys.

Supplement 4. There was no significant change in the expression of LC3 between Rpt3<sup>Control</sup> and Rpt3<sup>pdKO</sup> mice.

Supplement 5. The expressions of p62 and LC3 were analyzed using primary culture podocytes.

Supplement 6. Rhodamine phalloidin staining showed that the treatment of BTZ altered the actin formation of cultured podocytes.

Supplement 7. Proteasome activity was decreased in cultured podocytes by the treatment of BTZ.

Supplement 8.

Immunofluorescence intensity of 8-OHdG was increased in cultured podocytes by the treatment of BTZ.

Supplement 9.

Rapamycin induced autophagic activity of podocytes in Rpt3<sup>pdKO</sup> mice.

Supplement 10.

ER stress did not participate in podocyte injury via the impariment of proteasome function.



Supplemental Figure 1. Periodic acid–Schiff (PAS) staining of NPHS-Cre-Rpt3 flox/flox and NPHS-Cre+Rpt3 flox/WT (A) NPHS-Cre-Rpt3 flox/flox and NPHS-Cre+Rpt3 flox/WT showed no pathologic phenotypes at 2 months of age when examined using histologic methods. (B) Rpt3 flox/flox, Rpt3 WT/WT and Rpt3 flox/+WT were distinguished by genotyping with cut tails. Scale bars: 100 µm (upper), 50 µm (lower).



Supplemental Figure 2. The expression of Rpt3 was decreased in glomeruli of Rpt3<sup>pdKO</sup> mice at 4 weeks of age.



Supplemental Figure 3. The number of WT1 was decreased in the glomeruli of Rpt3<sup>pdKO</sup> kidneys compared with Rpt3<sup>Control</sup> kidneys.

In IF staining, the number of WT1 which is a nuclear marker for podocytes was decreased in the kidney of Rpt3 <sup>pdKO</sup> mice than in the kidney of Rpt3<sup>Control</sup> mice at 4 and 8 weeks of age. Scale bars: 20 µm.



Supplemental Figure 4. There was no significant change in the expression of LC3 between Rpt3<sup>Control</sup> and Rpt3<sup>pdKO</sup> mice.

(A and B) There was no significant change in the LC3-II / LC3-I ratio between Rpt3<sup>Control</sup> and Rpt3<sup>pdKO</sup> kidneys at 4 weeks of age in WB by glomerular lysate (n=3, P=0.39, Welch's test).

(A)



Green : Synaptopodin Blue : DAPI

(B)



## (C)



Supplemental Figure 5. The expressions of p62 and LC3 were analyzed using primary culture podocytes from 4 weeks of age mice.

(A) Synaptopodin-positive cellular outgrowths from glomeruli were distinguished as podocytes. (B) The accumulation of p62 was detected in primary cultured podocytes from Rpt3<sup>pdKO</sup> mice. (C) Three days after primary culture, podocytes were treated with lysosomal protease inhibitors E64d (10 μg/ ml) and pepstatin A (25 μg/ ml) (E/P) for 24 hours. Under the effect of E/P, the expression of LC3 was lower in the podocytes from Rpt3<sup>pdKO</sup> mice than from Rpt3<sup>Control</sup> mice. Scale bars: 100 μm (A), 25 μm (B and C).

Rhodamine phalloidin staining of cultured podocytes treated with bortezomib



Supplemental Figure 6. Rhodamine phalloidin staining showed that the treatment of BTZ altered the actin formation of cultured podocytes. Bortezomib (BTZ) was used at concentration of 100nM. Scale bars: 20 µm



Supplemental Figure 7. Proteasome activity was decreased by proteasome inhibition in podocytes at 6 hours after treatment of bortezomib to cultured podocytes. The proteasome activity of podocytes was reduced to about 13% compared with that of DMSO treated podocytes.

## 8OHdG intensity

	80HdG b	inary image of 80Hd	G 80HdG-DAPI
control			
BTZ (100nM) 3h			
9h			
15h			

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Supplemental Figure 8. Immunofluorescence intensity of 8-OHdG was increased in podocytes in a time dependent manner after treatment wtih 100nM of BTZ. Left side shows the 8-OHdG staining, middle shows the binary image of 8-OHdG staining and right side shows co-staining of 8-OHdG and nuclei; Green : 8HdG, Blue : DAPI. Scale bars: 50 µm



Supplemental Figure 9. Rapamycin induced autophagic activity of podocytes in Rpt3<sup>pdKO</sup> mice.

(A and B) The LC3 dots in podocytes increased by rapamycin treatment in control and Rpt3<sup>pdKO</sup> mice.

(C and D) The p62 particles in podocytes decreased by rapamycin treatment in Rpt3<sup>pdKO</sup> mice.

(E and F) The ubiquitin accumulation in podocytes decreased by rapamycin treatment in Rpt3<sup>pdKO</sup> mice.

(P<0.05 by Mann-Whitney's U test)



Supplemental Figure 10. ER stress did not participate in podocyte injury via the impariment of proteasome function.

(A) There was no significant change in the expressions of BiP between Rpt3<sup>Control</sup> and Rpt3<sup>pdKO</sup> mice in WB by glomerular lysate (n=3). (B) The densitometry quantification of BiP expression in WB shown in (A) (n=3). (C) There was no significant change in the expression of BiP in cultured podocytes after the treatment with 100nM of BTZ in vitro (n=3). (D) The densitometry quantification of BiP expression in WB shown in (C) (n=3). \*P<0.05 ; \*\*P<0.01 ; \*\*\*P<0.001 by Welch's t-test.