

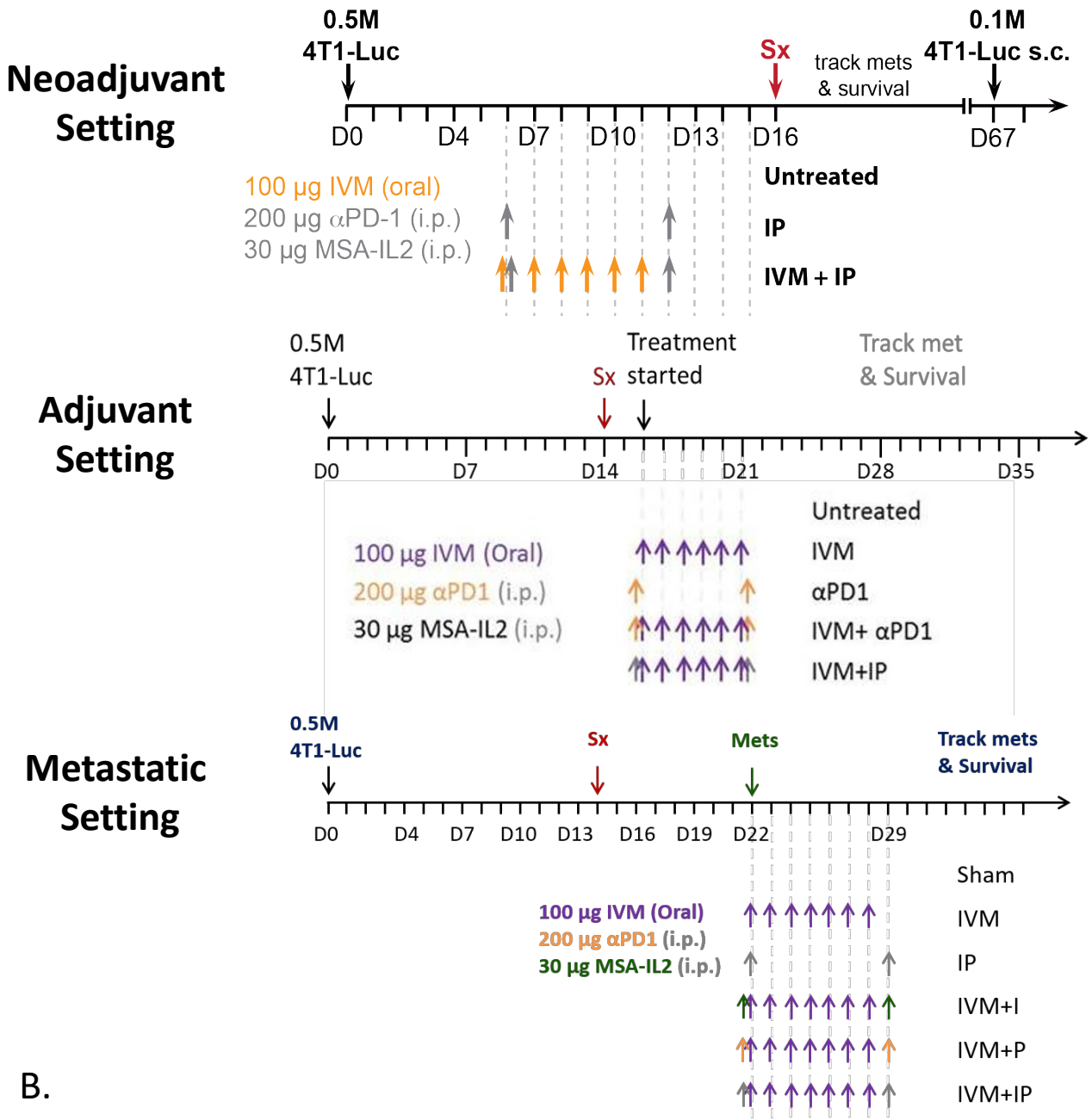
Supplementary Materials

Supplementary Table 1. Experimental Design, Treatment Settings

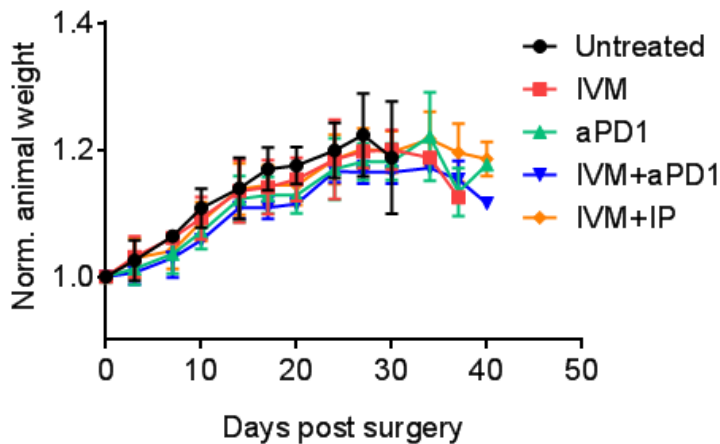
	Surgical Resection of Tumor	Ivermectin Alone	Anti-PD1 Antibody Alone	Combined Treatment	No Treatment	Days of Observation
Primary	No					
Experiment A		5	0	0	5	25
Experiment B		5	0	0	5	21
Experiment C		5	5	5	5	21
Experiment D		0	5	5	5	49
Experiment E		5	0	5	5	56
Total (N=70)		20	10	15	25	
Neoadjuvant With IL-2	Yes		5 (Anti-PD1 + MSA-IL-2)	5 [#]	5	84
Adjuvant With IL-2	Yes					84
Without IL-2		0	0	10	0	
Total (N=50)		10	10	10	10	
Metastatic With IL-2	Yes					82
Without IL-2		5	5	10	0	
Total (N=40)		10	10	15	5	

The IP+IVM treatment group shown in Fig. 3A had n=5 with 1 death from surgery complication — thus survival was shown for n=4.

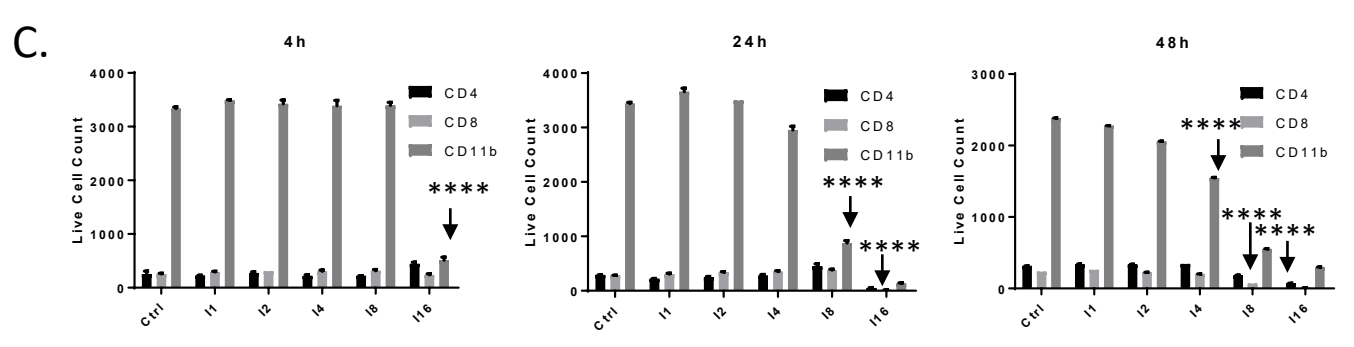
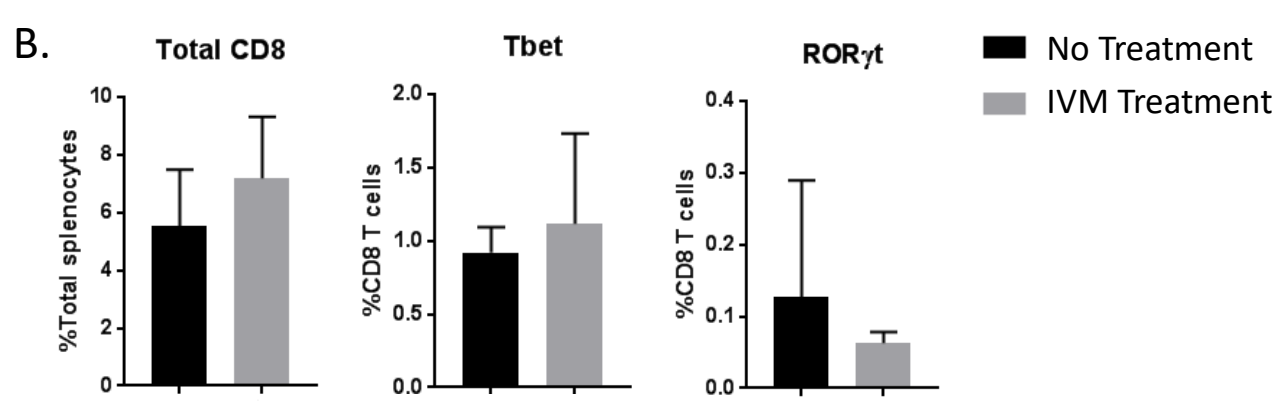
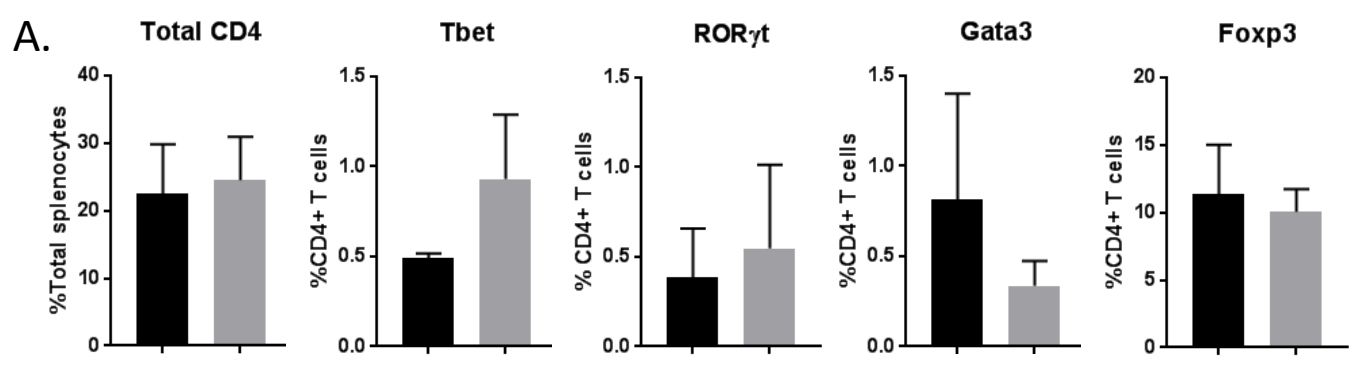
A.



B.

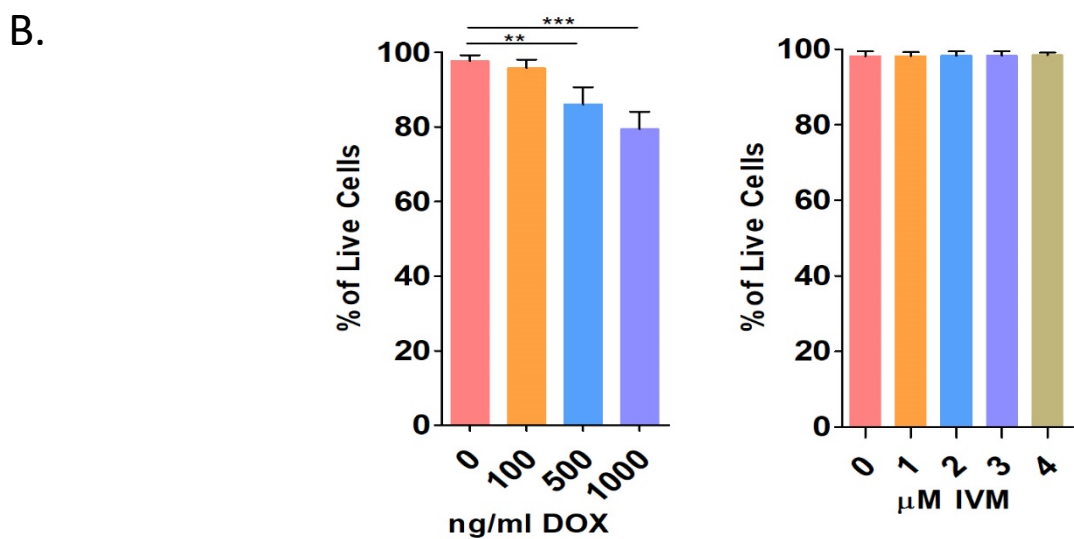
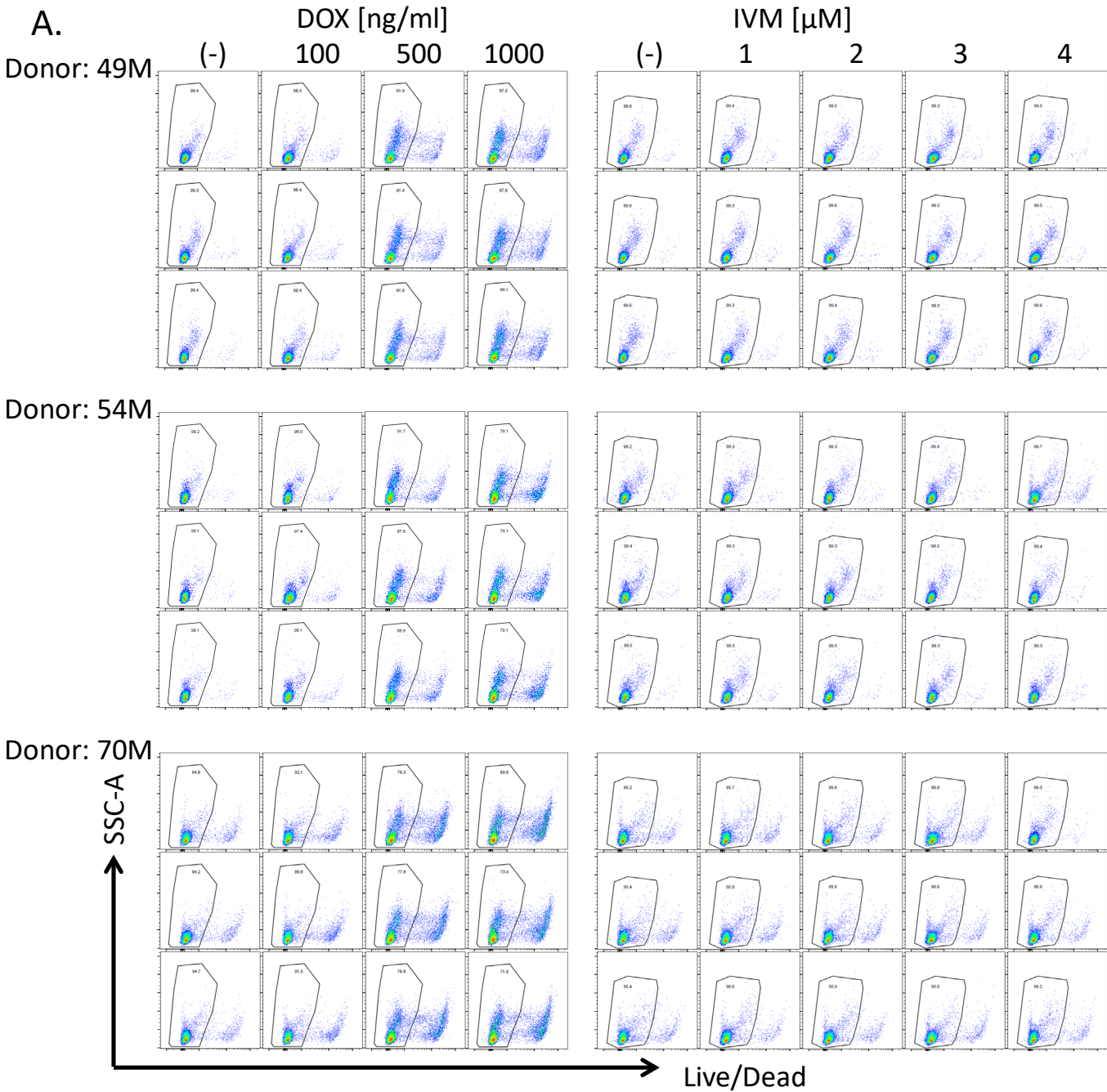


Supplementary Figure 1. (A) Treatment schedules in the neoadjuvant, adjuvant, and metastatic settings (surgical resection = **Sx**). (B) Body weight measurements of treated animals demonstrating the absence of significant synergistic toxicity associated with the combination of anti-PD-1 and Ivermectin in the adjuvant settings. Similar observations were seen in the metastatic treatment settings.



Supplementary Figure 2. Immunomodulatory effects of Ivermectin on immune cells *in*

vivo* and *ex vivo*.** (A) Flow cytometry analysis of splenocytes from 4T1 tumor-bearing animals treated with ivermectin, demonstrating the absence of significant changes *in vivo* of various CD4 (A) and CD8 (B) effector and regulatory T cell subpopulations, which were identified based on the expression of key transcriptional factors as indicated. The transcriptional factors used are well accepted as the most specific markers for different CD4 and CD8 T cell subsets: Tregs (Foxp3+ CD4 T cells), Th1 (Tbet+ CD4 T cells) / Th2 (Gata3+ CD4 T cells) / Th17 (RORgt+ CD4 T cells) / Tc1(Tbet+ CD8 T cells) / Tc17 (RORgt+ CD8 T cells). All comparisons were non-significant, NS. (C) Differential sensitivity of immune subpopulations in splenocytes isolated from 4T1 tumor-bearing mice exposed *ex vivo* to increasing (1-16 μ M) doses of ivermectin for 4h to 48h showing dose and time-dependent sensitivity. A linear mixed effects model of log cell count adjusted for cell type revealed that the CD11b+ myeloid cells were the most sensitive to ivermectin, showing significant reductions with as little as 4 μ M after 48 hours, or 8 μ M after 24 hours, or 16 μ M after 4 hours (each result, $p < 0.0001$). In contrast, achieving similar reductions in lymphocytes required higher doses and/or longer exposure to ivermectin, being observed in CD8+ cells only after 48 hours of 8 μ M or 24 hours of 16 μ M and in CD4+ cells only after the maximum exposure (48 hours of 16 μ M). Statistical significance versus (-) CTRL or as indicated was evaluated using the linear mixed effects model of log cell count adjusted for cell type, * $p \leq 0.0001$.



Supplementary Figure 3. Cytotoxic effects of physiologically relevant levels of

doxorubicin (Dox) and ivermectin (IVM) against human PBMC *ex vivo*. (A) Flow cytometry analysis of the cytotoxic effects of physiologically relevant doses of doxorubicin and Ivermectin, as reported in the literature, against freshly isolated human PBMC treated for 24h *in vitro*.

Peripheral blood from three healthy donors was obtained by venipuncture using heparin collection tubes, transported at room temperature from the clinic to the lab and processed within 6 hours of drawing. PBMCs were isolated via Ficoll-Paque gradient centrifugation (GE Healthcare) following the manufacturer's instruction. One million PBMCs in triplicate from each healthy donor were individually treated with 100, 500, or 1000 ng/ml of Doxorubicin (DOX) or 1, 2, 3 or 4 μ M of Ivermectin (IVM) in RPMI medium (Life Technologies, Thermo Fisher Scientific) containing 10% FBS for 24h at 37°C with 5% CO₂. After 24h of incubation cells were washed once with 1X PBS and stained with Live/Dead™ Fixable Far Red Dead Cell Stain Kit (L34974, Invitrogen, USA,) for 30 minutes in dark at 4°C. Before FACS analysis, cells were washed once with 1X PBS and fixed with 4% PFA. Samples were acquired using a BD Fortessa using FACSDiva 6.1.3. (B) Statistical analysis of the data presented in (A) was performed by using GraphPad Prism version 5 (GraphPad Software Company Incorporation). Values were expressed as mean \pm SEM. Statistical significance was determined by using two-way ANOVA followed by Bonferroni post-tests. Statistical differences were considered significant when the P-value was <0.05.