

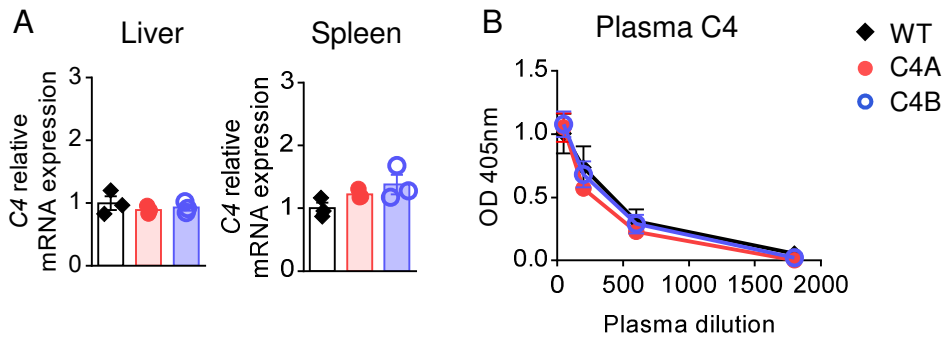
**Cell Reports, Volume 33**

**Supplemental Information**

**Complement C4A Regulates Autoreactive**

**B Cells in Murine Lupus**

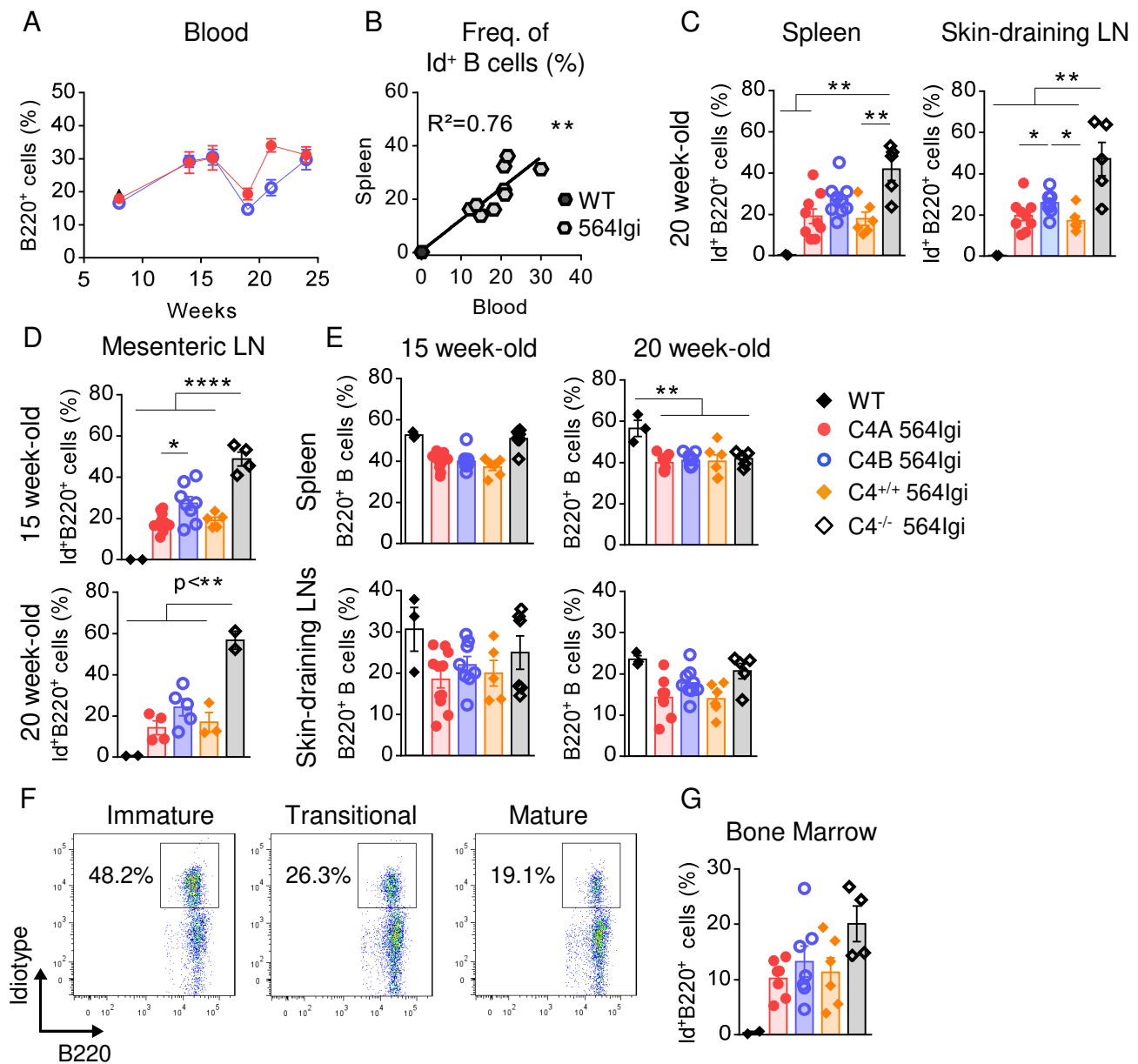
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**Figure S1: C4A and C4B strains present physiological levels of C4, Related to Figure 1.**

(A) *C4* mRNA expression quantification by ddPCR. Each sample was normalized to the endogenous control *eif4h* and relative to *C4* expression in WT. Mean  $\pm$  SEM, one dot represents one animal.  $n=3$  for each strain.

(B) C4 protein level measured in plasma from WT, C4A and C4B mice by ELISA. One dot represents the average value  $\pm$  SEM of  $n=4$  for each strain.



**Figure S2: Analysis of B220<sup>+</sup> B cells and Id<sup>+</sup> B cells at 8, 15 and 20 wo in periphery and in BM, correlation of Id<sup>+</sup> B cells in blood vs spleen, Related to Figure 2.**

(A) Analysis of B220<sup>+</sup> cells percentages in blood circulation by flow cytometry. Mean  $\pm$  SEM, one dot represents the average percentage of n=5 mice for each strain.

(B) Correlation of Id<sup>+</sup> B cells in spleen vs blood by flow cytometry in 564Igi animals. One dot represents one animal, Spearman test, n=9.

(C) Frequency of Id<sup>+</sup> B cell proportions in spleen and skin-draining LN at 20 wo determined by flow cytometry. Mean  $\pm$  SEM, one dot represents one animal, n>5. One-way ANOVA with Tukey's test.

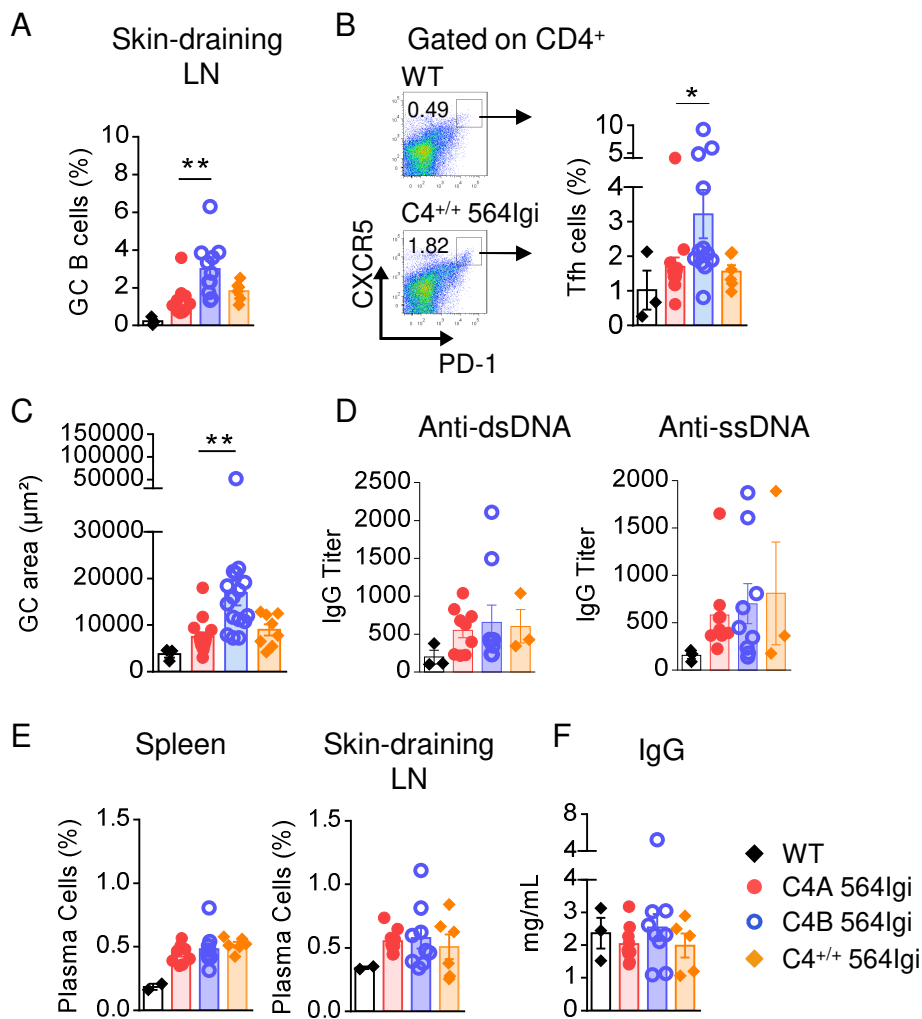
(D) Same as (C) in mesenteric LN at 15 wo and 20 wo, n>4.

(E) Same as (A) in spleen and skin-draining LN at 15 wo and 20 wo, n>5. One-way ANOVA with Tukey's test.

(F) Representative dot plots of Id<sup>+</sup> B220<sup>+</sup> cell population (gated on B220<sup>+</sup> cells) within immature (B220<sup>+</sup> AA4.1<sup>+</sup>), transitional (B220<sup>+</sup> AA4.1<sup>int</sup>) and mature (B220<sup>+</sup> AA4.1<sup>-</sup>) subpopulations of 15 wo 564Igi animals.

(G) Same as (C) in bone marrow at 15 wo, n>4.

(\*<0.05, \*\*<0.01, \*\*\*< 0.001, \*\*\*\*<0.0001)



**Figure S3: Analysis of GC and PC, measurement of anti-DNA autoantibody titers and IgG levels, Related to Figure 3.**

(A) GC B cell population (B220<sup>+</sup> CD38<sup>dim/-</sup> GL7<sup>high</sup>) analyzed by flow cytometry in skin-draining LN from 15 wo C4A, C4B and C4<sup>+/+</sup>564Igi. Mean ± SEM, one dot represents one mouse, n>6. One-way ANOVA with Tukey's test.

(B) Splenic Tfh cells (CD4<sup>+</sup> PD-1<sup>high</sup> CXCR5<sup>+</sup>) from 15 wo C4A, C4B and C4<sup>+/+</sup> 564Igi mice, analyzed by flow cytometry. Mean ± SEM, one dot represents one mouse, n>6. One-way ANOVA with Tukey's test.

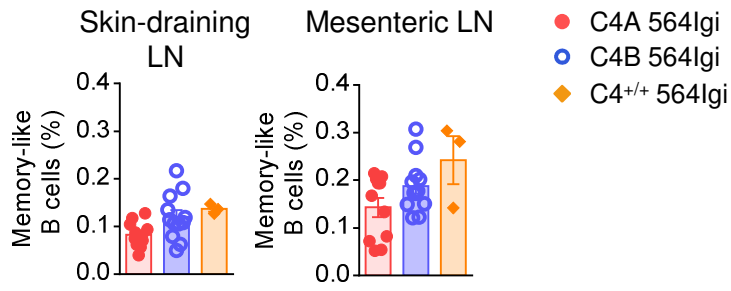
(C) Splenic GL7<sup>+</sup> area quantification. Spleens were stained for marginal zone macrophages: CD169, B cell zone: B220, T cell zone: CD3 and GC: GL7. Individual GL7<sup>+</sup> areas were measured using ImageJ software. Mean ± SEM, one dot represents the average value obtained for one mouse, n>8. One-way ANOVA with Tukey's test.

(D) Serum anti-dsDNA and -ssDNA IgG titers measured by ELISA. Mean ± SEM, one dot represents one mouse, n>3.

(E) Plasma cells (B220<sup>-</sup> CD138<sup>+</sup>) from 15 wo C4A, C4B and C4<sup>+/+</sup>564Igi mice (spleen (left) and skin-draining LNs (right)) analyzed by flow cytometry. Mean ± SEM, one dot represents one mouse, n>6 .

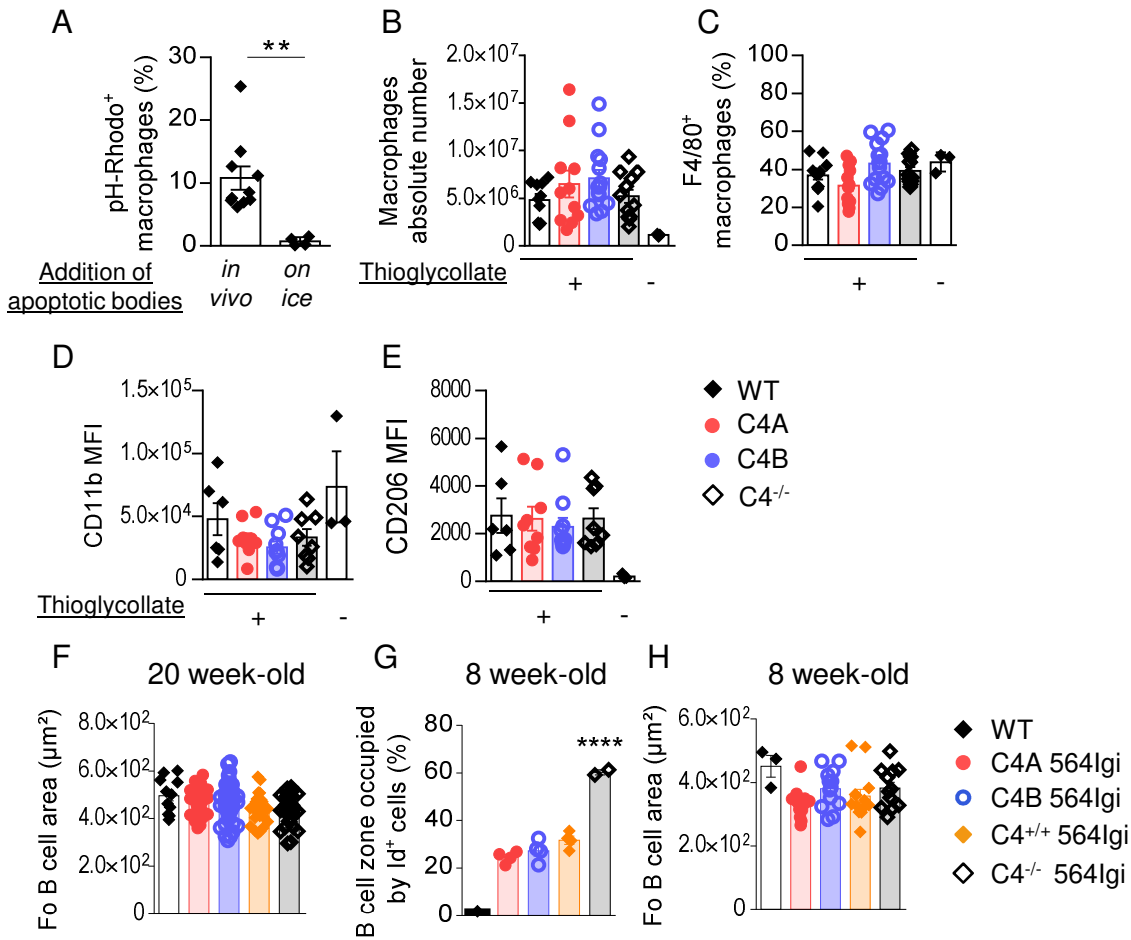
(F) Serum IgG levels measured by ELISA. Mean ± SEM, one dot represents one mouse, n>5.

(\*<0.05, \*\*<0.01)



**Figure S4: Analysis of autoreactive memory-like B cells, Related to Figure 4.**

Analysis of autoreactive memory-like B cell population (B220<sup>+</sup> YFP<sup>+</sup> CD138<sup>-</sup> GL7<sup>-</sup>) at 15 wo by flow cytometry. Mean ± SEM, one dot represents one mouse, n>3.



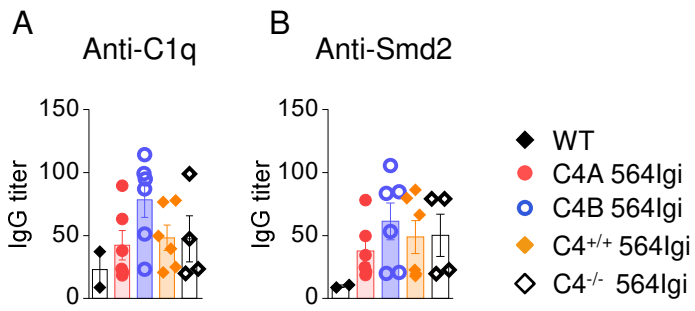
**Figure S5: Elicited peritoneal macrophages phenotype and follicular exclusion of Id<sup>+</sup> B cells analysis, Related to Figure 5.**

(A) Assessment of phagocytosis activity on ice. Macrophages were incubated on ice for 3 hours (time needed to perform the experiment) with apoptotic cells and then analyzed for apoptotic cell uptake. Mean  $\pm$  SEM, one dot represents one mouse,  $n > 4$ . Mann and Whitney test,  $** < 0.005$ .

(B-E) Characterization of elicited macrophages. The absolute numbers of F4/80<sup>+</sup> peritoneal macrophages (B), their frequency (C), and the expression of CD11b (D) and CD206 (E) at the surface were analyzed by flow cytometry. Mean  $\pm$  SEM, one dot represents one mouse,  $n > 10$ .

(F) Follicular B cell zone area analyzed per field of view of splenic sections from 20 wo mice. Mean  $\pm$  SEM, each dot represents one field of view,  $n > 31$  per strain.

(G-H) Analysis of Id<sup>+</sup> B cell follicular exclusion at 8 wo using cell profiler software. (G) Quantification of follicular exclusion expressed as % of follicular B cell zone occupied by Id<sup>+</sup> B cells. Mean  $\pm$  SEM, each dot represents one mouse,  $n = 4$  for all groups except  $n = 2$  for C4<sup>-/-</sup> 564lgi. One-way ANOVA with Tukey's test,  $**** < 0.001$ ; (H) Follicular B cell zone area analyzed per field of view of splenic sections from 8 wo mice. Mean  $\pm$  SEM, each dot represents one field of view,  $n > 12$ .



**Figure S6: Measurement of anti-C1q and anti-Smd2 IgG titers, Related to Figure 6.**

(A-B) Dosage of anti-C1q and anti-Smd2 IgG titers in serum of 15-20 wo C4<sup>-/-</sup>, C4A, C4B and C4<sup>+/+</sup> 564Igi by ELISA. Mean  $\pm$  SEM, one dot represents one mouse, n>4. One-way ANOVA with Tukey's test.

**Table S1. Cas9 DNA templates used to generate C4A and C4B gene-edited strains, Related to STAR Methods.**

Cas9 DNA template	
C4A	ccaggaacagggtgggcaactccccggagaagctgcaggagacggctagctggctgctggcccagcagctgggtgatggctccttcaggatccctgtccagtgtagacaggagcatgc agggtgtaacacgctggaggcacgtgtaggatgtcagcaggaagacgggctcctcacaactctggcctgacagcctctt
C4B	gagaagctgcaggagacggctagctggctgctggcccagcagctgggtgatggctccttcaggacctctctccggtgatacataggagcatgcaggtgtgaacacgctggaggcacgt gtaggatgtcagcaggaagacgg gctcctca



**Table S2. Primer sequences and reaction conditions used for PCR genotyping, Related to STAR Methods.**

Trans-genes	Primers (IDT)	Sequences (5'-3')	Amplification program				Fragment size (bp)	
			I. 94°C	II. Cycles		III. 72°C		IV. 8°C
				Description	Number			
564 Light Chain	564k	CCAGTGCAGATTTTCAGCTTC	4 min	94°C 1min 55°C 1min 72°C 1min	40 cycles	5 min	∞	724 bp
	Jk5	CAGCTTGGTCCCAGCACCGAA						
564 Heavy Chain	VH4emu	CACAGATTCTTAGTTTTTCAA	4 min	94°C 1min 50°C 1min 72°C 1min	40 cycles	5 min		542 bp
	VH564	TGGAGCTATATCATCCTCTTT						
C4 KO	C4 23F	ATAACTGGGTCCGACTTTGG	4 min	94°C 1min 56°C 1min 72°C 1min	30 cycles	10 min		Mutant: 800 bp WT: 600 bp
	C4 25R	TCTTCCGAAACTGCTGGATCC						
	C4 29R	TACCTGGGTACTGCGGAATGC						
	Neo4483	AAGCCGGTCTTGTCGATCAG						
C4Crispr_P CR1	3' mC4 ex22-28	GTCGACCAAGTCCCAGGTGCTCTGC C	2 min	95°C 30sec 65°C 30sec 72°C 1min30	35 cycles	5 min		N/A
	5' mC4 ex22-28	GTCGACCTGGCCTTGAACTTCAGTA ATC						
C4Crispr_P CR2	C4BF1	TCACCTGAAGTCCATGGACCCTG	2 min	95°C 30sec 65°C 30sec 72°C 2min	35 cycles	5 min	978 bp	
	C4BR1	CATCGTATCCTGGAAGACGTCCAA						

**Table S3. Post-PCR digestion conditions to genotype C4A and C4B mouse strains, Related to STAR Methods.**

Transgenes	Fragment size before digestion	Restriction enzyme	Digestion conditions	Fragment size after digestion
C4A	978 bp	BamHI	2 hours – 37°C	WT: 515 + 463 bp Mut: 293 + 222 + 463 bp
C4B	978 bp	BsAWI	2 hours – 60°C	WT: 978bp Mut: 765 + 213 bp

**Table S4. Primer sequences and reaction conditions used to measure *C4* mRNA expression by ddPCR, Related to STAR Methods.**

Transgenes	Primers	Sequences (5'-3')	Amplification program				
			I. 94°C	II. Cycles		III. 72°C	IV. 8°C
				Description	Number		
<i>C4</i>	Forward	AGCGTGTTTCCAGCTCAAAG	4 min	94°C 1min 55°C 1min 72°C 1min	40 cycles	5 min	∞
	Reverse	GTCCTAAGGCCTCACACCTG					
<i>eif4h</i>	Forward	TGCAGCTTGCTTGGTAGC		94°C 1min 50°C 1min 72°C 1min			
	Reverse	GTAATTGCCGAGACCTTGC					

**Table S5. ELISA conditions, Related to STAR Methods.**

Measurement	Coating conditions	Sample dilutions in PBS-T 0.1%	Standard	Detection
C4 level	1.5µg/mL rat anti-mouse C4 (gift of E. Kremmer, Munich, Germany)	1/50 to 1/5000	N/A	4µg/mL of rabbit anti-human C4c (Dako) + goat anti-Rabbit-AP (Southern biotech, 1/1000)
IgG level	1.5µg/mL goat anti-mouse IgG (southern Biotech)	1/500 to 1/250 000	Mouse IgG (sigma) from 1000ng/mL to 1.9ng/mL	Goat anti-mouse IgG-AP (Southern biotech, 1/1000)
IgG2a level	1.5µg/mL goat anti-mouse IgG 2a (southern Biotech)	1/500 to 1/250 000	Purified mouse IgG2a (southern biotech) from 1000ng/mL to 1.9ng/mL	Goat anti-mouse IgG2a-AP (Southern biotech, 1/1000)
Anti-nucleoli IgG titers	100µL of purified nucleoli per 96-well plate, spun 10min at 1300g	1/25 to 1/200	N/A	Goat anti-mouse IgG-AP (Southern biotech, 1/10000)
Anti-Ro60 IgG titers	1.5µg/mL Ro60 protein (Diarect)	1/20 to 1/1280	N/A	Goat anti-mouse IgG-HRP (Southern biotech, 1/10000)
Anti-BPI IgG titers	1.5µg/mL BPI protein (Diarect)			
Anti-CTGF IgG titers	1.5µg/mL CTGF protein (R&D system)			
Anti-Smd2 IgG titers	1µg/mL Smd2 protein (Diarect)			
Anti-C1q IgG titers	1µg/mL C1q protein (Meridian Life Science)			
Anti-APRIL IgG titers	1.5µg/mL APRIL protein (PEPROTECH)			

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## Antigens

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Human IgG from serum  
Anti-Human IgG Fc fragment Specific  
Anti-Human IgG (H+L)  
Anti-Human IgG F(ab') fragment specific  
La/SSB  
Ro 52/SSA  
CENP B  
EBV EBNA-1  
EGFR  
FBL (Fibrillarin/U3RNP)  
Hepatitis B Surface Antigen HBSAg  
U11/U12  
Ku, p70/p80  
Nucleolin  
PDC-E2  
PM/Scl 100  
POLR3H  
Pyruvate dehydrogenase (PDH)  
Ro 60/SSA (bovine)  
Ro 60/SSA (recombinant)  
RPP14 (Th/To)  
Scl-70, full-length  
Scl-70, truncated  
Sm/RNP (SRC-3000)  
Smith (SMA-3000)  
ssDNA  
TG  
TPO (Thyropoxidase)  
U1-snRNP 68  
U1-snRNP A  
U1-snRNP C  
RPP25 (Th/To)  
MDC  
TNF-B  
GBM  
APRIL  
CTGF  
IFN-g  
IL17-A  
Catalase  
SSA/Ro60  
BPI  
Smd2  
TPO  
OVA  
C1q

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**Table S6. List of self-antigens and controls tested to measure sera autoreactivity,** Related to Figure 6 and STAR Methods.