

Fig. S1 The map of SARS-CoV-2 specific IgG binding epitopes revealed from convalescent sera directly. (A) The epitopes (**Table S3**) of each patient (P1-P55) were plotted alongside the known SARS-CoV-2 proteins from N terminal to C terminal. The total number of the identified epitopes for each patient and each protein were defined as frequency and sum_epitopes, respectively. (**B**) The histogram of the epitope frequency (bin = 4). (**C**) The sum_epitopes of each protein, length of proteins and ratio of sum_epitopes and length.



Fig. S2 Protein specific antibodies enrichment from COVID 19 convalescent sera. COVID-19 convalescent sera were used as input. Terminal biotinylated proteins were used as baits to enrich specific antibodies. SA conjugated magnetic beads were adopted to isolate the antibodies.



Fig. S3. Protein based antibody enrichment is necessary for AbMap. S protein specific epitopes identified from sera directly (black dot), and S1+S2 protein enriched antibodies (red dot).

4				E	3
No	. ID	Sequence	Position	Frequency	28-YTNSFTR-34 577-RDPQTLE-583
1	Epitope-S1-1	AYTNSFTR	27-34	12	
2	Epitope-S1-2	FRSSVLHS	43-50	4	64-WFHAIH-69 558-KFLPFQQ-56
3	Epitope-S1-3	VTWFHAIH	62-69	3	
4	Epitope-S1-4	KQGNFKNL	182-189	4	
5	Epitope-S1-5	I <mark>Y</mark> QTSNFR	312-319	4	
6	Epitope-S1-6	KCVNFNFN	537-544	3	629-LTPTWRVY-636 550-GVLTESN-556
7	Epitope-S1-7	GVL TESNK	550-557	7	818-IEDLLFNK-825
8	Epitope-S1-8	KFLPFQQF	558-565	34	695-YTMSL-699
9	Epitope-S1-9	VRDPQTLE	576-583	6	805-ILPDPSK-811
10	Epitope-S1-10	DQLTPTWR	627-634	5	795-KDFGGFNF-802
11	Epitope-S1-11	QLTPTWRV	628-635	10	
12	Epitope-S1-12	LTPTWRVY	629-636	15	at 3 to
13	Epitope-S1-13	TPTWRVYS	630-637	4	
14	Epitope-S2-1	IIAYTMSL	692-699	3	() 120°
15	Epitope-S2-2	AYTMSLGA	694-701	5	Ť
16	Epitope-S2-3	Q <mark>E</mark> VFAQV <mark>K</mark>	779-786	5	
17	Epitope-S2-4	Y <mark>KTPP</mark> IKD	789-796	6	537-KCVNFNFN-544
18	Epitope-S2-5	KTPP IKDF	790-797	3	43-FRSSVLH-49
19	Epitope-S2-6	KDFG <mark>GF</mark> NF	795-802	7	64-WFHAIH-69
20	Epitope-S2-7	QILPDPSK	804-811	7	
21	Epitope-S2-8	ILPDPSKP	805-812	9	Martin Branch Bar
22	Epitope-S2-9	IEDLLFNK	818-825	4	818-IEDLLFNK-825
23	Epitope-S2-10	CLGDIAAR	840-847	3	
24	Epitope-S2-11	FNSAIGKI	927-934	3	003-ILPDPSR-011
25	Epitope-S2-12	FKEELDKY	1148-1155	4	780-EVFAQVK-786- 928-NSAIG-932
26	Epitope-S2-13	DKYFKNHT	1153-1160	5	
27	Epitope-S2-14	EPVLKGVK	1262-1269	3	790-KTPP-793
28	Epitope-S2-15	KGVKLHYT	1266-1273	4	- 795-KDFGGFNF-802
29	Epitope-RBD-1	RDISTEIY	466-473	2	

Fig. S4. Detailed information of the significant epitopes on S protein. (**A**) The list of the significant epitopes (frequency >=3), (**B**) The distribution of the significant epitopes (frequency>=3) on the trimer 3D structure of the S protein. Red marked amino acids represent critical residues of epitopes.



Fig. S5

RBD

Fig. S5. Functional analysis of several key sites of the significant epitopes of S protein. (A) One relatively significant epitope (frequency >= 2) was identified for RBD enriched antibody. (B) Match the critical epitope residues to the structure of RBD. (C) The critical epitope residues locate adjacent to but not at the binding interfaces of RBD and ACE2. (D-J) ¹⁻⁶ The critical epitope residues locate adjacent to but not at the binding interfaces of RBD strong binding antibodies or highly potent neutralization antibodies, CR3022-Fab (D), S309-Fv (E), P2B-2F6-Fab (F), BD23-Fab (G), CB6-Fab (H), S2H13 (I) and S2H14 (J).



Fig. S6. Match the critical epitope residues with naturelly existing mutants of Spike protein. (**A**) Match the epitopes on S protein with the critical naturally existing mutants ^{7,8}. (**B**) The Substitution mutants of Spike protein collected by China National Center for Bioinformation were plotted alongside the Spike protein with the mutated frequency. (The data were collected by 2021-1-12)



100 150 200 250 300 350 400 450

CTD

Position

T

SRNSSRNS

FPPTEPKKDKKKKADETQALPQRQKK QQTVTLLPAADLDDFSKQLQQSMSSA DS

No.	ID	Sequence	Position	Frequency
1	Epitope-N-1	SRNSSRNS	190-197	8
2	Epitope-N-2	FPPTEPKK	363-370	3
3	Epitope-N-3	QALPQRQK	380-387	3
4	Epitope-N-4	KQQTVTLL	388-395	3
5	Epitope-N-5	TLLPAADL	393-400	4
6	Epitope-N-6	KQLQQ <mark>SMS</mark>	405-412	3



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4-								BtCoV-R
3-								SARS-Co
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ARS-CoV-2	FPPTEPKK
tCoV-RaTG13	B FP <mark>PTEPKK</mark>
ARS-CoV	FP <mark>PTEPKK</mark>
ERS-CoV	FPKK <mark>E-</mark> K <mark>K</mark>
CoV-OC43	QQDGMM
CoV-HKU1	SNQNTDSD
CoV-NL63	PSSIKEMQ
CoV-229E	EMQQHP

363

370

Ş	SARS-CoV-2 BtCoV-RaTG1 SARS-CoV MERS-CoV HCoV-OC43	388 KQQTVI 3 KQQTVI KQPTVI EQRVQQ ENDNIS	395 LL LL LL SSI SVA
N.	HCoV-OC43 HCoV-HKU1	ENDNIS QFDSLN	3VA JLS
A	HCoV-NL63	ESKPLA	ADD
N 80	HCoV-229E	TAEP <mark>v</mark> f	١DE

	393 400	0		405 412
ARS-CoV-2	TLLPAADL		SARS-CoV-2	KQLQQ <mark>SMS</mark>
CoV-RaTG13	3 TLLPAADL	4-	BtCoV-RaTG1	3 <mark>K</mark> QLQQ <mark>SMS</mark>
ARS-CoV	TLLPAADM	3-	SARS-CoV	RQLQN <mark>SMS</mark>
ERS-CoV	GSITQRTR		MERS-CoV	VQPGPMID
CoV-OC43	SVAVPKSRVQQNKSREL		HCoV-OC43	ISLLKK <mark>M</mark> D
CoV-HKU1	NLSAGTQHISNDF		HCoV-HKU1	HSLLATLD
CoV-NL63	ADDDS		HCoV-NL63	IEIVNEVL
CoV-229E	RDEVS	0 - 0 0 7 8 0 0 - 0	HCoV-229E	TDIIDEVN
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					SARS-CoV-2	190 RSR <mark>N</mark> S	197 SRNS
4-			1		BtCoV-RaTG	13 rsr <mark>n</mark> s	<mark>S</mark> RNS
3-					SARS-CoV	RSRGN	<mark>S</mark> RNS
s			ΓΙ	M	MERS-CoV	VSR <mark>N</mark> S	SRSS
ii 2-		N		N	HCoV-OC43	RTSSR	ASSA
1-			4 I		HCoV-HKU1	RSQSR	GP <mark>N</mark> N
		=	×	⊉	HCoV-NL63	RNNSR	DSSR
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Frequency

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16-

8

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RNAPRITF

50

NTD

		380	387
	SARS-CoV-2	QALPQR	QK
	BtCoV-RaTG13	3 QA <mark>lp</mark> Qr	Q <mark>K</mark>
	SARS-CoV	QPLPQR	QK
	MERS-CoV	DQMSEF	P.
V	HCoV-OC43	RGHKNG	ŞQG
Ν	HCoV-HKU1	RGVKQI	PE
B	HCoV-NL63	TVLNAS	JP
æ	HCoV-229E	NPSOTS	βPA



Fig. S7. Significant epitopes on N protein. (**A**) The distribution of the sequence-matched epitopes on N protein. (**B**) The list of significant epitopes (frequency >= 3). (**C**) Homology analysis of the significant epitopes among three deadly coronaviruses, i. e., SARS-CoV-2, SARS-CoV and MERS-CoV, four common human coronaviruses, i. e., HCoV-OC43, HKU-1, NL63 and 229E and the highly homologous bat coronavirus BtCoV-RaTG13.

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	LV-1	I-K5	-F5{	I-L56	-P56	-F56	-Q5	-Q5	-F56		LM-	5-L62	2-T63	-Pep	2-T63	2-W6	-R6;	5-V6;	2-Y6;	Σ	BSA
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Pep3-T1160A

DKYFKNH**A**

Pep4-Y4T3A

RDISTEIA

Fig. S8. The layout of the peptide microarray of 4 significant epitopes. (**A**) Quality control of the peptide microarray. All the peptides were chemically synthesized and conjugated to BSA. The BSA conjugated peptides were printed and immobilized on the microarray. The microarray was stained with an anti-BSA antibody and followed with a Cy5 conjugated secondary antibody. (**B**) The layout of the peptide microarray. (**C**) The sequences of the peptides that were included on the peptide microarray.



Α

Fig. S9

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Fig. S9. Validation of critical residues in epitopes by a peptide microarray. (A) Selected samples were subjected for peptide microarray validation. The summary of the validation results. (B) The microarray results of selected samples against three representative epitopes. Sample with red label indicates the corresponding epitope was identified from this sample by AbMap. While other samples were included as negative controls for corresponding epitopes.



Fig. S10. Validation of epitope identification procedures by a peptide microarray for more samples: the comparison of sera and protein enriched antibodies. Peptide 1 (Pep 1), which corresponds to Epitope-S1-8 (KLFPFQQF) was selected as an example. Sample with red label indicates the corresponding epitope was identified from this sample by AbMap. The ratio of signal_intensity (S1 enriched)/ signal_intensity (serum) for each residue was plotted (right).

C(OVID-19 patients	n = 55				
Gender	Male Female	27 28				
Age	Mean ± s.d. < 60 ≥ 60	41.5 ± 14.8 48 7				
Severity	Mild Severe and critical	55 0				
Outcome	Recovered Death	55 0				
Days after s	ymptom onset for sampling	27.5 ± 7.7				
	Control group	n = 25				
ŀ	Healthy controls	5				
	Lung cancer	20				
Gender	Male Female	15 10				
Age	Mean ± s.d. < 60 ≥ 60	51.7 ± 12.7 18 7				

Table S1. Serum samples tested in this study.