



Supplementary Information for

A Tandem Activity-Based Sensing and Labeling Strategy Enables Imaging of Transcellular Hydrogen Peroxide Signaling

Hidefumi Iwashita^a, Erika Castillo^e, Marco S. Messina,^a Raymond A. Swanson^{d,e}, and Christopher J. Chang^{a,b,c*}

^aDepartment of Chemistry, ^bDepartment of Molecular and Cell Biology, and ^cHelen Wills Neuroscience Institute, University of California, Berkeley, California 94720, United States

^dNeurology Service, San Francisco Veterans Affairs Health Care System, San Francisco, California 94121, United States

^eDepartment of Neurology, University of California San Francisco and San Francisco Veterans Affairs Medical Center, San Francisco, California 94143, United States

Corresponding Author: *E-mail: chrischang@berkeley.edu

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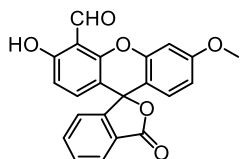
Supplementary Text
Figures S1 to S22
SI references

Synthesis

PF2. Synthesis of **PF2** was synthesized according to literature procedures^[1].

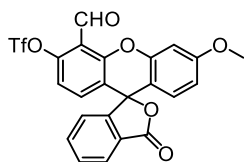
Compound 1. Compound **1** was synthesized according to literature procedures^[2].

Compound 2



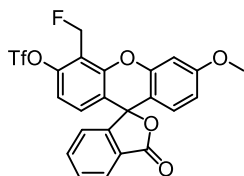
To a stirred solution of compound **1** (2.4 g, 6.9 mmol) in trifluoroacetic acid (70 mL) was added hexamethylenetetramine (1.0 g, 7.6 mmol, 1.1 equiv). The mixture was stirred at 90 °C for 16 h and then H₂O (70 mL) was added to the mixture. The resulting mixture was stirred at 95 °C for another 1 h, and then cooled to room temperature. The mixture was extracted with CH₂Cl₂ three times, dried over Na₂SO₄, and evaporated. The crude product was purified by silica-gel column chromatography (eluent: CH₂Cl₂/MeOH = 100/0 to 100/1) to yield 900 mg (34%) of **2** as a white solid. ¹H NMR (300 MHz, CDCl₃) δ 12.17 (s, 1H), 10.68 (s, 1H), 8.10 – 7.97 (m, 1H), 7.67 (ddd, *J* = 8.8, 7.2, 1.3 Hz, 2H), 7.21 – 7.12 (m, 1H), 6.91 (d, *J* = 8.9 Hz, 1H), 6.86 – 6.80 (m, 1H), 6.80 – 6.49 (m, 3H), 3.89 – 3.80 (m, 4H); ¹³C NMR (101 MHz, CDCl₃) δ 193.38, 169.20, 164.56, 161.65, 153.26, 152.66, 151.40, 137.15, 135.42, 130.20, 129.27, 126.80, 125.36, 123.93, 113.92, 112.78, 111.12, 109.81, 109.01, 101.05, 82.09, 55.81. LRMS: calcd for [M+H]⁺, 375.0; found 375.0.

Compound 3



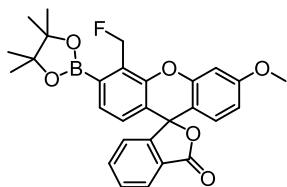
A mixture of compound **2** (900 mg, 2.4 mmol), *N*-phenyl-bis(trifluoromethanesulfonimide) (1.28 g, 3.6 mmol, 1.5 equiv) and Cs₂CO₃ (1.56 g, 4.8 mmol, 2.0 equiv) in MeCN (30 mL) was stirred at ambient temperature for 1 h. After the filtration, the filtrate was evaporated under the reduced pressure. The resulting residue was purified by silica-gel column chromatography with CH₂Cl₂ eluent to yield 1.2 g (98%) of **3** as a pale yellow solid. ¹H NMR (300 MHz, CDCl₃) δ 10.79 (d, *J* = 0.7 Hz, 1H), 8.11 – 8.02 (m, 1H), 7.80 – 7.63 (m, 2H), 7.20 (dt, *J* = 7.6, 0.9 Hz, 1H), 7.13 (d, *J* = 8.8 Hz, 1H), 7.00 (d, *J* = 8.8 Hz, 1H), 6.88 (dd, *J* = 2.2, 0.7 Hz, 1H), 6.76 – 6.72 (m, 2H), 3.87 (s, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 185.57, 168.87, 162.08, 153.68, 152.27, 151.27, 148.34, 135.80, 134.87, 130.65, 129.73, 129.05, 127.52, 126.22, 125.70, 123.95, 123.64, 121.41, 117.78, 117.32, 113.54, 110.32, 101.19, 80.97, 77.48, 77.36, 77.16, 76.84, 55.93; ¹⁹F NMR (376 MHz, CDCl₃) δ -72.38. LRMS: calcd for [M]⁺, 506.0; found 506.9.

Compound 4



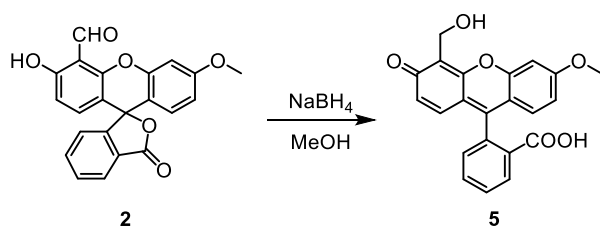
To a solution of compound **3** (50 mg, 0.098 mmol) in THF-MeOH (2 mL/2 mL) was added AcOH (6 mL, 0.098 mmol, 1.0 equiv) and NaBH(OAc)₃ (25 mg, 0.118 mmol, 1.2 equiv). The mixture was stirred for 1 h at ambient temperature under N₂ atmosphere, diluted with sat. NH₄Cl aq., and then extracted with EtOAc three times. The combined organic layer was dried over Na₂SO₄, and concentrated. The crude product was dissolved in CH₂Cl₂ (10 mL) and cooled to -20 °C. (Dimethylamino)sulfur trifluoride (DAST, 15 mL, 0.118 mmol, 1.2 equiv) was added to the mixture in one portion and stirred at ambient temperature for 1 h. After addition of MeOH to quench excess of DAST, the mixture was evaporated. The crude product was purified by silica-gel column chromatography with CH₂Cl₂ eluent to yield 40 mg (80%) of **4** as a white solid. ¹H NMR (400 MHz, CDCl₃) δ 8.05 (dq, *J* = 7.4, 0.9 Hz, 1H), 7.76 – 7.62 (m, 2H), 7.19 (dt, *J* = 7.5, 0.9 Hz, 1H), 7.07 (d, *J* = 8.9 Hz, 1H), 6.97 (ddd, *J* = 8.9, 2.3, 0.7 Hz, 1H), 6.88 (d, *J* = 2.4 Hz, 1H), 6.76 – 6.65 (m, 2H), 5.95 – 5.61 (m, 2H), 3.87 (d, *J* = 0.7 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 169.00, 161.93, 152.58, 151.75, 151.26, 149.13, 135.61, 131.20, 130.43, 129.04, 126.40, 125.54, 124.03, 120.35, 117.21, 116.64, 113.14, 110.49, 101.13, 81.66, 73.90, 72.23, 55.87, 30.45, 29.84.; ¹⁹F NMR (376 MHz, CDCl₃) δ -72.43, -210.36. LRMS: calcd for [M+H]⁺, 511.0; found 511.0.

Peroxy Green 1- Fluoromethyl (PG1-FM)



A mixture of **4** (160 mg, 0.31 mmol), bis(pinacolato)diboron (102 mg, 0.40 mmol, 1.3 equiv), Pd(dppf)Cl₂ (22 mg, 0.031 mmol, 0.1 equiv), and potassium acetate (91 mg, 0.93 mmol, 3.0 equiv) in dioxane (3 mL) was stirred at 90 °C for 3 h. After being cooled, the reaction mixture was diluted with CH₂Cl₂, filtered over Celite pad and concentrated. The crude product was purified by silica-gel column chromatography with CH₂Cl₂ eluent and washing with MeOH to yield 90 mg (59%) of **PG1-FM** as a white solid. ¹H NMR (300 MHz, CDCl₃) δ 8.03 (dd, *J* = 6.1, 2.1 Hz, 1H), 7.63 (ddd, *J* = 6.8, 4.7, 1.4 Hz, 2H), 7.46 (d, *J* = 7.9 Hz, 1H), 7.11 (dd, *J* = 6.4, 2.1 Hz, 1H), 6.91 – 6.79 (m, 2H), 6.72 (d, *J* = 8.8 Hz, 1H), 6.63 (dd, *J* = 8.8, 2.5 Hz, 1H), 6.03 (d, *J*_{H-F} = 47.1 Hz, 2H), 3.86 (s, 3H), 1.35 (d, *J* = 1.3 Hz, 13H); ¹³C NMR (101 MHz, CDCl₃) δ 169.57, 161.65, 153.55, 152.16, 149.98, 135.24, 130.19, 130.17, 129.94, 129.35, 129.20, 128.91, 128.48, 126.29, 125.28, 123.92, 121.74, 112.40, 110.82, 101.07, 84.42, 82.53, 78.62, 55.77, 24.91; ¹⁹F NMR (376 MHz, CDCl₃) δ -204.00. HRMS: calcd for [M+H]⁺, 489.1879; found, 489.1883 (+0.0004 mmu).

Compound 5



To a solution of compound **2** (40 mg, 0.106 mmol) in THF-MeOH (2 mL/2 mL) was added NaBH₄ (5 mg, 0.128 mmol, 1.2 equiv). The mixture was stirred for 16 h at ambient temperature under N₂ atmosphere, diluted with 2N HCl, and then extracted with EtOAc three times. The combined organic layer was dried over Na₂SO₄, and concentrated. The crude product was purified by silica-gel column chromatography (eluent: CH₂Cl₂/MeOH = 100/2.5) and washing with MeOH to yield 20 mg (50%) of compound **5** as a yellow solid. ¹H NMR (300 MHz, CDCl₃) δ 8.04 – 7.95 (m, 1H), 7.72 – 7.54 (m, 2H), 7.21 – 7.09 (m, 1H), 6.72 (d, *J* = 2.4 Hz, 1H), 6.69 – 6.51 (m, 4H), 5.28 – 5.14 (m, 2H), 3.79 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 170.02, 161.54, 158.72, 152.80, 152.41, 149.11, 135.25, 129.91, 129.07, 128.24, 126.92, 125.23, 124.20, 113.29, 112.00, 111.75, 111.13, 110.50, 100.94, 58.24, 55.73. LRMS: calcd for [M+H]⁺, 377.0; found 377.0.

ROS Selectivity Analysis

PG1-FM was dissolved in DMSO to obtain a 1 mM stock solution. The fluorescence spectra of **PG1-FM** (1 μ M) were measured in PBS (-) containing 25 μ M of H₂O₂, OCl⁻, TBHP, O₂⁻, ONOO⁻, or NO. OCl⁻, O₂⁻, ONOO⁻, or NO were generated by using NaOCl (Fluka), potassium superoxide (SIGMA), Peroxynitrite (Cayman), and PROLI NONOate (Cayman), respectively. The fluorescence spectra of **PG1-FM** were collected an emission window between 500 nm to 650 nm with excitation at 488 nm.

HPLC Analyses of the Reaction Between PG1-FM and H₂O₂

PG1-FM (10 μ M) and compound **5** (10 μ M) were diluted with eluent (A/B = 50/50(v/v)) or PBS (-) to prepare 100 μ M solutions containing 1% DMSO. The reaction mixture of **PG1-FM** (100 μ M) and H₂O₂ (10 μ M) in PBS (-) was incubated at 37 °C for 30 min, and then analyzed by HPLC. LC analyses were performed on an Agilent Technologies 1200 system using eluent A (H₂O containing 0.05% formic acid) and eluent B (acetonitrile containing 0.05% formic acid), and monitored at 254 nm and 470 nm.

SDS-PAGE Experiments in the Presence of BSA

PG1-FM (10 μ M) and BSA (0.5 mg/mL) were incubated at 25 °C for 1 h with or without H₂O₂ (100 μ M). Afterward, Ebselen (100 μ M) was added to the mixture to quench excess H₂O₂. The obtained samples were separated with NovexTM Mini Gel (Thermo). The gel was first analyzed by ChemiDocTM MP (Bio-Rad) to obtain fluorescence images (488 nm excitation) and stained with Coomassie brilliant blue (CBB) solution.

SDS-PAGE Experiment in the Presence of Cell Lysate

PG1-FM (10 μ M) or **Ctrl-PG1** (10 μ M) were gently mixed with MFP 231 cell lysate (1 mg/mL) and treated with varying concentrations of H₂O₂ (200-1000 μ M). Samples were incubated at 25 °C for 1 h. The obtained samples were separated with NovexTM Mini Gel (Thermo). The gel was first analyzed by ChemiDocTM MP (Bio-Rad) to obtain fluorescence images (488 nm excitation) and stained with Coomassie brilliant blue (CBB) solution.

Supplementary figures and legends

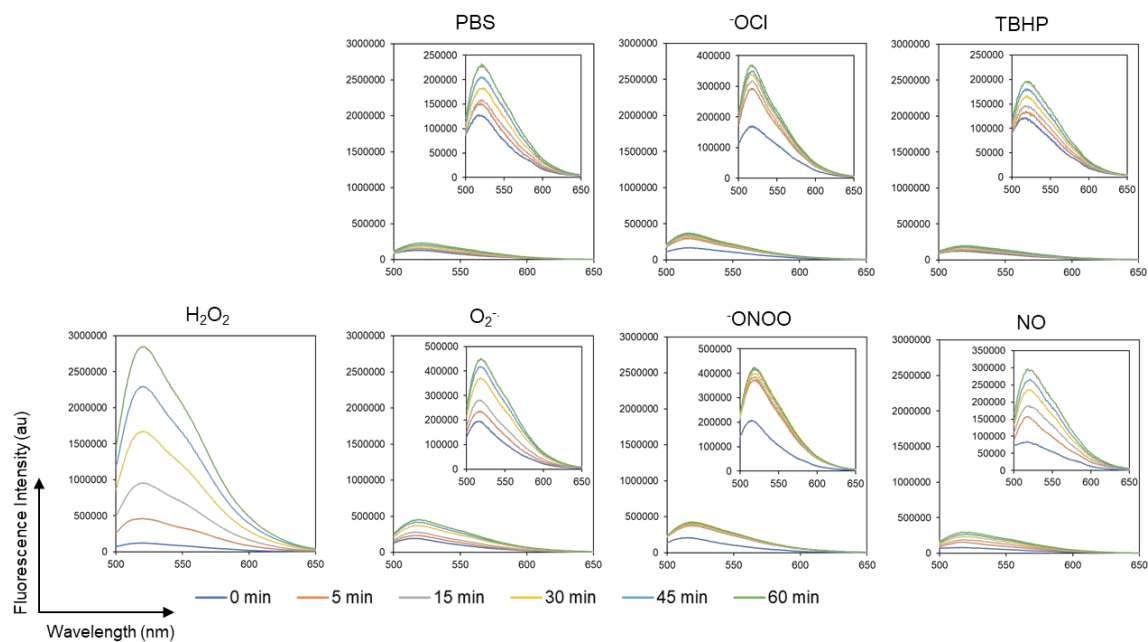


Figure S1. Fluorescence spectra of **PG1-FM** reacted with H_2O_2 , OCl^- , TBHP, O_2^- , ONOO^- , NO or PBS (control). **PG1-FM** (1 μM) was incubated with 25 μM of H_2O_2 , OCl^- , TBHP, O_2^- , ONOO^- , or NO at 37 $^\circ\text{C}$ in PBS (-). The mixtures were measured using a fluorimeter at 0, 5, 15, 30, 45, and 60 min.

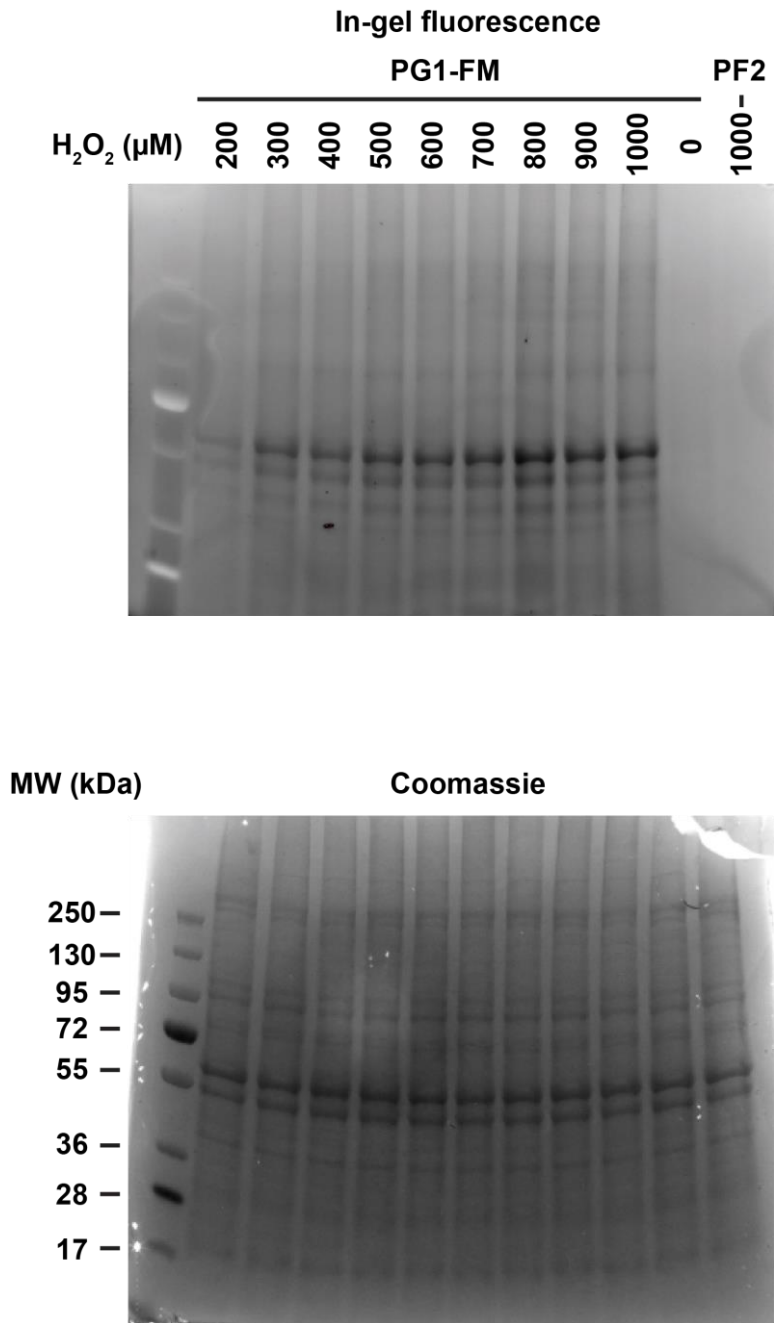


Figure S2. H_2O_2 -dependent activity-based labeling of **PG1-FM** or **PF2** in an MFP 231 cell lysate solution.

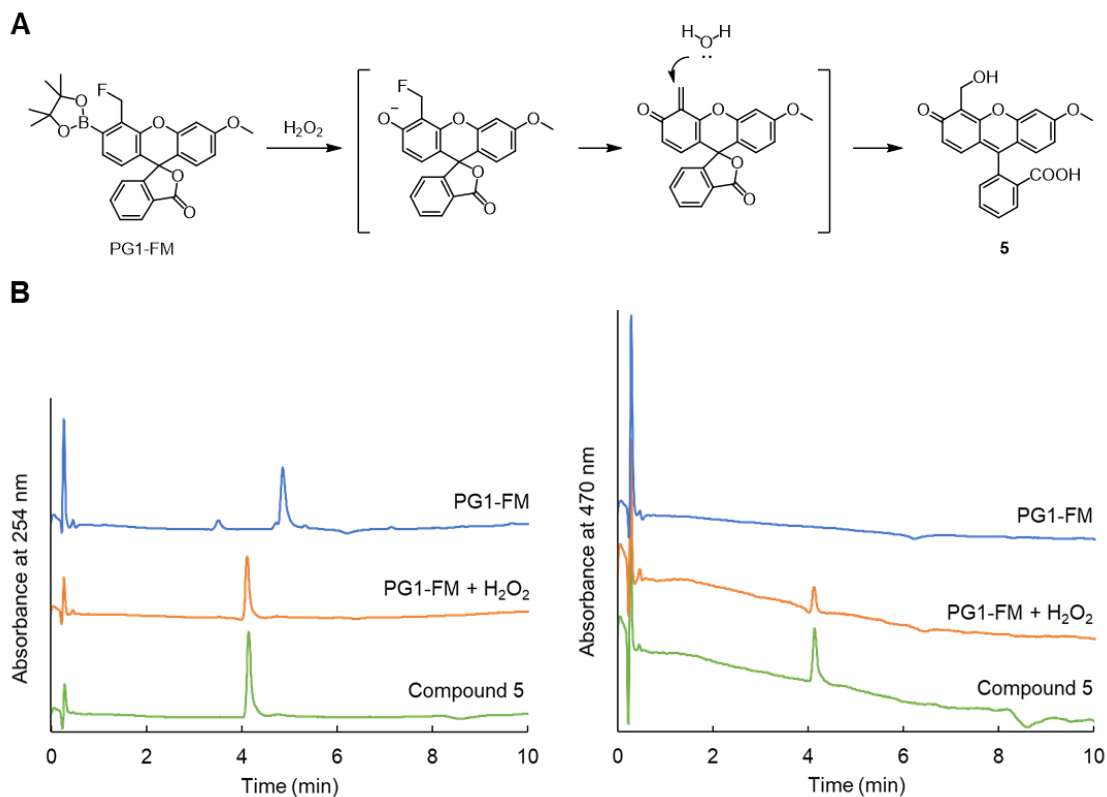


Figure S3. The reaction mechanism (A) and HPLC analyses (B) of the reaction product using **PG1-FM** and H₂O₂. 10 μM **PG1-FM** was treated with 10 μM H₂O₂ in PBS (-), containing 1% DMSO, and the mixture was incubated at 37 °C for 30 min. HPLC analysis was performed under a gradient condition (A: H₂O containing 0.05% formic acid; B: acetonitrile containing 0.05% formic acid; A/B = 75%/25% to 25%/75% in 15 min). Absorbance at 254 nm and 470 nm were monitored. The spectra show before (blue) and after (orange) reaction with H₂O₂, and its putative product, hydroxymethyl compound **5** (green), which is obtained by boronate deprotection and quenching of the quinone methide by water.

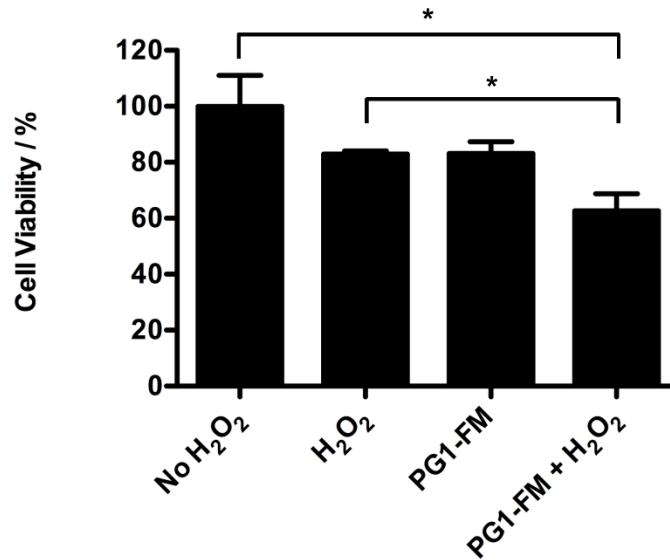


Figure S4. Cell viability assay of **PG1-FM** treated HeLa cells in the presence of H₂O₂. HeLa cells (96-well plate, 20,000 cells/well) were treated with **PG1-FM** (10 μM, 100 μL in HBSS) and placed in a humidified 5% CO₂ incubator at 37 °C for 30 minutes. The solution in wells were replaced with a 100 μM solution of H₂O₂ in HBSS (100 μL) and placed in a humidified 5% CO₂ incubator at 37 °C for one hour. The cells were washed once with HBSS (100 μL) at which point 100 μL of a 1:2000 dilution of 1 mM HOECHST 33342 stain in MilliQ water was added to each sample. The cells were placed in a humidified 5% CO₂ incubator at 37 °C for 10 minutes. Measurements λ_{ex}=350 nm/ λ_{em}= 460 nm were performed using a microplate reader. Mean ± SEM, normalized to the untreated control condition. N = 3. *P < 0.05.

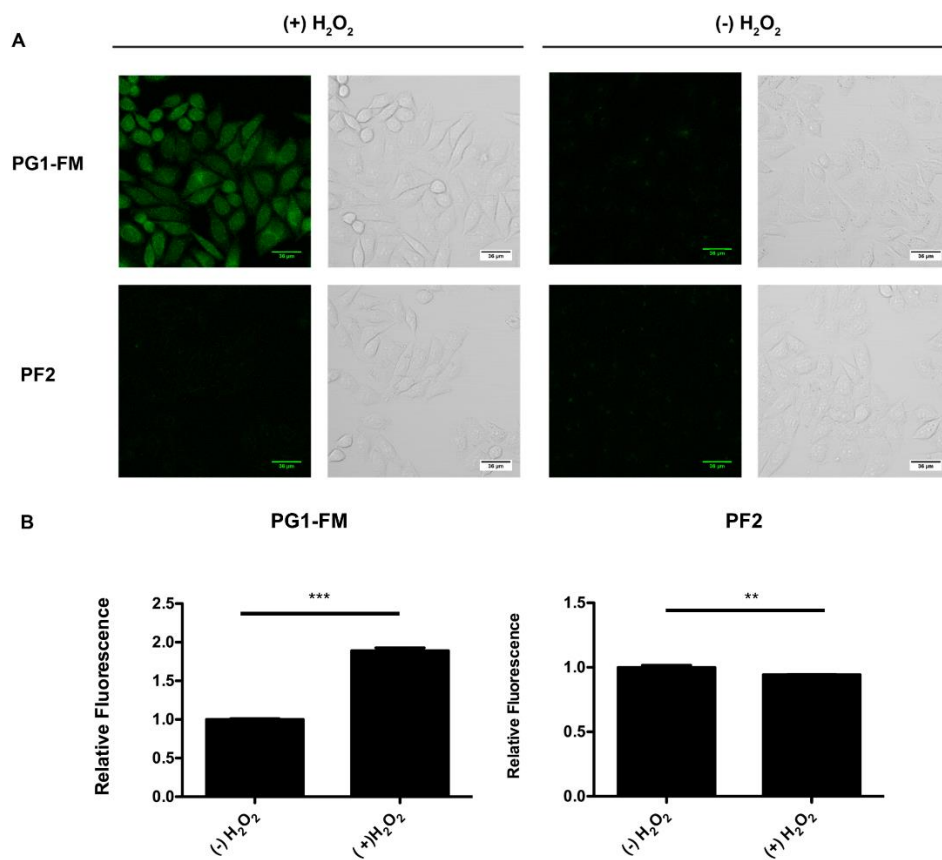


Figure S5. (A) CLSM images of HeLa cells stained with **PG1-FM** or **PF2** in the presence or absence of H_2O_2 (100 μM). Scale bar: 36 μm . (B) Quantification of **PG1-FM** and **PF2** fluorescence intensity. Mean \pm SEM, normalized to the untreated control condition. N = 3, with 4 fields examined per repeat. ** $P < 0.01$, *** $P < 0.001$.

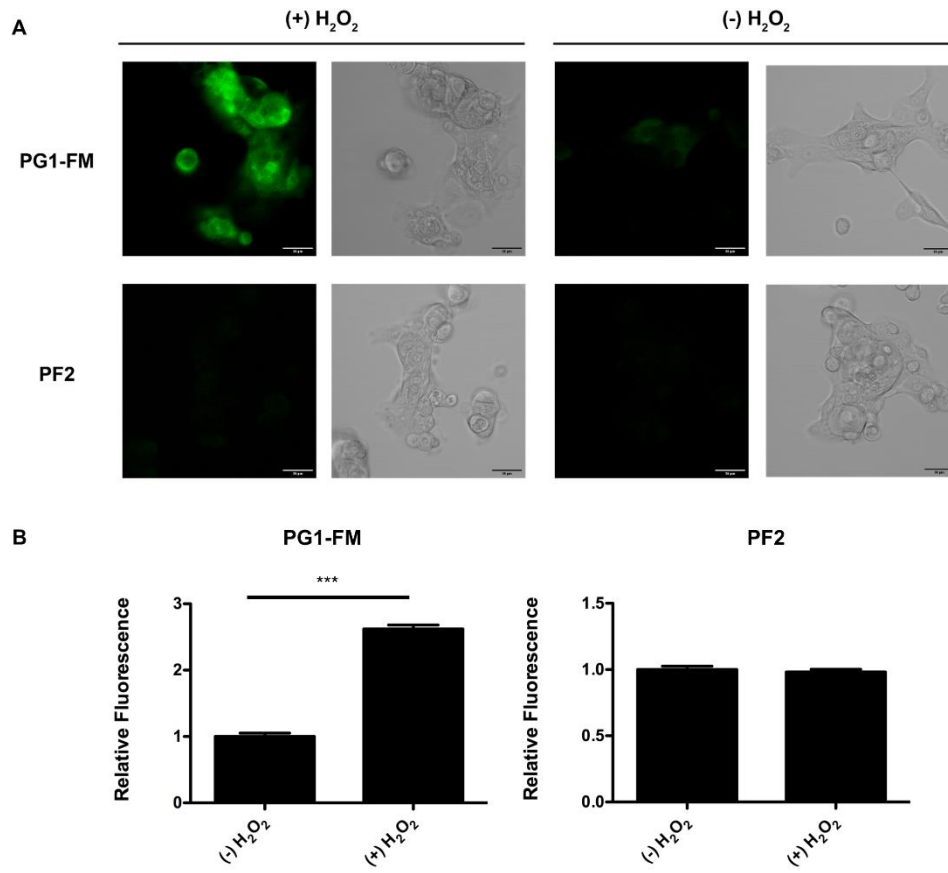


Figure S6. (A) CLSM images of MCF10A cells stained with **PG1-FM** or **PF2** in the presence or absence of H_2O_2 (100 μM). Scale bar: 36 μm . (B) Quantification of **PG1-FM** and **PF2** fluorescence intensity. Mean \pm SEM, normalized to the untreated control condition. N = 3, with 4 fields examined per repeat. *** $P < 0.001$.

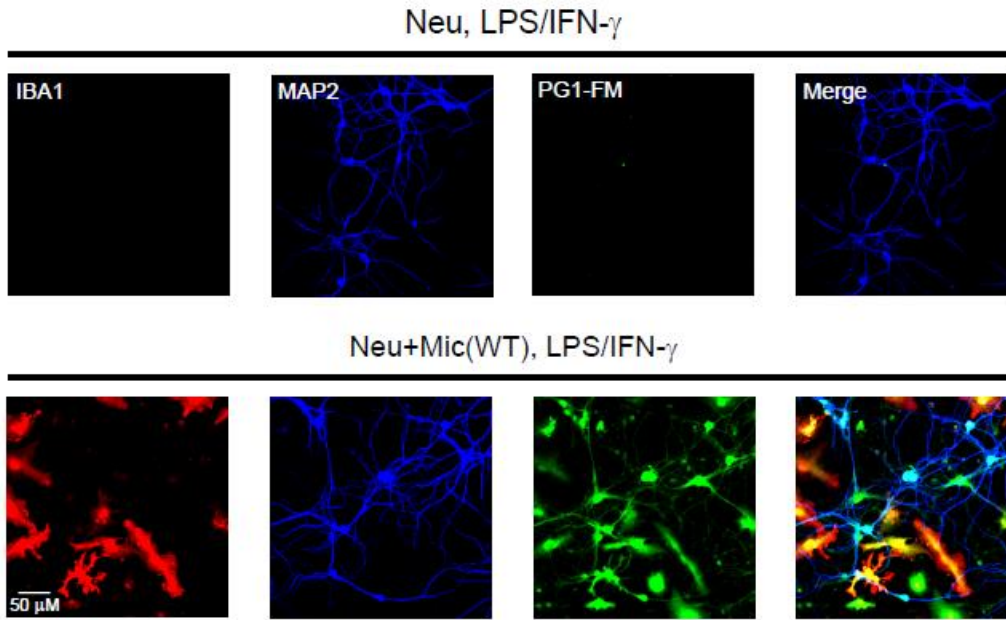


Figure S7. H₂O₂ signal in neurons in triple-labeled cultures. Representative images of cultures treated overnight with 500 ng/mL LPS + 50 ng/mL IFN- γ and incubated with 10 μ M **PG1-FM** (green) for 1 h. Cultures were immunostained for the neuronal marker MAP2 (blue) and the microglia marker Iba-1 (red). Neu: neurons, Mic: microglia. Scale bar = 50 μ m.

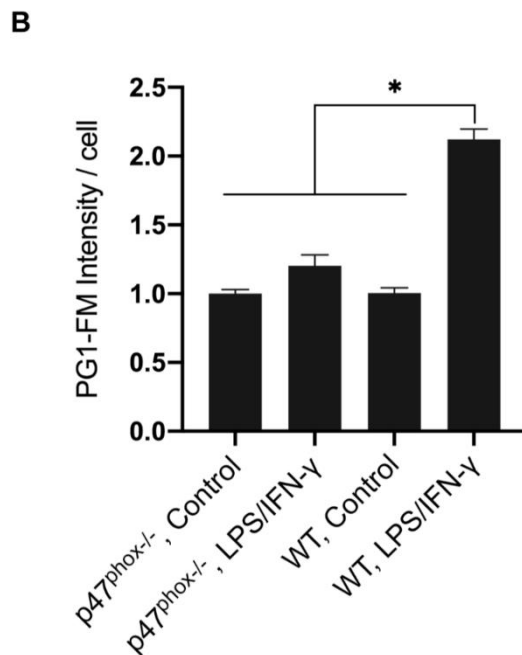
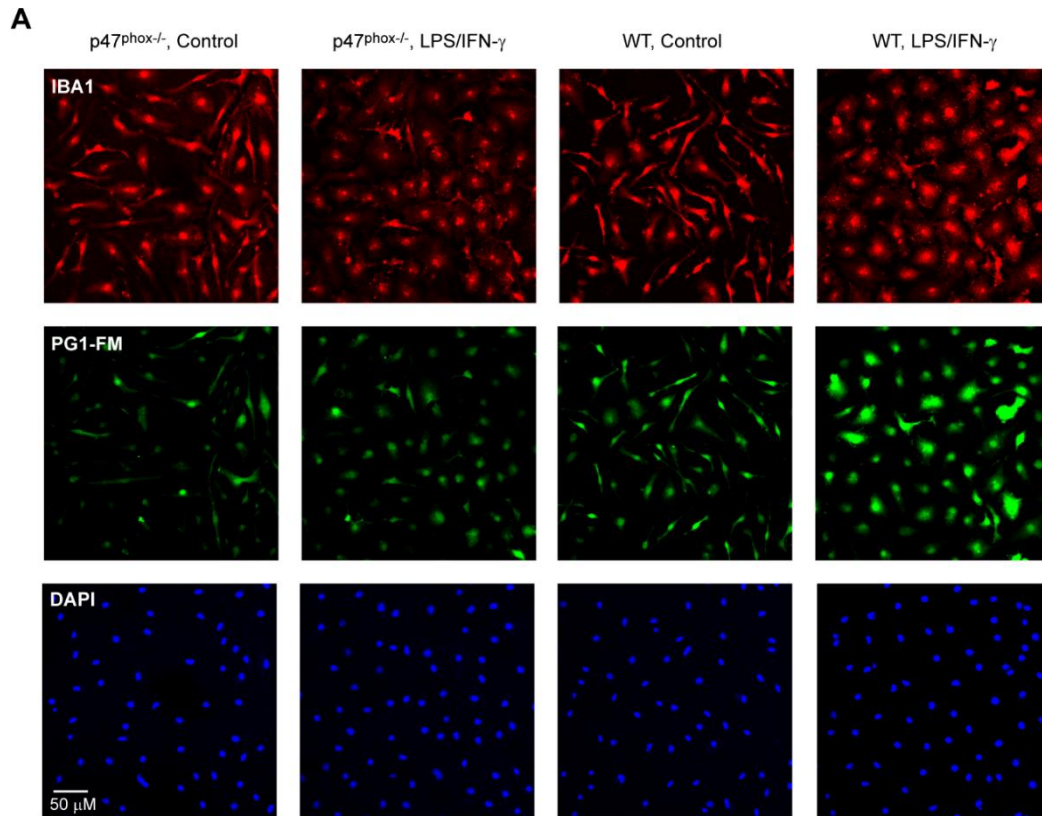


Figure S8. H₂O₂ production in WT and p47^{phox}^{-/-} microglia in response to LPS and IFN γ . (A) Representative images of microglia treated overnight with 500 ng/mL LPS + 50 ng/mL IFN- γ . The cultures were incubated with 10 μ M **PG1-FM** (green) for 1 h and immunostained for the microglial marker IBA1 (red), with nuclei labeled with DAPI (blue). Scale bar = 50 μ m. (B) Values were

plotted as **PG1-FM** fluorescence intensity \pm SEM, normalized to p47^{phox}^{-/-} control condition. N = 3, with > 200 cells evaluated per repetition. * P < 0.05.

Small Molecule Characterization

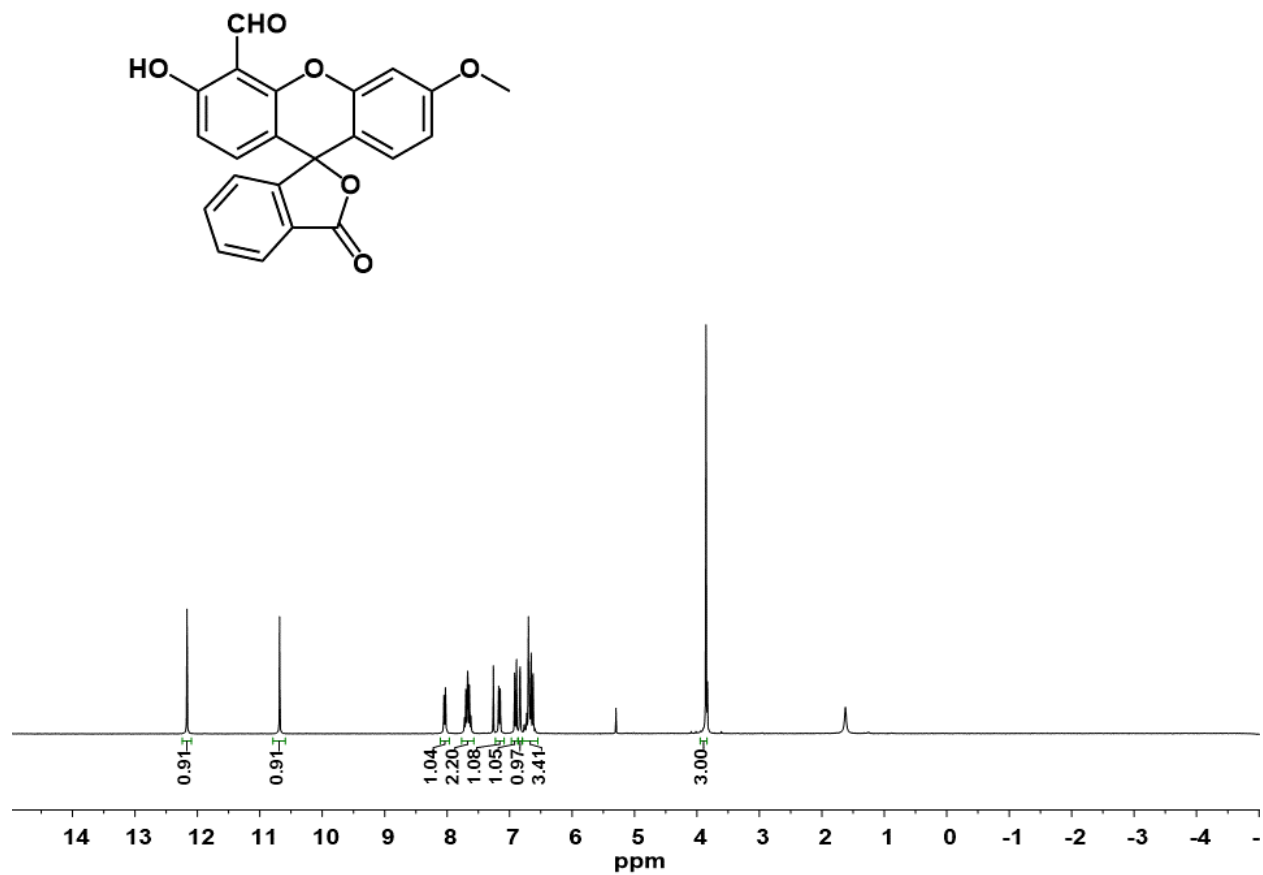


Figure S9. ¹H NMR spectrum of 2.

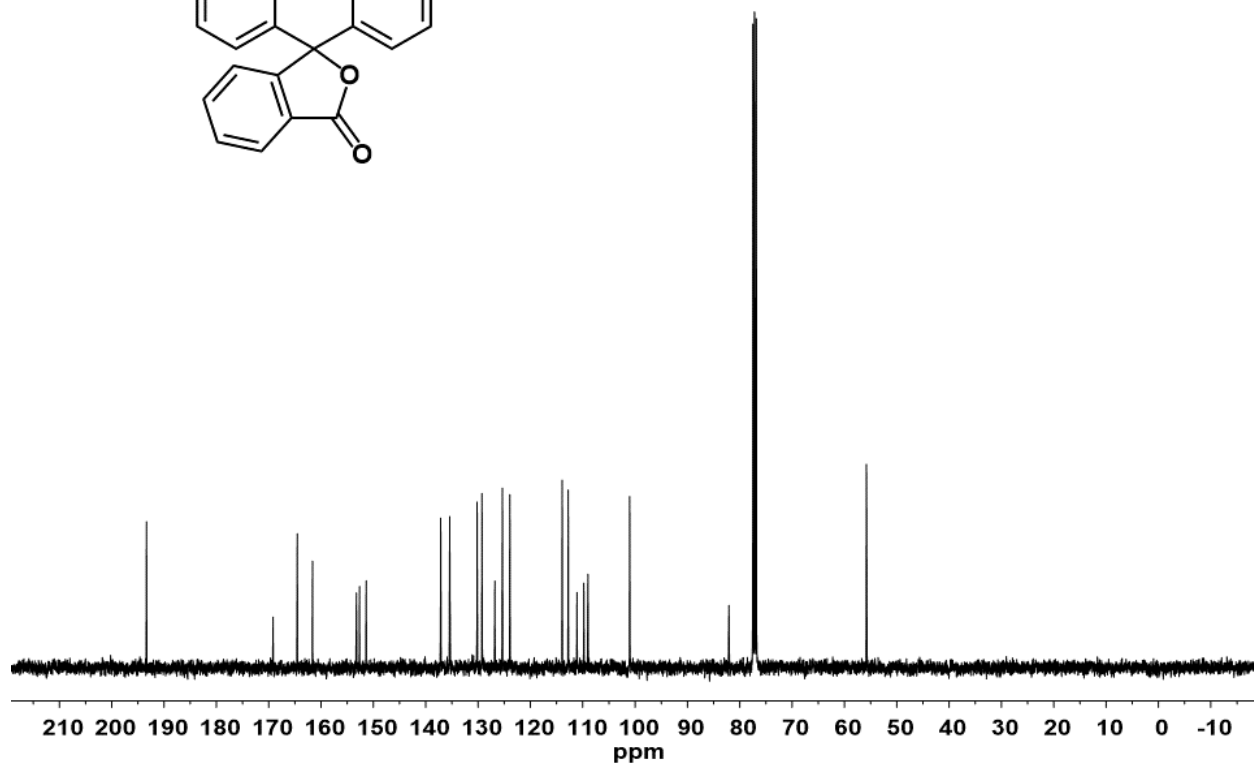
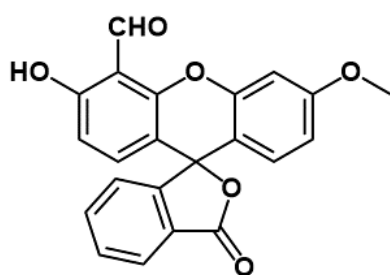


Figure S10. ¹³C NMR spectrum of 2.

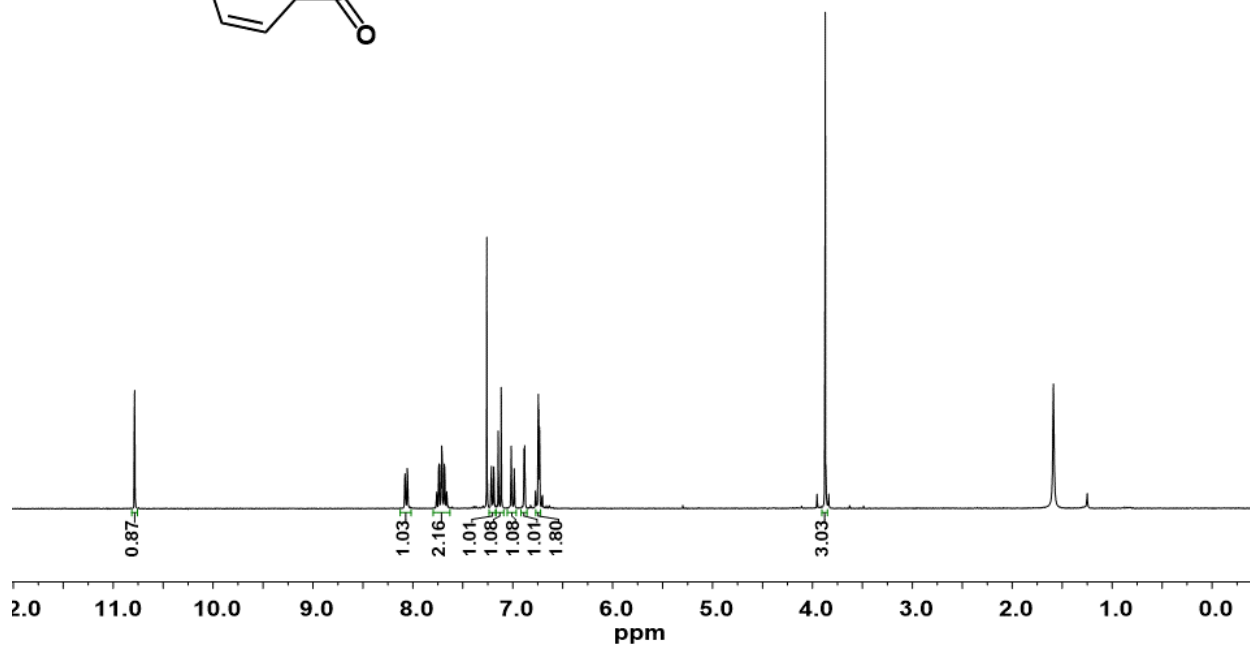
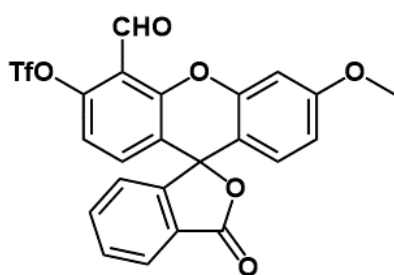


Figure S11. ¹H NMR spectrum of 3.

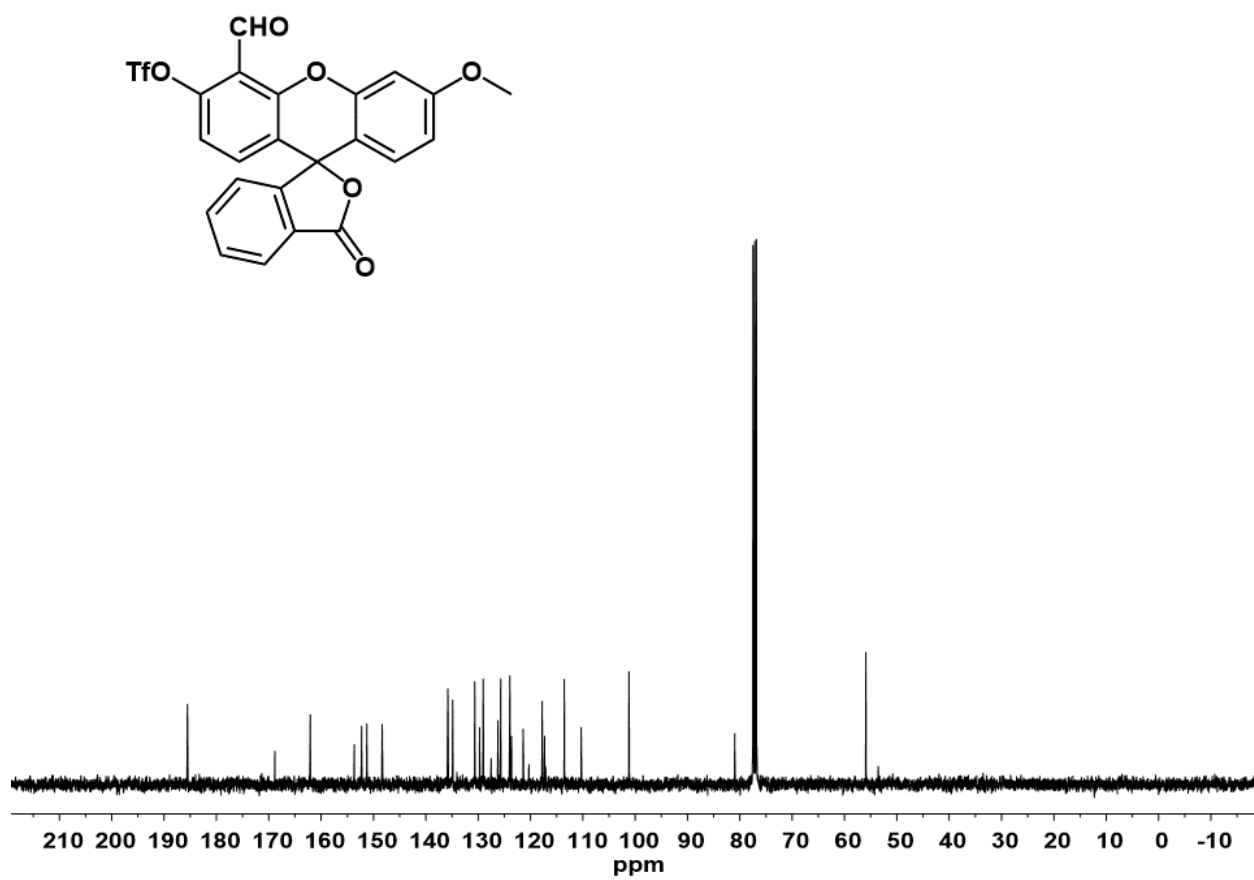


Figure S12. ¹³C NMR spectrum of 3.

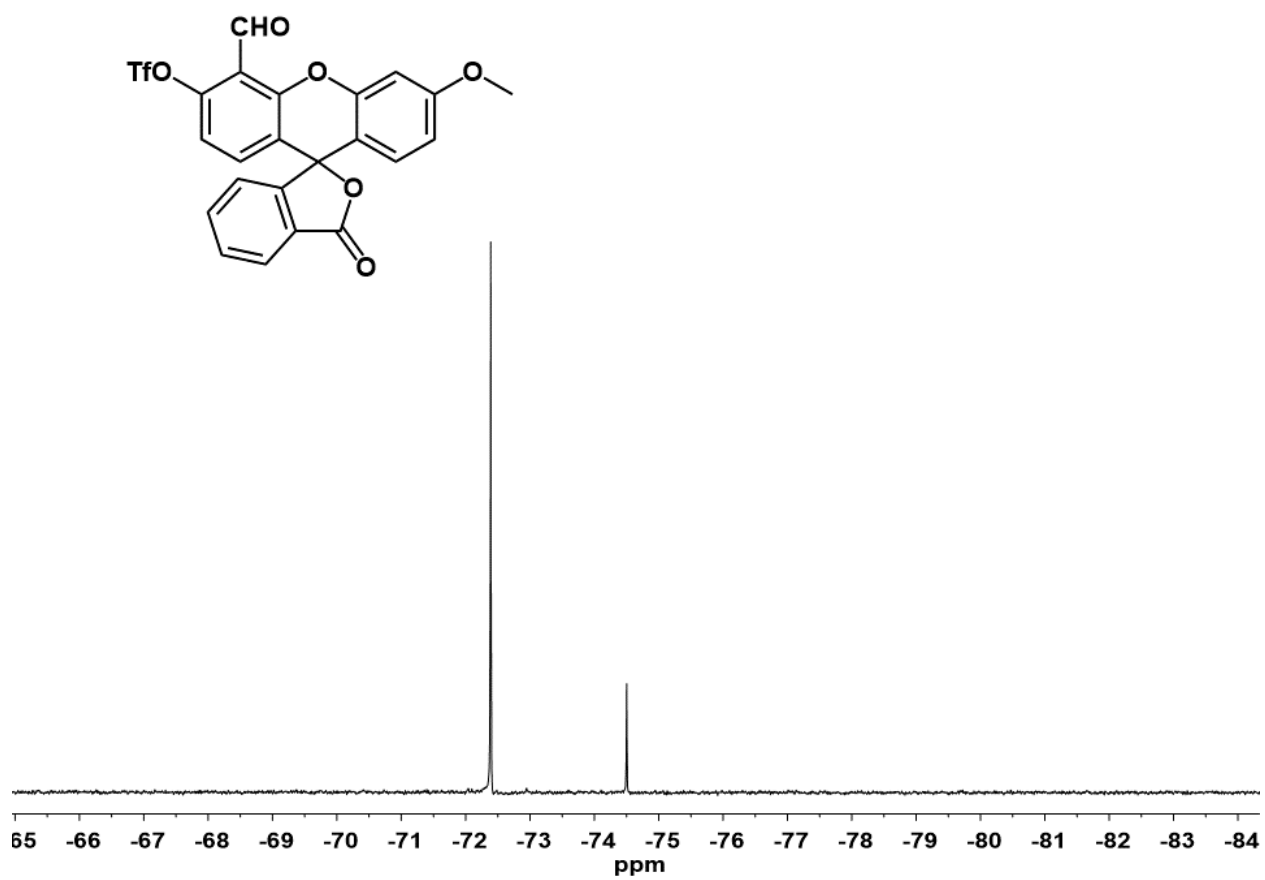


Figure S13. ^{19}F NMR spectrum of **3**.

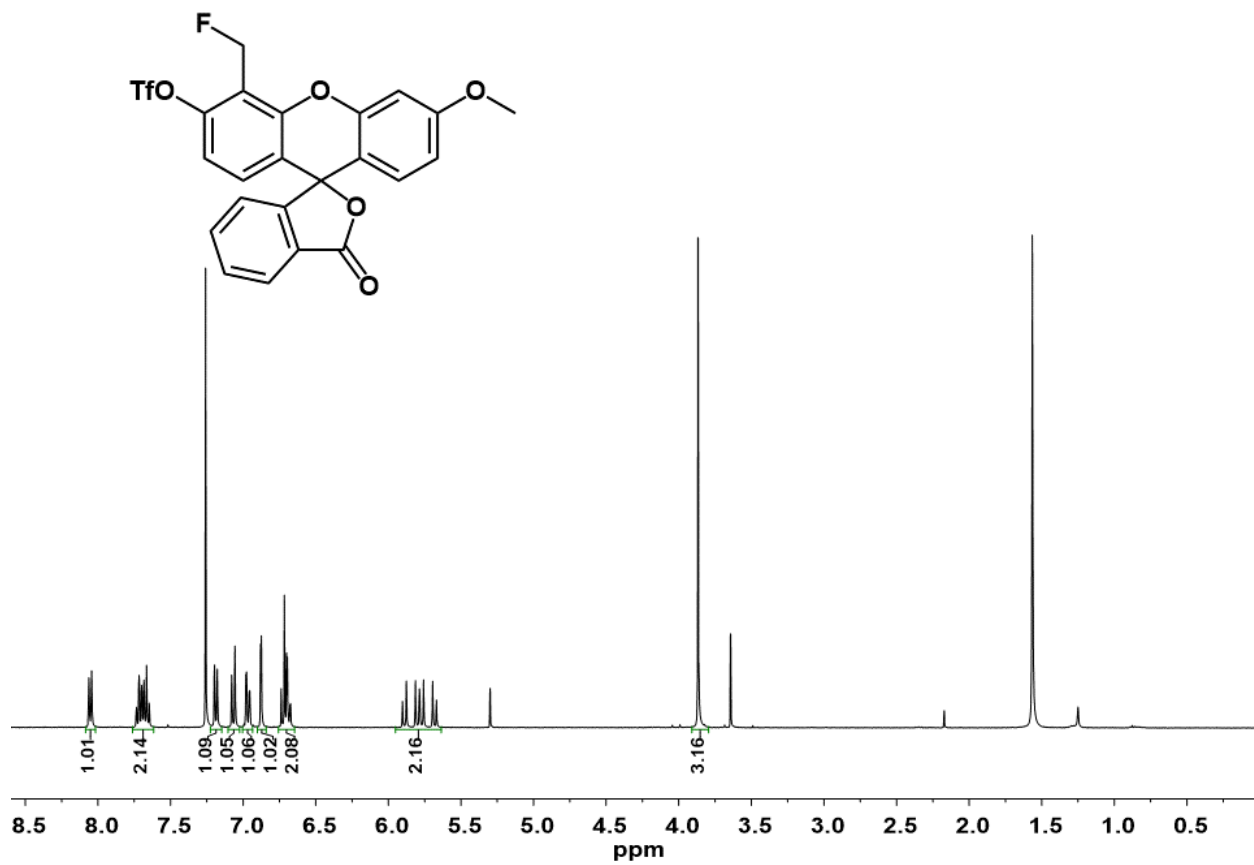


Figure S14. ¹H NMR spectrum of 4.

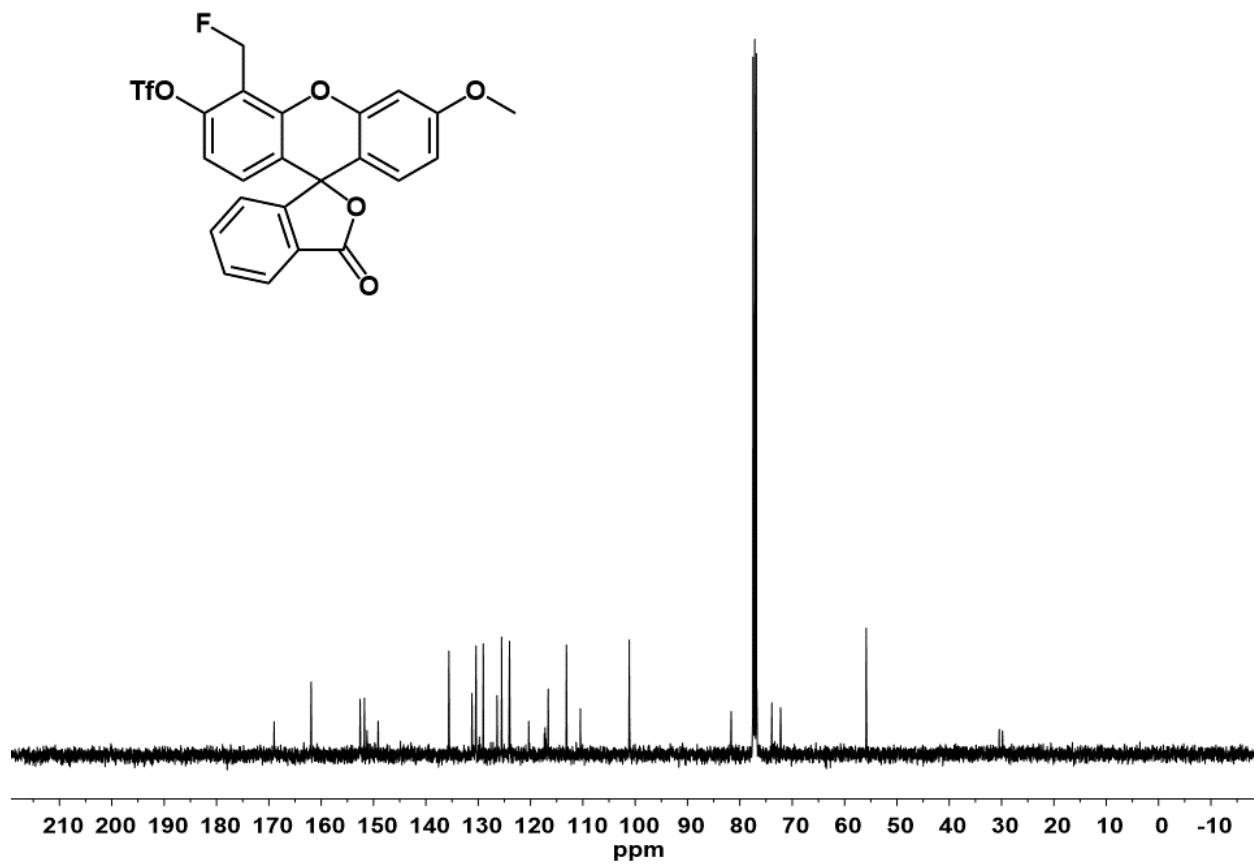


Figure S15. ¹³C NMR spectrum of 4.

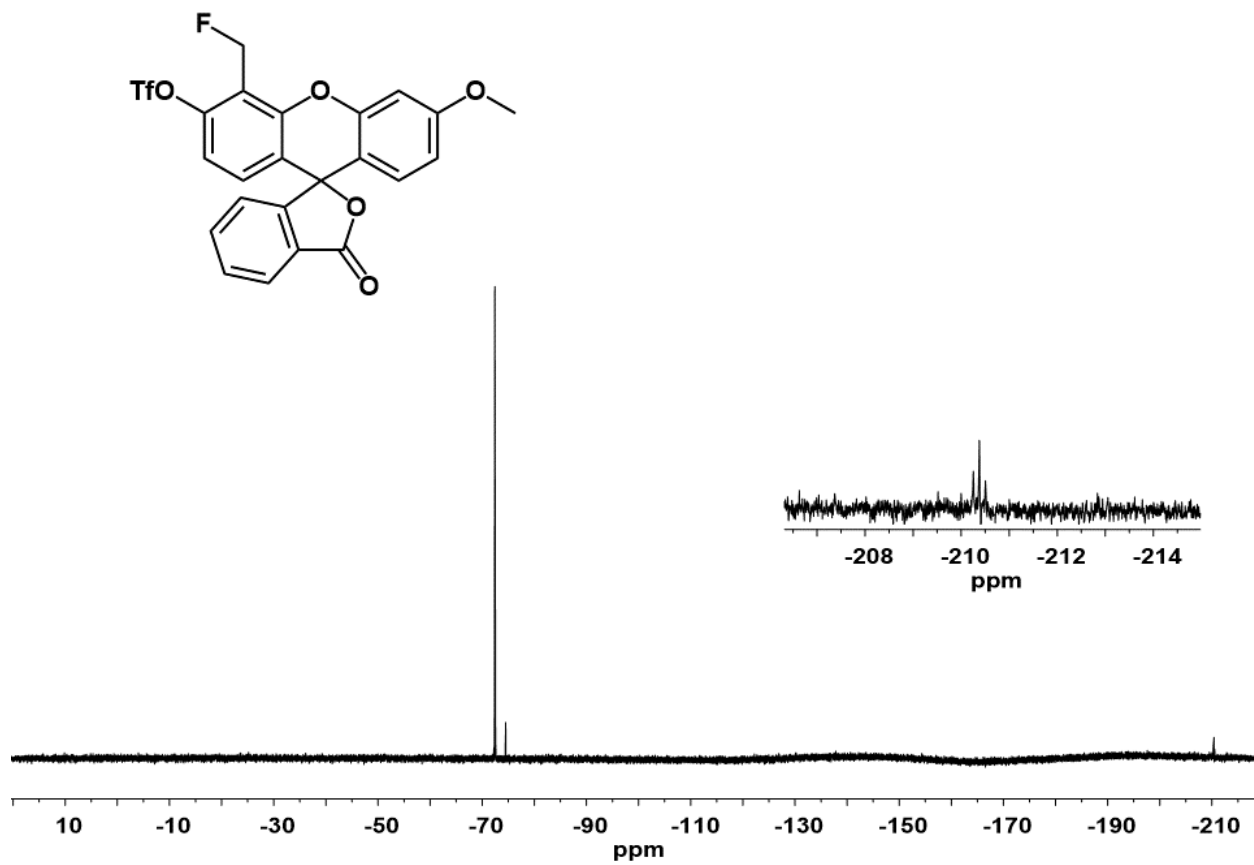


Figure S16. ^{19}F NMR spectrum of 4.

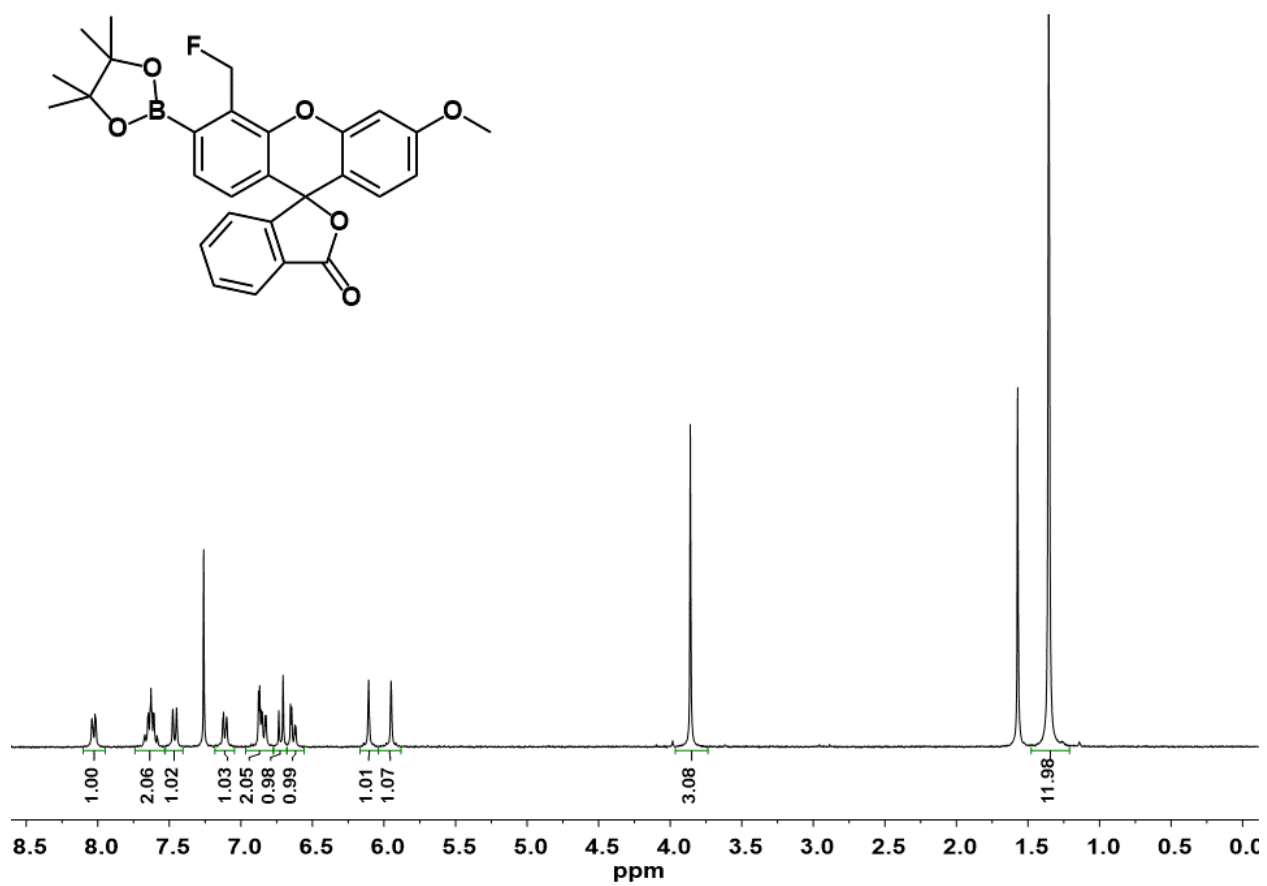


Figure S17. ¹H NMR spectrum of **PG1-FM**.

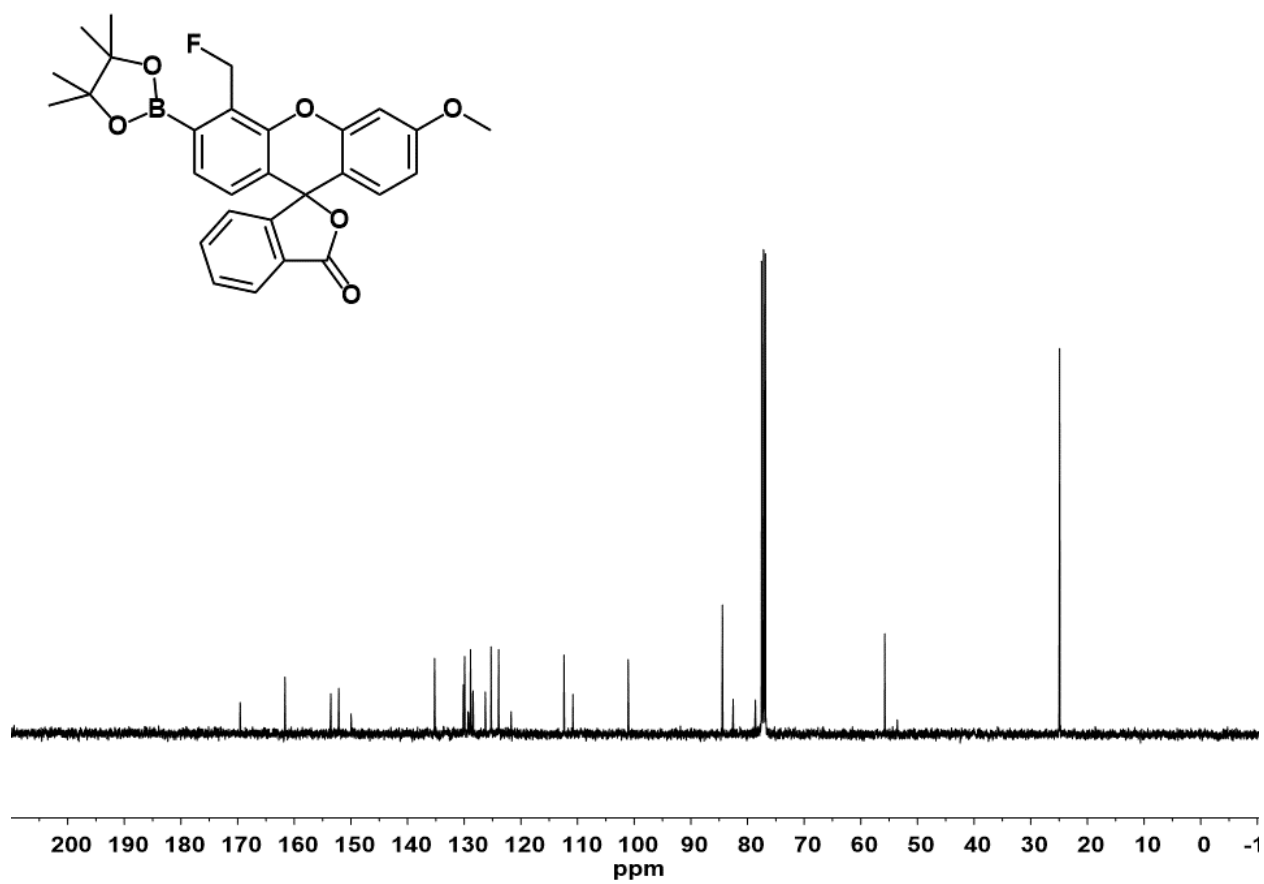


Figure S18. ¹³C NMR spectrum of **PG1-FM**.

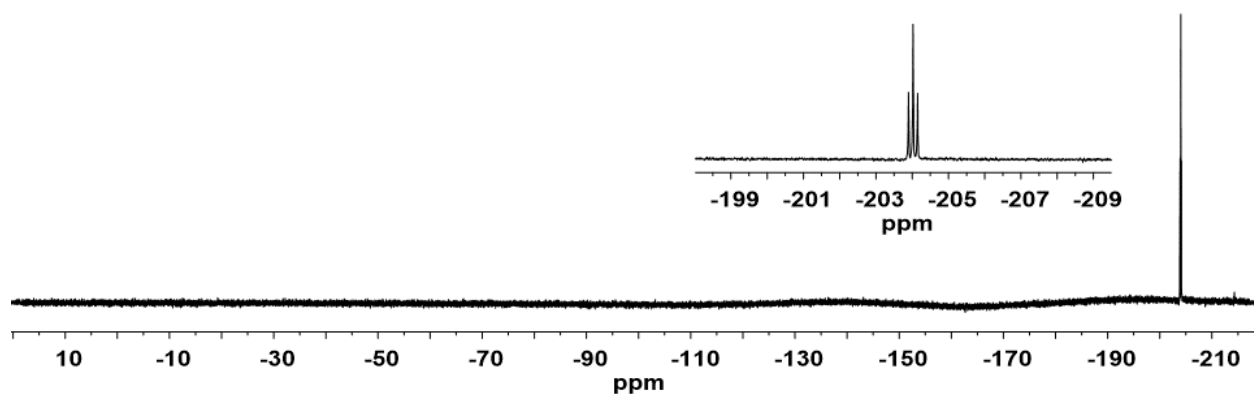
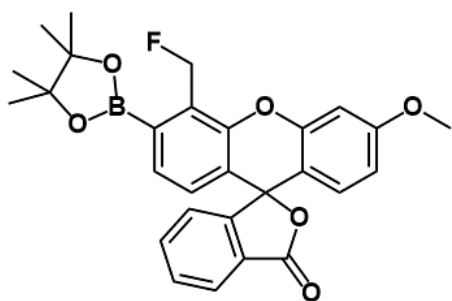


Figure S19. ^{19}F NMR spectrum of PG1-FM.

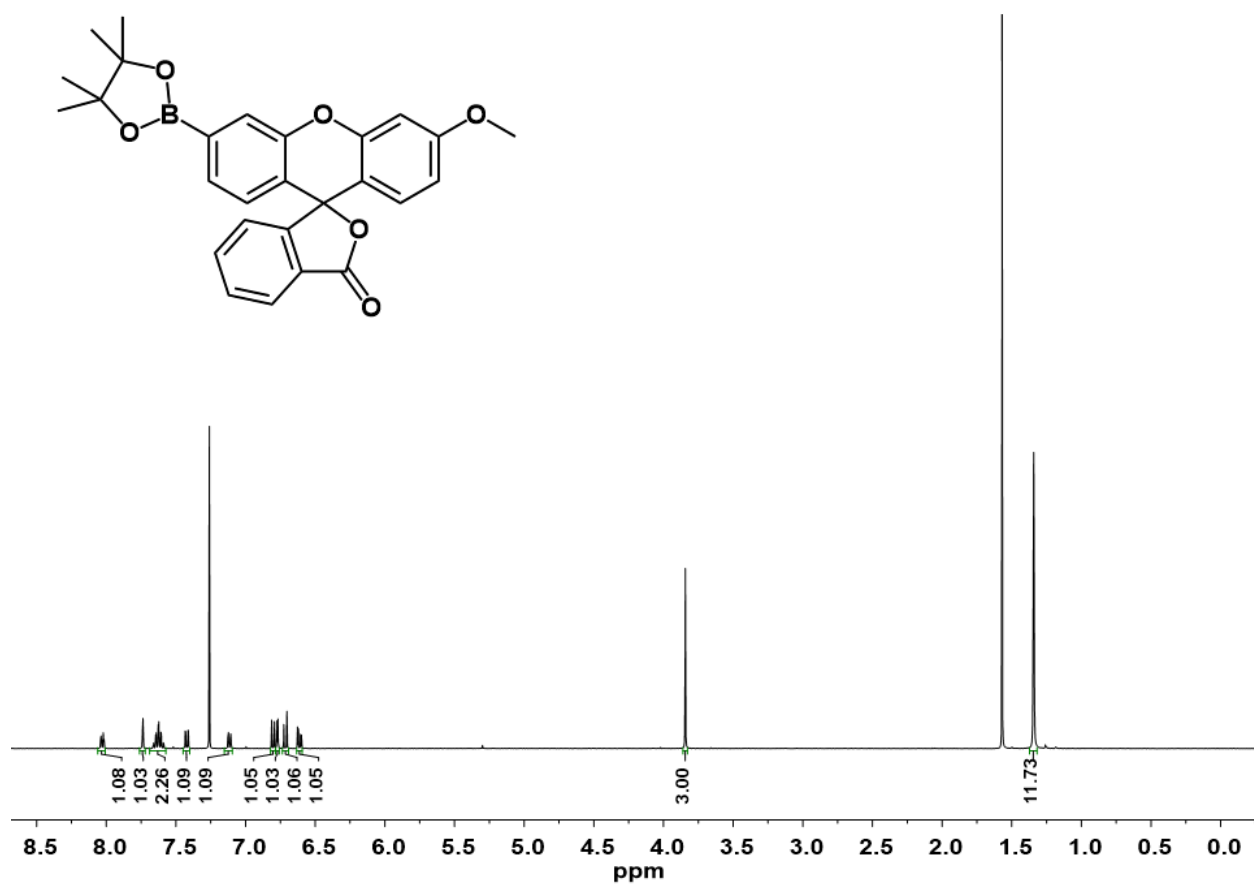


Figure S20. ¹H NMR spectrum of PF2.

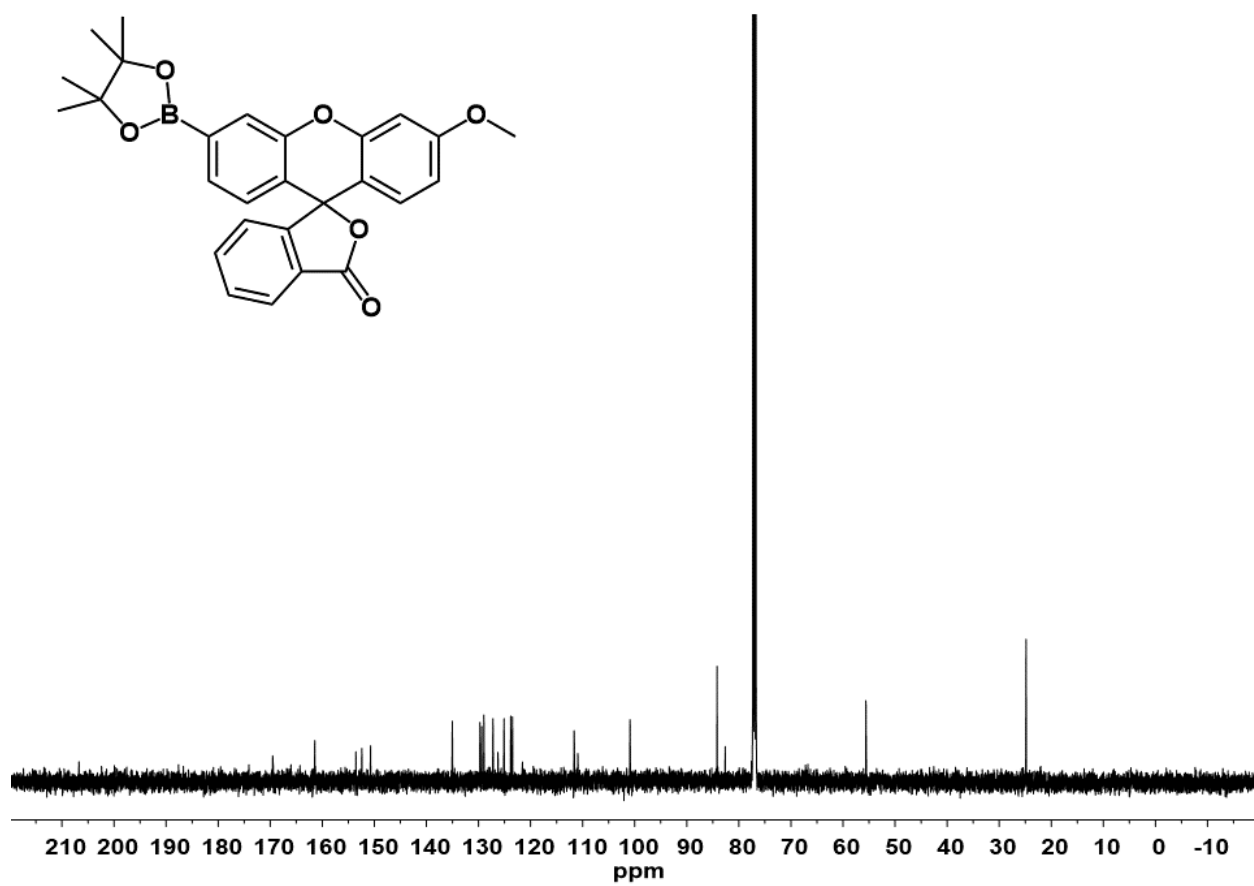


Figure S21. ¹³C NMR spectrum of **PF2**.

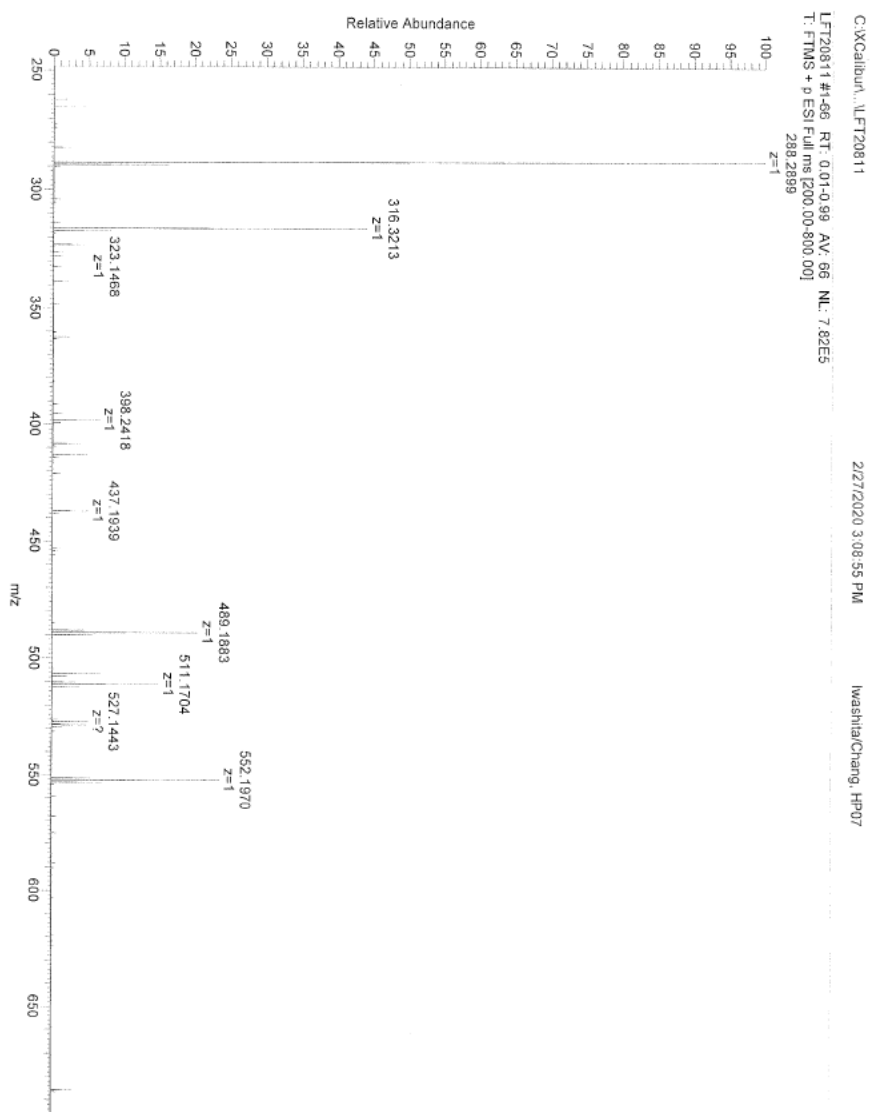


Figure S22. HRMS spectrum of **PG1-FM**.

References

1. Dickinson, B. C., Huynh, C., Chang, C. J., A Palette of fluorescent probes with varying emission colors for imaging hydrogen peroxide signaling in living cells. *J. Am. Chem. Soc.* **132**, 5906-5915.
2. Zhang J, Sun YQ, Liu J, Shi Y, Guo W (2013) Fluorescent probe for biological signaling molecule H₂S based on a specific H₂S trap group. *Chem. Commun* 49:11305-11307.
3. Lian H, Roy E, Zheng H (2016) Protocol for Primary Microglial Culture Preparation. *Bio-Protocol* 6 (21):1-10.
4. Yu AC, Neil SE, Quandt JA (2017) High yield primary microglial cultures using granulocyte macrophage-colony stimulating factor from embryonic murine cerebral cortical tissue. *J Neuroimmunol* 307:53-62.