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Supporting information for article:

Crystal structure of the nonclassical cadherin-17 N-terminus and implications for its adhesive binding mechanism

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#### **S1. Supplementary Methods**

### **S1.1. Sequence Alignments**

Alignments of the EC1-2 repeats of hs CDH17 (NP 004054.3, UniProt ID: Q12864), CDH16 (NP 004053.1, UniProt ID: O75309), CDH1 (NP 004351.1, UniProt ID: P12830), CDH2 (NP\_001783.2, UniProt ID: P19022), and CDH5 (NP\_001786.2, UniProt ID: P33151) were performed in MUSCLE (Edgar, 2004). The alignments were arranged in JalView (Waterhouse et al., 2009), and colored using the % identity for sequence identity with a conservation threshold cutoff of 40% for all alignments unless otherwise noted. The numbering on these alignments corresponds to the residue numbering in the human construct without signal peptide. Signal peptide cleavage sites were determined using the SignalP-5.0 server (Almagro Armenteros et al., 2019). Protein sequence alignments of individual CDH17 EC repeats were performed as above using sequences from 20 different species from NCBI (Table S3). Protein sequence alignments of 37 fish species (Fig. S2) were generated for the insertion seen in EC2 using MUSCLE (Edgar, 2004) and JalView (Waterhouse et al., 2009), as described above.

# **Supplementary Tables**

Macromolecule production information Table S1

Source organism	HOMO SAPIENS		
DNA source	Harvard PlasmID: HsCD0041924, UniProt ID: Q12864		
Forward primer	GGGAATTC* CATATG† CAAGAGGGGAAGTTTAGTGGACCCCT		
Reverse primer	CGGCGG* CTCGAG‡ TGCTTTCCAAATATTCTCTGTCACTATGATATCCAC		
Expression vector	pET21a		
Expression host	E. coli Rosetta2 (DE3)		
Complete amino acid sequence of the construct produced	MQEGKFSGPLKPMTFSIYEGQEPSQIIFQFKANPPAVTFELTGETDNIFVIER		
	EGLLYYNR ALDRETRSTHNLQVAALDANGIIVEGPVPITIEVKDINDNRPT		
	FLQSKYEGSVRQNSRPGKPFLYVNATDLDDPATPNGQLYYQIVIQLPMINN		
	VMYFQINNKTGAISLTREGSQELNPAKNPSYNLVISVKDMGGQSENSFSDT		
	TSVDIIVTENIWKALEHHHHHH		

<sup>\*</sup> Spacer nucleotides for the restriction enzyme site, † *NdeI* restriction enzyme sequence, ‡ *XhoI* restriction enzyme sequence

# Table S2 Crystallization

Method	Sitting drop vapour diffusion	
Plate type	96-well	
Temperature (K)	277	
Protein concentration	8 mg/mL	
Buffer composition of protein solution	20 mM Tris pH 8.0, 2 mM CaCl <sub>2</sub> , 150 mM KCl, 50 mM NaCl	
Composition of reservoir solution	0.1 M HEPES pH 7, 0.1 M KCL, 15% PEG 5000 MME	
Volume and ratio of drop	1.2 μL (1:1 protein:reservoir solution)	
Volume of reservoir	75 μL	

 Table S3
 EC1-2 Alignment Species

Accession #	UniProt ID	Species Name	Common Name
NP_004054.3	Q12864	Homo sapiens	Human
XP_016815174.1		Pan troglodytes	Chimpanzee
NP_446429.1	P55281	Rattus norvegicus	Norway rat
NP_062727.1	Q9R100	Mus musculus	Mouse
XP_544179.3		Canis lupus familiaris	Dog
XP_011289700.1		Felis catus	Cat
XP_026920025.1		Acinonyx jubatus	Cheetah
NP_001092372.1		Bos taurus	Cow
XP_013852061.1		Sus scrofa	Pig
XP_022440252.1		Delphinapterus leucas	Beluga whale
NP_001186424.1		Gallus gallus	Chicken
XP_010403277.2		Corvus cornix cornix	Crow
XP_009273778.1		Aptenodytes forsteri	Emperor Penguin
XP_016848299.1		Anolis carolinensis	Green anole
XP_025055189.1		Alligator sinensis	Chinese alligator
NP_919403.1		Danio rerio	Zebrafish
XP_022535364.1		Astyanax mexicanus	Mexican tetra
NP_001133585.1		Salmo salar	Atlantic salmon
XP_007886135.1		Callorhinchus milii	Australian ghostshark
NP_001135580.1		Xenopus tropicalis	Tropical clawed frog

#### Supplementary Figures

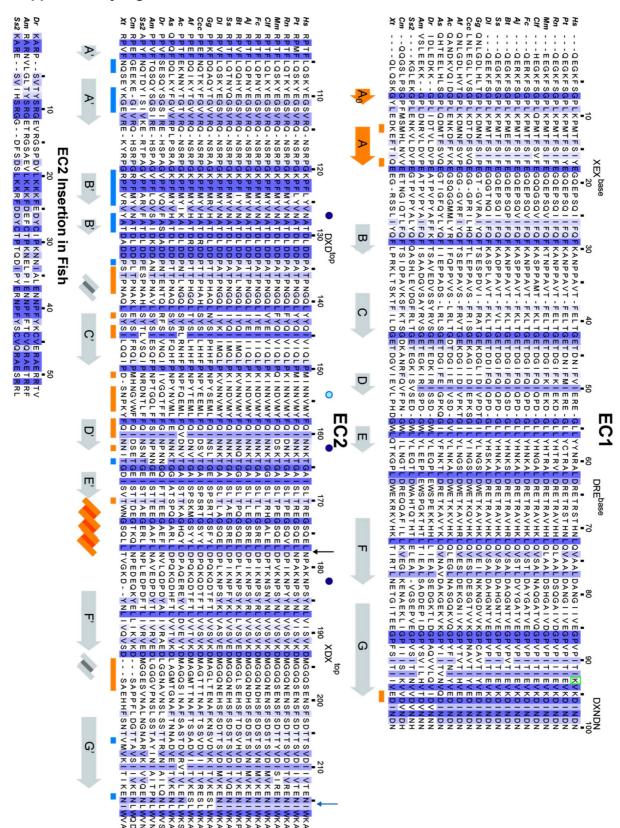
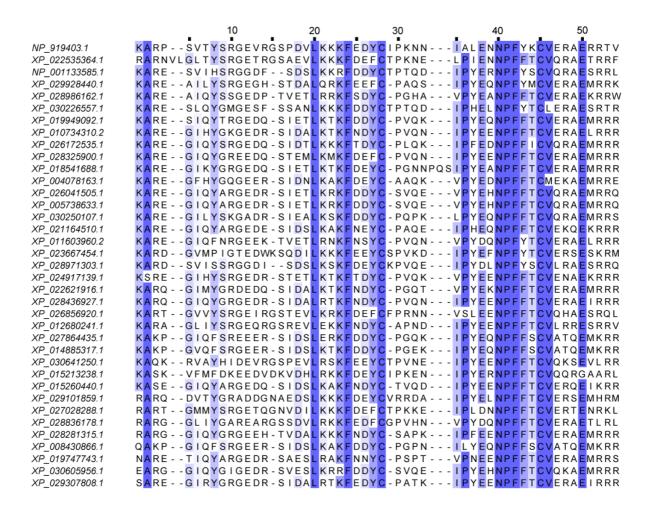
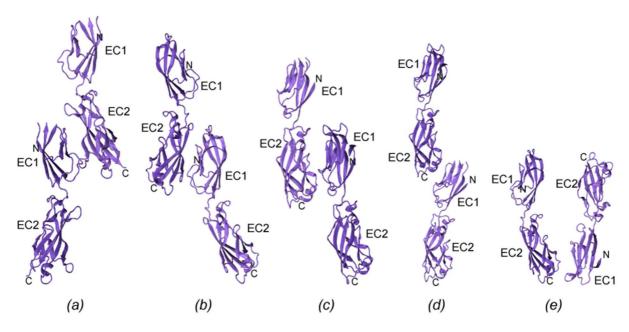


Figure S1 Multiple sequence alignments comparing repeats EC1 and EC2 of CDH17 from 20 different species (Table S3). Each alignment is colored by percent identity, with white being the lowest percent identity and dark blue being the highest. Sites of predicted N-linked glycosylation are denoted by a dark blue circle and site of the natural variant Lys93E is denoted by a green box in the human sequence. Site of the Asn154 residue implicated in disease is marked with a blue circle. Secondary structure elements observed in the crystal structures of hs CDH17 EC1-2 are illustrated below the respective repeats. Calcium-binding motifs are indicated above the sequences as in (a). Residues with ≥ 20% buried surface area in the EC1-2 cis interface (Fig. 6d) are denoted by orange bars and the same is done for the EC2-EC2 trans interface in blue (Fig. 6e). An arrow in EC2 indicates the location of a ~50 amino acid insertion observed in all fish species (Fig. S2), which has been aligned for included species below EC2. An arrowhead in blue in EC2 denotes the beginning of EC3 based on the hs CDH17 structure. Species are abbreviated as follows: Homo sapiens (hs), Pan troglodytes (Pt), Rattus norvegicus (Rn), Mus musculus (Mm), Canis lupus familiaris (Clf), Felis catus (Fc), Acinoyx jubatus (Aj), Bos taurus (Bt), Sus scrofa (Ss), Delphinapterus leucas (Dl), Gallus gallus (Gg), Corvus cornix cornix (Ccc), Aptenodytes forsteri (Af), Anolis carolinensis (Ac), Alligator sinensis (As), Danio rerio (Dr), Astyanax mexicanus (Am), Salmo salar (Ss2), Callorhinchus milli (Cm), and Xenopus tropicalis (Xt). Species were chosen based on sequence availability and taxonomical diversity. Accession numbers and species can be found in Table S3.



**Figure S2** Sequence alignment of CDH17 EC2 insertion in fish species. Alignment is shown as in Fig. S1.



**Figure S3** Additional crystallographic interfaces in the *hs* CDH17 EC1-2 structure. (a-e) Crystal contacts between two monomers of *hs* CDH17 EC1-2 as identified by PISA (Krissinel & Henrick, 2007). Interface areas are 464.2 Å<sup>2</sup> (a), 454.0 Å<sup>2</sup> (b), 452.0 Å<sup>2</sup> (c), 183.4 Å<sup>2</sup> (d), and 100.0 Å<sup>2</sup> (e) respectively.