Exploitation of DHODH and p53 activation as therapeutic targets - a case study in polypharmacology (SUPPLEMENTAL DATA)

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в

С

6

3Å





5 Å

(A) Predicted binding mode of tenovin 6 with the enzyme Sirtuin1 (SirT1). Sirtuin1 is shown as cartoon (grey colour) and the cofactor NAD (yellow carbon) and bound tenovin 6 (green carbon) are shown as thick lines. The tenovin binding pocket residues are shown as thin lines with the hydrogen bond interactions between SirT1 and the tenovin shown as dashed lines (magenta). (B) Binding free energies between SirT1 and the tenovins calculated using the MMPBSA approach averaged over the conformations sampled during the MD simulations. The error bars (standard deviation) are indicated. (C) The position of either tenovins 1, 6 or 39OH at each unit of distance (Å) in the pulled ligand simulation that has been graphed in figure 1E.



## **Supplemental Figure 2**

Values obtained using a kinetic DHODH enzyme assay for each tenovin indicated. Values correspond to the average of 3 independent repeats  $\pm$  SD with 3 technical repeats each.



#### **Supplemental Figure 3**

(A) p53 transcriptional activity assay (CPRG) in ARN8 cells treated with a low dose of tenovin 1, tenovin 6 or 5  $\mu$ M nutlin 3a as a positive control and either supplemented with vehicle, DHO, OA or uridine. This panel supplements figure 4a. (B) SRB assay in ARN8 cells treated with low dose tenovin 1 or tenovin 6 for 72 h supplemented with DHO, OA or uridine. This panel supplements figure 4c.

Figure 3a whole blots



Figure 3d whole blots



#### **Supplemental Figure 4 continued**

Figure 4b whole blots



#### **Supplemental Figure 4 continued**

Figure 4b whole blots



Figure 4b whole blots continued



Figure 6b whole blots



DMSO Trw 1 Trw 6 Trw 3 Trw 39 Trw 39 CH Trw 50 CH

Total blot images for figures 3a, 4b and 6b. Areas highlighted are the regions of the blot that have been used in the figures. Antibodies were used sequentially on each blot without stripping the blot between primary antibodies. For figure 4b blots, each blot has its own DMSO control from the same treatment plate next to it, though for clarity only one control was shown on the figure. The total protein loading controls have been shown here in full. The total protein loading controls have been processed as per manufacturer's instructions (1).

**Table S1:** Summary of structural determination of DHODH in complex withtenovin 6.

Ligands	FMN, L-DHO, Tenovin, DDQ, glycerol, sulphate, acetic acid, chloride
Resolution (Å)	20-1.85 (1.89-1.85)
Wavelength (Å)	1,0000
Space Group	P3221
Unit Cell (Å)	A=90.5, b=90.5, c=122.8
Completeness (%)	99.8 (99.5)
Number of observations/unique reflections	223 553 / 50 141
<i σ(i)=""></i>	5.4 (1.2)
CC(1/2) (%)	97.9 (41.3)
R <sub>merge</sub> (I) (%)	17.6 (105.8)
R <sub>cryst</sub> (F) (%)	16.5
R <sub>free</sub> (F) (%)	19.7
No. of non-hydrogen atoms	3209
No. of water molecules	228
Rms deviations from ideal geometry: Bond lengths (Å)	0.022
Bond angles (deg)	2.1
Mean B-factor protein (Ų)	26.0
Mean B-factor (solvent, Ų)	32.9
Mean B-factor (ligand tenovin, Ų)	53.9
Mean B-factor (other ligands, Ų)	37.2

# Supplementary References

1. Gurtler A, Kunz N, Gomolka M, Hornhardt S, Friedl AA, McDonald K, et al. Stain-Free technology as a normalization tool in Western blot analysis. Anal Biochem. 2013;433(2):105-11.