

Supporting Information

Synthesis and Structure—Activity Relationship Study of Antimicrobial Auranofin against ESKAPE Pathogens

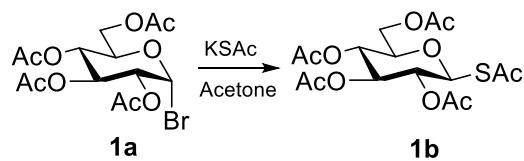
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Synthesis of 1b.

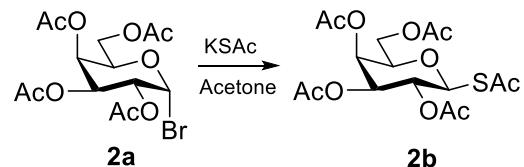


Scheme S1. Synthesis of compound **1b**.

2,3,4,6-Tetra-O-acetyl-1-S-acetyl- β -D-glucopyranose (**1b**)

This compound was synthesized according to general procedure A from **1a** (1.52 g, 3.70 mmol) and purified by flash column chromatography (ethyl acetate:hexanes = 1:1) to give **1b** as a white solid (1.50 g, 86%). ^1H NMR (500 MHz, CDCl_3) δ 5.24–5.14 (m, 2H, H-3, H-1), 5.08 – 4.97 (m, 2H, H-2, H-4), 4.18 (dd, J = 12.5, 4.5 Hz, 1H, H-6a), 4.01 (dd, J = 12.5, 2.1 Hz, 1H, H-6b), 3.77 (ddd, J = 10.1, 4.4, 2.1 Hz, 1H, H-5), 2.30 (s, 3H, SAc), 1.98 (s, 3H, OAc), 1.94 (s, 3H, OAc), 1.93 (s, 3H, OAc), 1.91 (s, 3H, OAc). The NMR spectrum is consistent with published data.¹

Synthesis of 2b.



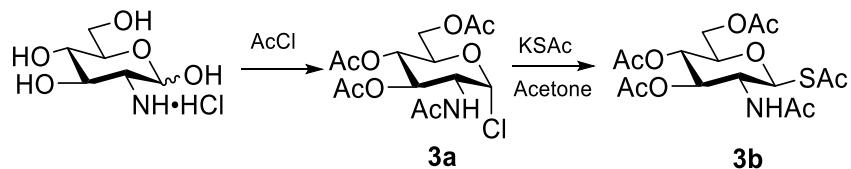
Scheme S2. Synthesis of compound **2b**.

2,3,4,6-Tetra-O-acetyl-1-S-acetyl- β -D-galactopyranose (**2b**)

This compound was synthesized from compound **2a** (2.24 g, 3.22 mmol) according to general procedure A and purified by flash column chromatography (ethyl acetate:hexanes = 1:1) to give **2b** (1.18 g, 90%) as a pale-yellow solid. ^1H NMR (500 MHz, CDCl_3) δ 5.46 (d, J = 3.3 Hz, 1H, H-4), 5.35 – 5.29 (t, J = 3.3 Hz, 1H, H-2), 5.25 (d, J = 10.4 Hz, 1H, H-1), 5.11 (dd, J = 9.7, 3.4 Hz, 1H, H-3), 4.18 – 4.02 (m, 3H, H-6a, H-6b, H-5), 2.39 (s, 3H),

2.15 (s, 3H, SAc), 2.04 (s, 3H, OAc), 2.03 (s, 3H, OAc), 1.98 (s, 3H, OAc). The NMR spectrum is consistent with published data.¹

Synthesis of 3b.



Scheme S3. Synthesis of compound 3b.

3,4,6-Tri-O-acetyl-2-N-acetylamido-2-deoxy- α -D-glucopyranosyl chloride (3a)

Prepared according to reported protocol.² Acetyl chloride (2.0 mL, 28.0 mmol) was added dropwise into a round-bottom flask containing *N*-acetylglucosamine (1.0 g, 4.52 mmol) at 0 °C. The mixture was stirred for 48 h under Ar at rt. DCM (200 mL) was added and the organic layer was washed by water, saturated sodium bicarbonate, brine and dried over MgSO₄. After removing the solvent, the residue was purified by flash column chromatography (ethyl acetate:hexanes = 3:1) to afford **3a** as a white solid (951 mg, 57%).

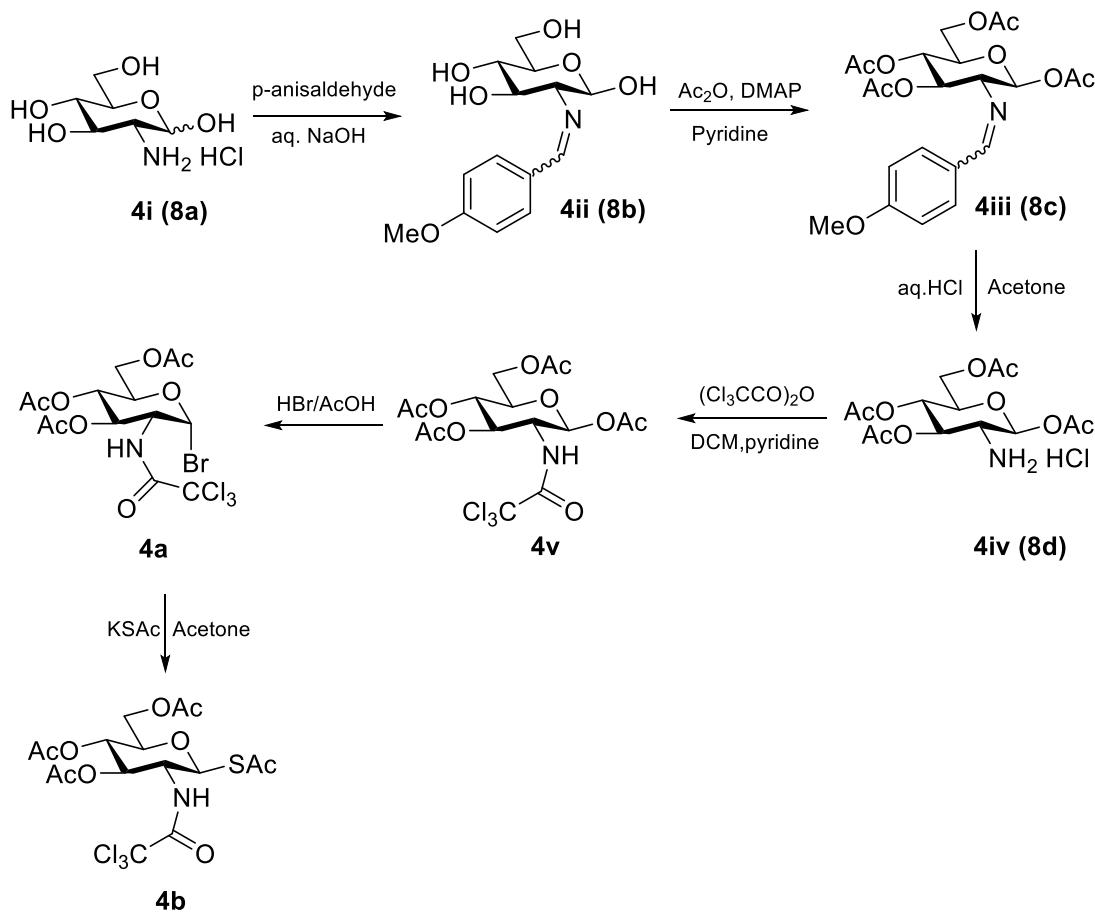
¹H NMR (500 MHz, CDCl₃) δ 6.19 (d, *J* = 3.7 Hz, 1H, H-1), 5.79 (d, *J* = 8.7 Hz, 1H, NH), 5.32 (dd, *J* = 10.5, 9.6 Hz, 1H, H-3), 5.22 (t, *J* = 9.8 Hz, 1H, H-4), 4.53 (ddd, *J* = 10.7, 8.7, 3.7 Hz, 1H, H-2), 4.32 – 4.23 (m, 2H, H-6a, H-5), 4.17 – 4.10 (m, 1H, H-6b), 2.10 (s, 3H, OAc), 2.06 (s, 3H, OAc), 2.05 (s, 3H, OAc), 1.99 (s, 3H, NHAc).

3,4,6-Tri-O-acetyl-2-N-acetyl-S-acetyl-1-thio- β -D-glucosamine (3b)

This compound was synthesized according to general procedure A from **3a** (951 mg, 2.60 mmol), and purified by flash column chromatography (ethyl acetate:hexanes = 2:1) to give **3b** as a pale yellow solid (925 mg, 88%). ¹H NMR (500 MHz, CDCl₃) δ 5.58 (d, *J* = 9.7

Hz, 1H, NH), 5.15 (d, $J = 10.8$ Hz, 1H, H-1), 5.13 (t, $J = 9.3$ Hz, 1H, H-3), 5.09 (t, $J = 9.5$ Hz, 1H, H-4), 4.35 (q, $J = 10.0$ Hz, 1H, H-2), 4.24 (dd, $J = 12.5, 4.5$ Hz, 1H, H-6a), 4.10 (dd, $J = 12.5, 2.2$ Hz, 1H, H-6b), 3.78 (ddd, $J = 9.6, 4.5, 2.2$ Hz, 1H, H-5), 2.37 (s, 3H, SAC), 2.08 (s, 3H, OAc), 2.03 (s, 6H, 2 \times OAc), 1.92 (s, 3H, NHAc). The NMR spectrum is consistent with published data.¹

Synthesis of 4b.



Scheme S4. Synthesis of compound 4b.

2-Deoxy-2-[(4-methoxyphenyl)methylene]amino- β -D-glucopyranose (4ii)

Prepared according to a reported procedure.³ D-Glucosamine hydrochloride **4i** (3.0 g, 13.9 mmol) was dissolved in 1 M NaOH (15 mL), and *p*-anisaldehyde (2.0 mL, 16.7 mmol) was added. The mixture was stirred at RT for 0.5 h then was briefly sonicated for 1 min, and was stirred for another 0.5 h. The white precipitate was filtered off, washed with cold water (30 mL) and EtOH/Et₂O (30 mL, 1:1, v/v), and dried under high-vacuum, yielding **4ii** as a white powder (3.71 g, 89%). ¹H NMR (500 MHz, DMSO-*d*₆) δ 8.11 (s, 1H), 7.71 – 7.66 (m, 2H), 7.02 – 6.96 (m, 2H), 6.51 (d, *J* = 6.8 Hz, 1H), 4.90 (d, *J* = 5.3 Hz, 1H), 4.79 (d, *J* = 5.6 Hz, 1H), 4.69 (dd, *J* = 7.7, 6.7 Hz, 1H), 4.52 (t, *J* = 5.8 Hz, 1H), 3.80 (s, 3H), 3.73 (ddd, *J* = 11.6, 5.6, 2.2 Hz, 1H), 3.48 (dt, *J* = 11.8, 6.0 Hz, 1H), 3.45 – 3.37 (m, 1H), 3.23 (ddd, *J* = 9.7, 5.9, 2.2 Hz, 1H), 3.14 (ddd, *J* = 9.7, 8.7, 5.3 Hz, 1H), 2.79 (dd, *J* = 9.3, 7.6 Hz, 1H).

1,3,4,6-Tetra-*O*-acetyl-2-deoxy-2-[(4-methoxyphenyl)-methylene]amino]- β -D-glucopyranose (4iii**)**

Prepared according to an adapted procedure.³ To a cold mixture of **4ii** (4.0 g, 13.5 mmol,) and DMAP (0.06 g, 0.5 mmol) in pyridine (22 mL) was added acetic anhydride (12 mL) at 0 °C. The reaction was stirred overnight at RT. The reaction mixture was then poured into ice water. A large amount of precipitate formed. The precipitate was collected by filtration, and washed with water and Et₂O. After dried in vacuum overnight, **4iii** was obtained as a white powder (4.6 g, 74%). ¹H NMR (500 MHz, DMSO-*d*₆) δ 8.28 (s, 1H), 7.74 – 7.60 (m, 2H), 7.07 – 6.93 (m, 2H), 6.06 (d, *J* = 8.2 Hz, 1H), 5.44 (t, *J* = 9.7 Hz, 1H), 4.97 (t, *J* = 9.7 Hz, 1H), 4.33 – 4.16 (m, 2H), 4.01 (dd, *J* = 12.3, 2.1 Hz, 1H), 3.80 (s, 3H), 3.44 (dd, *J* = 9.5, 8.6 Hz, 1H), 2.02 (s, 3H), 1.98 (s, 3H), 1.98 (s, 3H), 1.82 (s, 3H).

1,3,4,6-Tetra-*O*-acetyl-2-amino-2-deoxy- β -D-glucopyranose hydrochloride (4iv**)**

Prepared according to a reported procedure.³ To a solution of **4iii** (3.6 g, 7.73 mmol) in refluxing acetone (20 mL), 5 M aqueous solution of HCl (2 mL) was added dropwise. After 5 min, a white precipitate started to form. After vigorous stirring for 30 min, the reaction was cooled to RT, and the precipitate was filtered off, and washed successively with acetone and Et₂O. After drying in vacuum overnight, **4iv** was obtained as white powder (2.68 g, 90%). ¹H NMR (500 MHz, DMSO-*d*₆) δ 8.75 (s, 3H, NH₃Cl), 5.90 (d, *J* = 8.6 Hz, 1H, H-1), 5.35 (dd, *J* = 10.4, 9.2 Hz, 1H, H-3), 4.93 (dd, *J* = 10.1, 9.2 Hz, 1H, H-4), 4.19 (dd, *J* = 12.5, 4.5 Hz, 1H, H-6a), 4.05 (ddd, *J* = 10.1, 4.5, 2.3 Hz, 1H, H-5), 4.00 (dd, *J* = 12.5, 2.3 Hz, 1H, H-6b), 3.57 (dd, *J* = 10.4, 8.7 Hz, 1H, H-2), 2.17 (s, 3H, OAc), 2.03 (s, 3H, OAc), 2.00 (s, 3H, OAc), 1.98 (s, 3H, OAc).

1,3,4,6-Tri-*O*-acetyl-2-*N*-trichloroacetyl- β -D-glucosamine (4v**)**

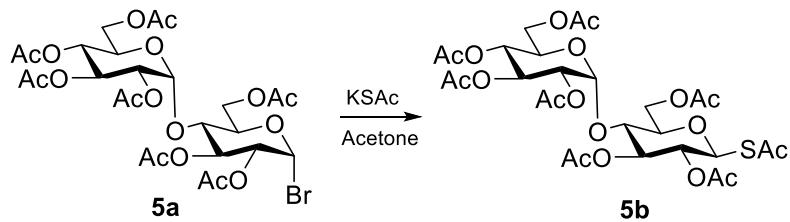
To a solution of **4iv** (1.00 g, 2.61 mmol) in 10 mL of DCM, pyridine (0.84 mL, 10.4 mmol) was added. The solution was brought to 0 °C. Trichloroacetic anhydride (1.21 g, 3.91 mmol) was added. The reaction was stirred overnight, then was poured into 50 mL of 1 M HCl solution followed by extraction with DCM (50 mL×3). The combined organic phase was washed by saturated NaHCO₃, brine, and dried over MgSO₄. After removal of the solvent, the residue was purified by flash column chromatography (ethyl acetate/hexanes=1:2) to afford **4v** as a white solid (1.25 g, 97%). ¹H NMR (500 MHz, CDCl₃) δ 7.21 (d, *J* = 9.5 Hz, 1H, NH), 5.82 (d, *J* = 8.7 Hz, 1H, H-1), 5.40 (dd, *J* = 10.9, 9.4 Hz, 1H, H-3), 5.17 (t, *J* = 9.7 Hz, 1H, H-4), 4.37 – 4.25 (m, 2H, H-2, H-6a), 4.17 (dd, *J* = 12.5, 2.2 Hz, 2H, H-6b), 3.91 (ddd, *J* = 10.0, 4.9, 2.2 Hz, 1H, H-5), 2.12 (s, 3H, OAc),

2.11 (s, 3H, OAc), 2.08 (s, 3H, OAc), 2.07 (s, 3H, OAc). The NMR spectrum is consistent with published data.⁴

3,4,6-Tri-O-acetyl-2-N-trichloroacetyl-S-acetyl-1-thio- β -D-glucosamine (4b)

To a solution of **4v** (350 mg, 0.71 mmol) in 3 mL of dry DCM, 4 mL of 33% wt HBr/AcOH solution was added dropwise at 0 °C. The reaction was brought to RT and was further stirred for 2 h. After diluting with 30 mL of DCM, the reaction mixture was poured into 60 mL of ice water and was extracted with DCM 3 times. The combined organic phase was washed by saturated NaHCO₃ solution, water, brine, and dried over Na₂SO₄. After removing the solvent by rotovap evaporation, the crude bromide product **4a** was obtained and was used immediately in the next step without further purification. To a solution of this crude **4a** in 8 mL of acetone, KSAc (162 mg, 1.42 mmol) was added. The reaction mixture was stirred at RT for 3 h. After removing the solvent by rotovap evaporation, 30 mL of DCM, 10 mL of water and 20 mL of brine were added to the residue. The mixture was extracted by DCM 2 times. The combined organic phase was dried over Na₂SO₄ then was purified by flash column chromatography (ethyl acetate/hexanes=1:2) to afford **4b** as an orange solid (285 mg, 79% over 2 steps). ¹H NMR (500 MHz, CDCl₃) δ 7.61 (d, *J* = 9.8 Hz, 1H, NH), 5.55 (dd, *J* = 10.4, 9.6 Hz, 1H, H-3), 5.37 (d, *J* = 10.7 Hz, 1H, H-1), 5.18 (t, *J* = 9.8 Hz, 1H, H-4), 4.38 (q, *J* = 10.4 Hz, 1H, H-2), 4.27 (dd, *J* = 12.6, 5.1 Hz, 1H, H-6a), 4.15 (dd, *J* = 12.5, 2.2 Hz, 1H, H-6b), 3.92 (ddd, *J* = 10.1, 5.1, 2.2 Hz, 1H, H-5), 2.38 (s, 3H, SAc), 2.11 (s, 3H, OAc), 2.10 (s, 3H, OAc), 2.06 (s, 3H, OAc). ¹³C NMR (126 MHz, CDCl₃) δ 193.14, 171.71, 170.68, 169.37, 162.20, 92.37, 80.88, 76.84, 73.42, 68.13, 62.15, 53.88, 30.85, 20.77, 20.63, 20.56.

Synthesis of 5b.

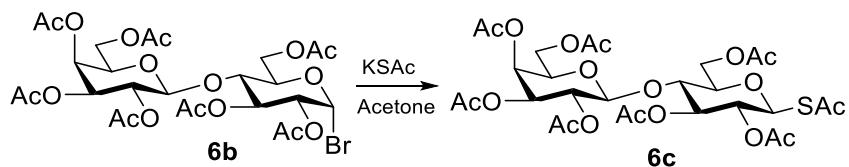


Scheme S5. Synthesis of compound **5b**.

2,3,4,6-Tetra-O-acetyl- α -D-glucopyranosyl (1 \rightarrow 4)-2,3,6-tri-O-acetyl-1-S-acetyl-1-thio- β -D-glucopyranose (**5b**)

This compound was synthesized according to general procedure A and purified by flash column chromatography (ethyl acetate: hexanes =1:1) to give **5b** as a viscous solid in 82% yield. ^1H NMR (500 MHz, CDCl_3) δ 5.38 (d, J = 3.7 Hz, 1H, H-1'), 5.37 – 5.23 (m, 3H, H-3', H-3, H-1), 5.12 – 4.90 (m, 2H, H-4', H-2), 4.84 (dd, J = 10.4, 4.0 Hz, 1H, H-2'), 4.43 (dd, J = 12.3, 2.3 Hz, 1H, H-6a), 4.30-4.12 (m, 2H, H-6b, H-6'a), 4.10 – 3.72 (m, 4H, H-6'b, H-4, H-5', H-5), 2.36 (s, 3H, SAc), 2.11 (s, 3H, SAc), 2.08 (s, 3H, OAc), 2.04 (s, 3H, OAc), 2.01 (s, 3H, OAc), 1.99 (s, 3H, OAc), 1.99 (s, 6H, OAc \times 2). The NMR spectrum is consistent with published data.¹

Synthesis of 6b.

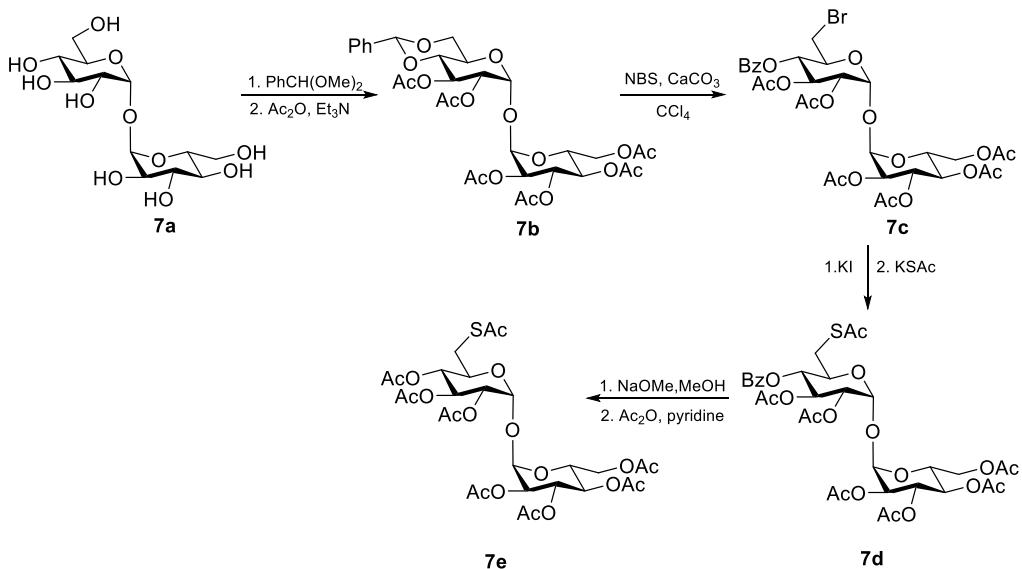


Scheme S6. Synthesis of compound **6b**.

2,3,4,6-Tetra-O-acetyl- β -D-galactopyranosyl (1 \rightarrow 4)-2,3,6-tri-O-acetyl-1-S-acetyl-1-thio- β -D-glucopyranose (**6b**)

This compound was synthesized according to general procedure A, and purified by flash column chromatography (ethyl acetate/hexanes =1:1) to give **6b** as a viscous solid in 85% yield. ¹H NMR (500 MHz, CDCl₃) δ 5.35 (dd, *J* = 3.4, 1.0 Hz, 1H, H-4'), 5.25 (t, *J* = 9.1 Hz, 1H, H-3), 5.21 (d, *J* = 10.5 Hz, 1H, H-1), 5.11 (dd, *J* = 10.4, 7.9 Hz, 1H, H-2'), 5.04 (dd, *J* = 10.4, 9.2 Hz, 1H, H-2), 4.94 (dd, *J* = 10.4, 3.5 Hz, 1H, H-3'), 4.46 (d, *J* = 7.9 Hz, 1H, H-1'), 4.45 (dd, *J* = 12.1, 1.9 Hz, 1H, H-6'a), 4.16 – 4.04 (m, 3H, H-6a, H-6b, H-6'b), 3.89 – 3.84 (m, 1H, H-5'), 3.82 (dd, *J* = 9.9, 9.0 Hz, 1H, H-4), 3.75 (ddd, *J* = 10.0, 4.7, 1.9 Hz, 1H, H-5), 2.37 (s, 3H, SAc), 2.15 (s, 3H, OAc), 2.11 (s, 3H, OAc), 2.07 (s, 3H, OAc), 2.05 (s, 3H, OAc), 2.04 (s, 3H, OAc), 2.02 (s, 3H, OAc), 1.96 (s, 3H, OAc). The NMR spectrum is consistent with published data.¹

Synthesis of **7e**.



Scheme S7. Synthesis of compound **7e**.

2,3,2',3',4',6'-O-Acetyl-4,6-di-O-benzylidene-*a*-D-trehalose (**7b**)

Prepared according to an adapted procedure.⁵ Benzaldehyde dimethyl acetal (1.62 mL, 10.8 mmol), trehalose (2.05 g, 6.0 mmol) and *p*-toluenesulfonic acid monohydrate (0.23 g, 1.2 mmol) were added to DMF (30 mL). The mixture was stirred for 12 hours at 40 °C. After cooling to 0 °C, triethylamine (23.60 g, 232.0 mol) and DMAP (122 mg, 1.00 mmol) were added followed by Ac₂O (13.0 g, 116.0 mmol) dropwise. The reaction mixture was slowly warmed to RT and stirred overnight, after which it was poured into water, and extracted by ethyl acetate 3 times. The combined organic phase was washed by brine twice, and dried over Na₂SO₄. After removing the solvent, the residue was purified by column chromatography (ethyl acetate/hexanes = 1:1.5 to 1:1) twice to give the desire product as a white amorphous solid (1.60 g, 40%). ¹H NMR (500 MHz, CDCl₃): δ 7.48 - 7.38 (m, 2H, Ar-H), 7.38 - 7.30 (m, 3H, Ar-H), 5.61 (t, *J* = 9.8 Hz, 1H, H-3'), 5.55 - 5.44 (m, 2H, H-3, PhCH), 5.37 (d, *J* = 3.7 Hz, 1H, H-1'), 5.27 (d, *J* = 3.7 Hz, 1H, H-1), 5.09 - 5.02 (m, 2H, H-2' and H-4'), 5.00 (dd, *J* = 10.2, 4.0 Hz, 1H, H-2), 4.25 (dd, *J* = 12.2, 5.6 Hz, 1H; H-6a'), 4.17 (dd, *J* = 10.5, 4.9 Hz, 1H, H-6a), 4.09 (ddd, *J* = 10.3, 5.7, 2.2 Hz, 1H, H-5'), 4.01 (dd, *J* = 12.2, 2.2, 1H, H-6b'), 3.97 (td, *J* = 9.9, 4.9 Hz, 1H, H-5), 3.75 (t, *J* = 10.4 Hz, 1H, H-6b), 3.69 (t, *J* = 9.6 Hz, 1H, H-4), 2.22 - 1.96 (m, 18H, OAc). ¹³C NMR (126 MHz, CDCl₃) δ 170.76, 170.22, 170.03, 169.97, 169.78, 169.78, 136.87, 129.32, 128.41, 126.34, 101.96, 93.49, 92.35, 79.16, 77.43, 70.78, 70.24, 70.16, 69.06, 68.70, 68.28, 63.34, 61.94, 21.01, 20.84, 20.79, 20.79.

4-O-Benzoyl-6-bromo-2,3,2',3',4',6'-penta-O-acetyl-6-deoxy- α , α -D-trehalose (7c)

Prepared according to an adapted procedure.⁵ Compound **7b** (1.44 g, 2.11 mmol) was added into 60 mL of CCl₄ containing NBS (413 mg, 2.32 mmol) and CaCO₃ (232 mg, 2.32 mmol). The mixture was refluxed at 77 °C for 3 hours. After cooling to room temperature,

the solution was washed with saturated NaHCO₃ and water. The organic phase was dried over Na₂SO₄, and the filtrate was concentrated in vacuum. The residue was purified by flash column chromatography (ethyl acetate/hexanes = 2:3) to give **7c** as a white solid (1.43 g, 91%). ¹H NMR (500 MHz, CDCl₃): δ 8.15 - 7.95 (d, *J* = 7.3 Hz, 2H, Ar-H), 7.63 (t, *J* = 7.3 Hz, 1H, Ar-H), 7.49 (t, *J* = 7.7 Hz, 2H, Ar-H), 5.71 (t, *J* = 9.8 Hz, 1H, H-3), 5.54 (t, *J* = 9.7 Hz, 1H, H-3'), 5.41 (d, *J* = 3.9 Hz, 1H, H-1), 5.38 (d, *J* = 3.9 Hz, 1H, H-1'), 5.21 - 5.00 (m, 4H, H-2, H-4', H-2' and H-4), 4.25 (m, 2H, H-5' and H-6a'), 4.16 - 3.97 (m, 2H, H-5 and H-6b'), 3.49 - 3.25 (m, 2H, H-6a and H-6b), 2.28 - 1.76 (m, 18H, OAc). ¹³C NMR (126 MHz, CDCl₃) δ 170.81, 170.81, 170.24, 169.73, 169.65, 169.65, 165.59, 134.10, 130.17, 130.17, 128.91, 128.91, 128.65, 92.39, 91.98, 71.82, 70.48, 70.40, 70.29, 69.43, 69.43, 68.80, 68.45, 61.99, 30.75, 21.17, 20.92, 20.90, 20.81, 20.80, 20.78, 20.76.

4-O-Benzoyl-6-S-acetyl-2,3,2',3',4',6'-penta-O-acetyl -α, α-D-trehalose (7d)

Compound **7c** (2.95 g, 3.87 mmol) and KI (1.93 g, 11.6 mmol) was added to a round bottle flask containing 30 mL of DMF. After stirring at 60 °C for 4 h, the mixture was cooled to RT, and KSAc (1.33 g, 11.6 mmol) was added to the mixture. The reaction was stirred under Ar protection overnight. The resulting mixture was poured into 100 mL brine/100 mL water and extracted with ethyl acetate (100 mL×3). The combined organic layer was further washed with 200 mL brine and dried on MgSO₄, concentrated in vacuum. The residue was purified by column chromatography (ethyl acetate/hexanes = 1 to 1) to afford the product as a viscous solid (2.93 g, 96%). ¹H NMR (500 MHz, CDCl₃) δ 8.04 (dd, *J* = 8.3, 1.2 Hz, 1H, Ar-H), 7.60 (t, *J* = 7.5 Hz, 1H, Ar-H), 7.47 (t, *J* = 7.8 Hz, 1H, Ar-H), 5.67 (dd, *J* = 9.8, 9.7 Hz, 1H, H-3), 5.52 (dd, *J* = 9.9, 9.6 Hz, 1H, H-3'), 5.34 (d, *J* = 3.9 Hz, 2H, H-1, H-1'), 5.20 (t, *J* = 9.7 Hz, 1H, H-4), 5.13 – 4.98 (m, 3H, H-2, H-2', H-4'), 4.20 (dd,

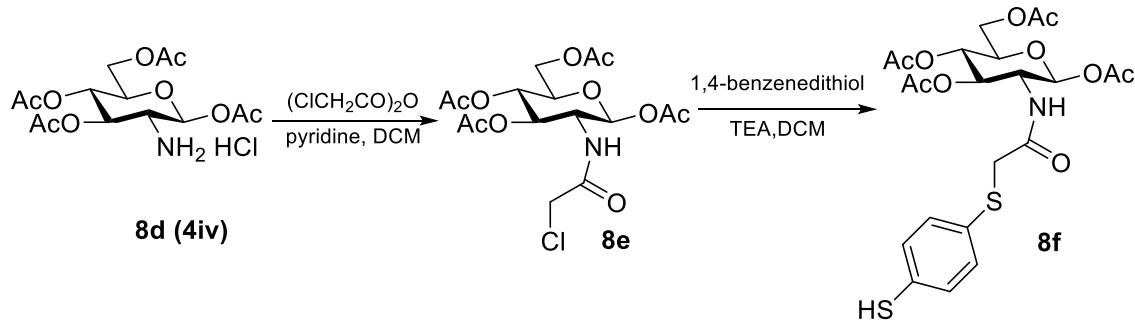
= 12.2, 6.0 Hz, 1H, H-6a'), 4.07 (dd, J = 12.2, 2.1 Hz, 1H, H-6b'), 4.03 – 3.90 (m, 2H, H-5, H-5'), 3.30 (dd, J = 14.2, 2.6 Hz, 1H, H-6a), 2.91 (dd, J = 14.5, 9.2 Hz, 1H, H-6b), 2.32 (s, 3H, SAC), 2.11 (s, 3H, OAc), 2.10 (s, 3H, OAc), 2.08 (s, 3H, OAc), 2.05 (s, 3H, OAc), 2.04 (s, 3H, OAc), 1.93 (s, 3H, OAc). ^{13}C NMR (126 MHz, CDCl_3) δ 194.64, 170.73, 170.01, 169.94, 169.85, 169.77, 169.70, 165.66, 133.78, 130.02, 128.95, 128.73, 91.26, 91.12, 72.03, 70.34, 70.31, 69.76, 69.60, 69.53, 68.72, 68.31, 61.93, 30.47, 30.27, 20.80, 20.72, 20.72, 20.72, 20.70, 20.70.

6-S-Acetyl-2,3,4,2',3',4',6'-hexa-O-acetyl- α,α -D-trehalose (7e)

Compound **7d** (1.44 g, 1.90 mmol) was dissolved in 50 mL methanol. NaOMe (113 mg, 2.09 mmol) in 3 mL methanol was added to the reaction under Ar protection. The reaction mixture was stirred overnight. After removing the solvent under vacuum, 7 mL of pyridine was added followed by addition of acetic anhydride (2.7 mL, 28.5 mmol) dropwise at 0° C. The reaction was stirred for 16 hours at RT. The reaction mixture was diluted by 100 mL ethyl acetate and poured into 100 mL of water. After extraction for 3 times by ethyl acetate, the combined organic layer was washed with 1 M HCl, saturated NaHCO_3 , water, and brine, dried over MgSO_4 . After removing the solvent, the residue was purified by column chromatography (ethyl acetate/hexanes = 1:1) to afford the product as a viscous solid (964 mg, 73% over 2 steps). ^1H NMR (500 MHz, CDCl_3) δ 5.44 (t, J = 9.7 Hz, 2H, H-4, H-4'), 5.27 (d, J = 3.9 Hz, 1H, H-1'), 5.24 (d, J = 3.8 Hz, 1H, H-1), 5.05 – 4.89 (m,

4H, H-2, H-2', H-3, H-3'), 4.16 (dd, $J = 12.2, 6.0$ Hz, 1H, H-6'a), 4.01 (dd, $J = 12.2, 1.9$ Hz, 1H, H-6'b), 3.94 (ddd, $J = 10.0, 5.9, 1.7$ Hz, 1H, H-5'), 3.89 – 3.80 (m, 1H, H-5), 3.15 (dd, $J = 14.2, 2.6$ Hz, 1H, H-6a), 2.93 (dd, $J = 14.2, 7.9$ Hz, 1H, H-6b), 2.32 (s, 3H, SAc), 2.07 (s, 3H, OAc), 2.06 (s, 3H, OAc), 2.05 (s, 3H, OAc), 2.04 (s, 3H, OAc), 2.02 (s, 3H, OAc), 2.00 (s, 3H, OAc), 2.00 (s, 3H, OAc). ^{13}C NMR (50 MHz, CDCl_3) δ 194.73, 170.69, 169.96, 169.85, 169.83, 169.66, 91.45, 91.31, 71.07, 70.20, 70.12, 69.93, 69.70, 69.48, 68.68, 68.30, 61.92, 30.47, 29.93, 20.76, 20.76, 20.73, 20.67.

Synthesis of 8f.



Scheme S8. Synthesis of compound **8f**. Compounds **8a–8d** are identical to compounds **4i–4iv** (see **Scheme S4** and the procedures for their syntheses).

1,2,3,4-Tetra-O-acetyl-2-chloroacetamido-2-deoxy- β -D-glucopyranose (8e)

Prepared according to a reported procedure.⁶ To a solution of **8d** (2.53 g, 6.51 mmol) in 30 mL of DCM, pyridine (0.84 mL, 10.4 mmol) was added. The solution was brought to 0 °C. Chloroacetic anhydride (1.67 g, 9.77 mmol) was added. The reaction solution was stirred overnight, then was poured into 50 mL 1 M HCl solution and was extracted by DCM (50

mL×3). The combined organic phase was washed by saturated NaHCO₃, brine, and dried over MgSO₄. After removing the solvent, the residue was purified by flash column chromatography (ethyl acetate/hexanes=2:3) to afford **8e** as a white solid (2.45 g, 89%).

¹H NMR (500 MHz, CDCl₃) δ 6.59 (d, *J* = 9.2 Hz, 1H, NH), 5.81 (d, *J* = 8.6 Hz, 1H, H-1), 5.28 (dd, *J* = 10.5, 9.3 Hz, 1H, H-3), 5.14 (t, *J* = 9.5 Hz, 1H, H-4), 4.29 (dd, *J* = 12.5, 4.7 Hz, 1H, H-6a), 4.22 (dt, *J* = 10.5, 9.0 Hz, 1H, H-2), 4.14 (dd, *J* = 12.5, 2.3 Hz, 1H, H-6b), 3.98 (d, *J* = 1.6 Hz, 2H, CH₂Cl), 3.85 (ddd, *J* = 9.8, 4.6, 2.3 Hz, 1H, H-5), 2.12 (s, 3H, OAc), 2.10 (s, 3H, OAc), 2.05 (s, 3H, OAc), 2.05 (s, 3H, OAc).

1,3,4,6-Tetra-*O*-acetyl-2-((4-mercaptophenyl) sulfanyl)acetamido-2-deoxy-β-D-glucopyranose (8f**)**

To a solution of compound **8e** (160 mg, 0.378 mmol) and 1,4-benzenedithiol (107 mg, 0.755 mmol) in 5 mL of DCM, TEA (42 mg, 0.415 mmol) was added. The reaction was stirred for 16 h. Then it was concentrated and directly purified by flash column chromatography (ethyl acetate/DCM = 1:1.2) to give the product as a colorless viscous solid (180 mg, 90%). ¹H NMR (500 MHz, CDCl₃) δ 7.22 – 7.17 (m, 2H, Ar-H), 7.15 – 7.09 (m, 2H, Ar-H), 6.81 (d, *J* = 9.2 Hz, 1H, NH), 5.73 (d, *J* = 8.7 Hz, 1H, H-1), 5.24 (dd, *J* = 10.5, 9.3 Hz, 1H, H-3), 5.09 (t, *J* = 9.6 Hz, 1H, H-4), 4.27 (dd, *J* = 12.5, 4.6 Hz, 1H, H-6a), 4.18 (dt, *J* = 10.5, 9.1 Hz, 1H, H-2), 4.11 (dd, *J* = 12.5, 2.2 Hz, 1H, H-6b), 3.82 (ddd, *J* = 9.9, 4.6, 2.3 Hz, 1H, H-5), 3.53 (q, *J* = 16.6 Hz, 2H, SCH₂), 3.45 (s, 1H, SH), 2.08 (s, 3H, OAc), 2.02 (s, 3H, OAc), 1.95 (s, 3H, OAc), 1.88 (s, 3H, OAc). ¹³C NMR (126 MHz, CDCl₃) δ 170.80, 170.74, 169.43, 169.30, 168.43, 131.72, 130.33, 130.06, 129.04, 92.28, 72.94, 72.18, 68.10, 61.76, 53.47, 37.60, 20.83, 20.81, 20.69, 20.57.

Determination of compound purity by absolute qHNMR with internal calibration

The purities of compounds **1—40** were determined by absolute qNMR following the “general guidelines for quantitative 1D ^1H NMR (qHNMR) experiments,” provided by the *Journal of Medicinal Chemistry*.

The internal calibrant dimethyl sulfone (DMSO_2) was purchased from Sigma-Aldrich (product number: M81705, batch number: WXBC7924V). The purity of the internal calibrant (P_{IC}) was 100.0% according to the “Certificate of Analysis” of this particular batch provided by the vendor. The absolute qHNMR with internal calibration was conducted on a Bruker Avance Spectrospin DRX500 spectrometer at 298K. The data were obtained at 90° pulse tip angle with an interpulse delay (D_1) of 60 s and an acquisition time of 3.2 s in a non-spinning mode. The FID was obtained following 64 scans of 256 K data points with a 20 ppm width spectral window. The data were processed with the MestReNova 9.0.1 software.

General procedure for qHNMR

Step 1: The weights of the sample (m_s) and the internal calibrant (m_{IC}) were measured on a semi-micro METTLER TOLEDO balance with 0.01 mg accuracy. Then they were fully dissolved in 650 μL of CDCl_3 or 600 μL of D_2O in a 2-mL vial. The resulting solution was transferred into a 5-mm standard NMR tube for analysis.

Step 2: After manual phase and baseline correction, the purest signals of the sample were integrated. The integral of the sample (Int_t) was calculated as the average of all integrated

protons by dividing the sum of all those integrals by the number of protons that give rise the signals. By this procedure, the total number of protons (n_t) is set to 1.

Step 3: The integral of the internal calibrant (DMSO₂, singlet at 3.0-3.2 ppm) and the number of protons that give rise to this signal were recorded as Int_{IC} and n_{IC} ($n_{IC} = 6$).

Step 4: Calculate the molecular weights of the sample (MW_t) and the internal calibrant ($MW_{IC} = 94.13$ g/mol).

Step 5: Calculate the purity (P) of the sample according to the following equation:

$$P [\%] = \frac{n_{IC} \cdot Int_t \cdot MW_t \cdot m_{IC}}{n_t \cdot Int_{IC} \cdot MW_{IC} \cdot m_s} \cdot P_{IC}$$

References

- Shu, P.; Zeng, J.; Tao, J.; Zhao, Y.; Yao, G.; Wan, Q., Selective S-deacetylation inspired by native chemical ligation: practical syntheses of glycosyl thiols and drug mercapto-analogues. *Green Chem.* **2015**, *17* (4), 2545-2551.
- Floyd, N.; Vijayakrishnan, B.; Koeppe, J. R.; Davis, B. G., Thiyil glycosylation of olefinic proteins: S-Linked glycoconjugate synthesis. *Angew. Chem., Int. Ed.* **2009**, *48* (42), 7798-7802.
- Dąbrowa, K.; Niedbała, P.; Jurczak, J., Engineering light-mediated bistable azobenzene switches bearing urea D-aminoglucose units for chiral discrimination of carboxylates. *J. Org. Chem.* **2016**, *81* (9), 3576-3584.
- Qin, C.; Schumann, B.; Zou, X.; Pereira, C. L.; Tian, G.; Hu, J.; Seeberger, P. H.; Yin, J., Total synthesis of a densely functionalized plesiomonas shigelloides serotype 51 aminoglycoside trisaccharide antigen. *J. Am. Chem. Soc.* **2018**, *140* (8), 3120-3127.
- Chen, X.; Wu, B.; Jayawardana, K. W.; Hao, N.; Jayawardena, H. S. N.; Langer, R.; Jaklenec, A.; Yan, M., Magnetic multivalent trehalose glycopolymers nanoparticles for the detection of mycobacteria. *Adv. Healthcare Mater.* **2016**, *5* (16), 2007-2012.
- Liu, F.; Vijayakrishnan, B.; Faridmoayer, A.; Taylor, T. A.; Parsons, T. B.; Bernardes, G. J. L.; Kowarik, M.; Davis, B. G., Rationally designed short polyisoprenol-Linked PglB substrates for engineered polypeptide and protein N-glycosylation. *J. Am. Chem. Soc.* **2014**, *136* (2), 566-569.

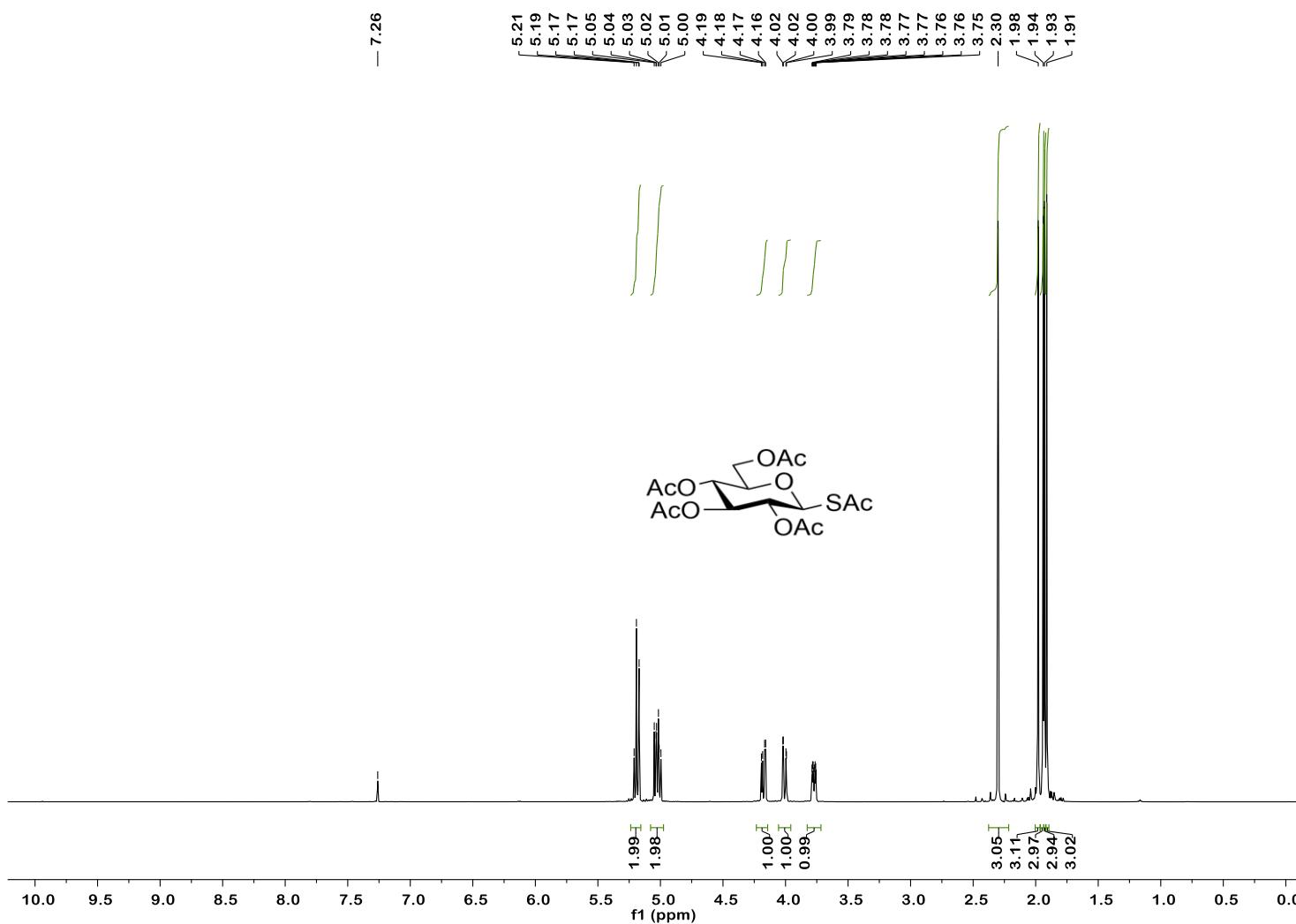


Figure S1. ^1H NMR spectrum of compound **1b** in CDCl_3 .

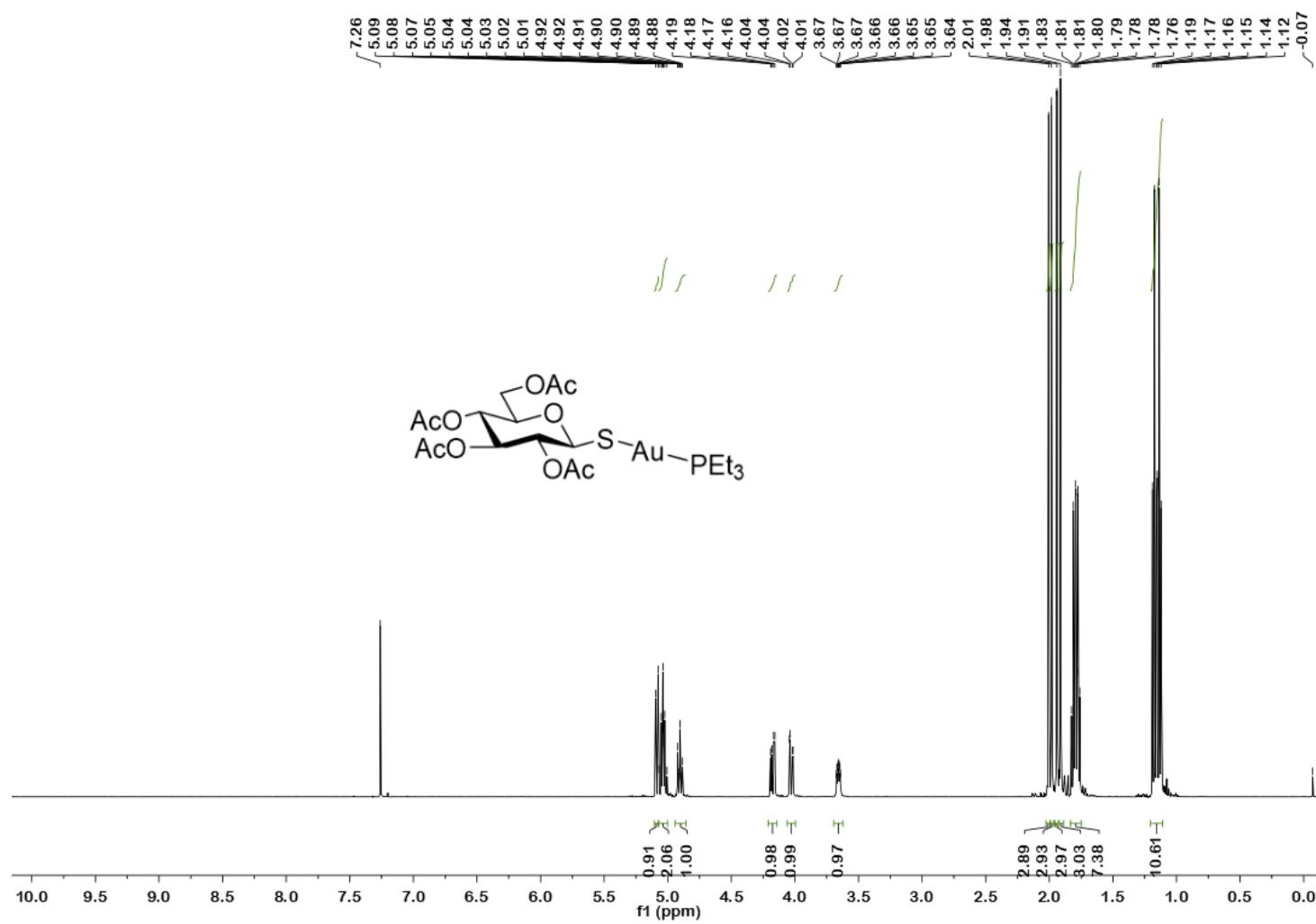


Figure S2. ^1H NMR spectrum of compound **1** in CDCl_3 .

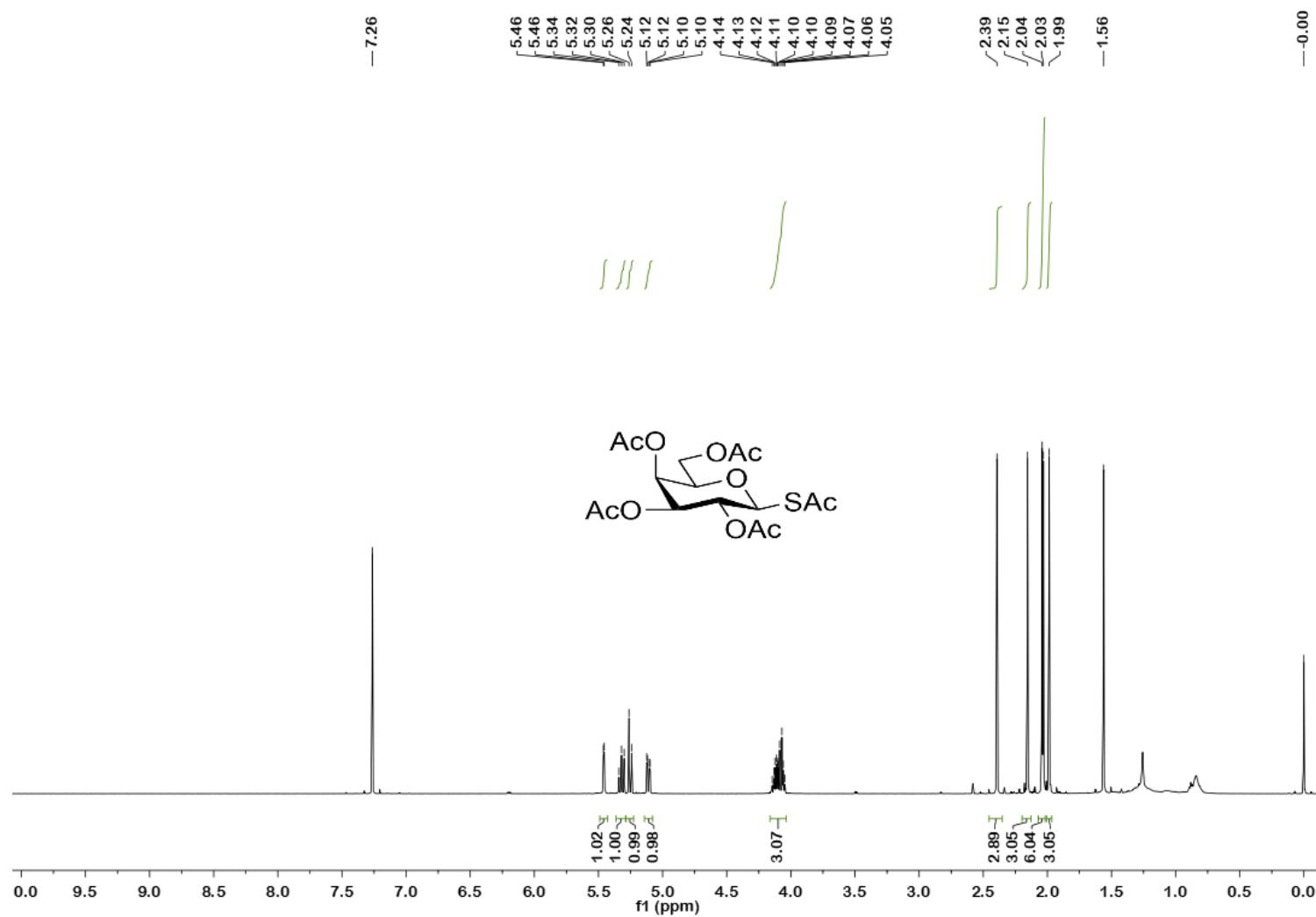


Figure S3. ^1H NMR spectrum of compound **2b** in CDCl_3 .

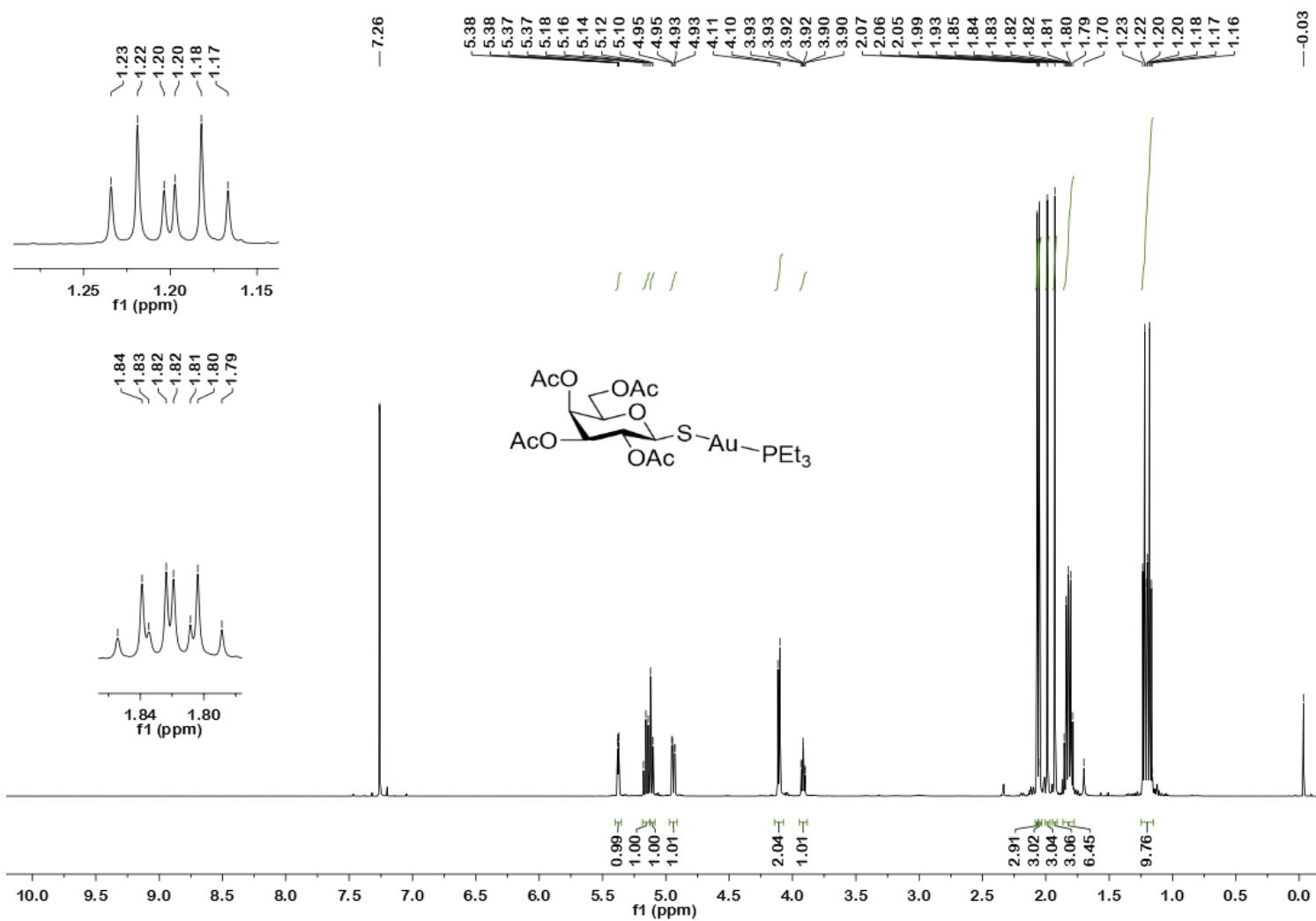


Figure S4. ^1H NMR spectrum of compound **2** in CDCl_3 .

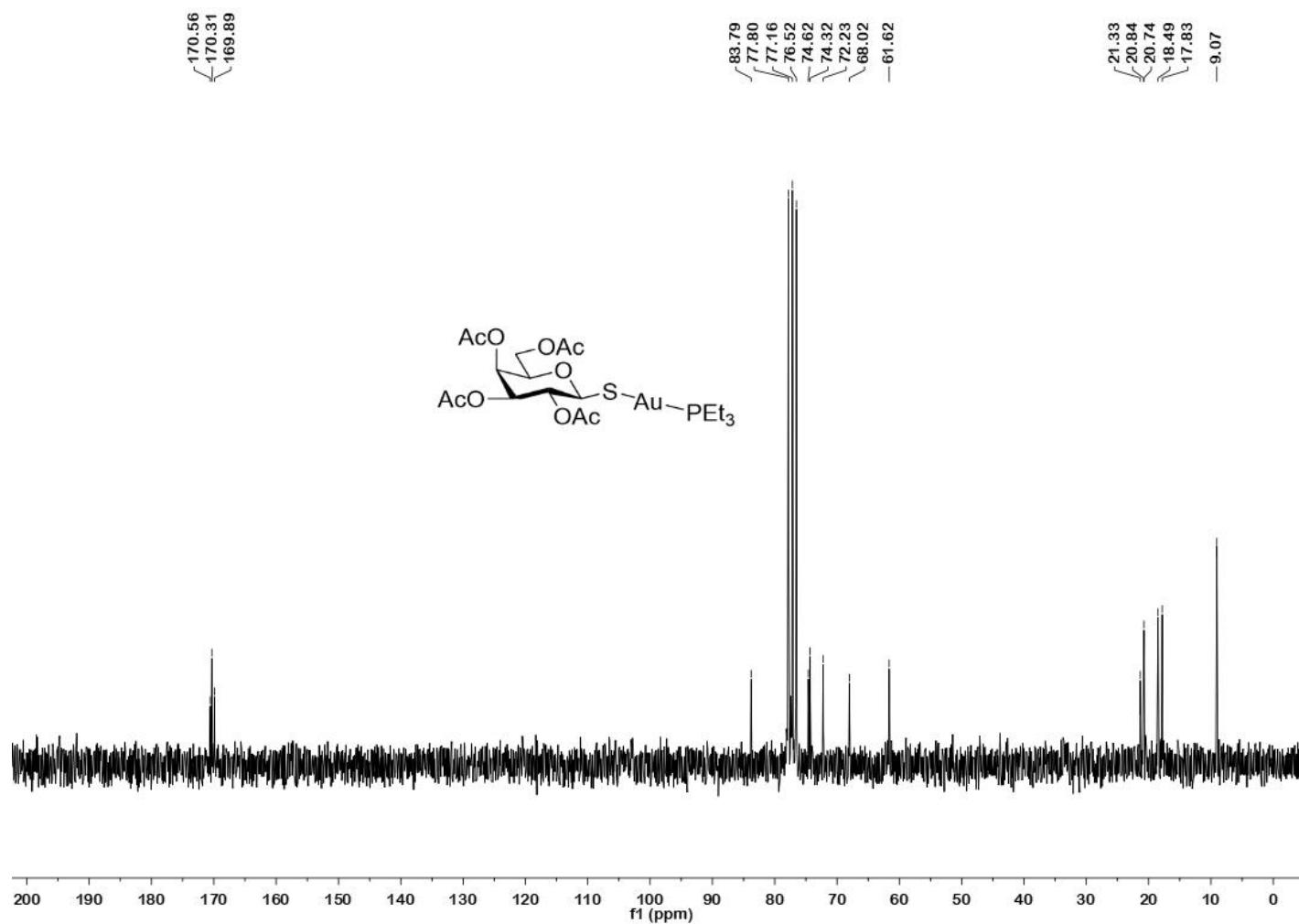


Figure S5. ^{13}C NMR spectrum of compound 2 in CDCl_3 .

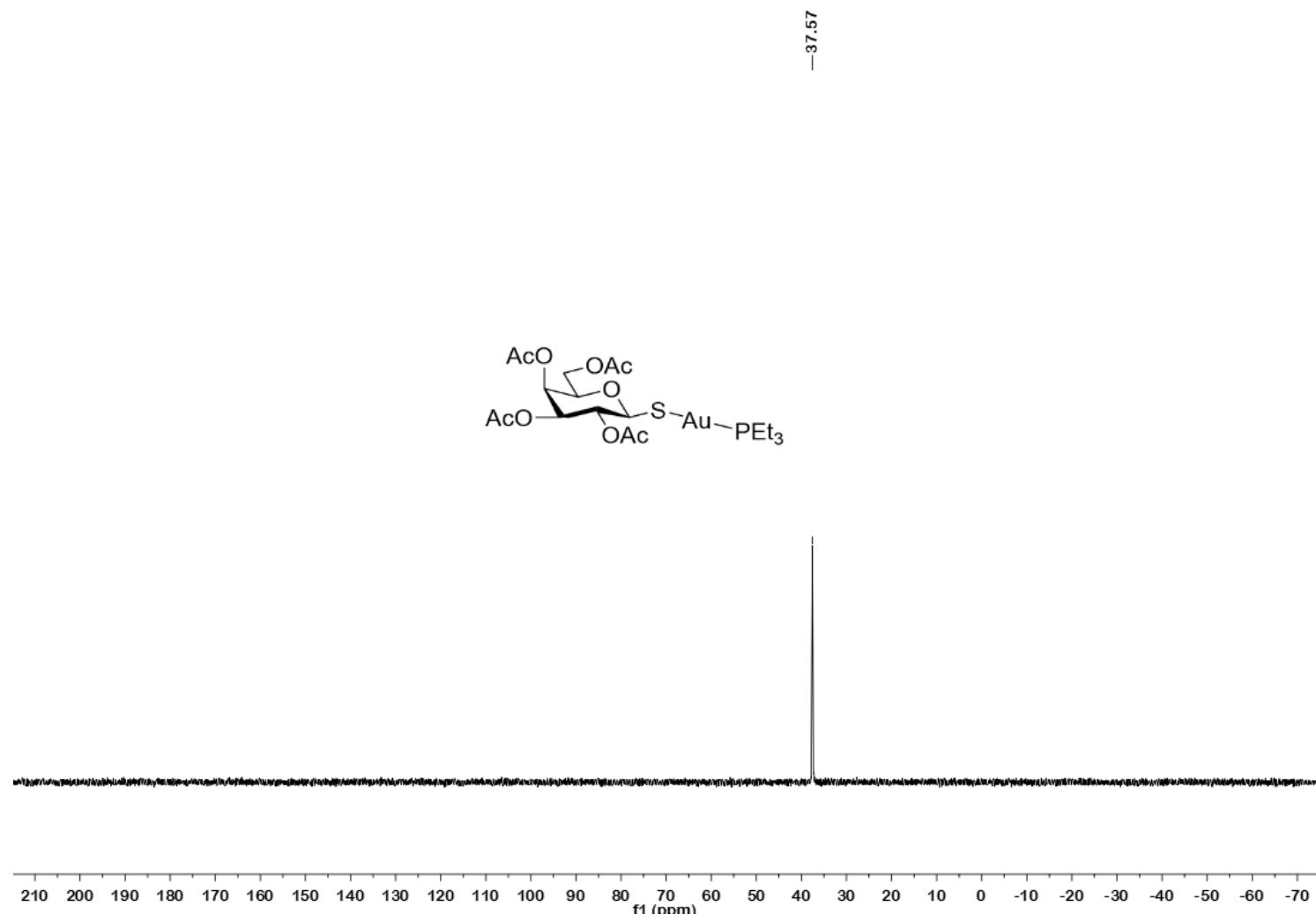


Figure S6. ^{31}P NMR spectrum of compound 2 in CDCl_3 .

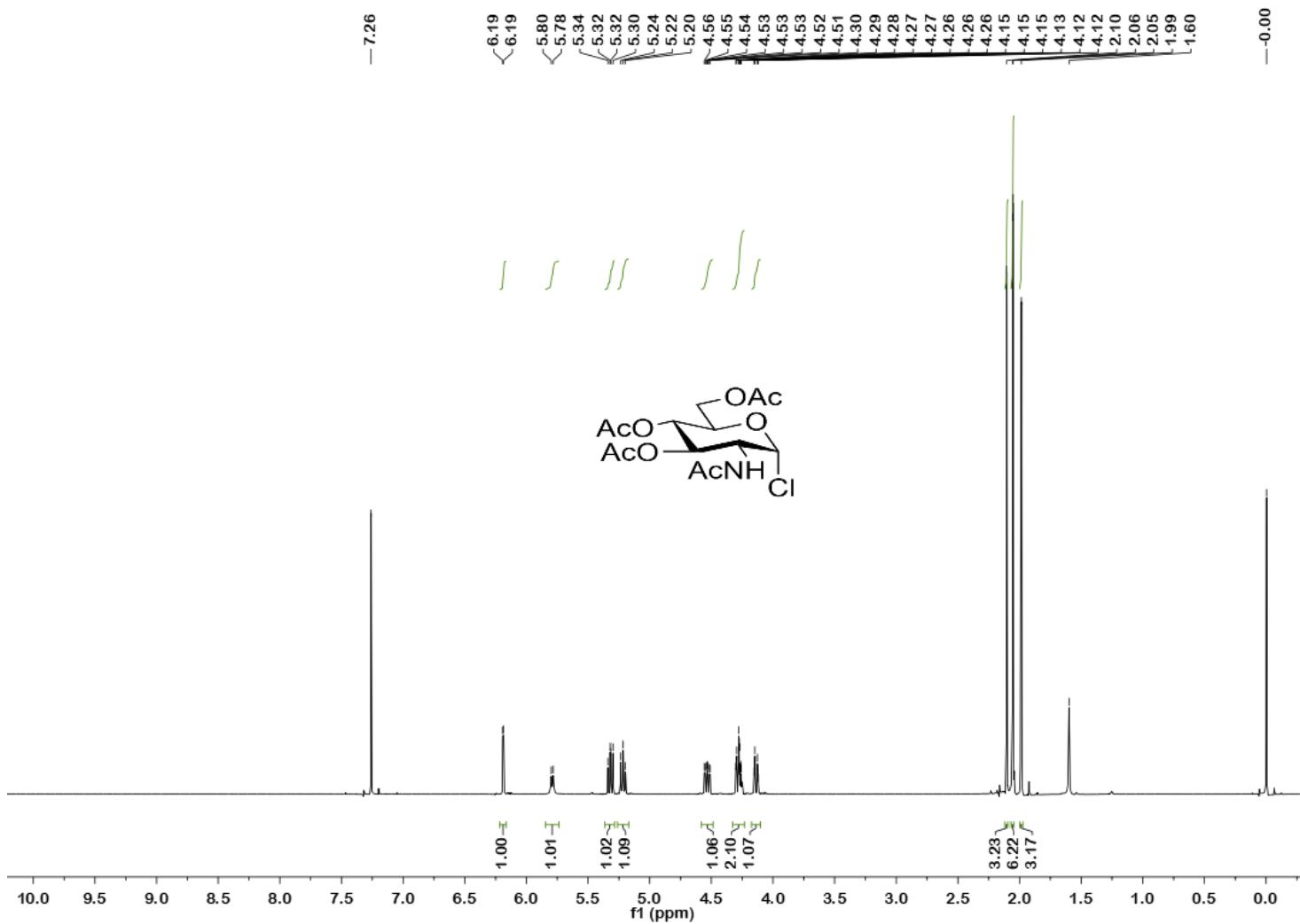


Figure S7. ^1H NMR spectrum of compound **3a** in CDCl_3 .

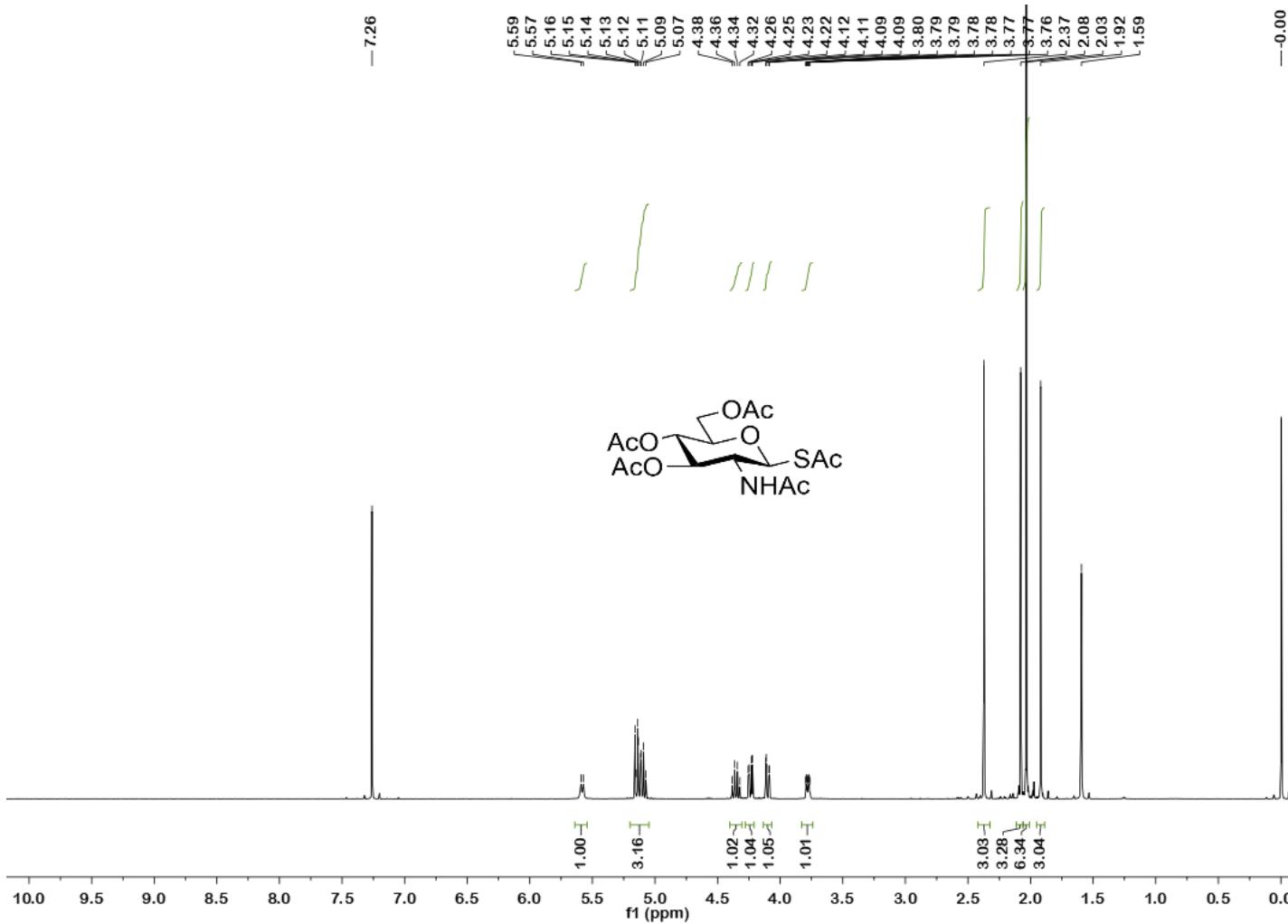


Figure S8. ^1H NMR spectrum of compound **3b** in CDCl_3 .

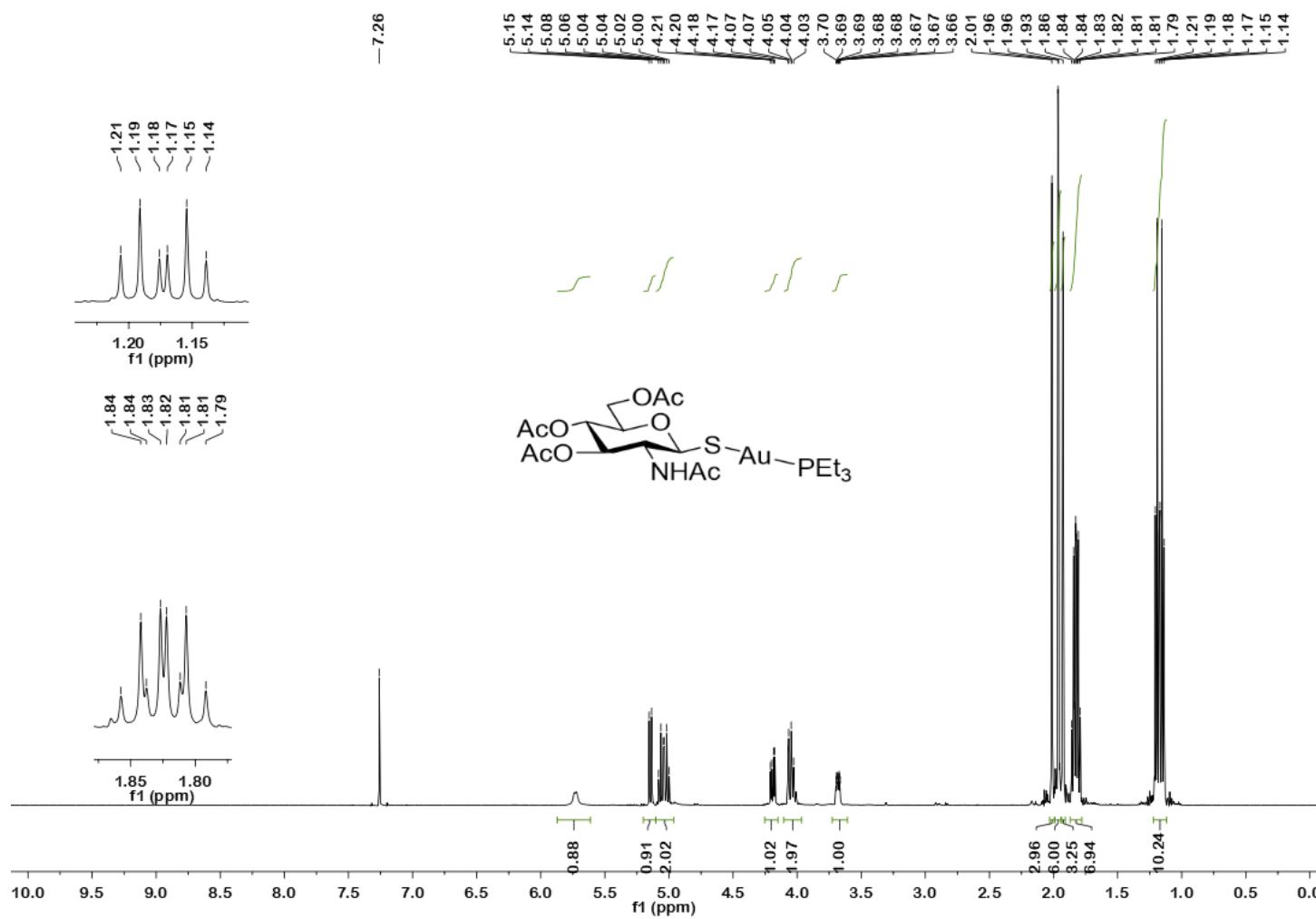


Figure S9. ^1H NMR spectrum of compound **3** in CDCl_3 .

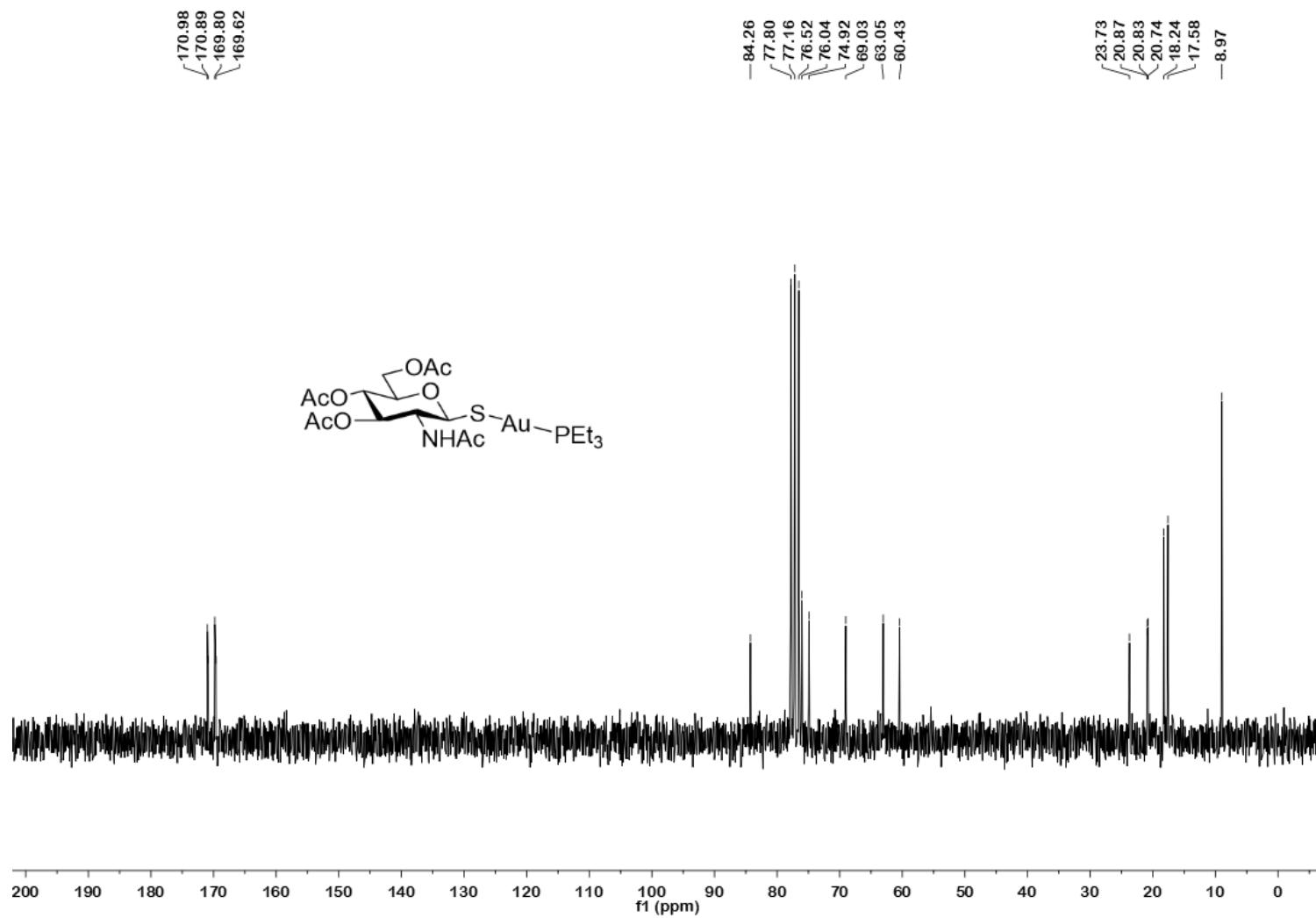


Figure S10. ¹³C NMR spectrum of compound 3 in CDCl₃.

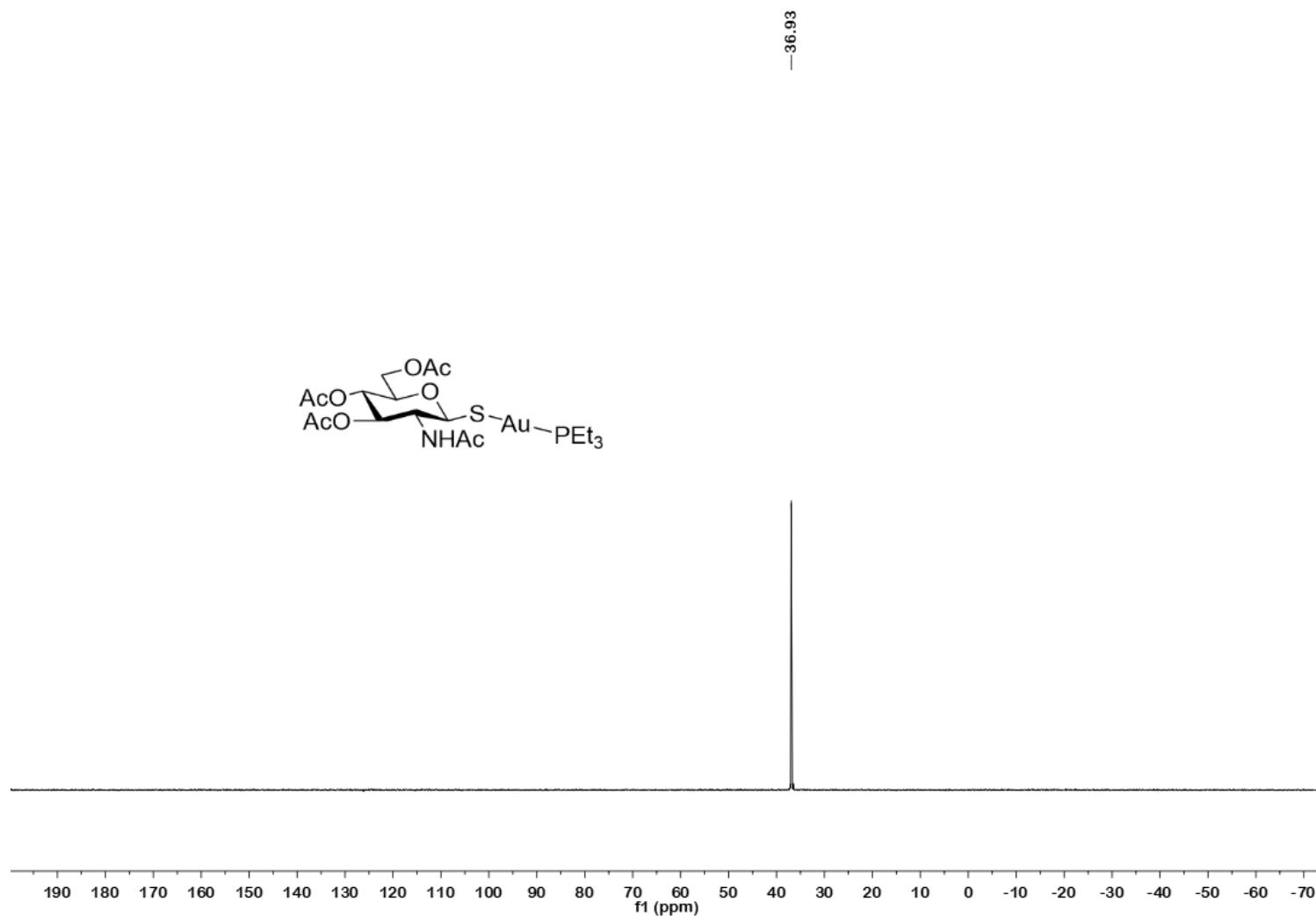
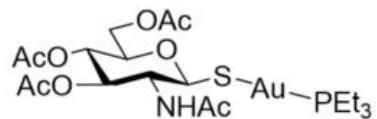


Figure S11. ^{31}P NMR spectrum of compound 3 in CDCl_3 .

Madanodaya Sundhor WB-08

Synapt_10293 26 (0.536) Cm (26-8)

100



Mass Spectrometry Lab, SCS, University of Illinois

SYNAPTG2-Si#UGA305

13:52:21

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992.2065

644.1696

645.1725

700.1380

740.0776

993.2096

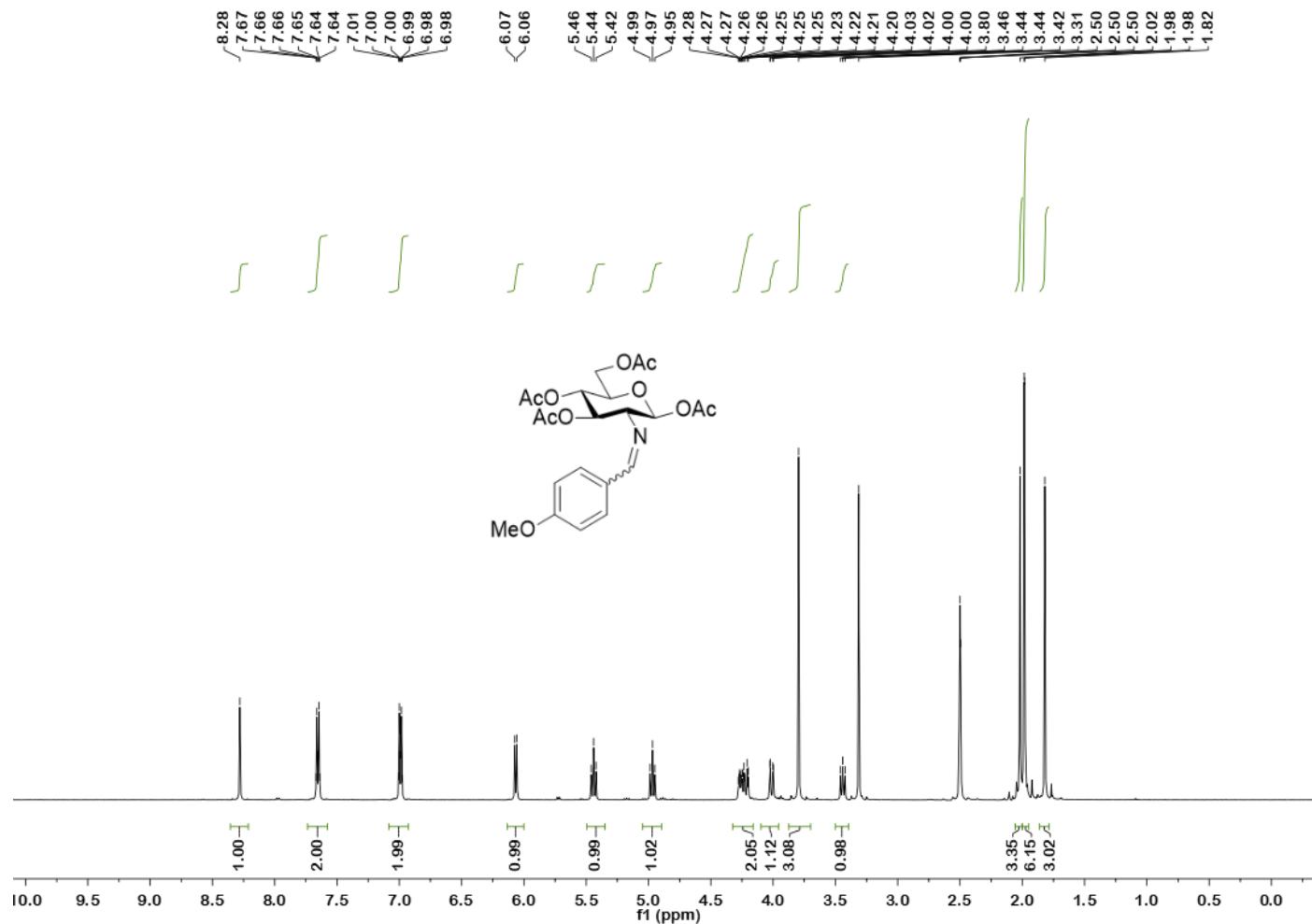
994.2076

977.1433

1052.2084

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Figure S12. HRMS of compound 3.



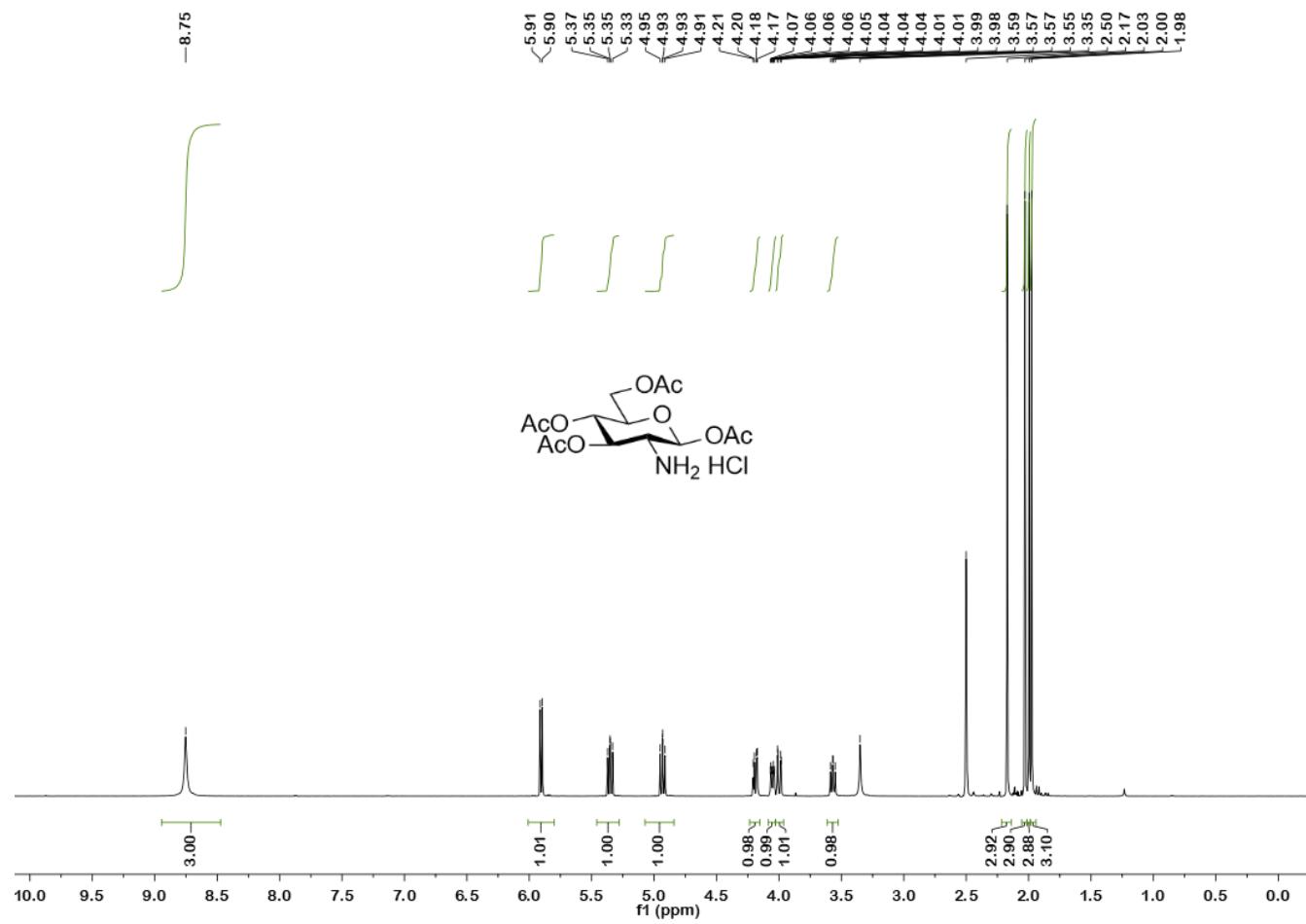


Figure S14. ^1H NMR spectrum of compound **4d** in DMSO-d_6 .

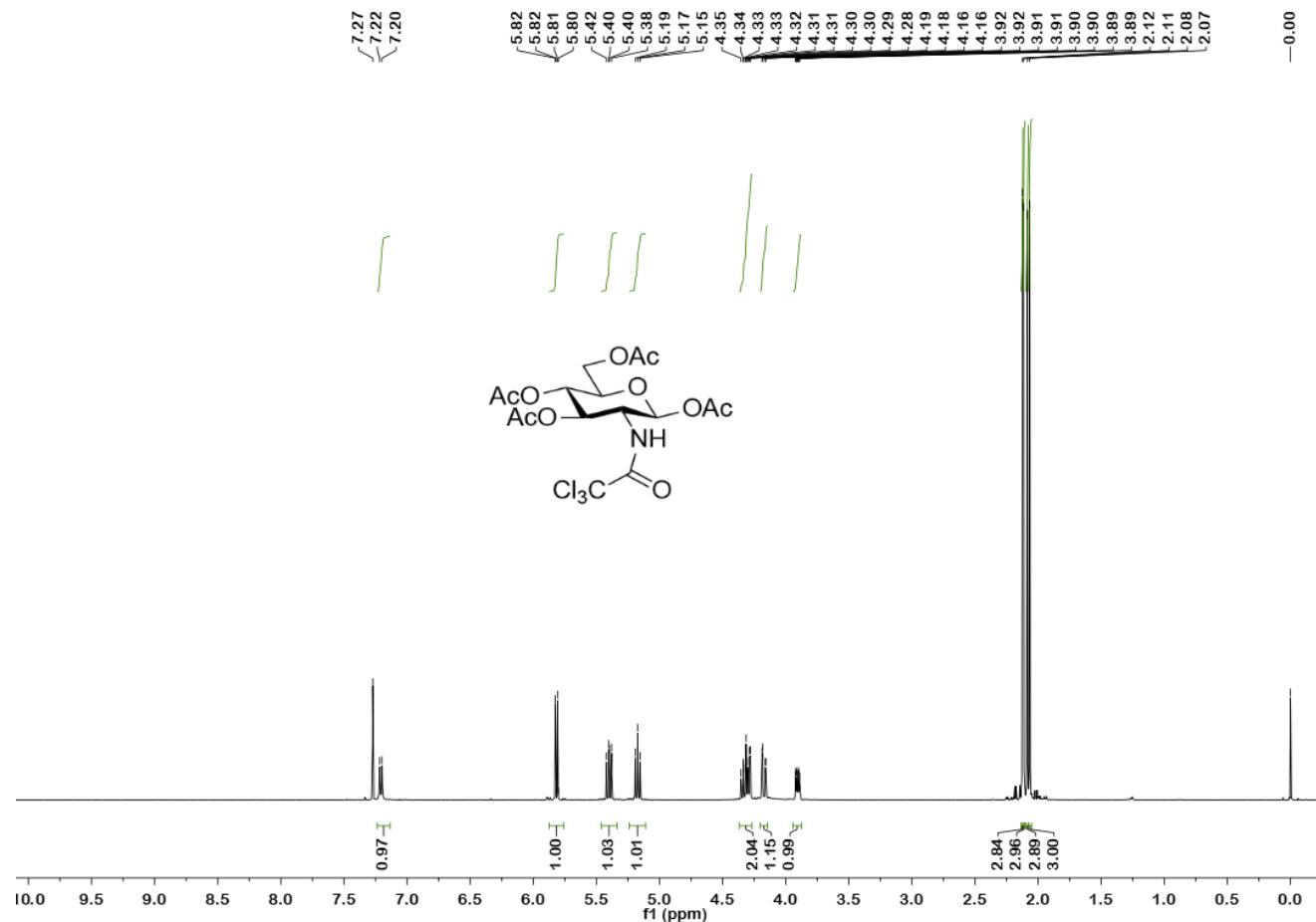


Figure S15. ^1H NMR spectrum of compound **4e** in CDCl_3 .

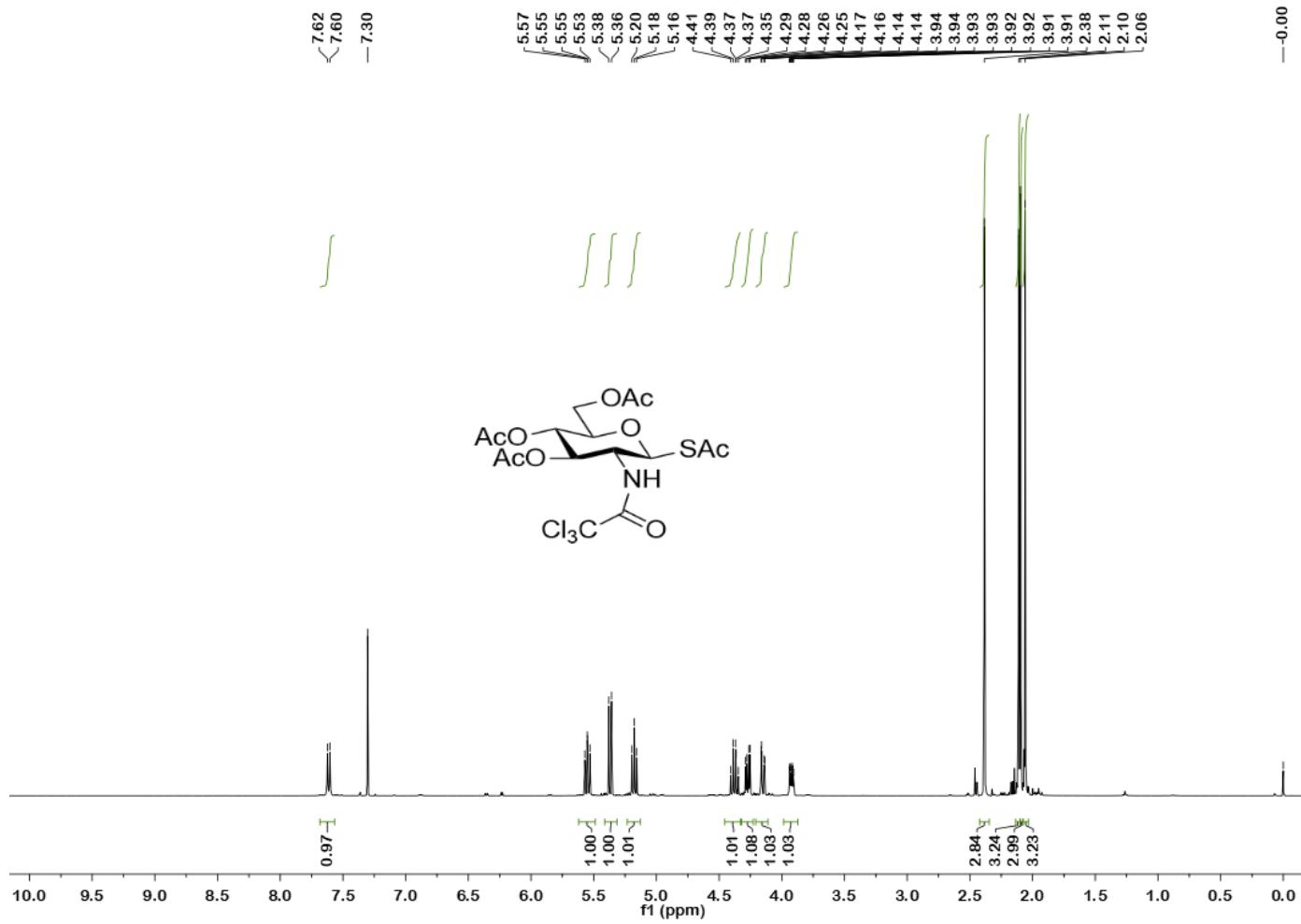


Figure S16. ^1H NMR spectrum of compound **4f** in CDCl_3 .

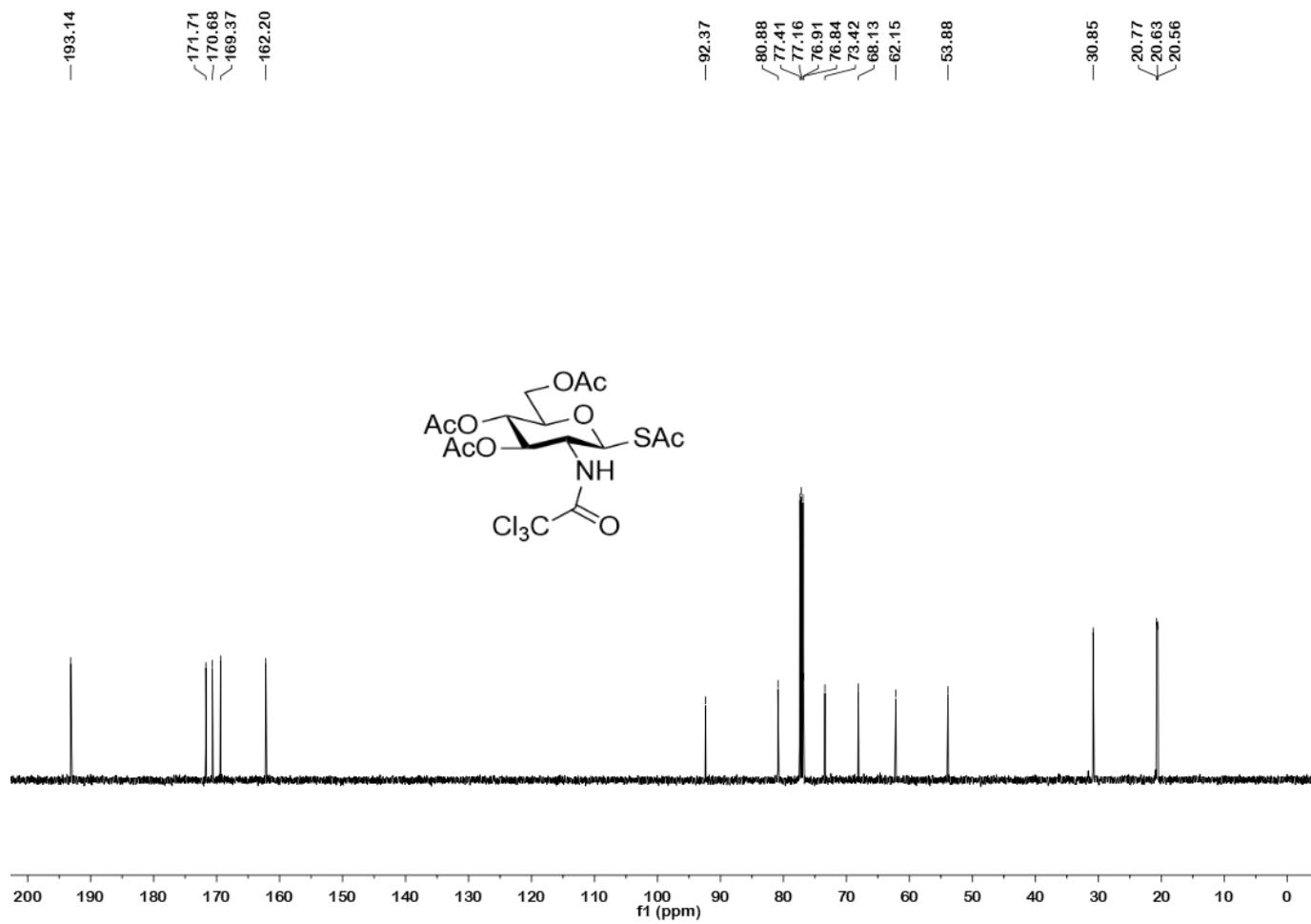


Figure S17. ^{13}C NMR spectrum of compound **4f** in CDCl_3 .

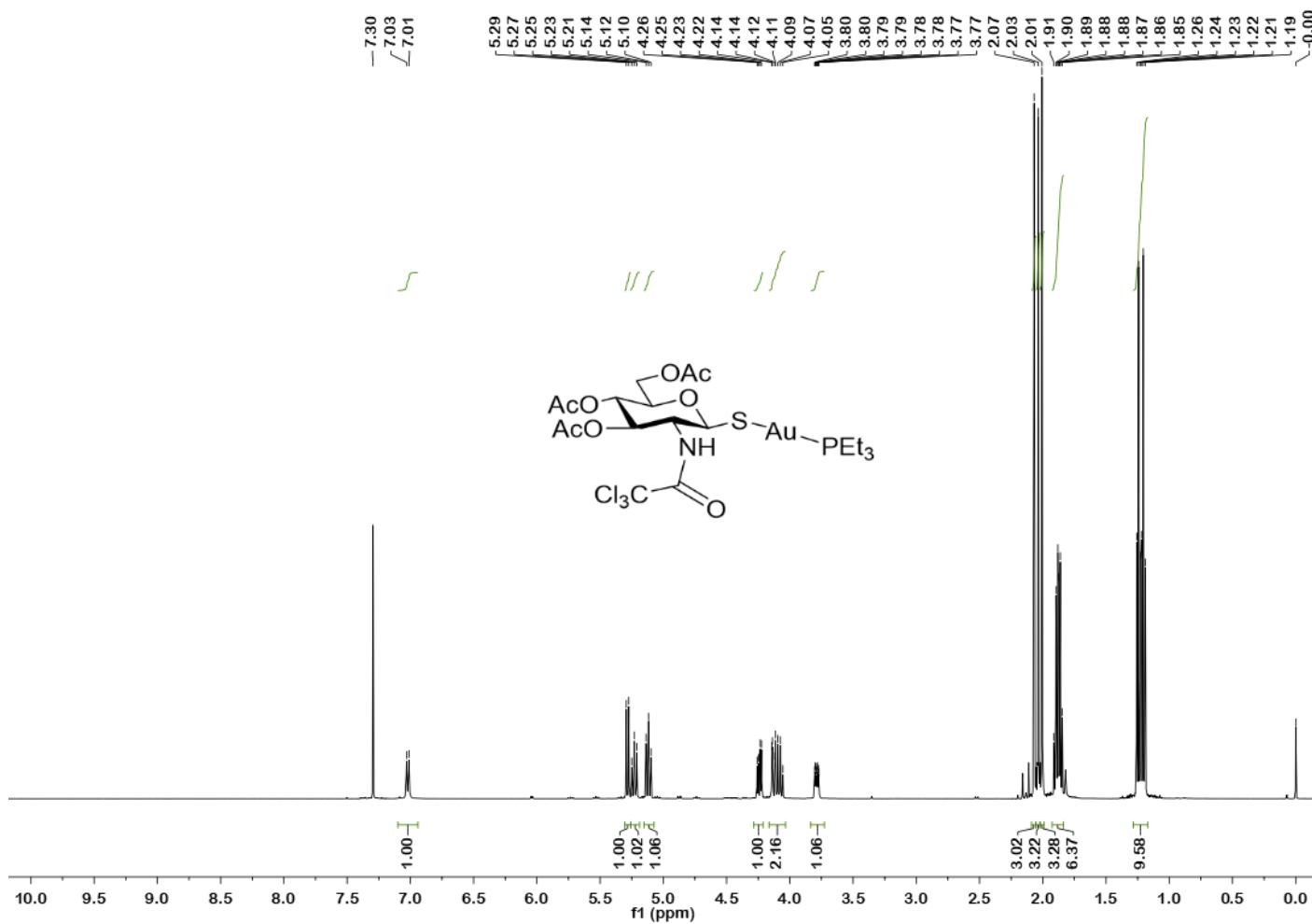


Figure S18. ¹H NMR spectrum of compound 4 in CDCl₃.

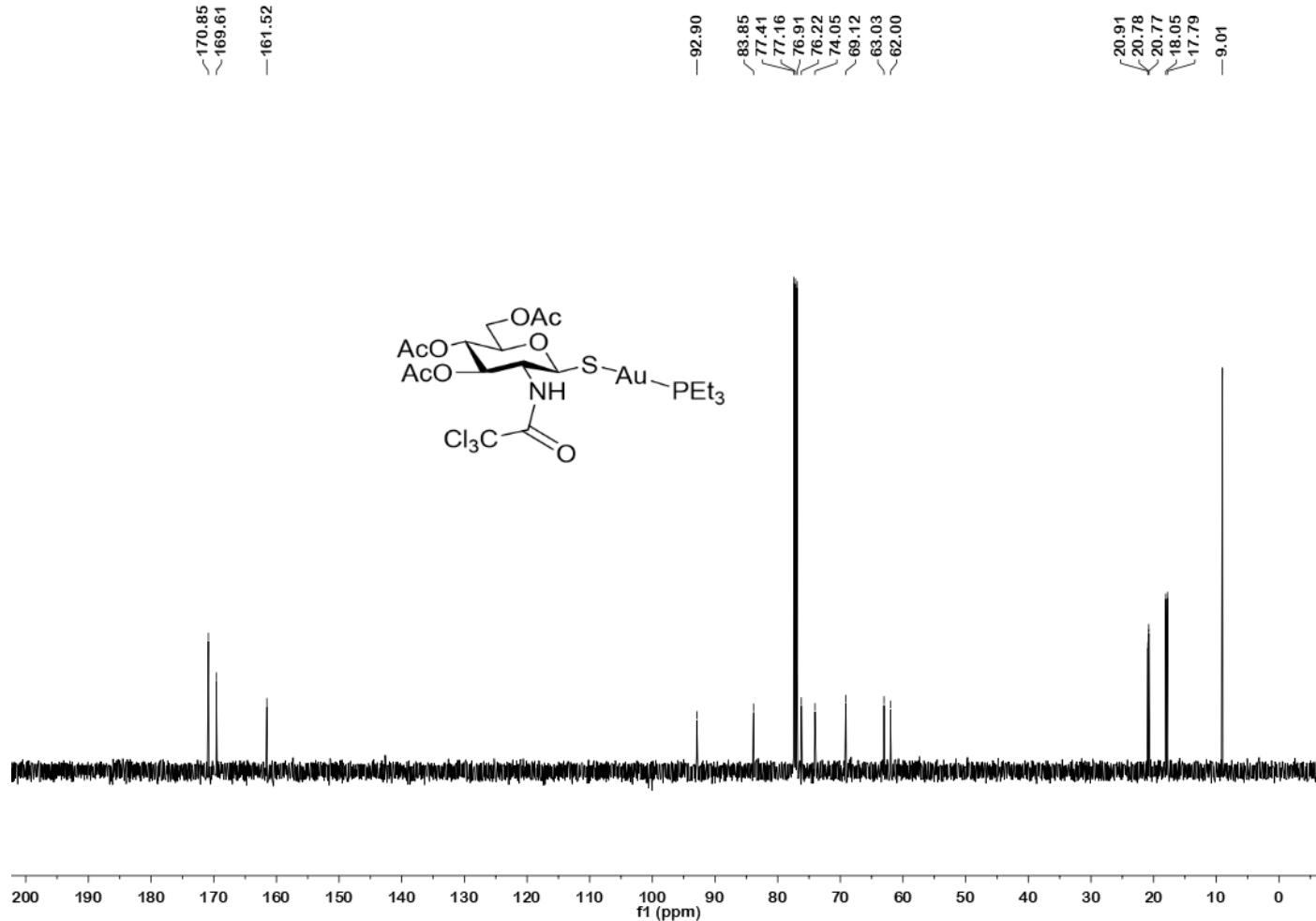


Figure S19. ^{13}C NMR spectrum of compound 4 in CDCl_3 .

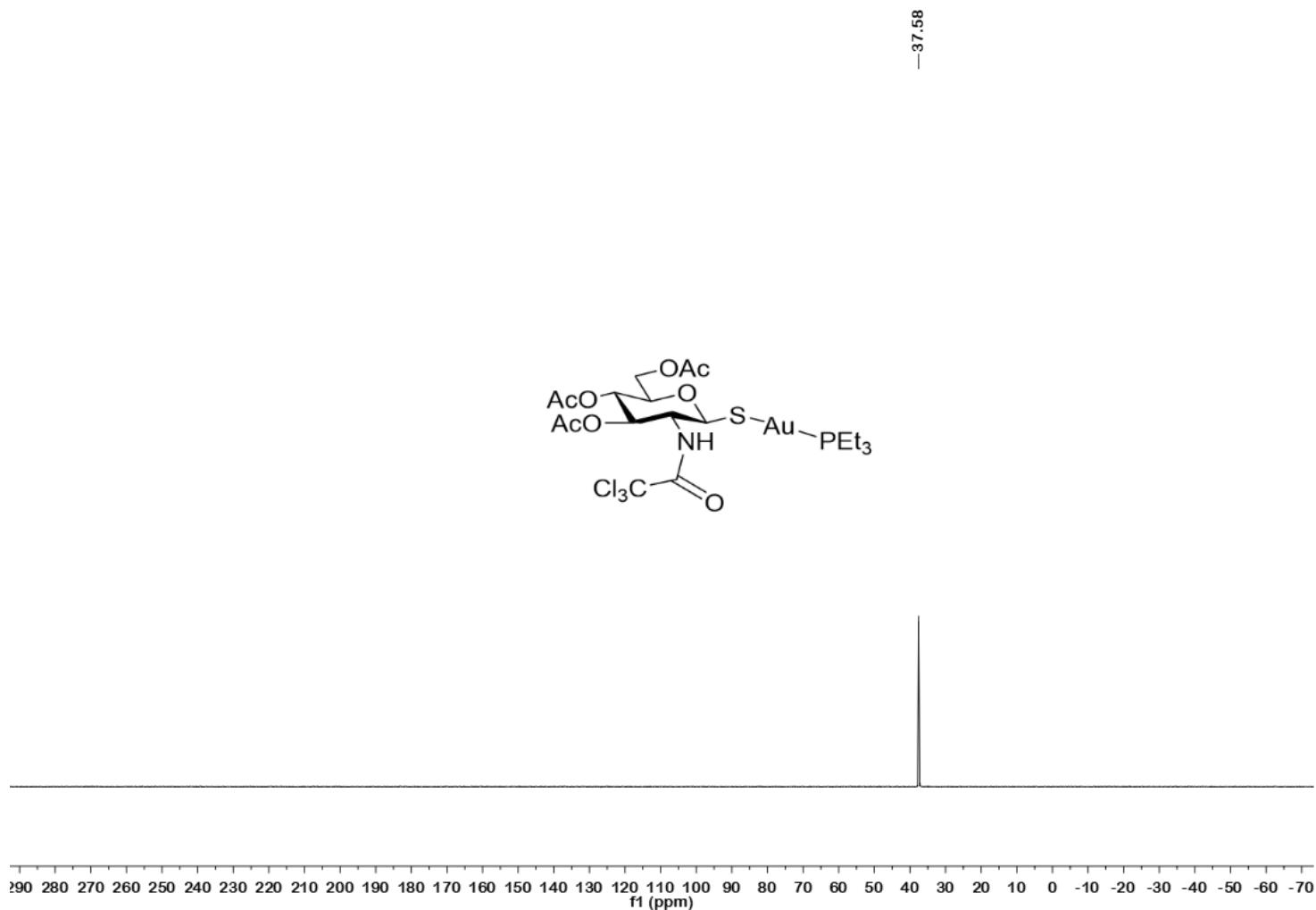


Figure S20. ^{31}P NMR spectrum of compound 4 in CDCl_3 .

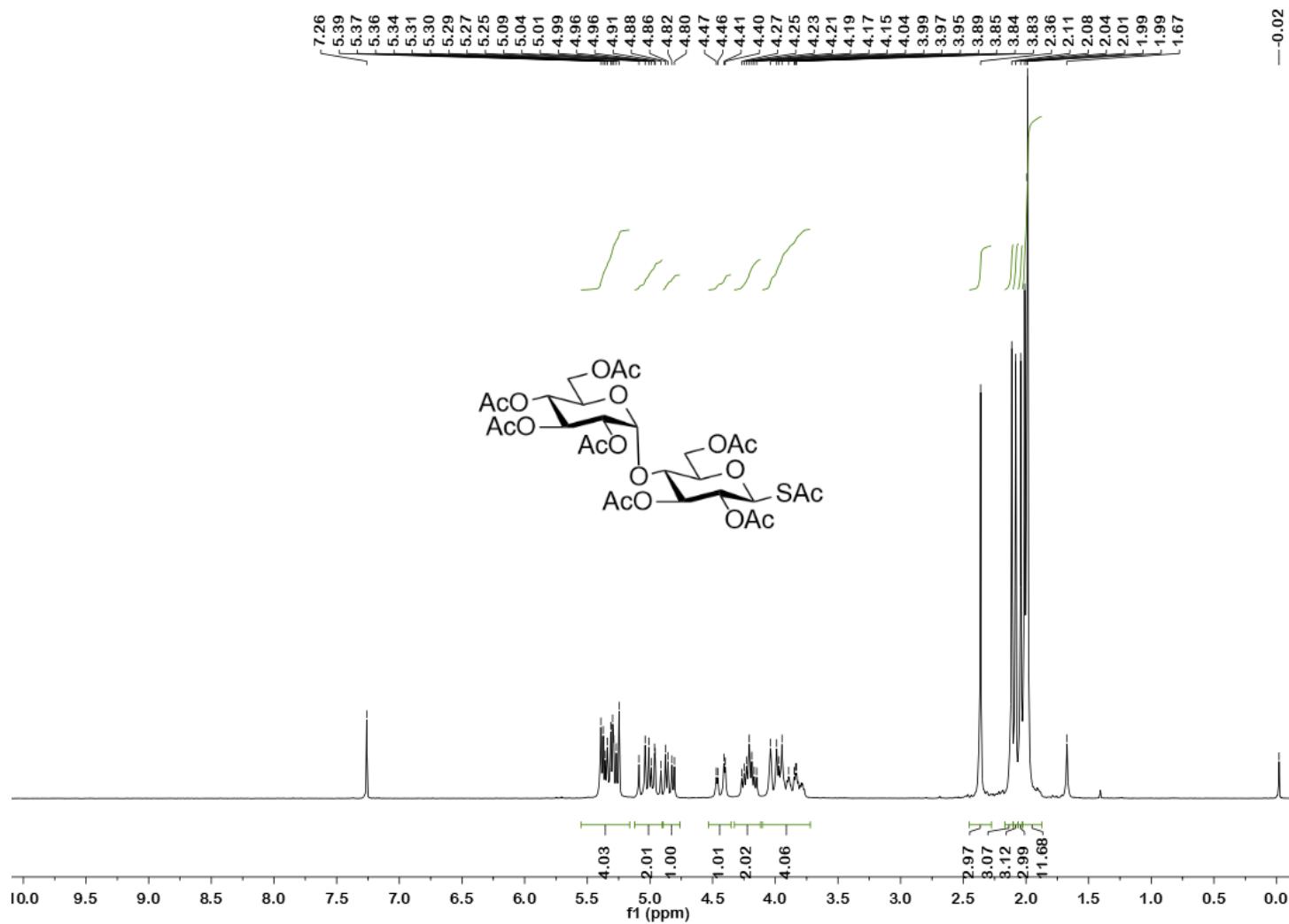


Figure S21. ^1H NMR spectrum of compound **5b** in CDCl_3 .

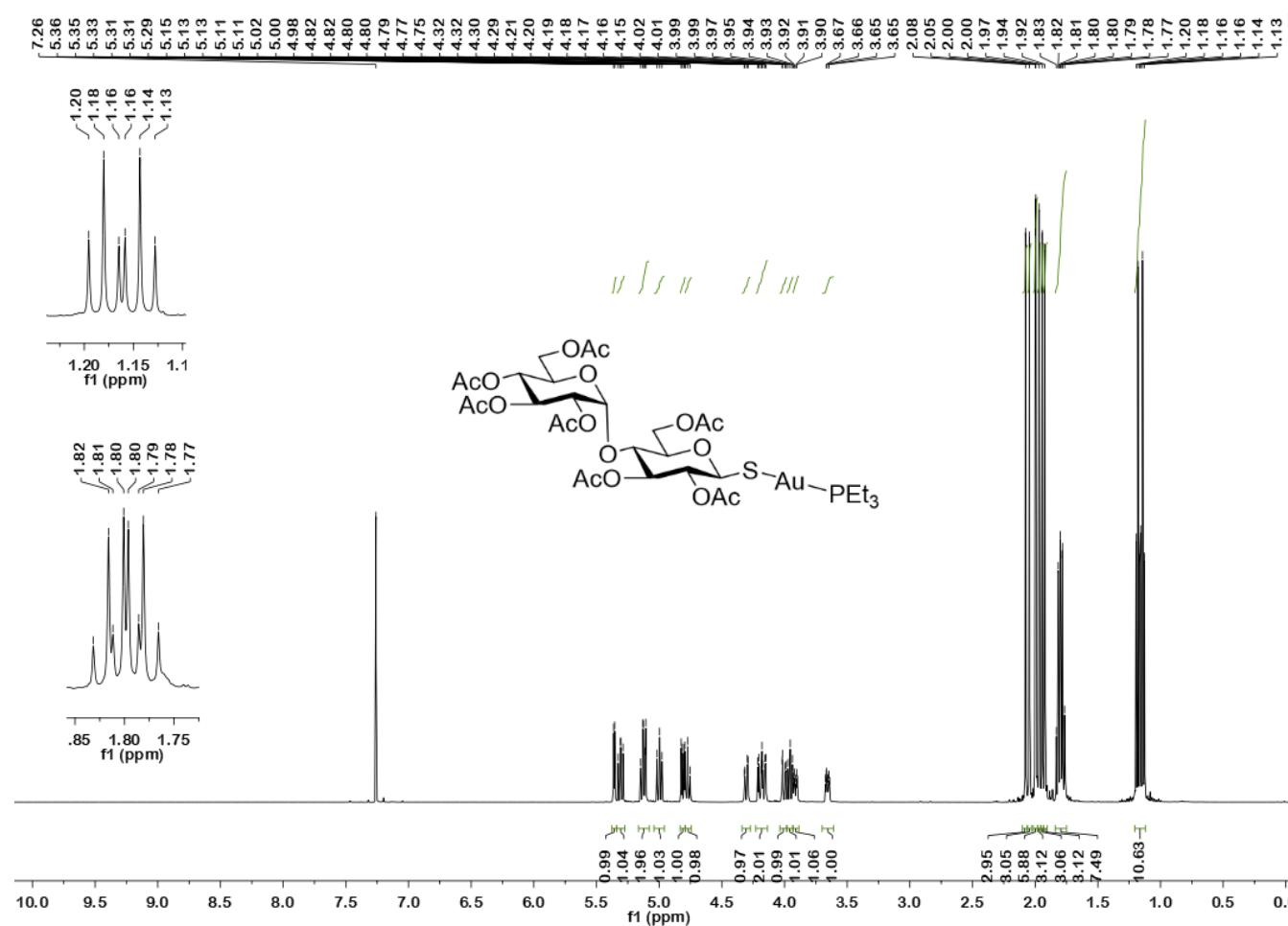


Figure S22. ^1H NMR spectrum of compound **5** in CDCl_3

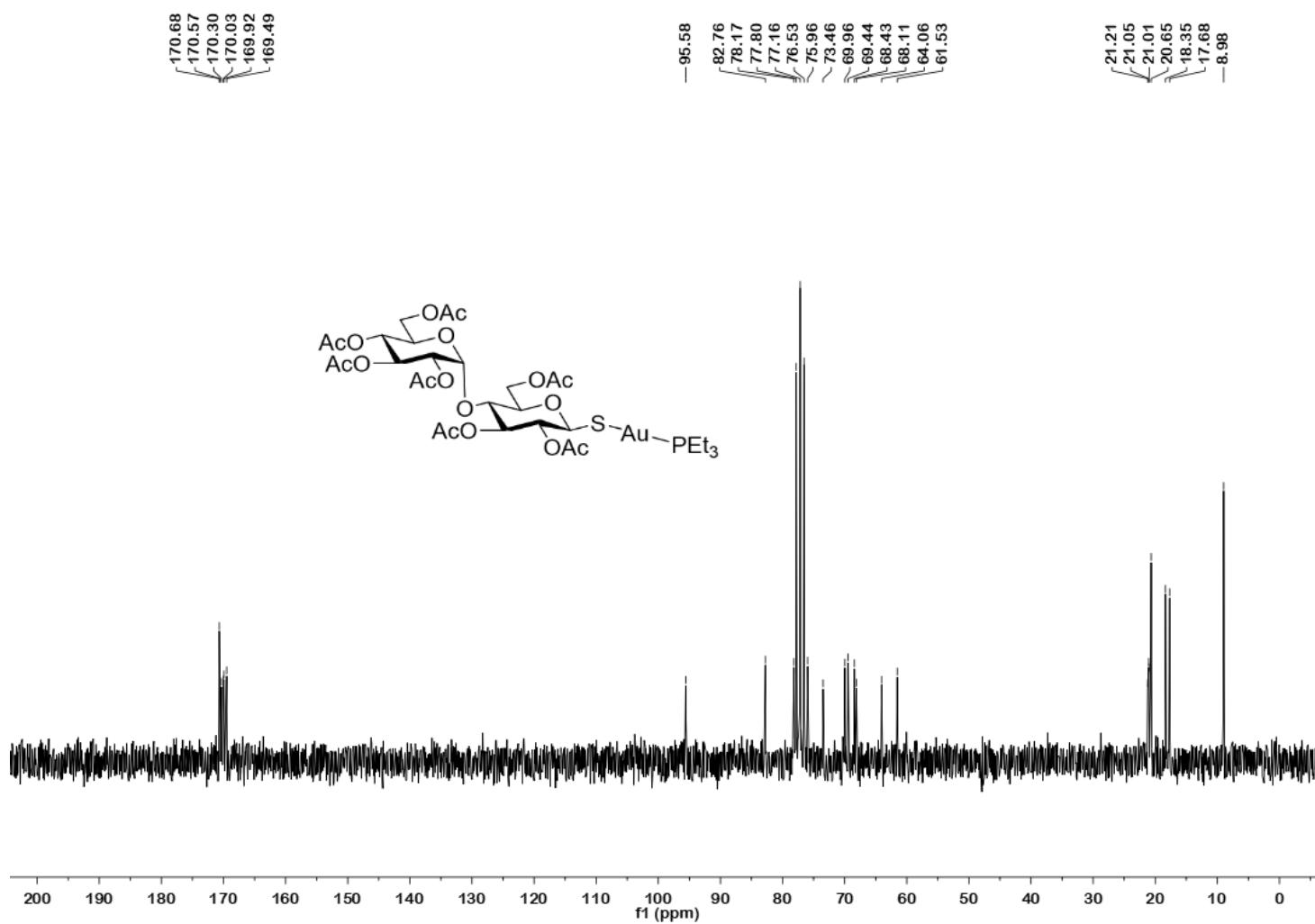


Figure S23. ^{13}C NMR spectrum of compound 5 in CDCl_3

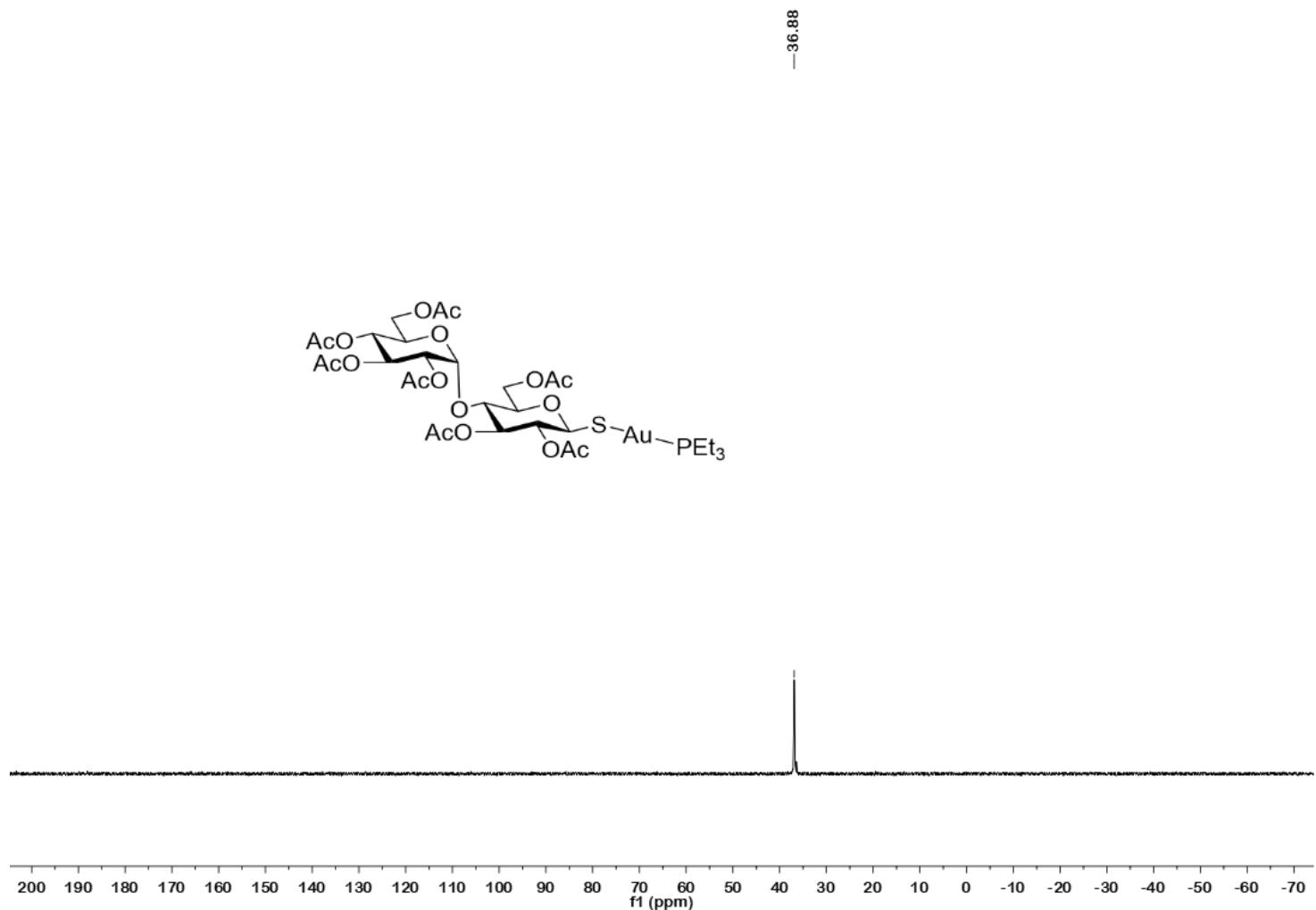


Figure S24. ^{31}P NMR spectrum of compound 5 in CDCl_3

Madanodaya Sundhor WB-07

Synapt_10292 28 (0.570) Cm (28-16)

100

50

0

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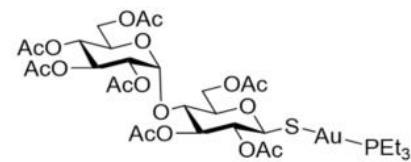
SYNAPTG2-Si#UGA305

13:50:21

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1281.2729



967.2233

356.0829

315.0565

514.2697

357.0857

619.1849

559.1669

675.1563

707.0912

907.1987

1029.1437

1031.1433

1032.1465

1033.1482

1282.2767

1283.2758

1284.2762

1215.2631

1370.2028

100 200 300 400 500 600 700 800 900 1000 1100 1200 1300 1400 1500 m/z

Figure S25. HRMS of compound 5.

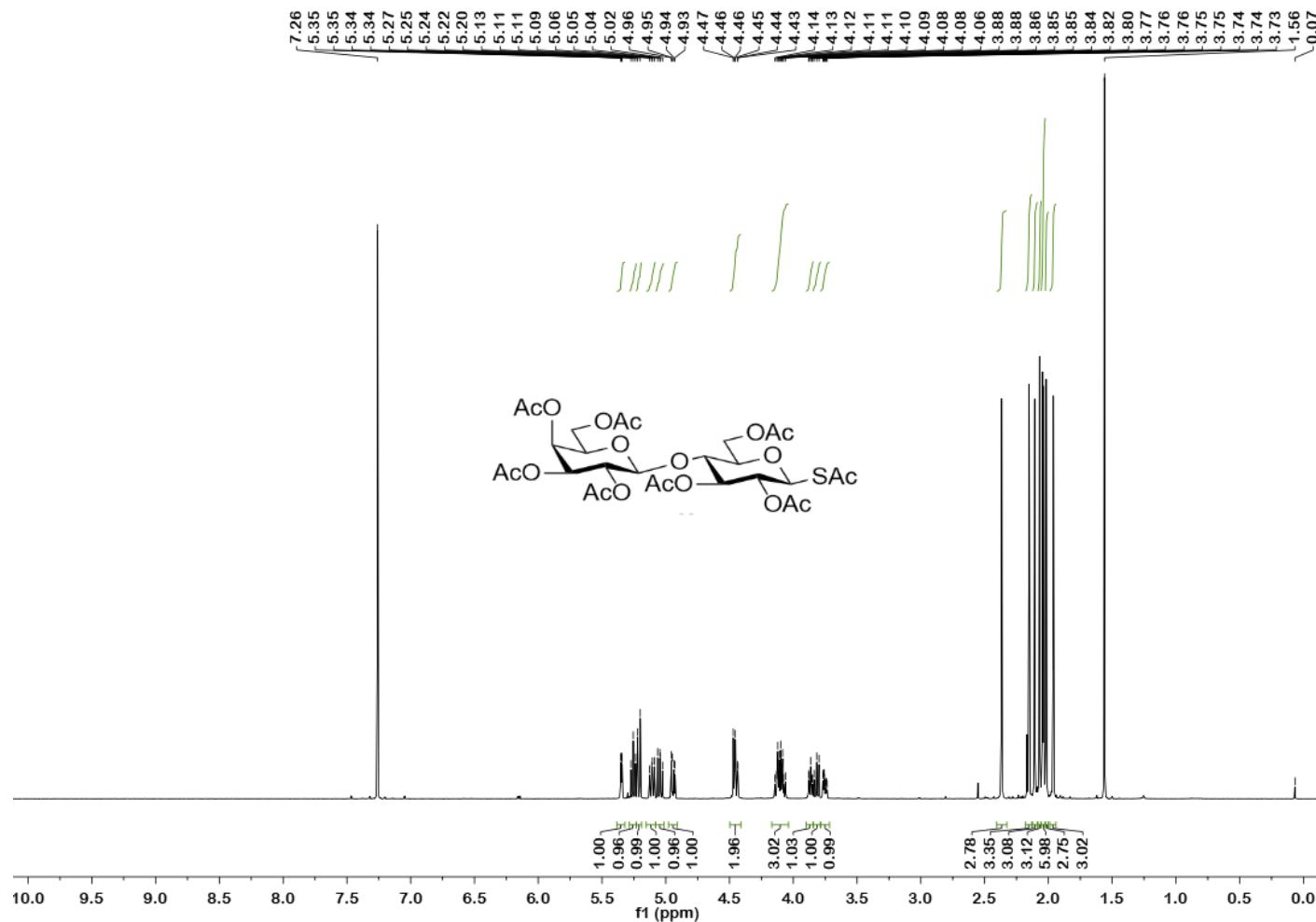


Figure S26. ^1H NMR spectrum of compound **6b** in CDCl_3 .

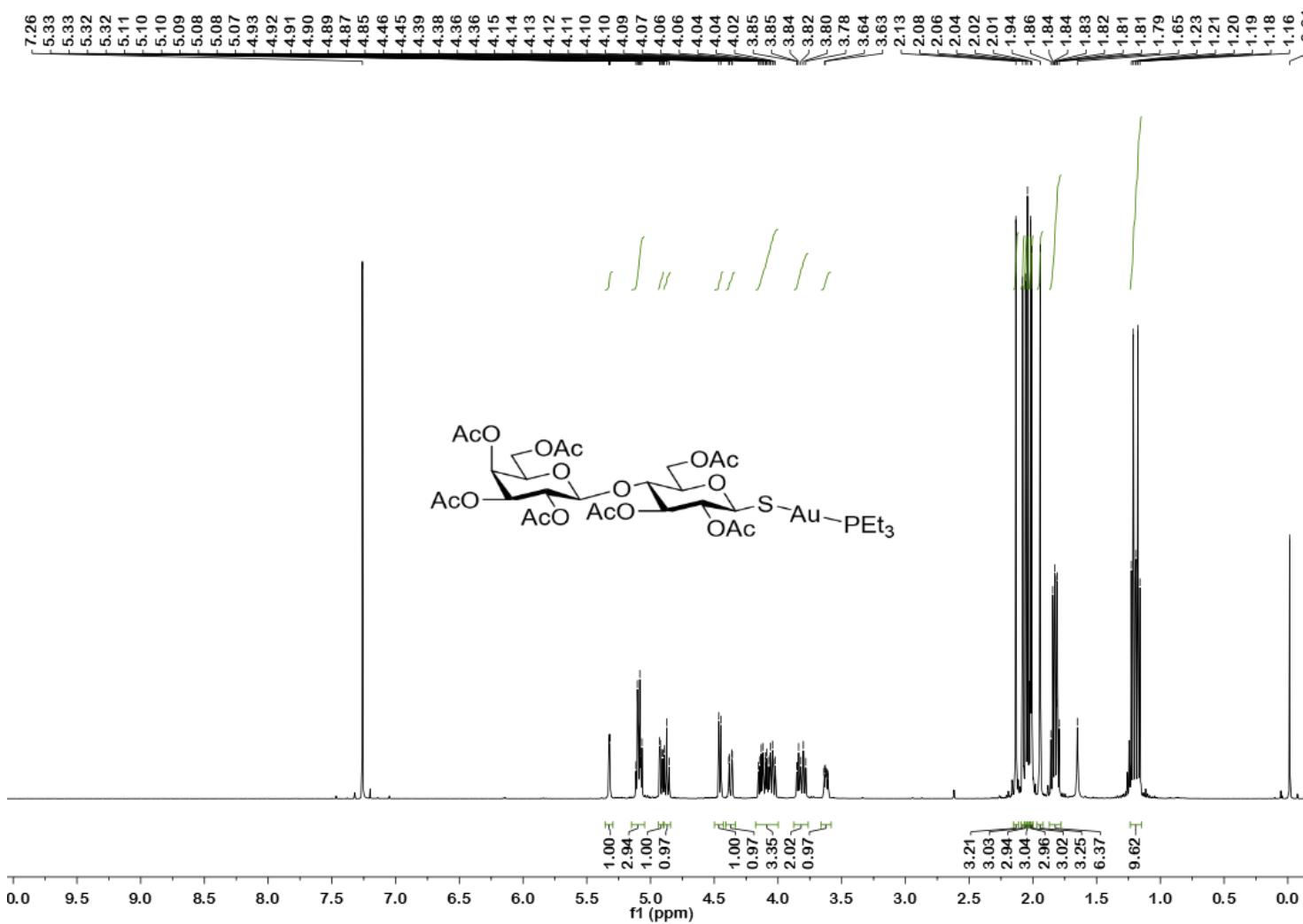


Figure S27. ¹H NMR spectrum of compound 6 in CDCl₃.

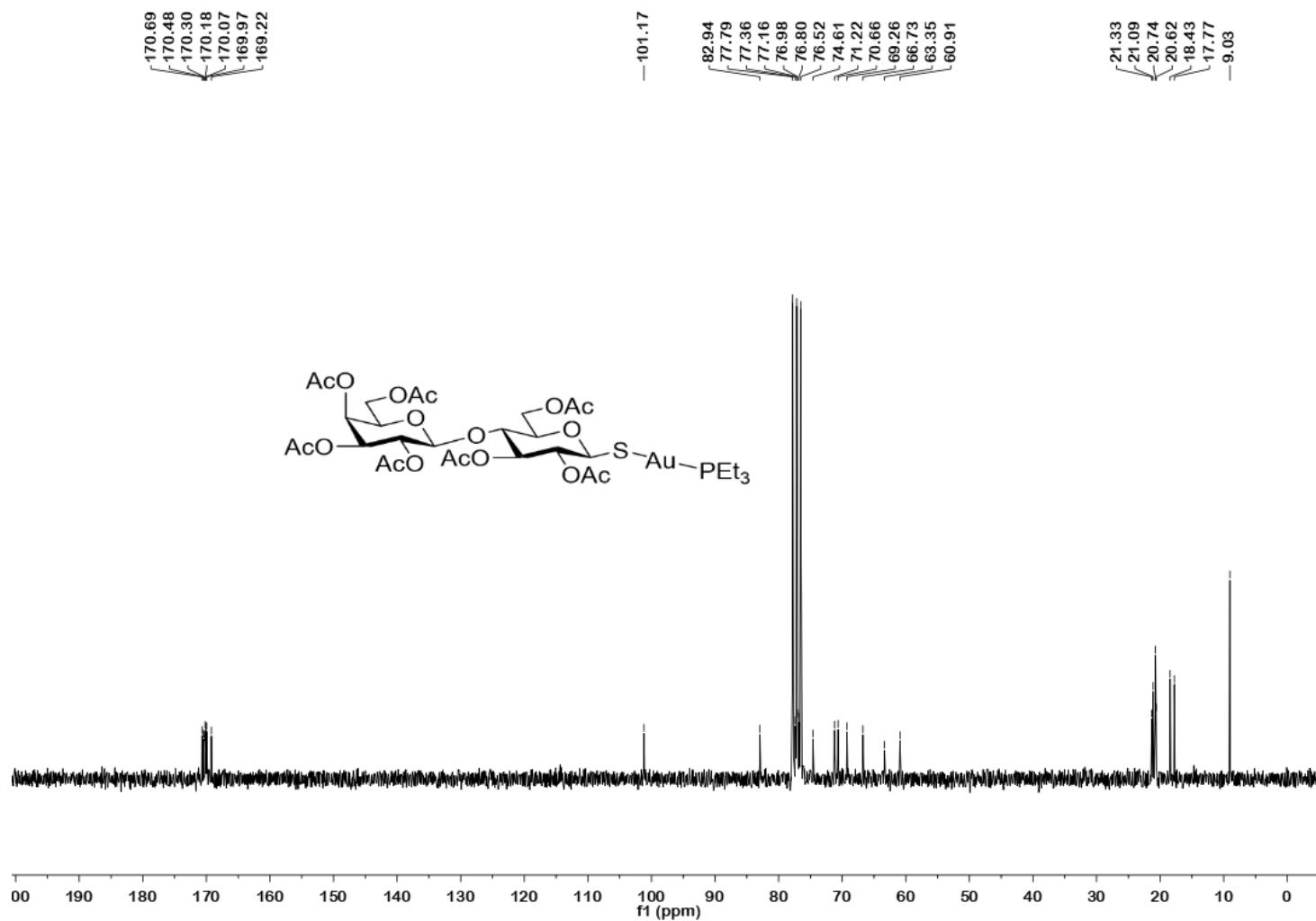


Figure S28. ^{13}C NMR spectrum of compound **6** in CDCl_3 .

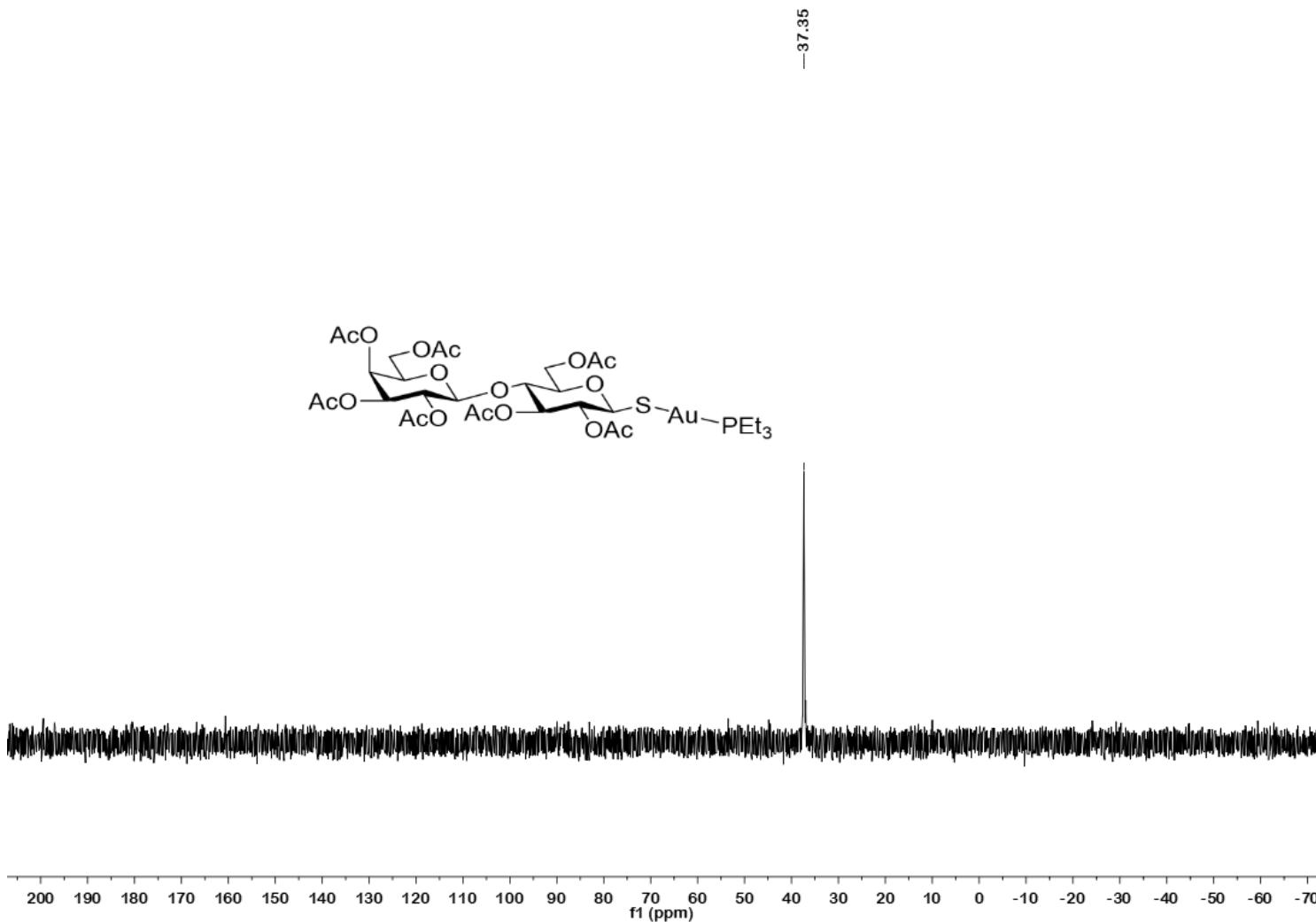


Figure S29. ^{31}P NMR spectrum of compound **6** in CDCl_3 .

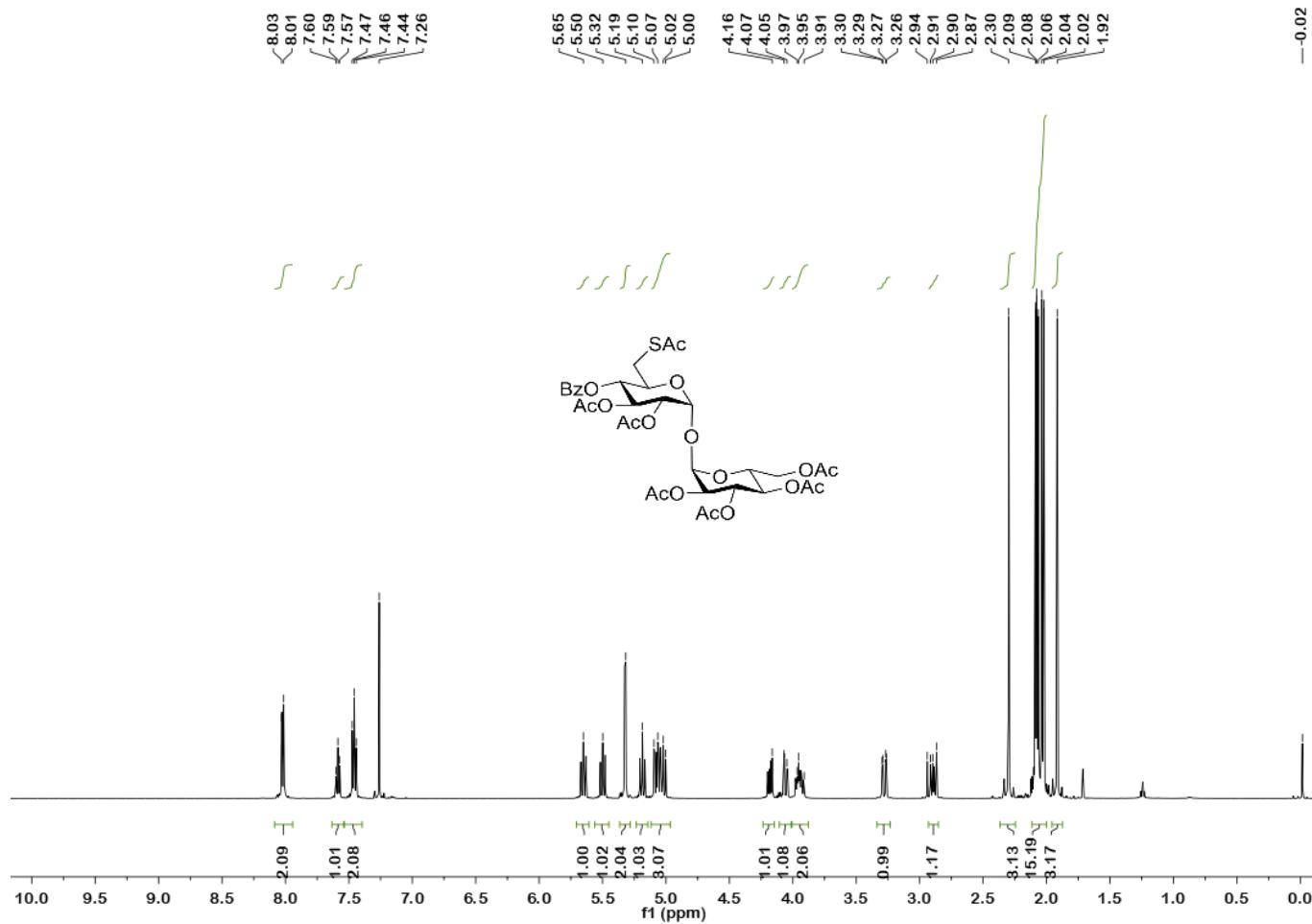


Figure S30. ^1H NMR spectrum of compound **7d** in CDCl_3 .

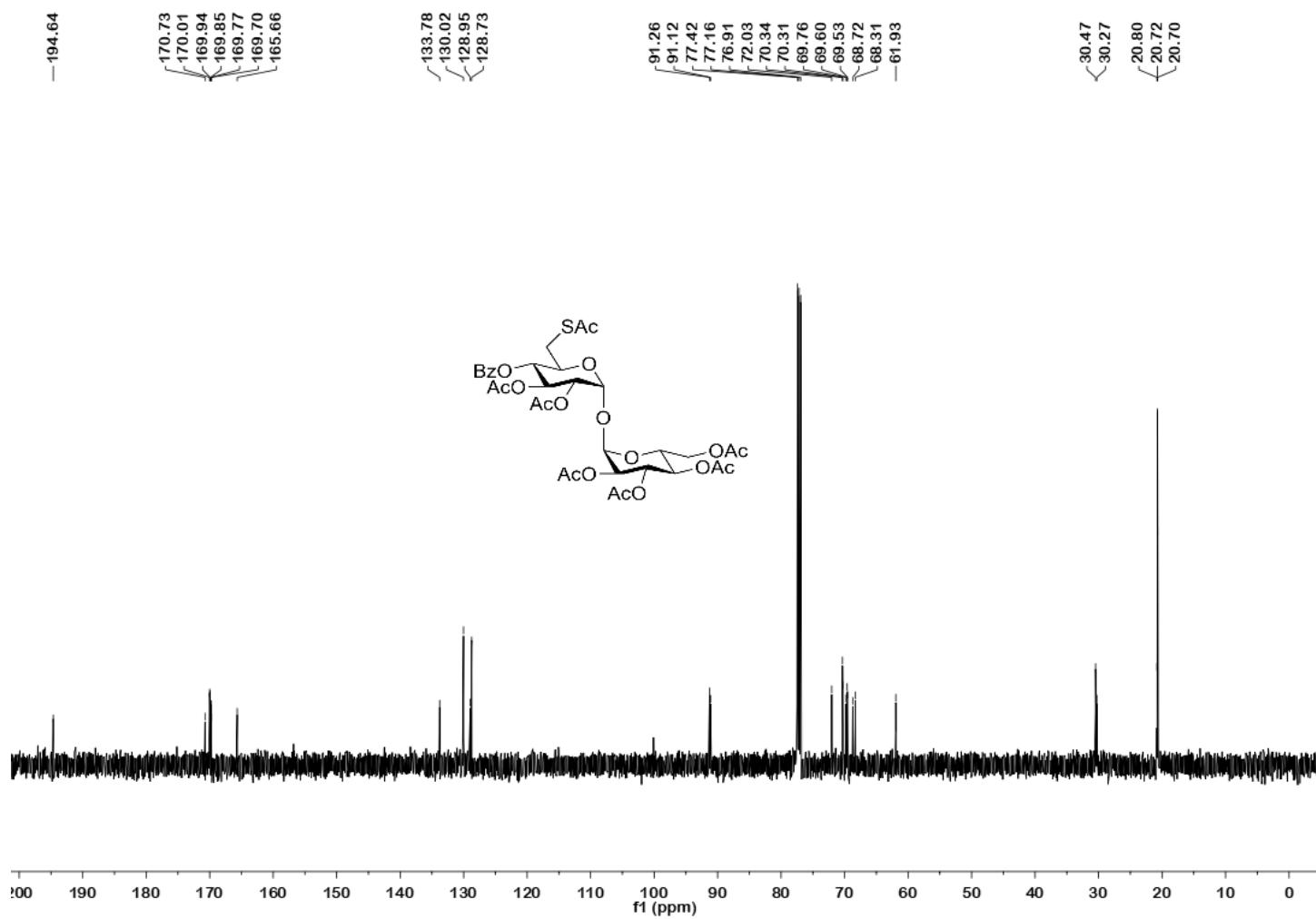


Figure S31. ^{13}C NMR spectrum of compound **7d** in CDCl_3 .

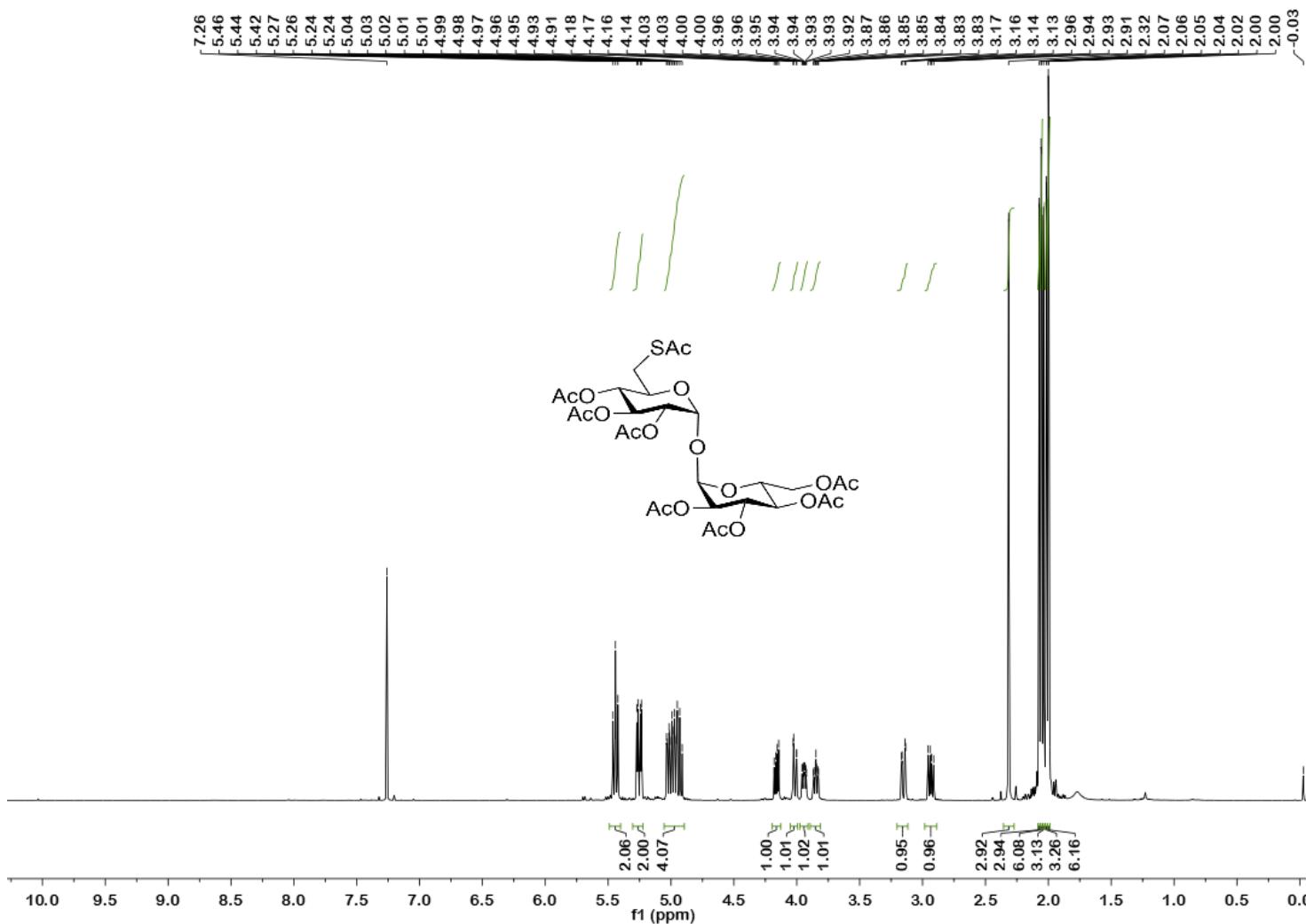


Figure S32. ^1H NMR spectrum of compound **7e** in CDCl_3 .

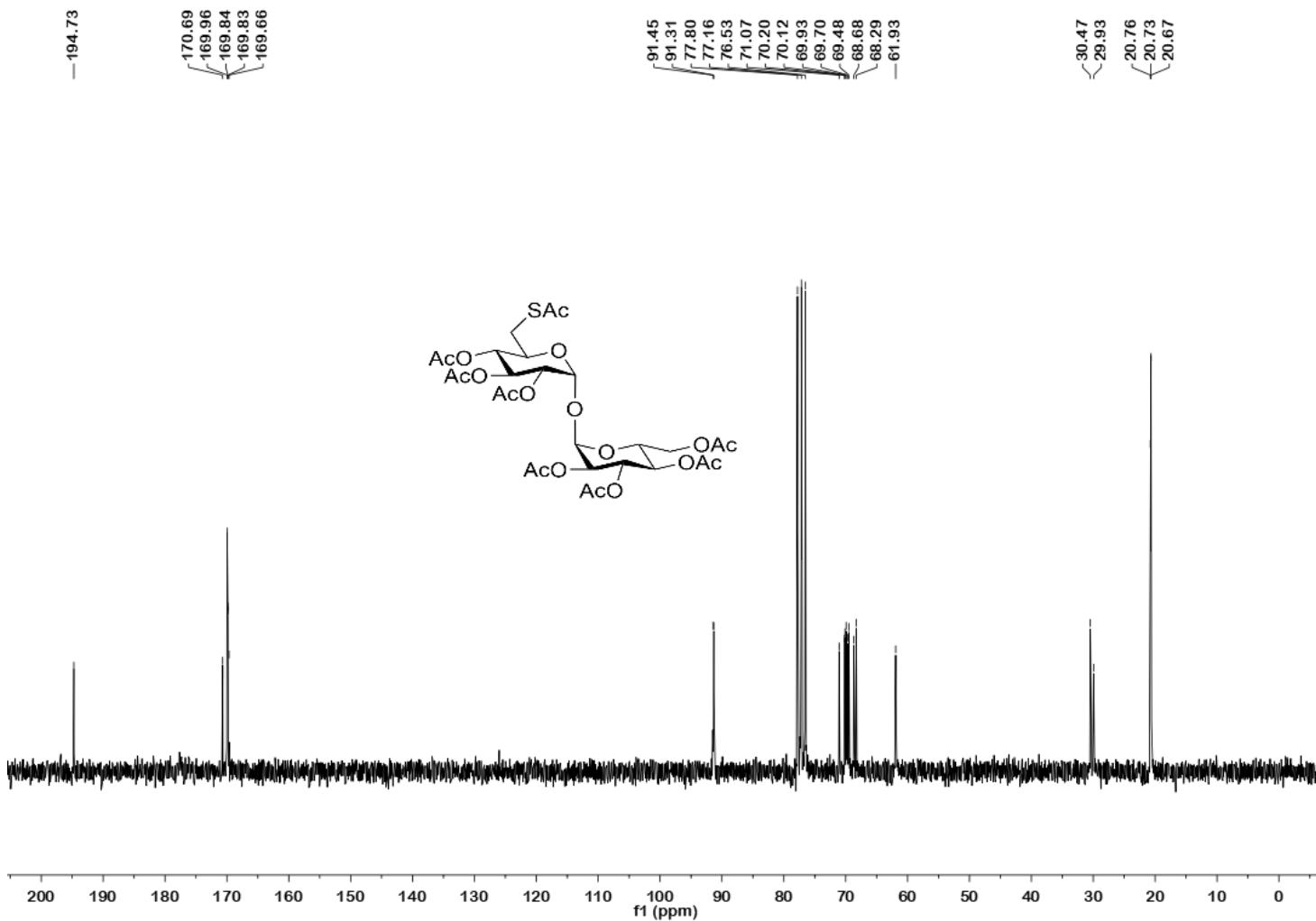


Figure S33. ^{13}C NMR spectrum of compound **7e** in CDCl_3 .

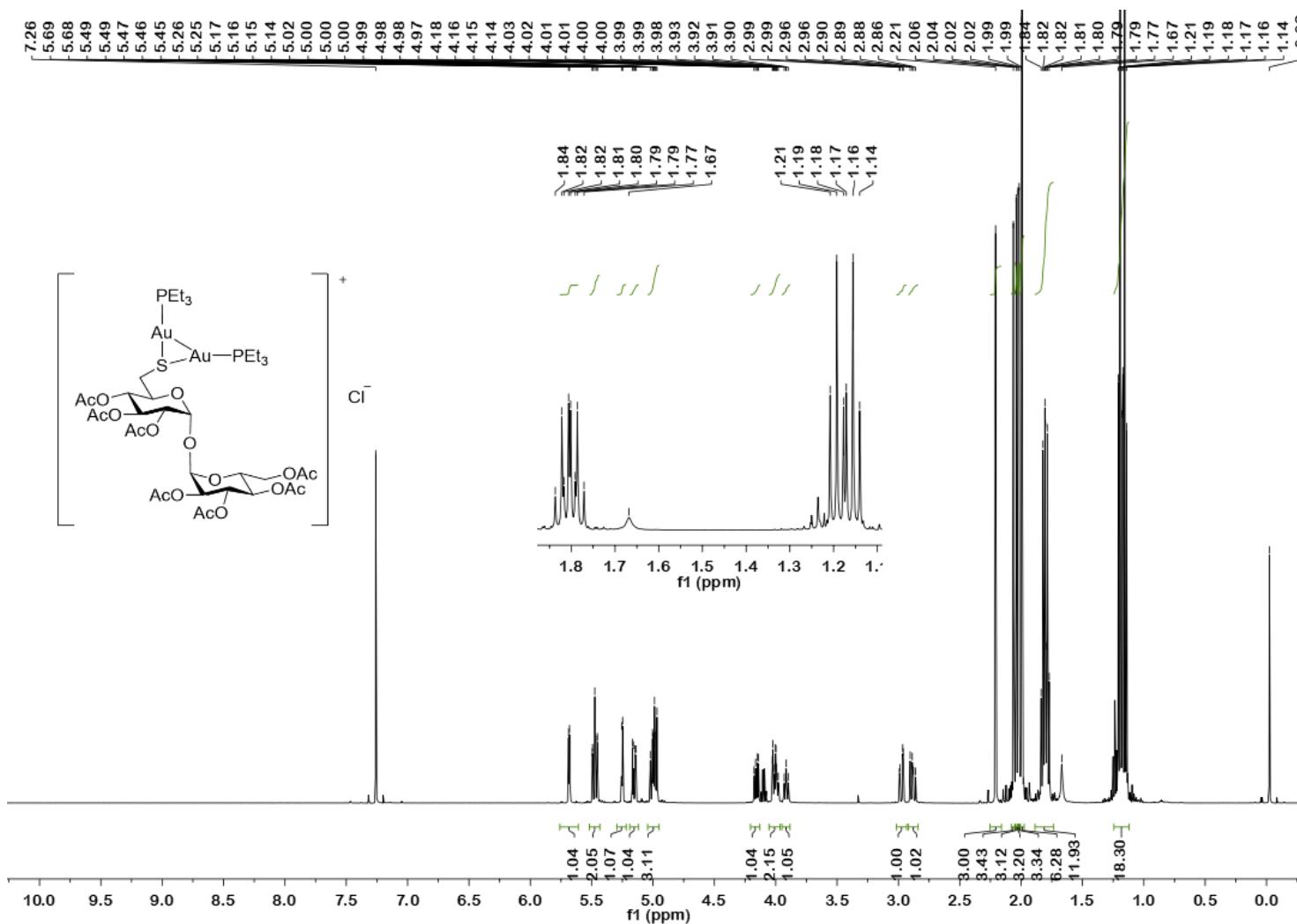


Figure S34. ^1H NMR spectrum of compound 7 in CDCl_3 .

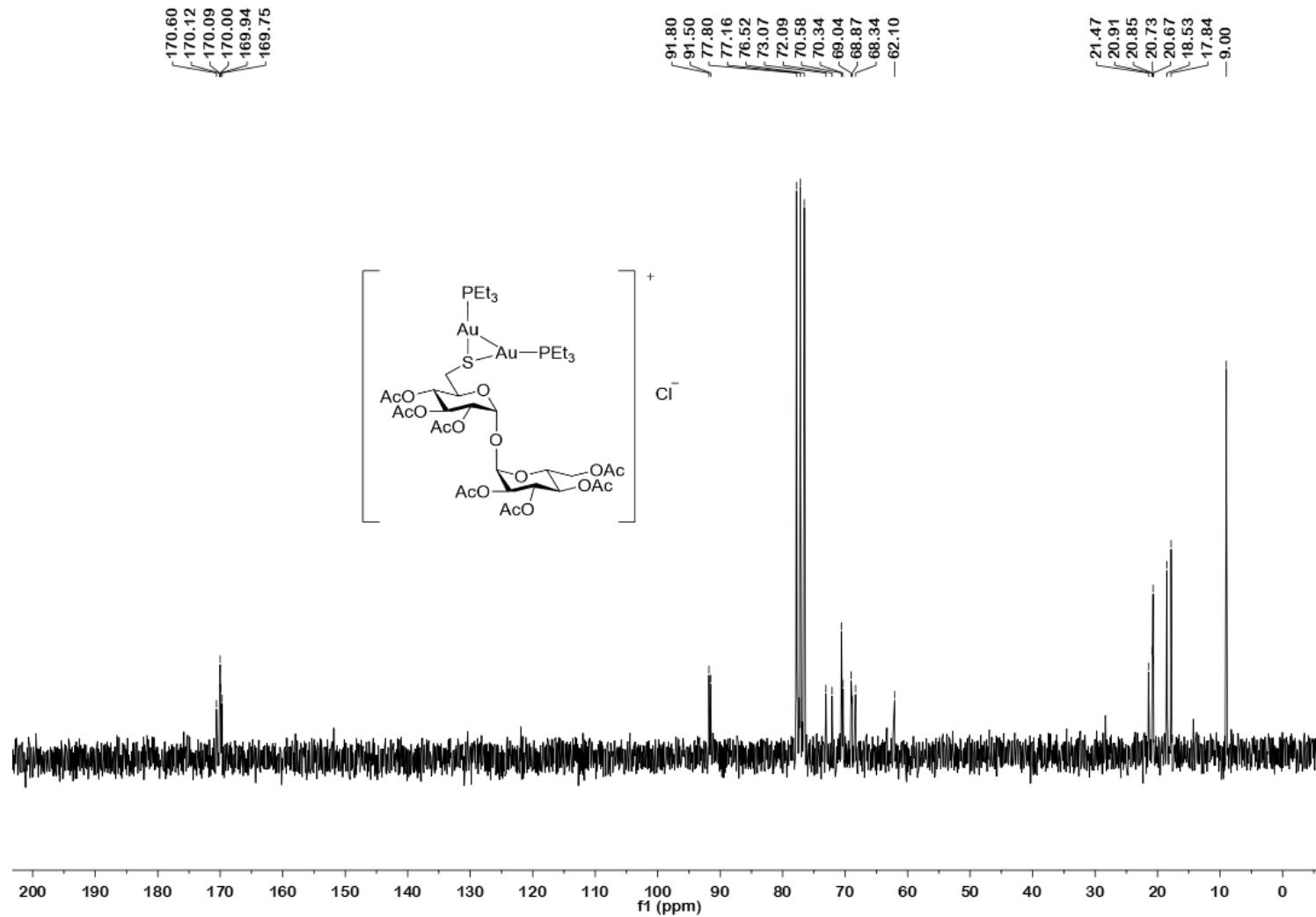


Figure S35. ^{13}C NMR spectrum of compound 7 in CDCl_3 .

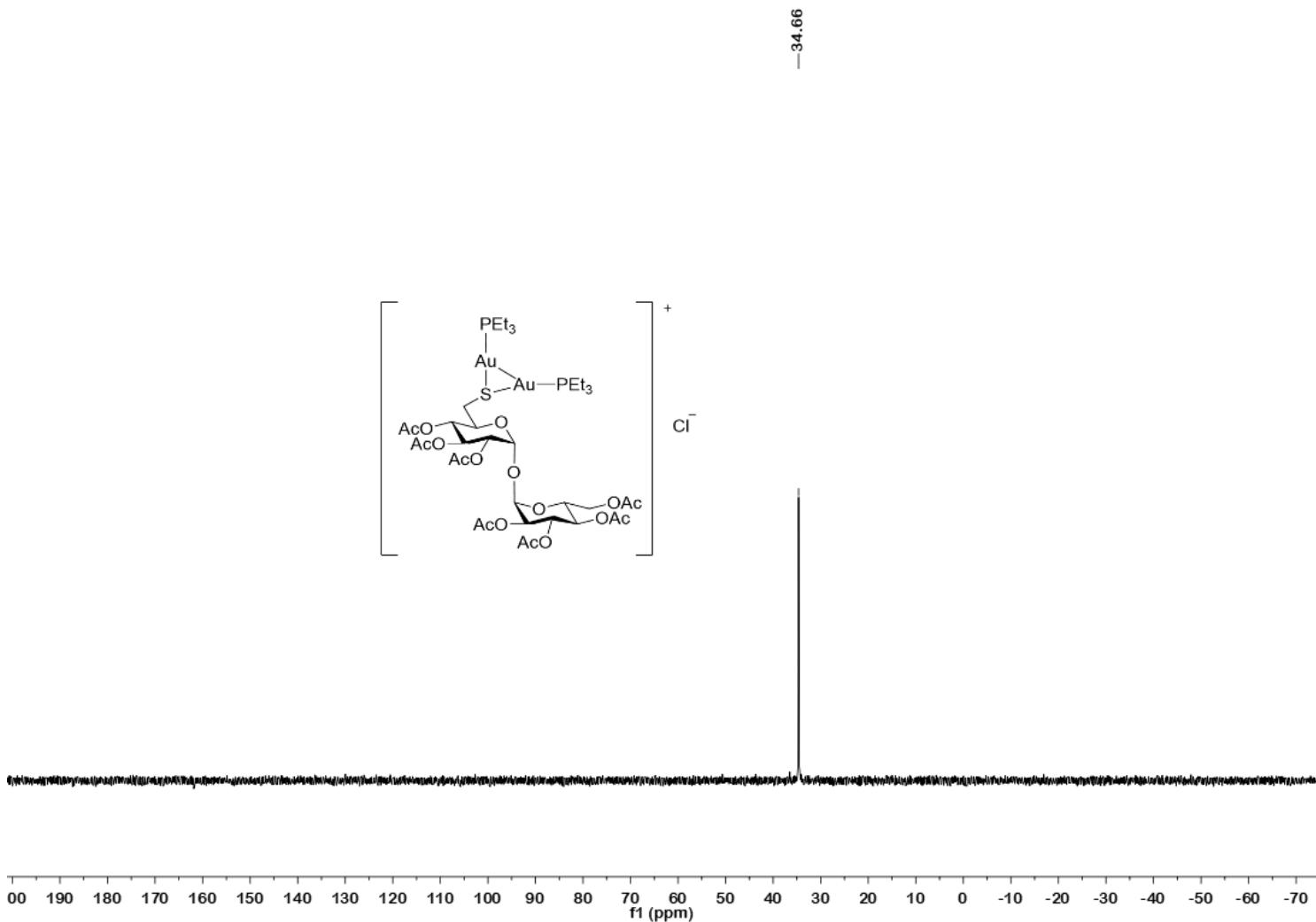


Figure S36. ^{31}P NMR spectrum of compound 7 in CDCl_3 .

Madanodaya Sundhor WB-09

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SYNAPTG2-Si#UGA305

13:54:23

1: TOF MS ES+
1.65e6

Synapt_10294 23 (0.465) Cm (22:24-7:10)

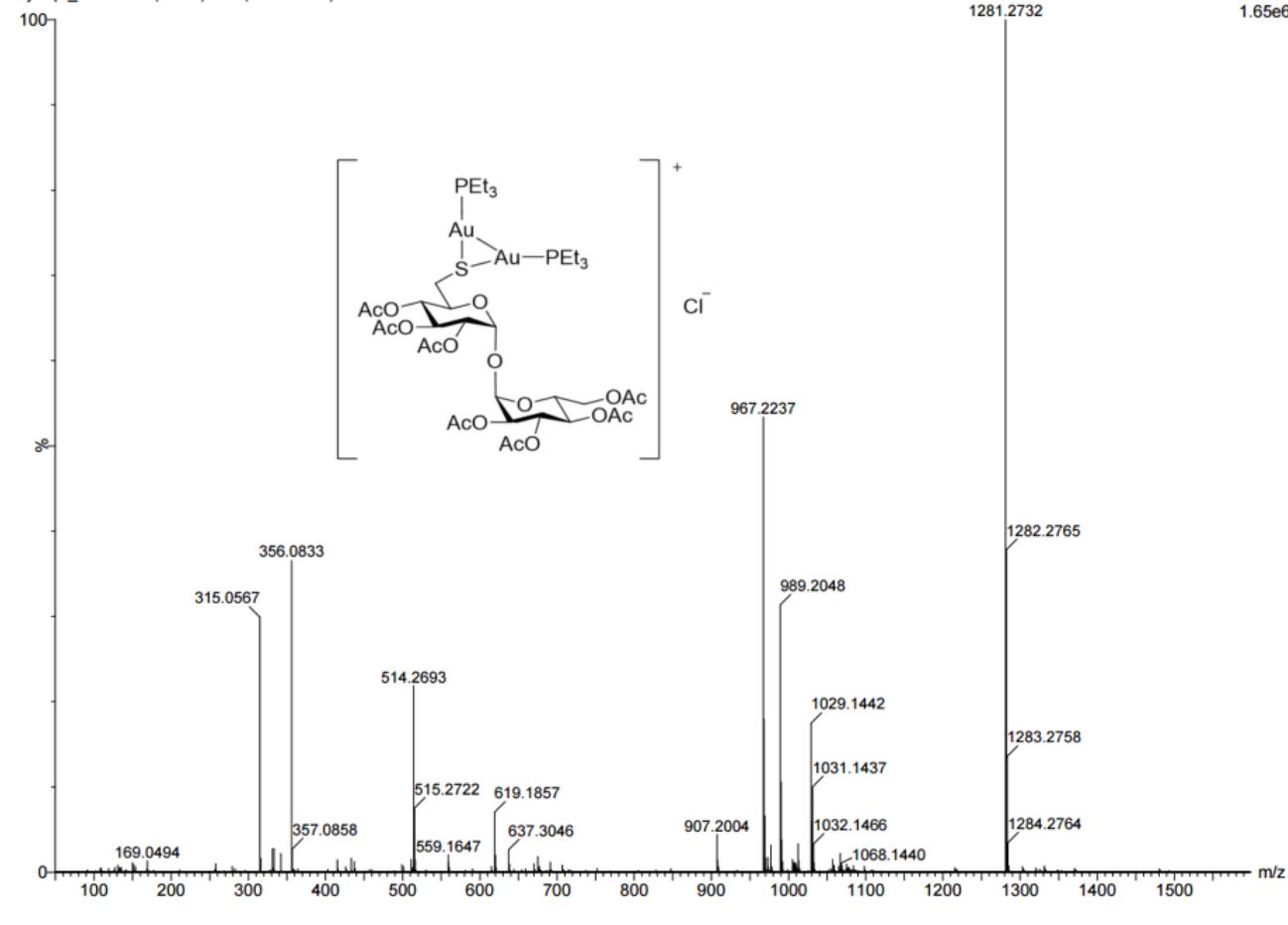


Figure S37. HRMS of compound 7.

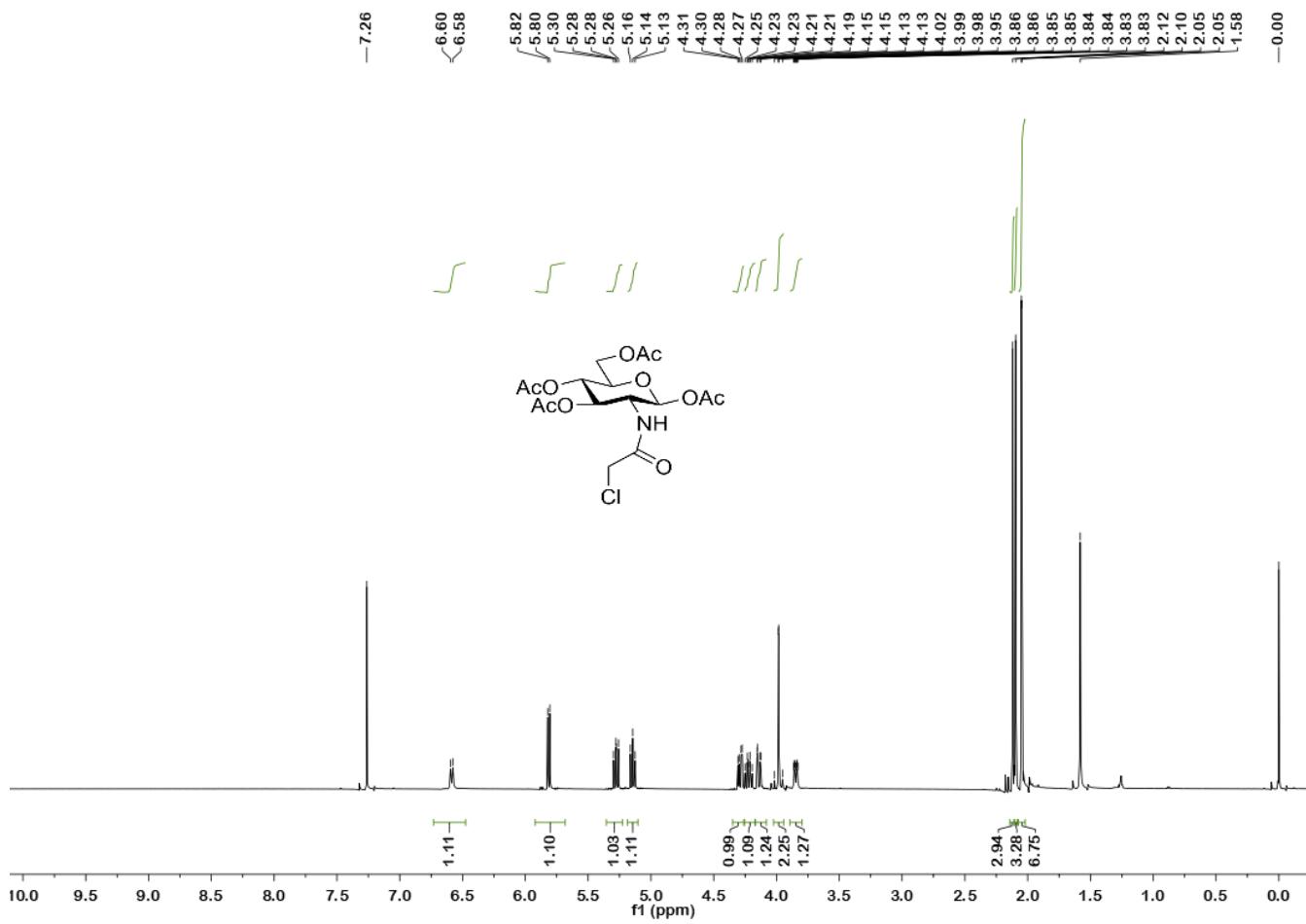


Figure S38. ^1H NMR spectrum of compound **8e** in CDCl_3

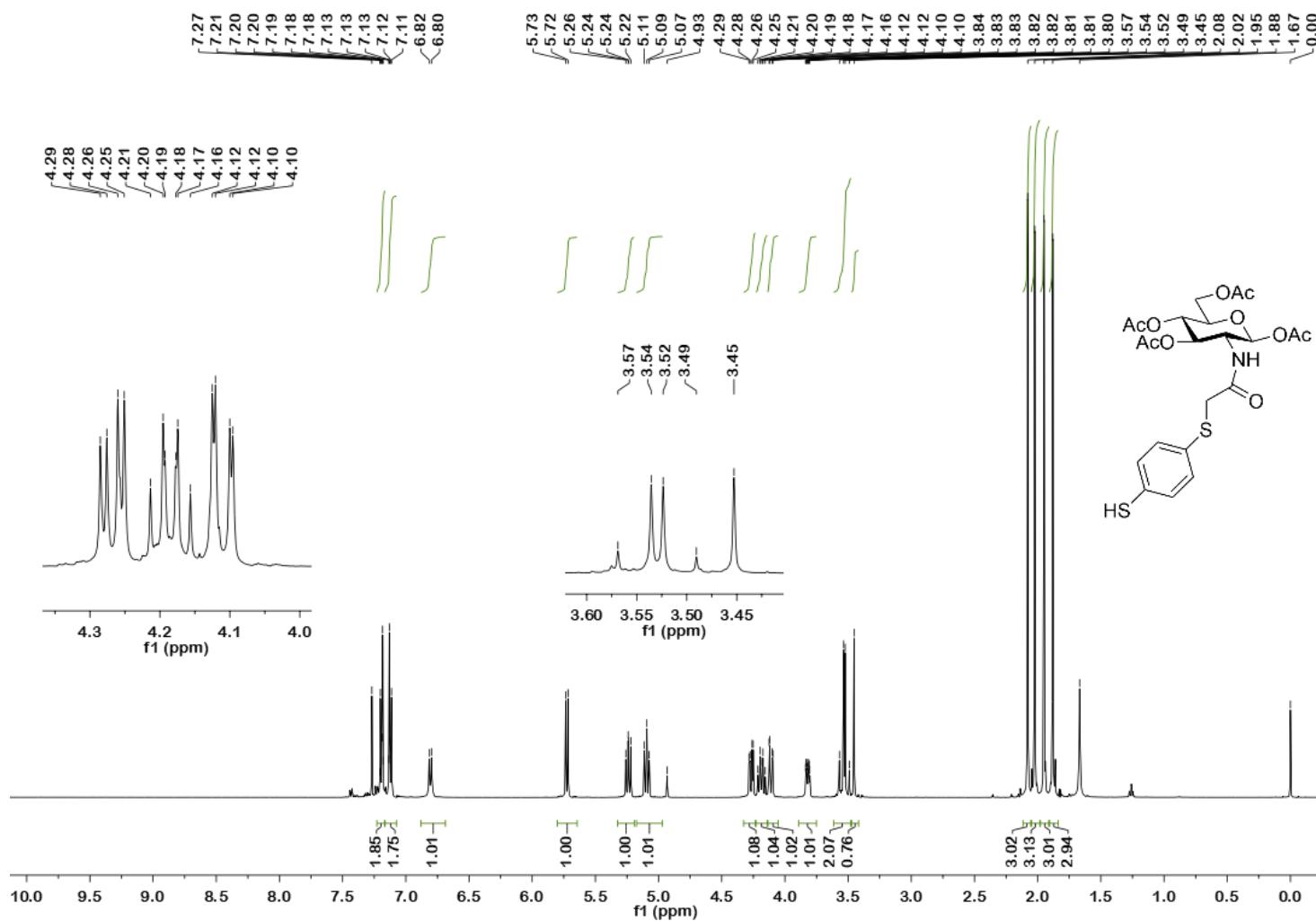


Figure S39. ^1H NMR spectrum of compound **8f** in CDCl_3

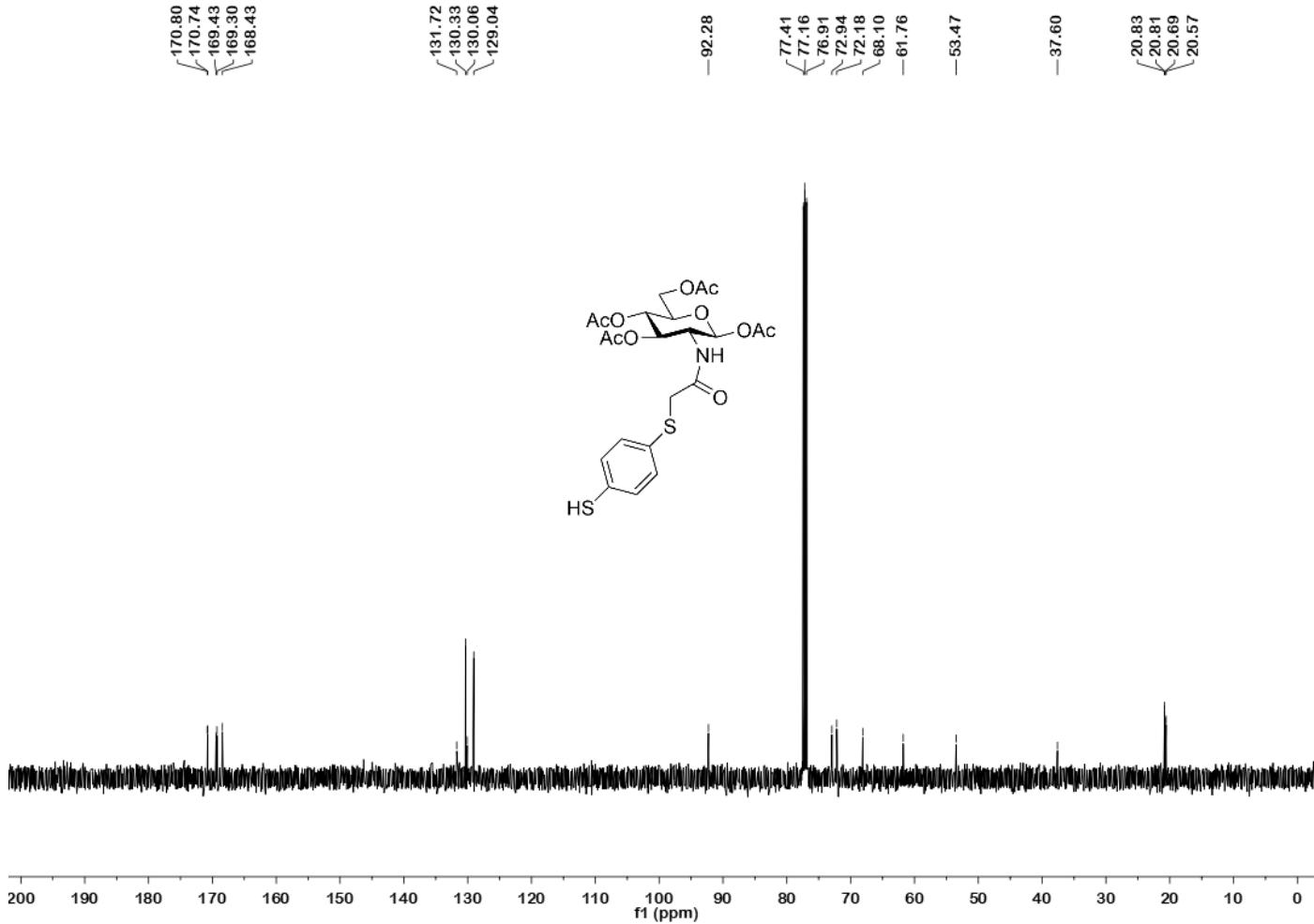


Figure S40. ^{13}C NMR spectrum of compound **8f** in CDCl_3

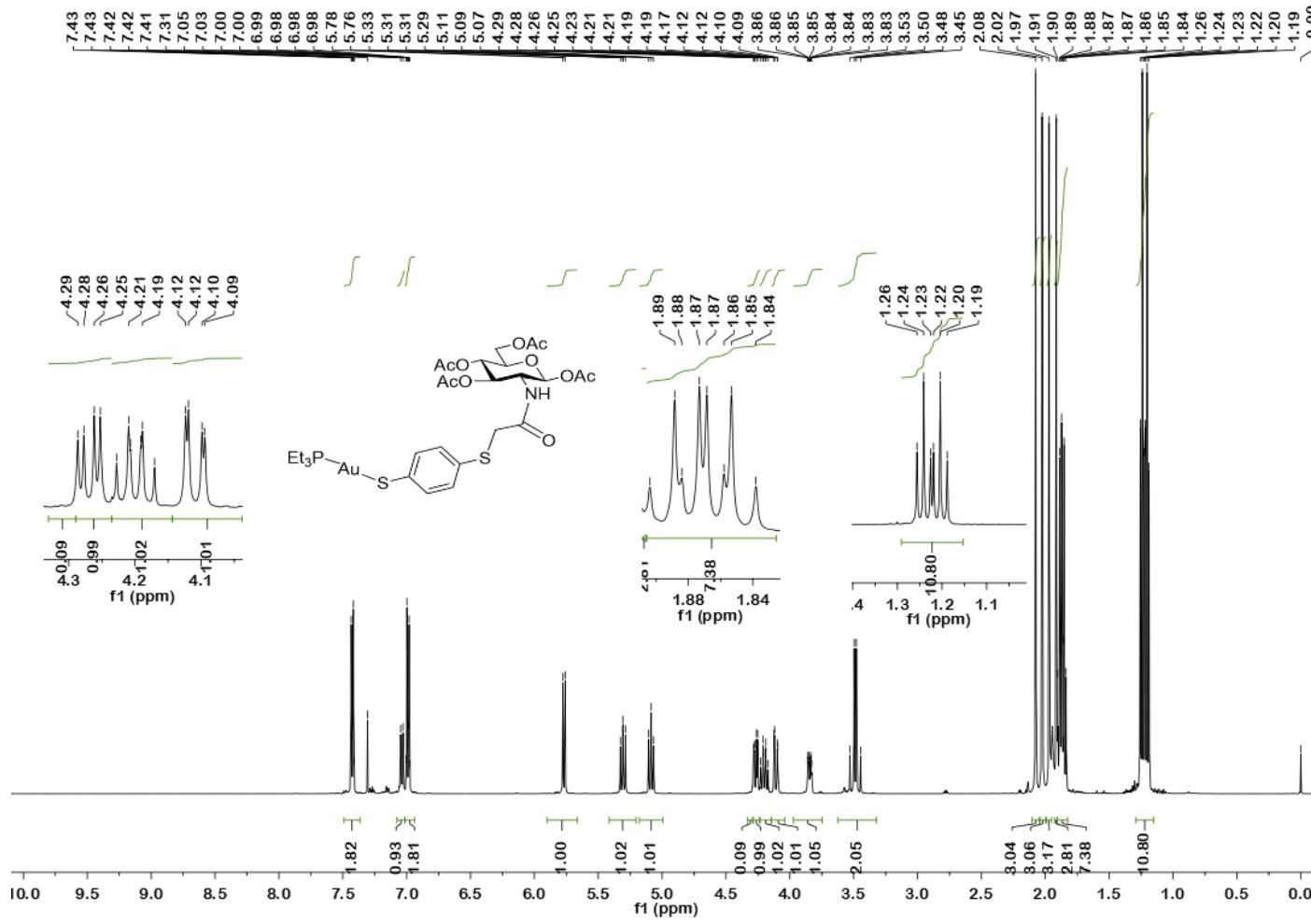


Figure S41. ^1H NMR spectrum of compound **8** in CDCl_3

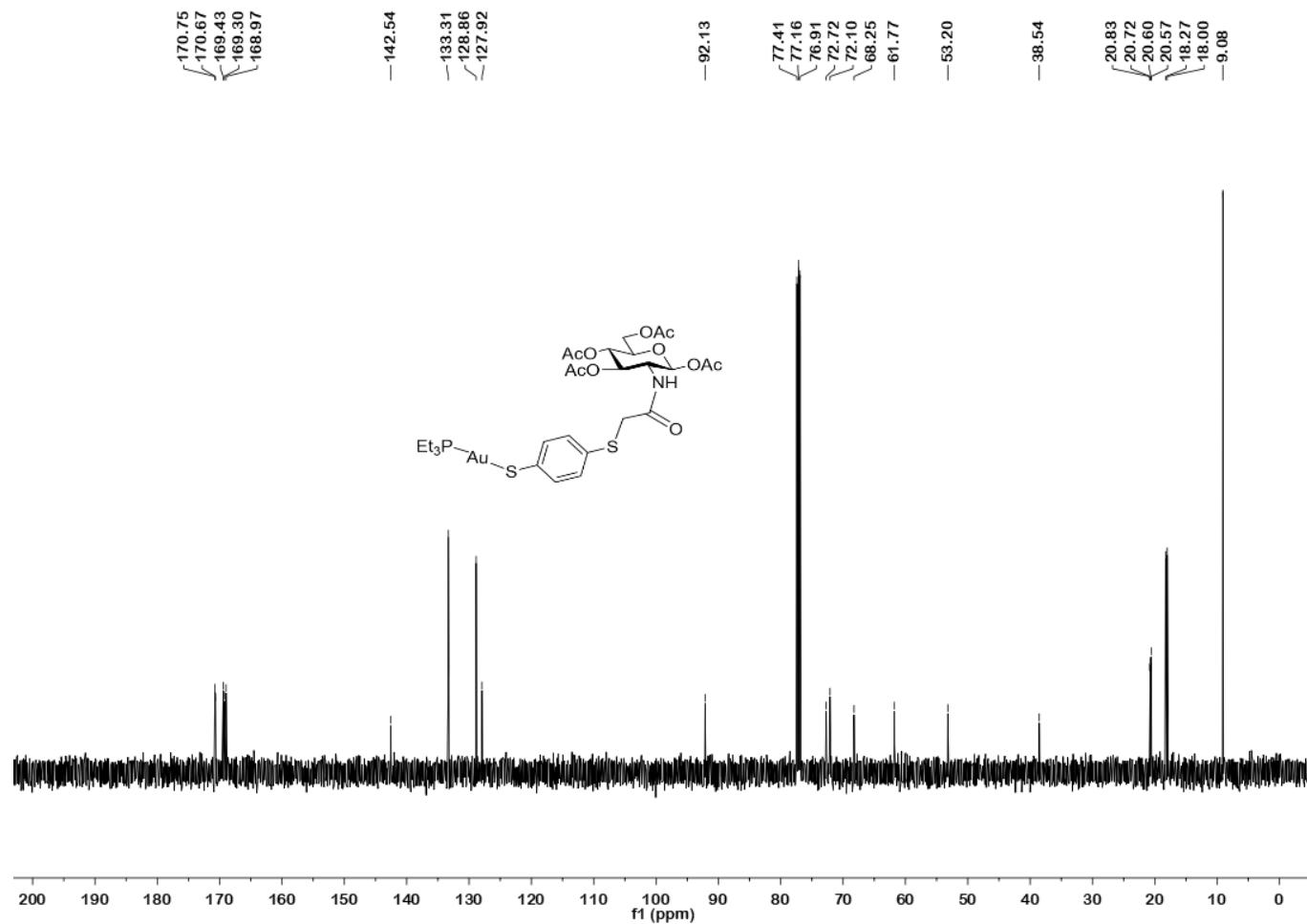


Figure S42. ^{13}C NMR spectrum of compound 8 in CDCl_3 .

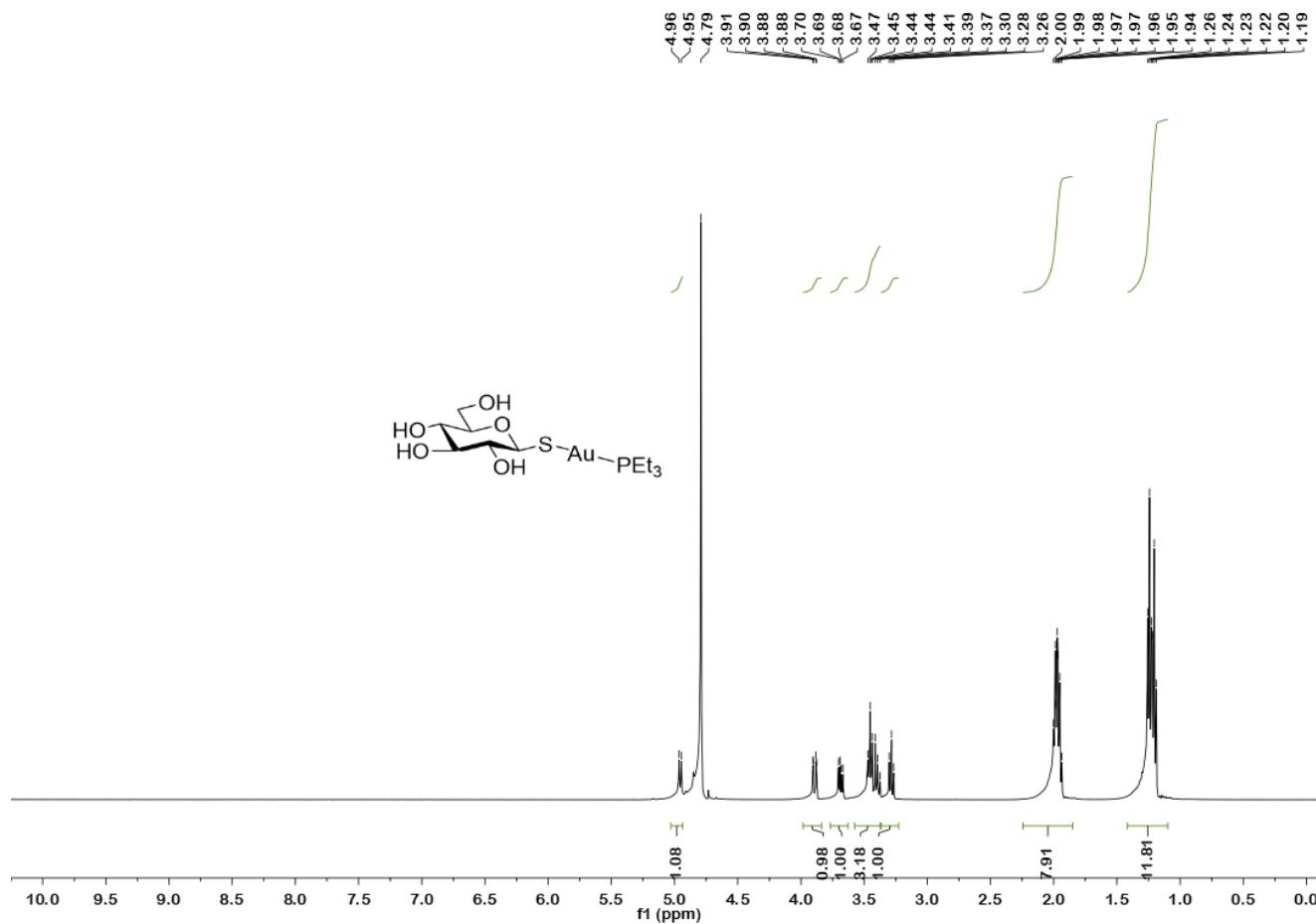


Figure S43. ¹H NMR spectrum of compound **9** in D_2O .

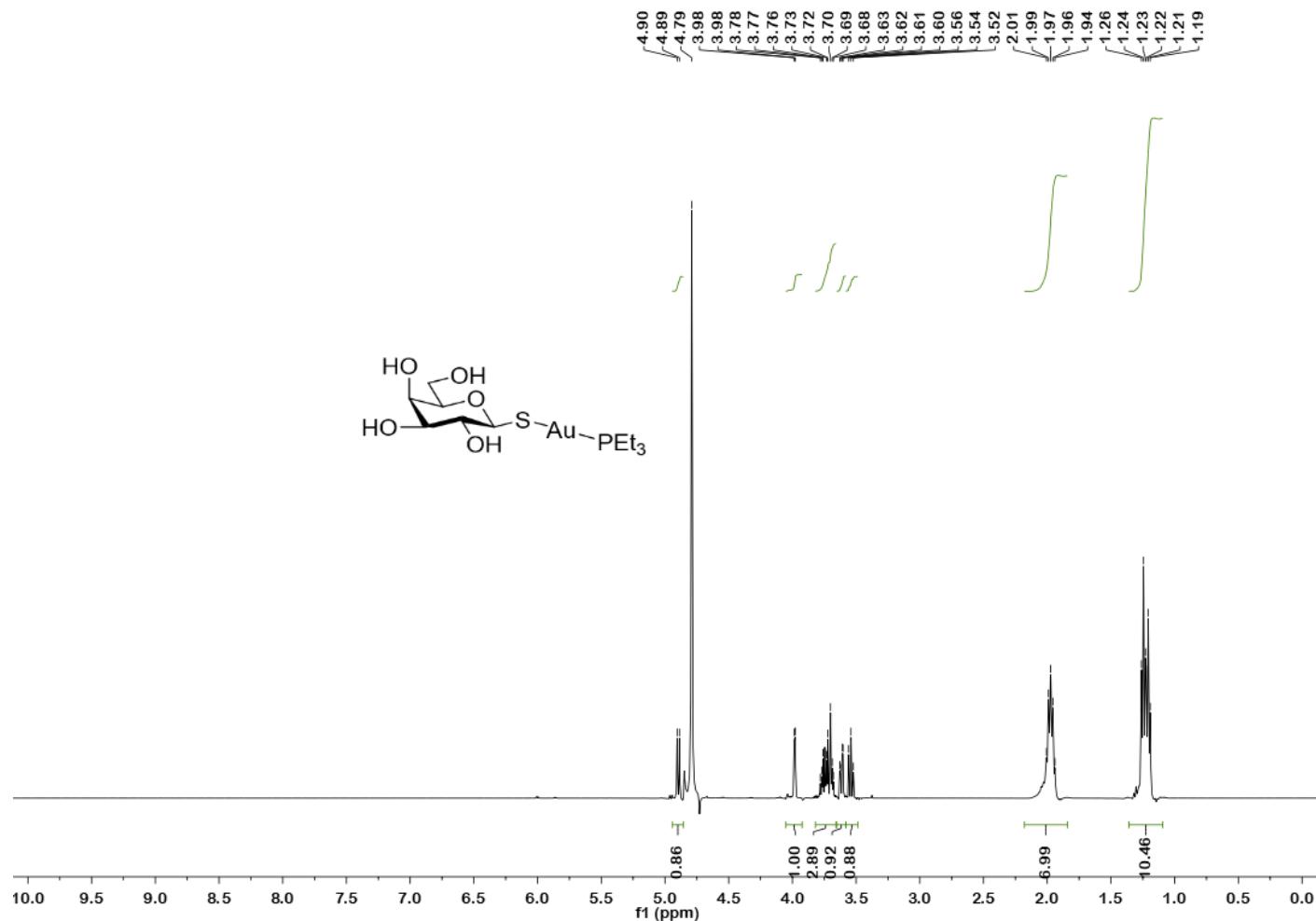


Figure S44. ¹H NMR spectrum of compound **10** in D_2O .

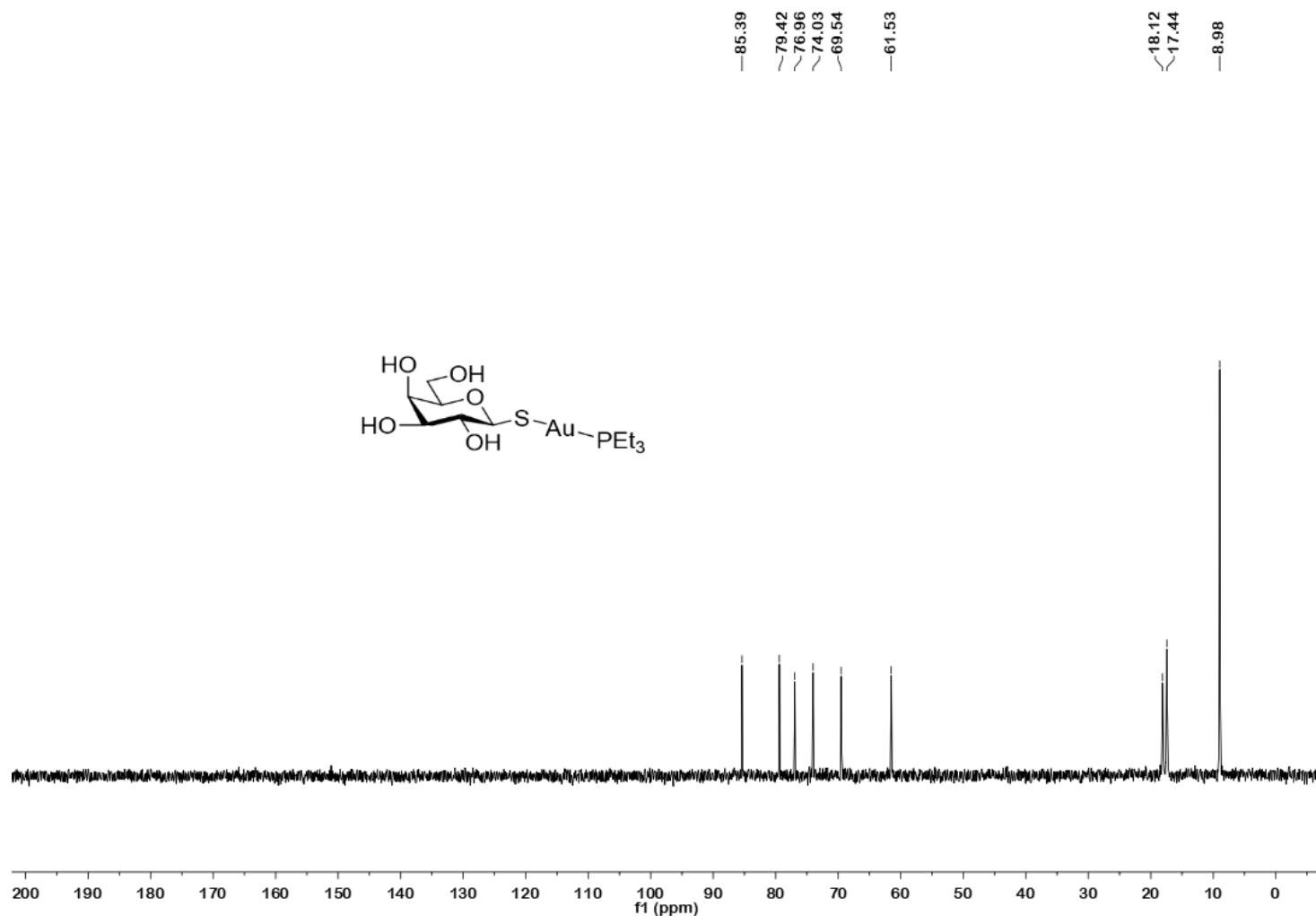


Figure S45. ^{13}C NMR spectrum of compound **10** in D_2O .

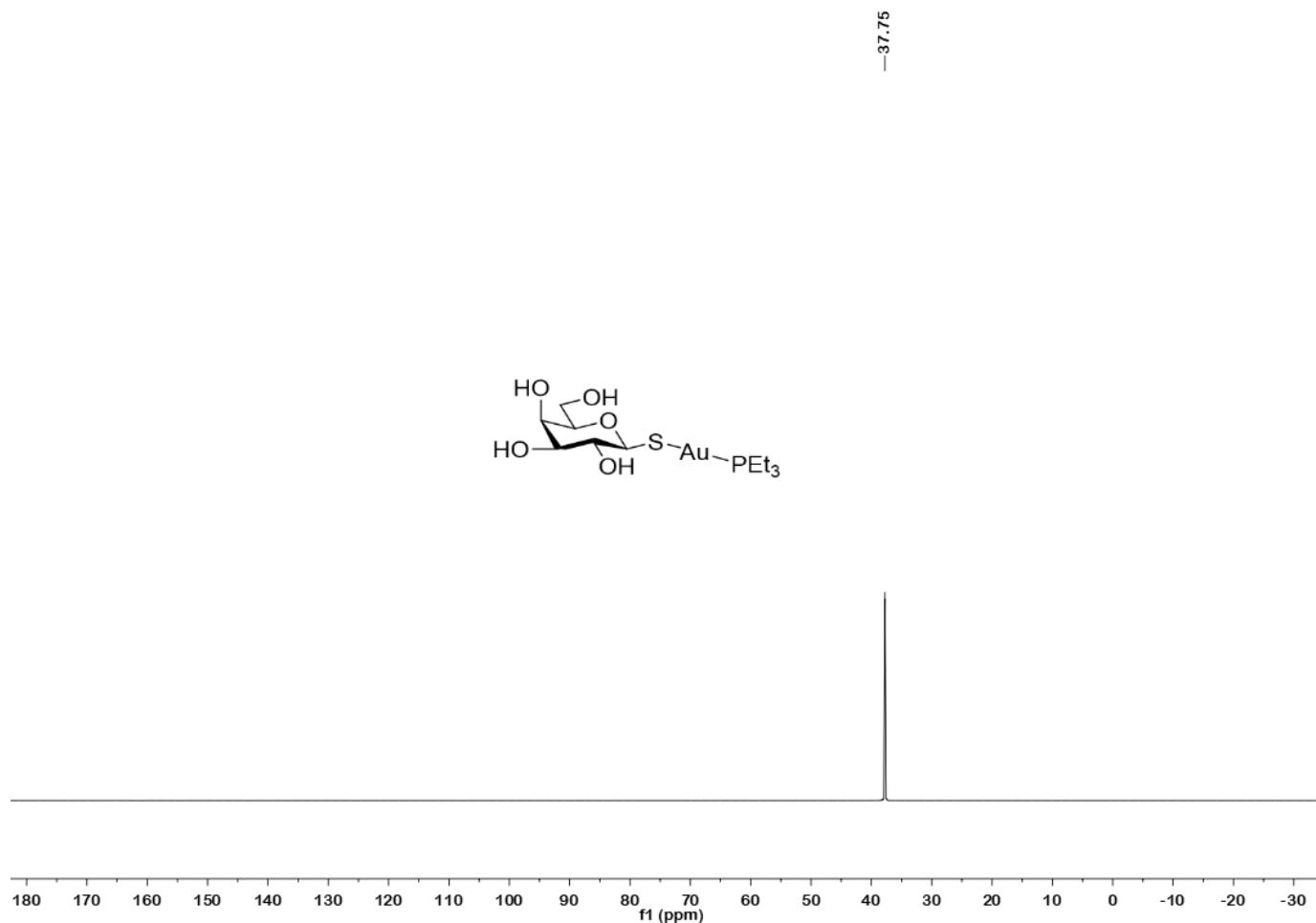


Figure S46. ^{31}P NMR spectrum of compound **10** in D_2O .

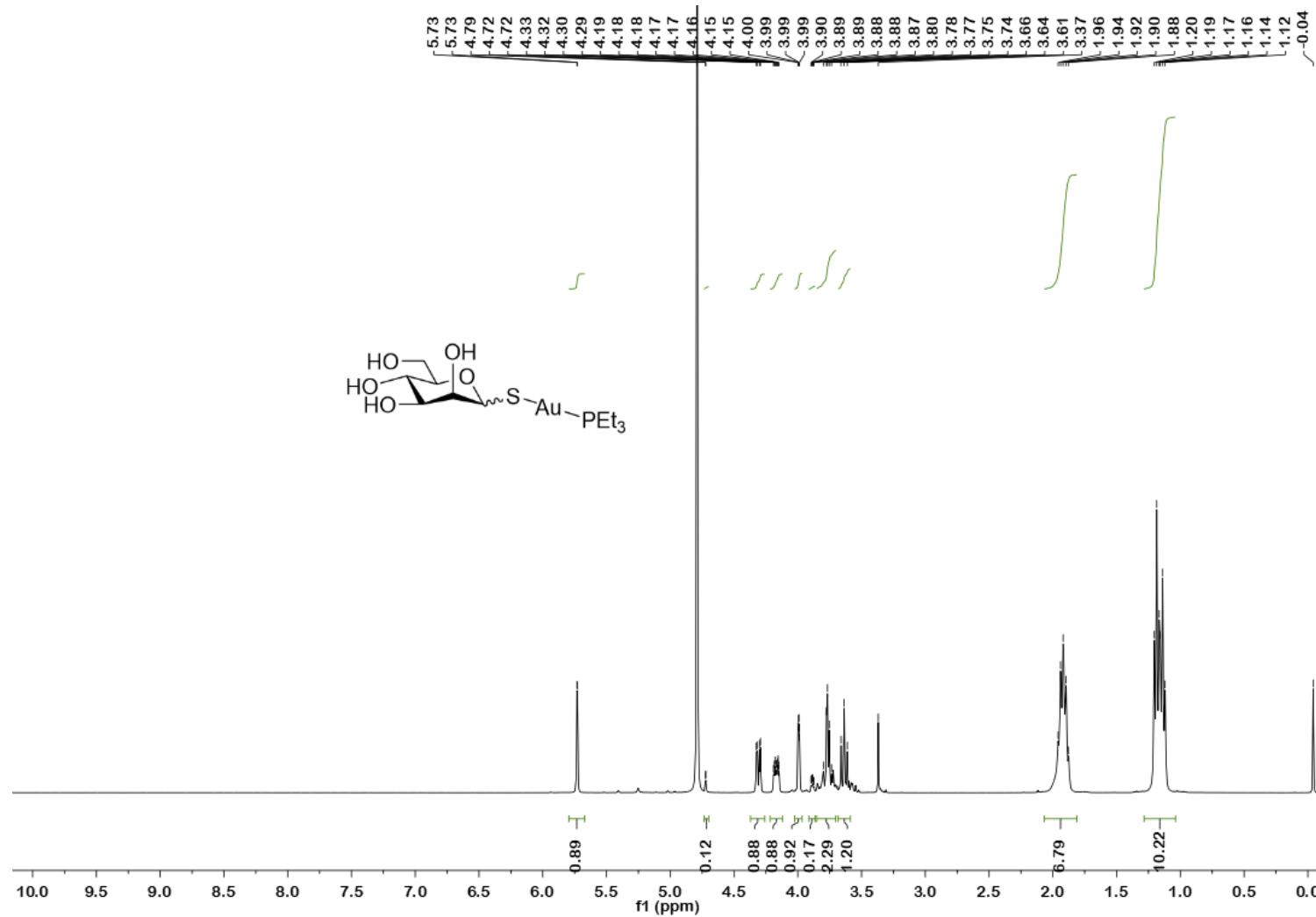


Figure S47. ¹H NMR spectrum of compound **11** in D₂O

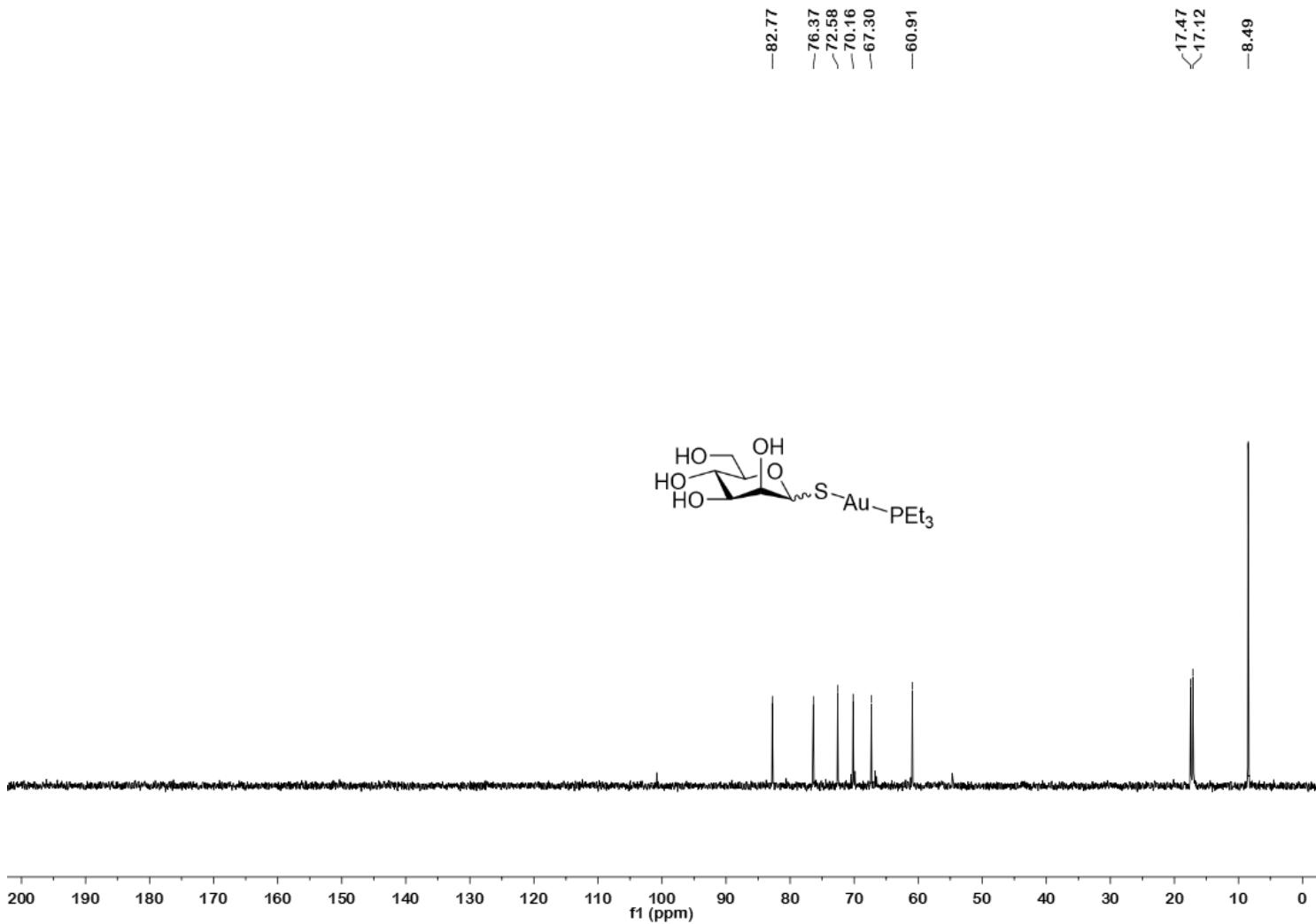


Figure S48. ^{13}C NMR spectrum of compound **11** in D_2O .

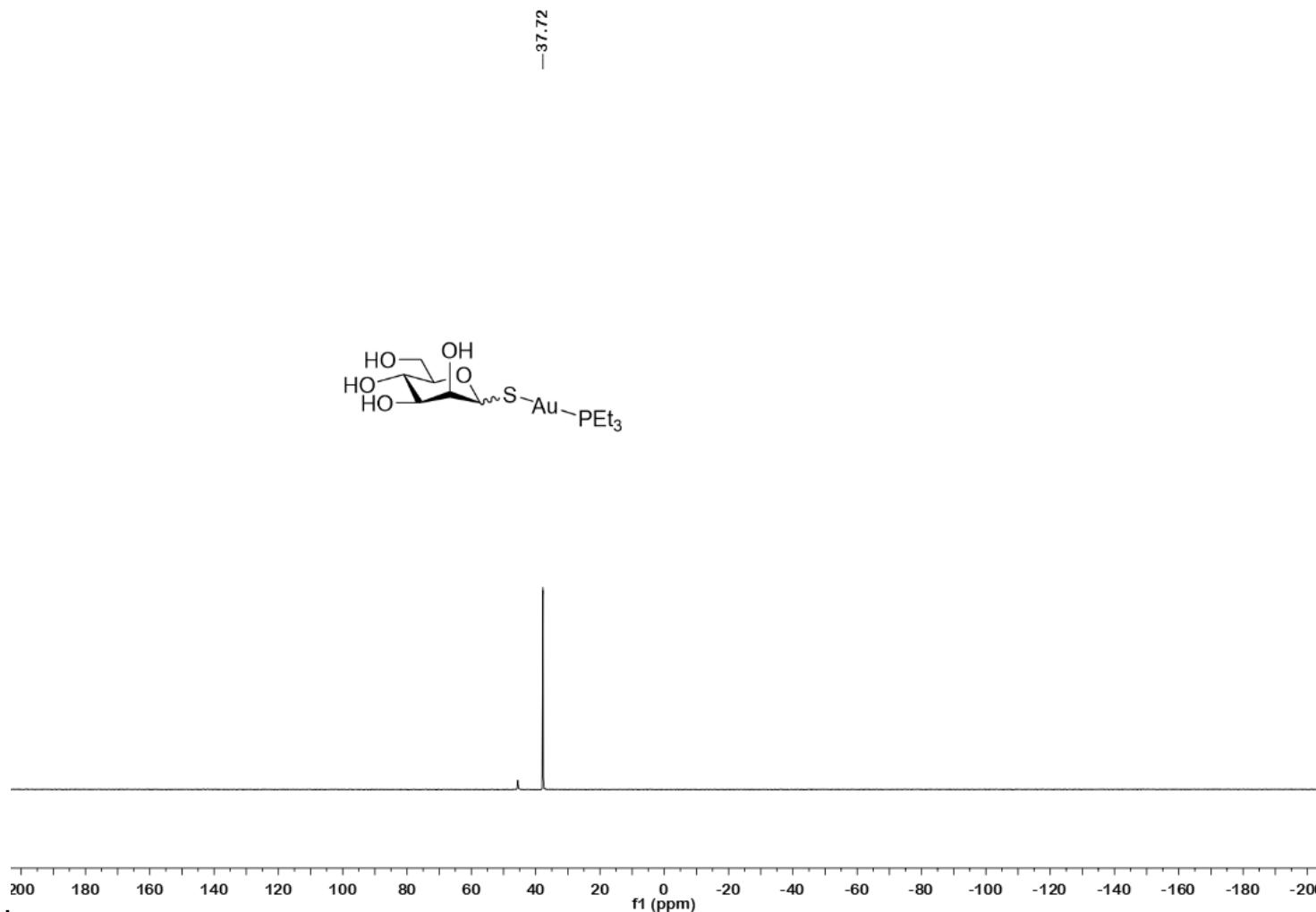


Figure S49. ^{31}P NMR spectrum of compound **11** in D_2O .

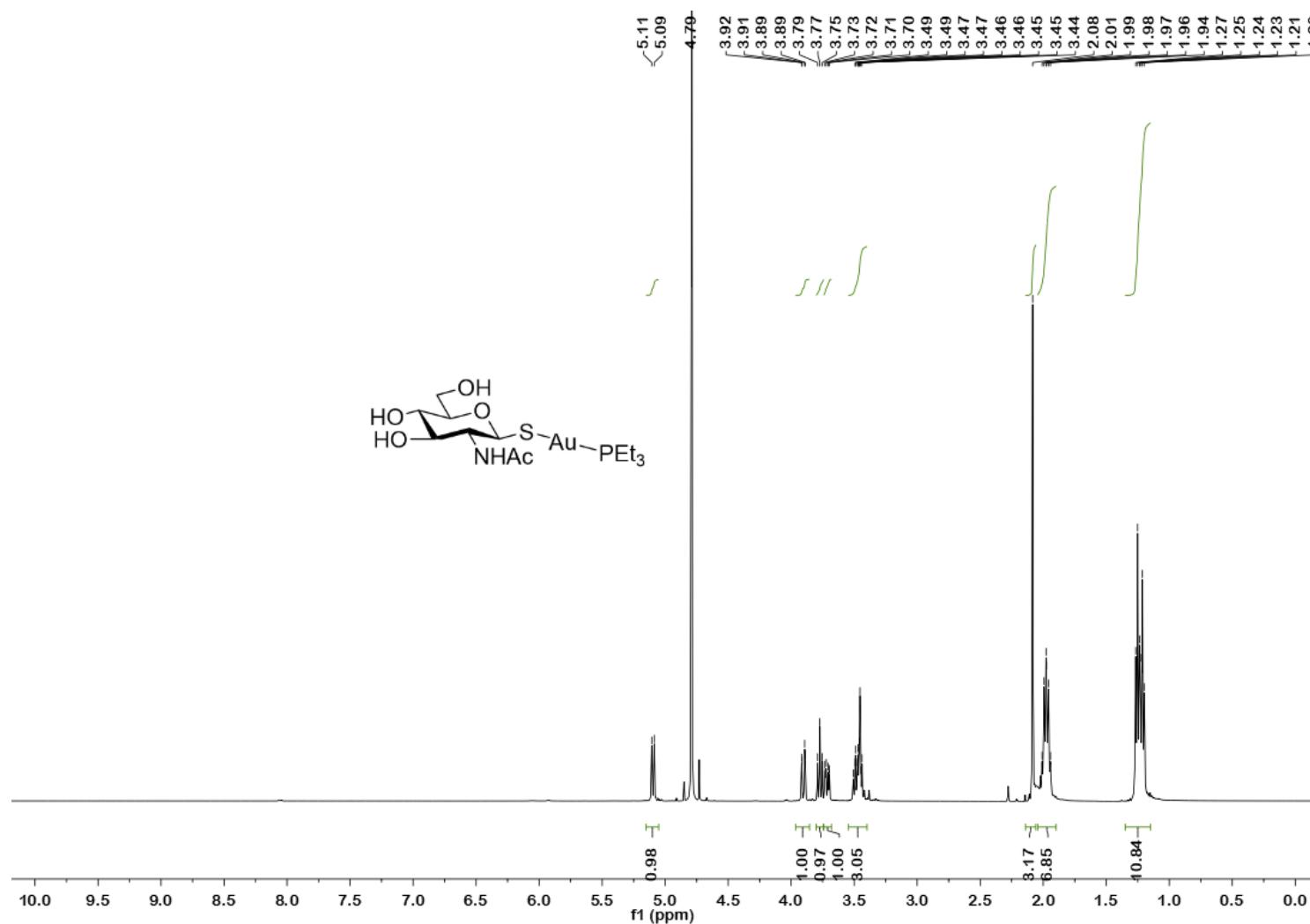


Figure S50. ^1H NMR spectrum of compound **12** in D_2O .

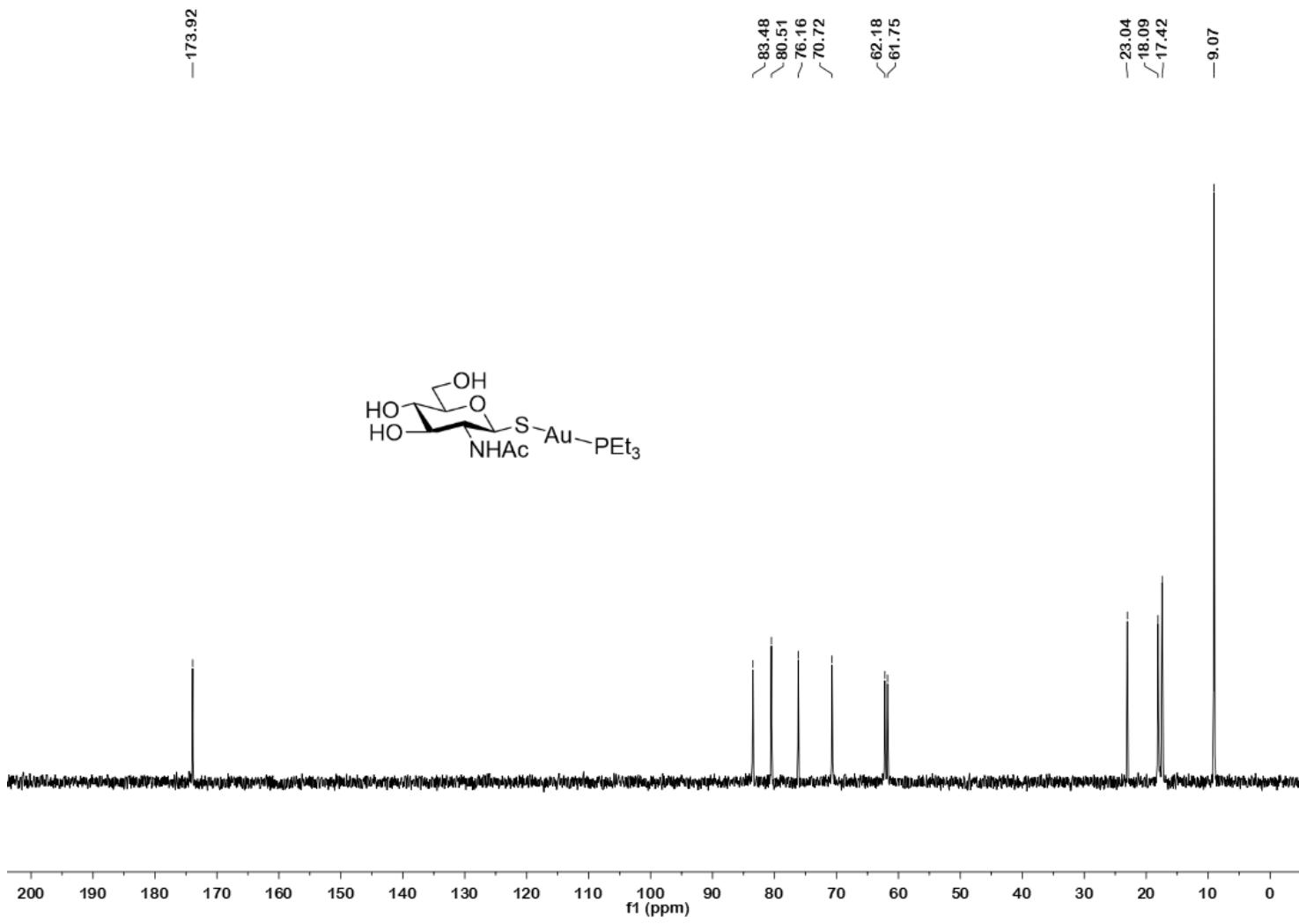


Figure S51. ^{13}C NMR spectrum of compound **12** in D_2O .

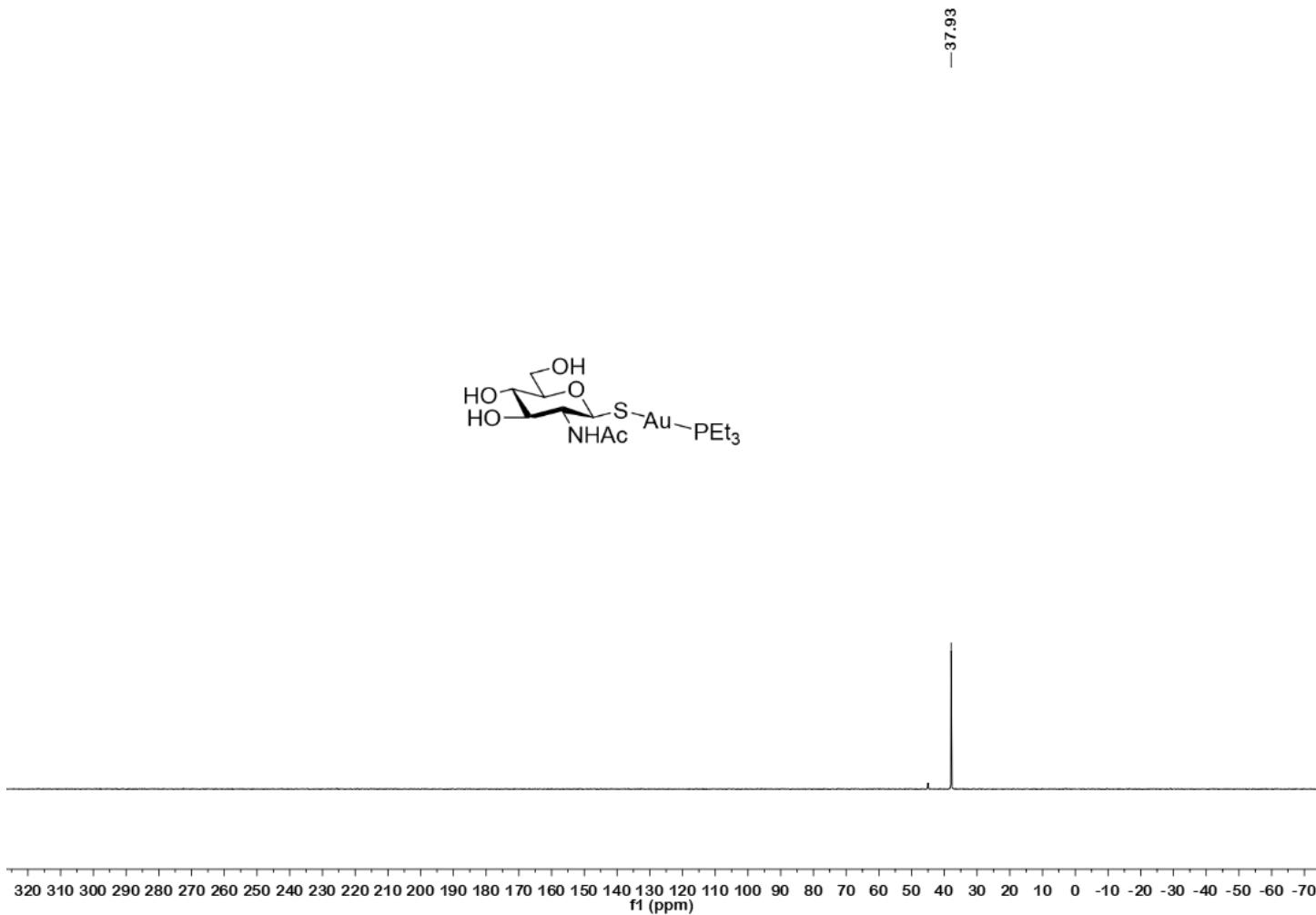


Figure S52. ${}^{31}\text{P}$ NMR spectrum of compound **12** in D_2O .

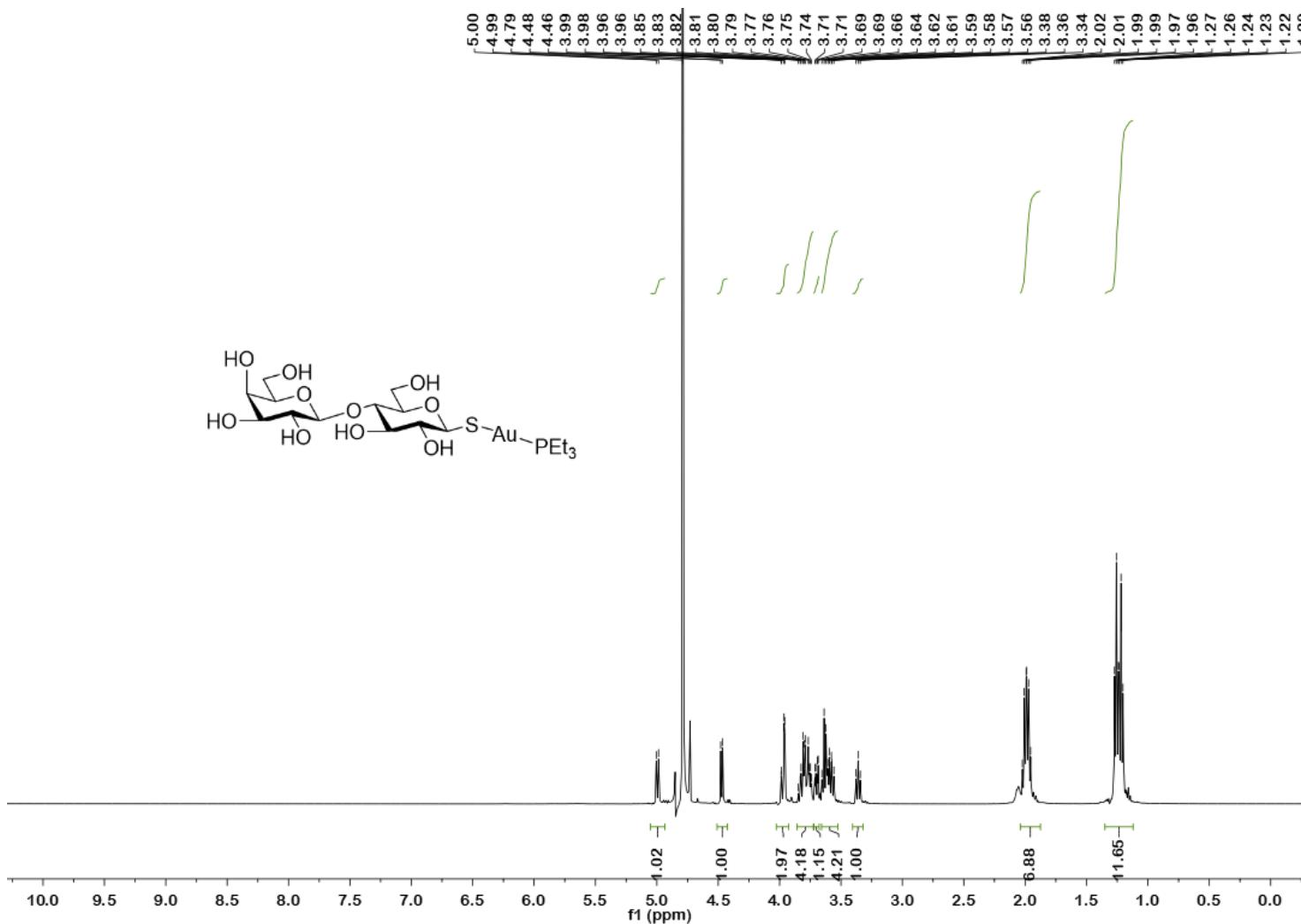


Figure S53. ^1H NMR spectrum of compound **13** in D_2O .

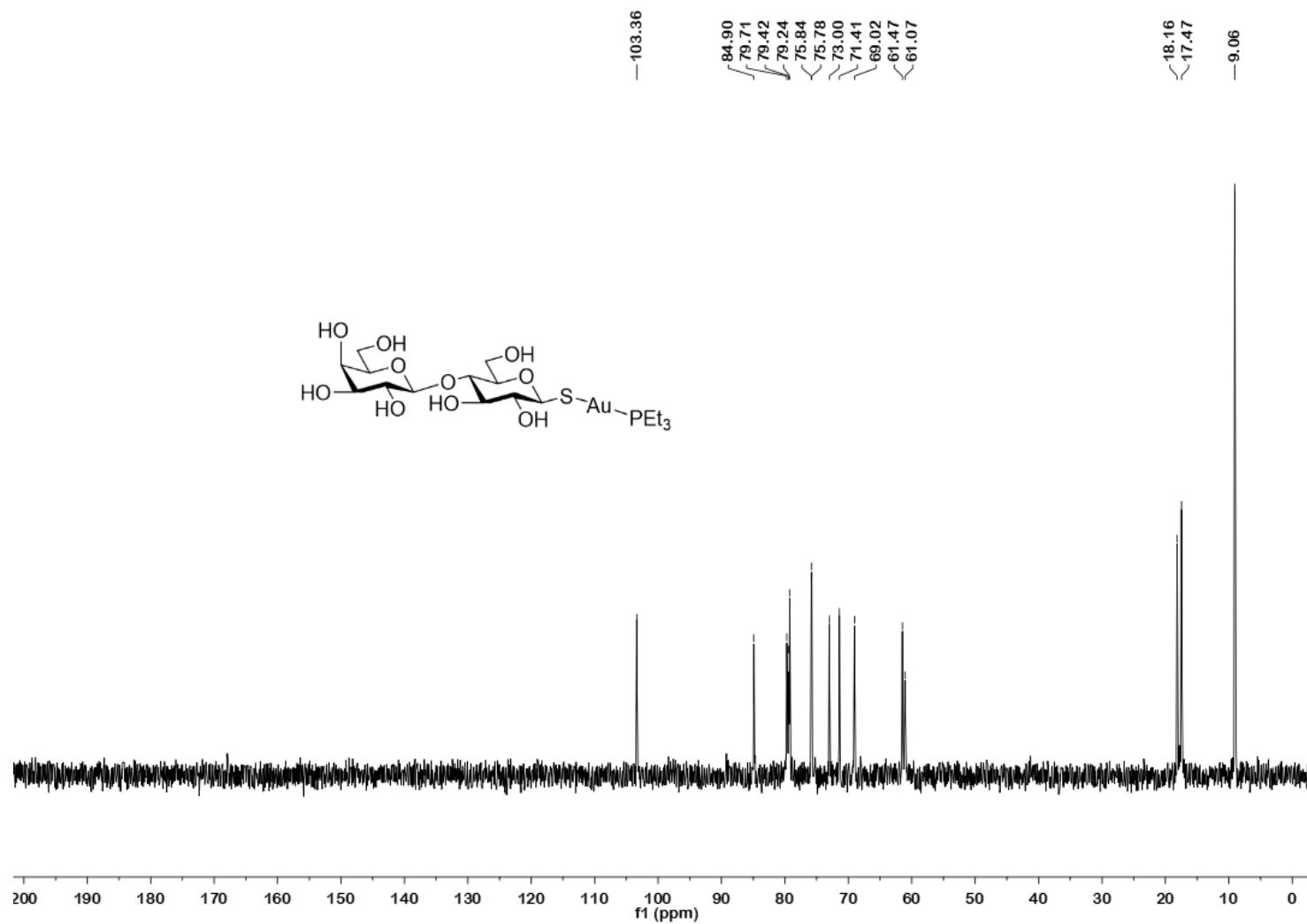


Figure S54. ^{13}C NMR spectrum of compound **13** in D_2O .

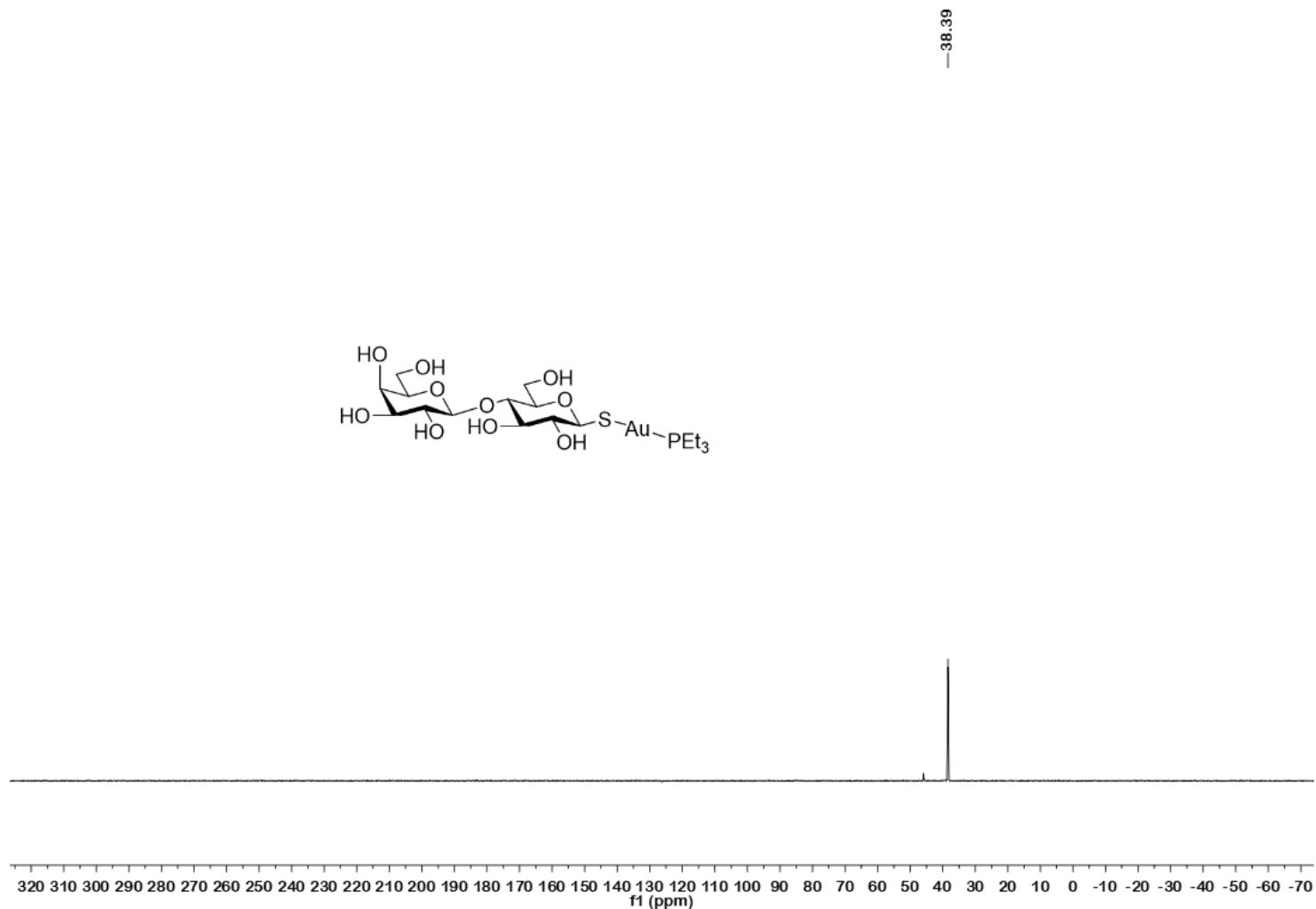


Figure S55. ^{31}P NMR spectrum of compound 13 in D_2O .

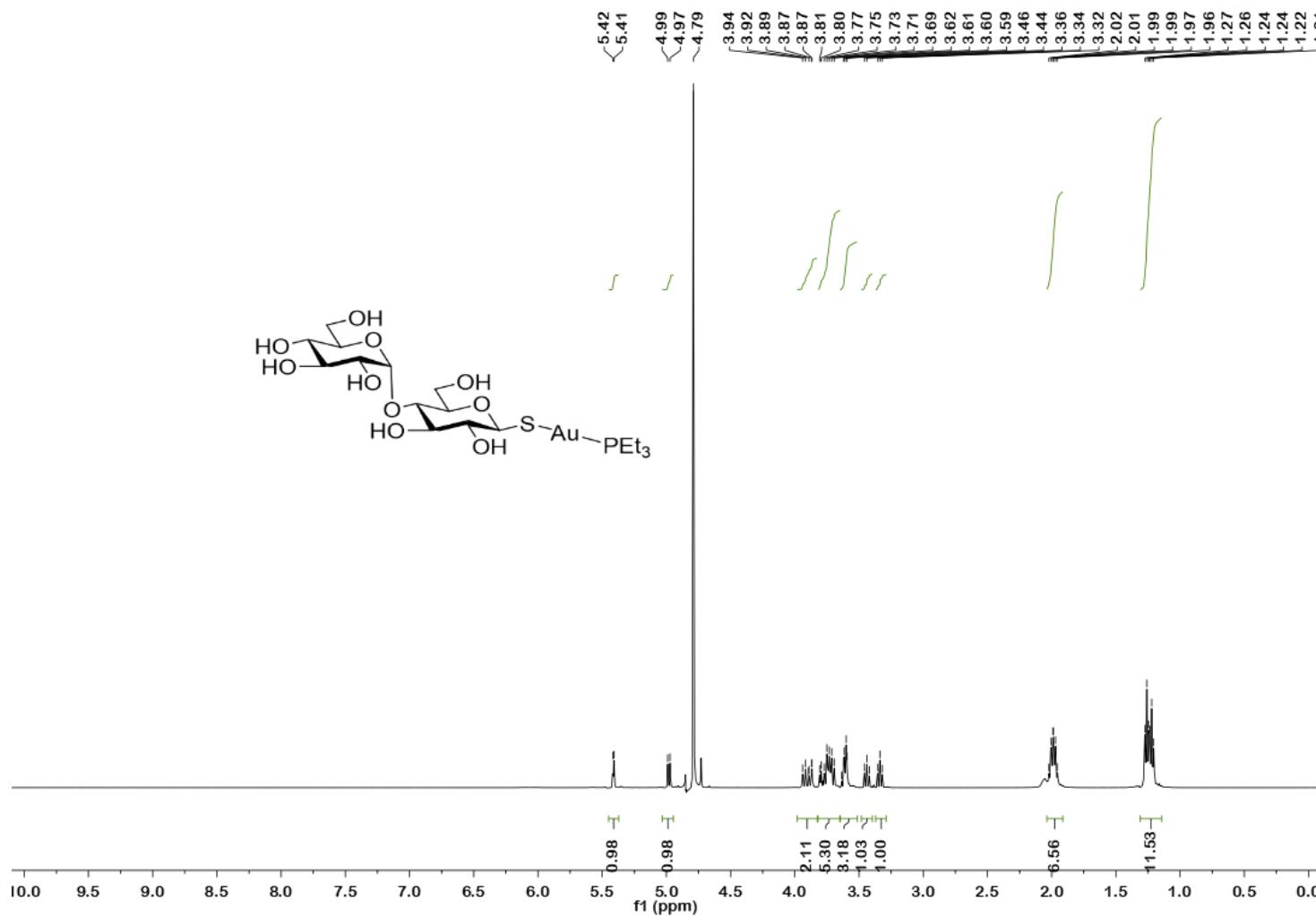


Figure S56. ^1H NMR spectrum of compound **14** in D_2O .

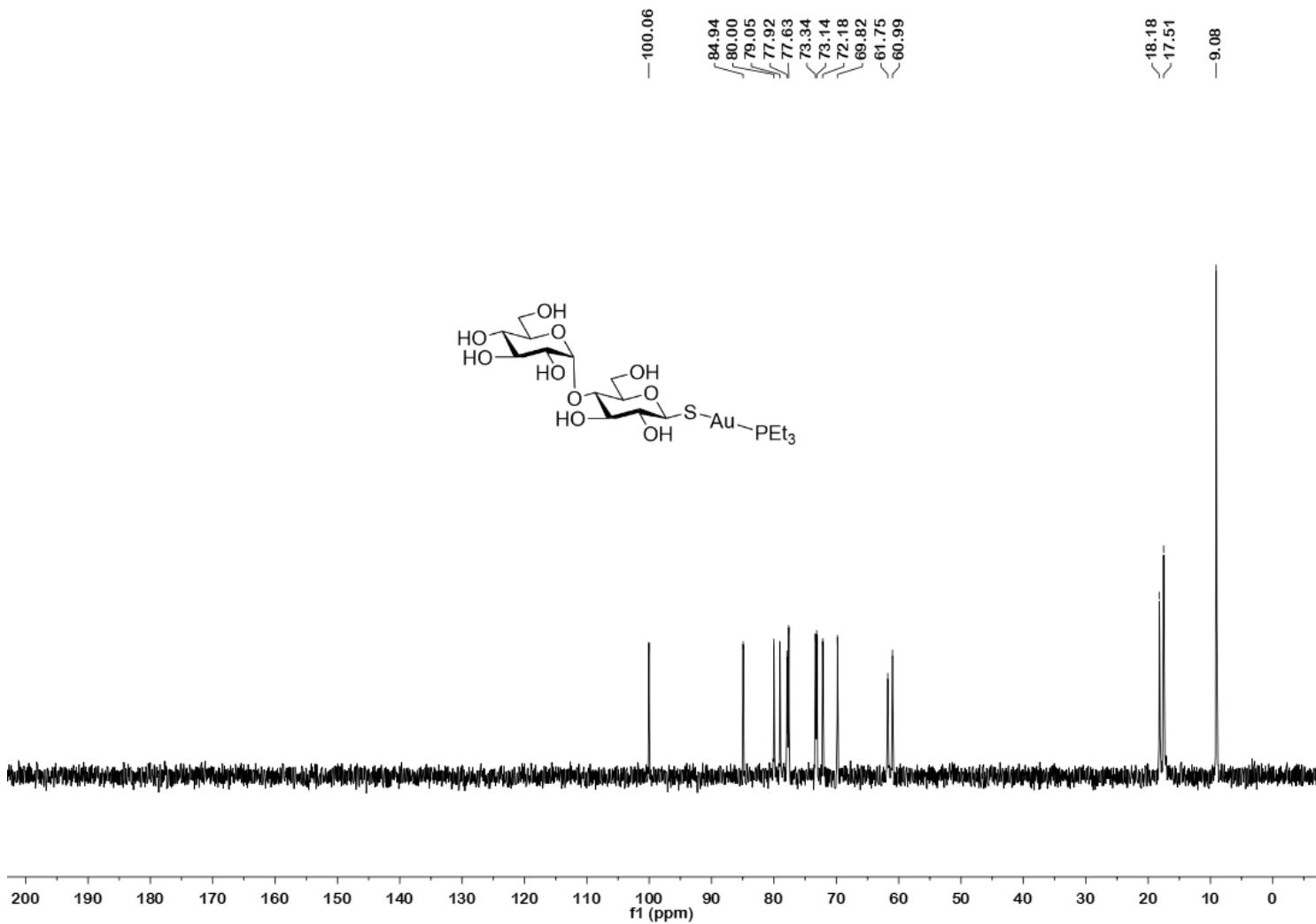


Figure S57. ^{13}C NMR spectrum of compound **14** in D_2O .

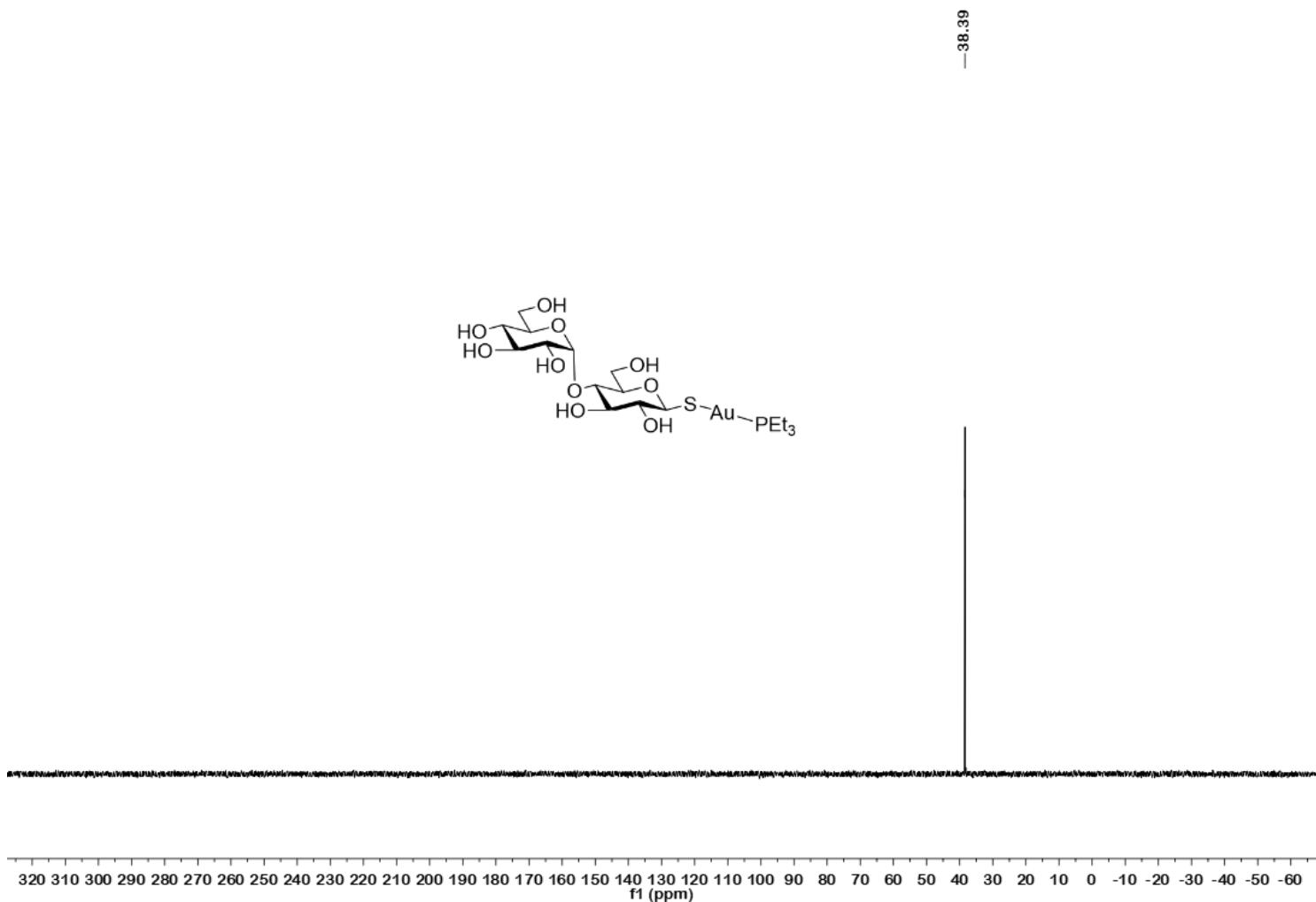


Figure S58. ^{31}P NMR spectrum of compound **14** in D_2O .

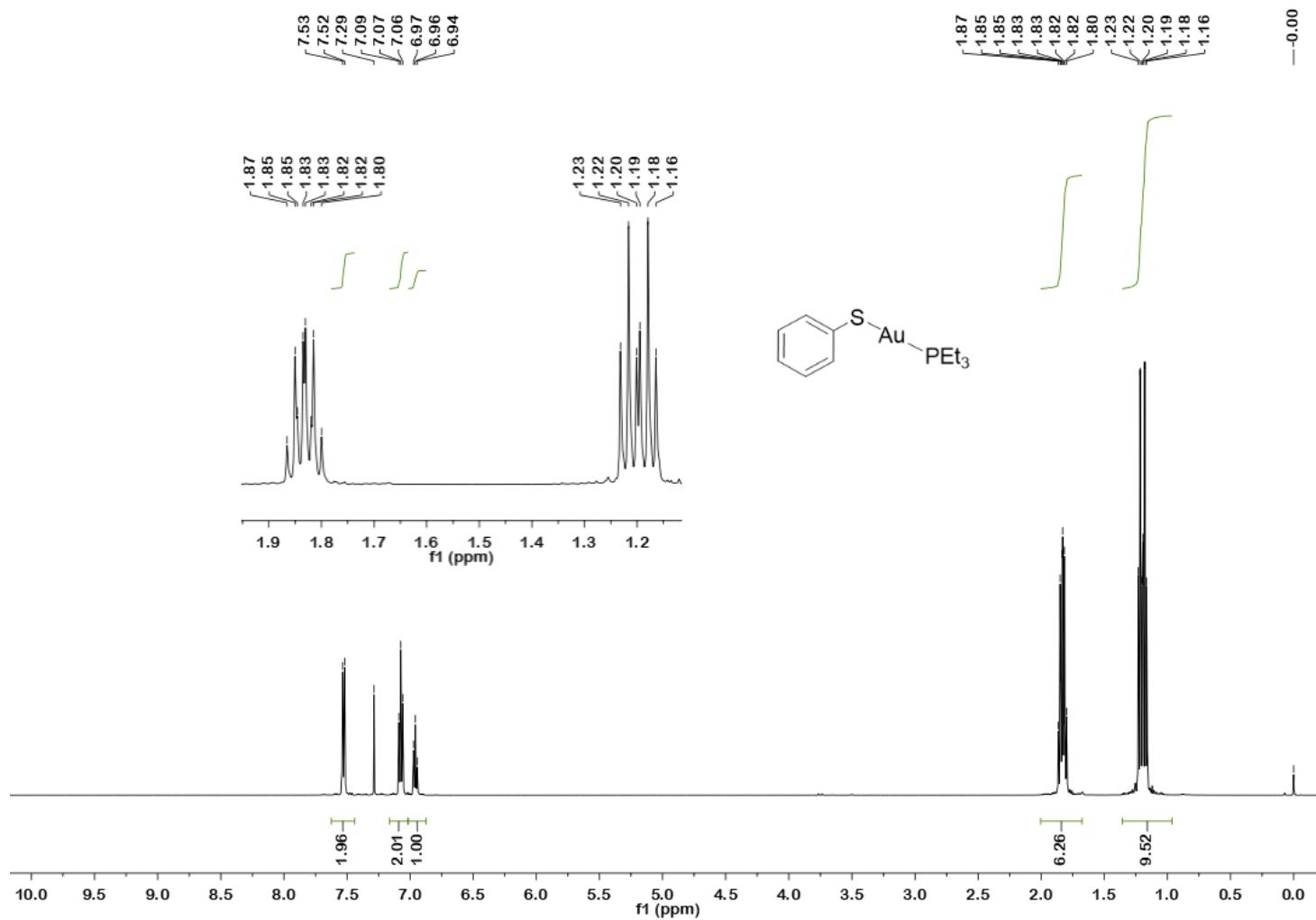


Figure S59. ¹H NMR spectrum of compound **15** in CDCl_3 .

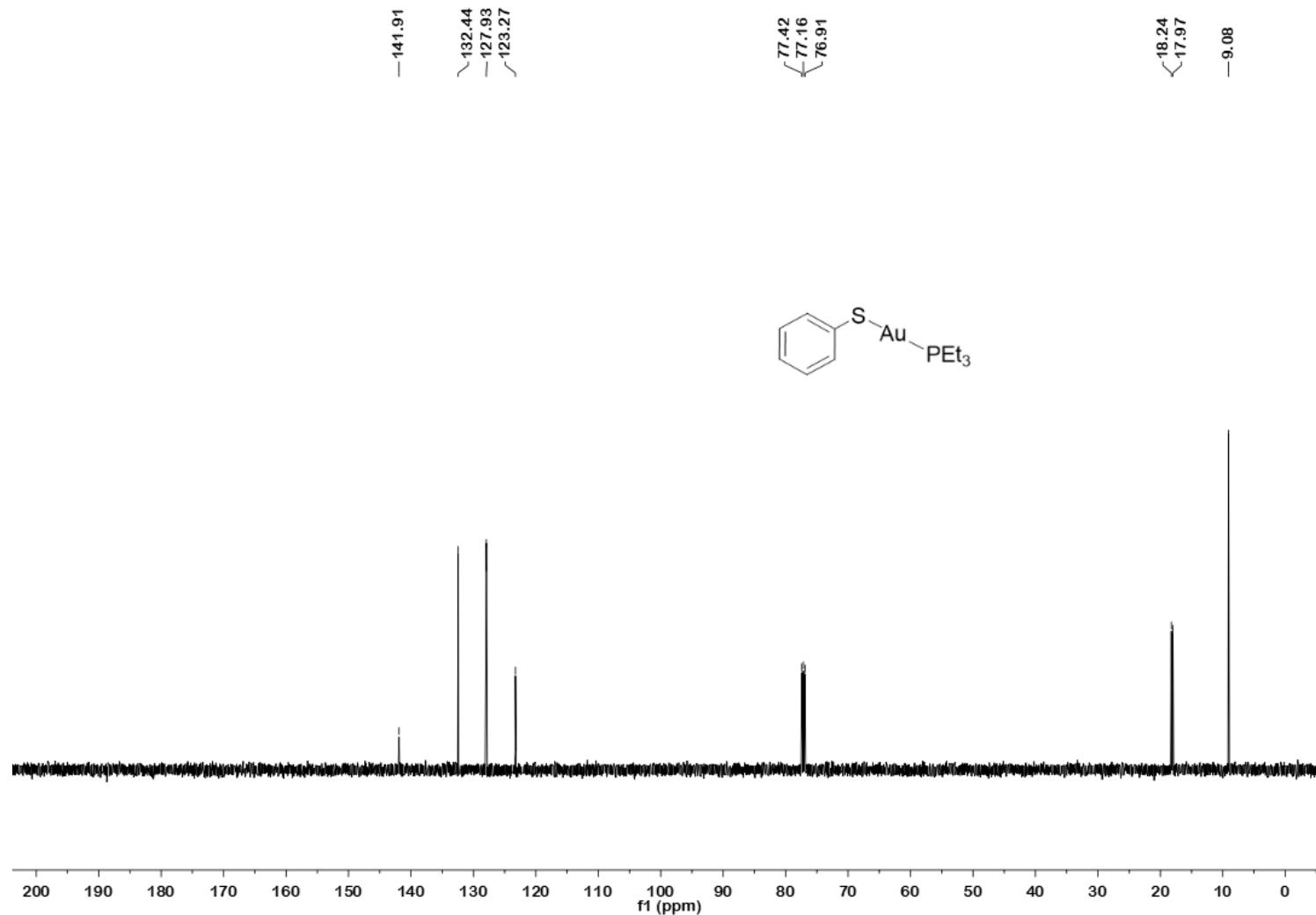


Figure S60. ^{13}C NMR spectrum of compound **15** in CDCl_3 .

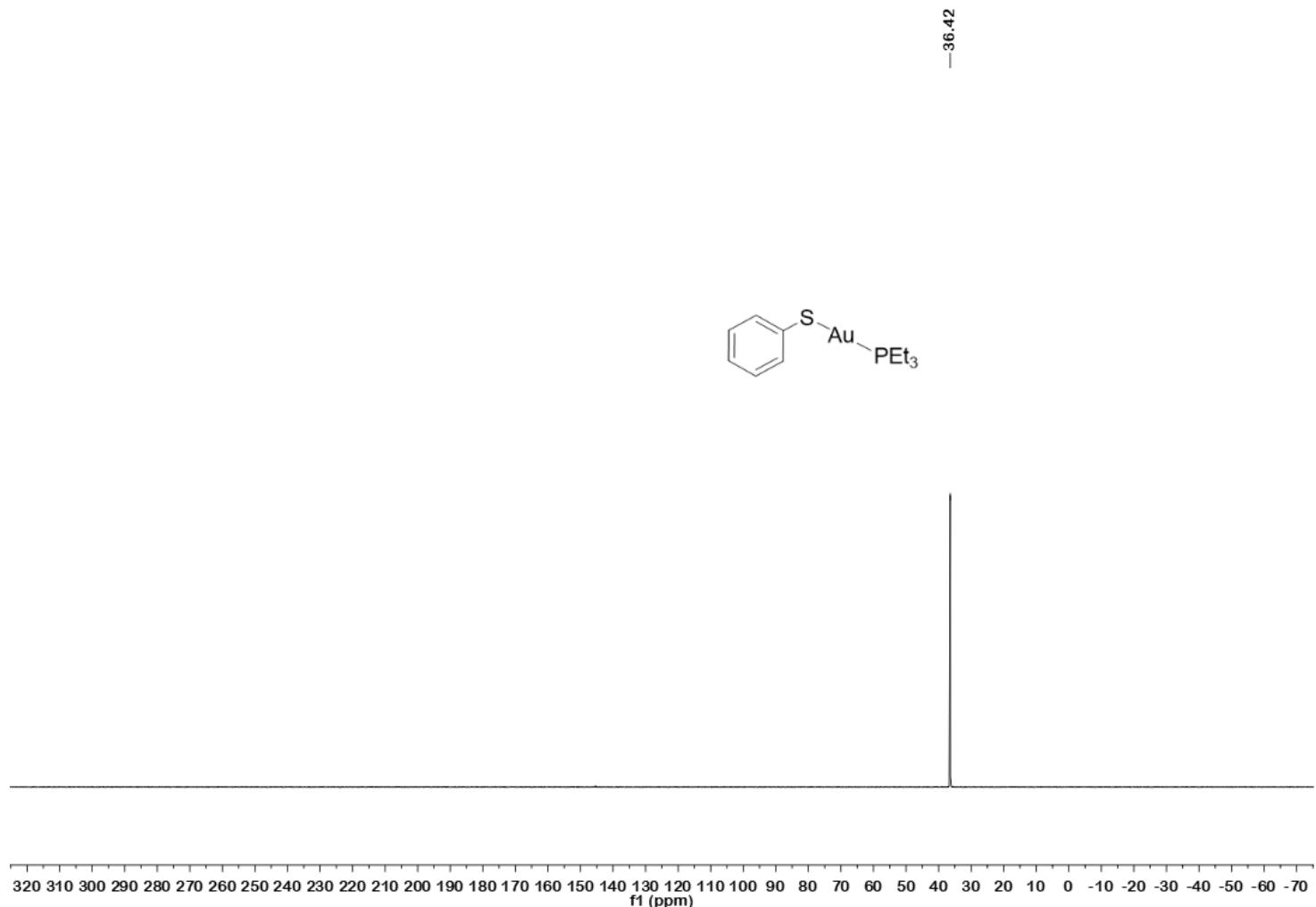


Figure S61. ^{31}P NMR spectrum of compound **15** in CDCl_3 .

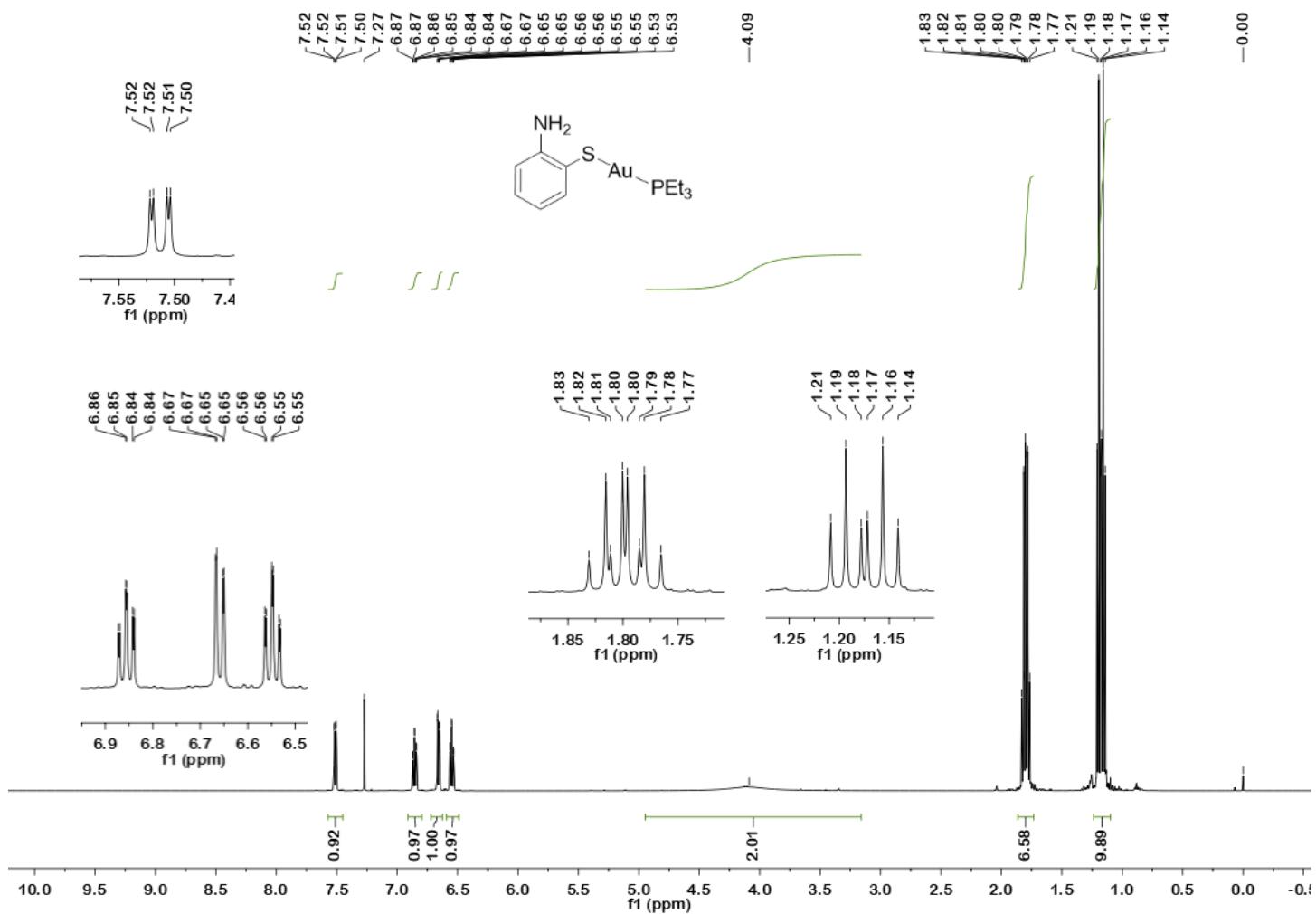


Figure S62. ^1H NMR spectrum of compound **16** in CDCl_3 .

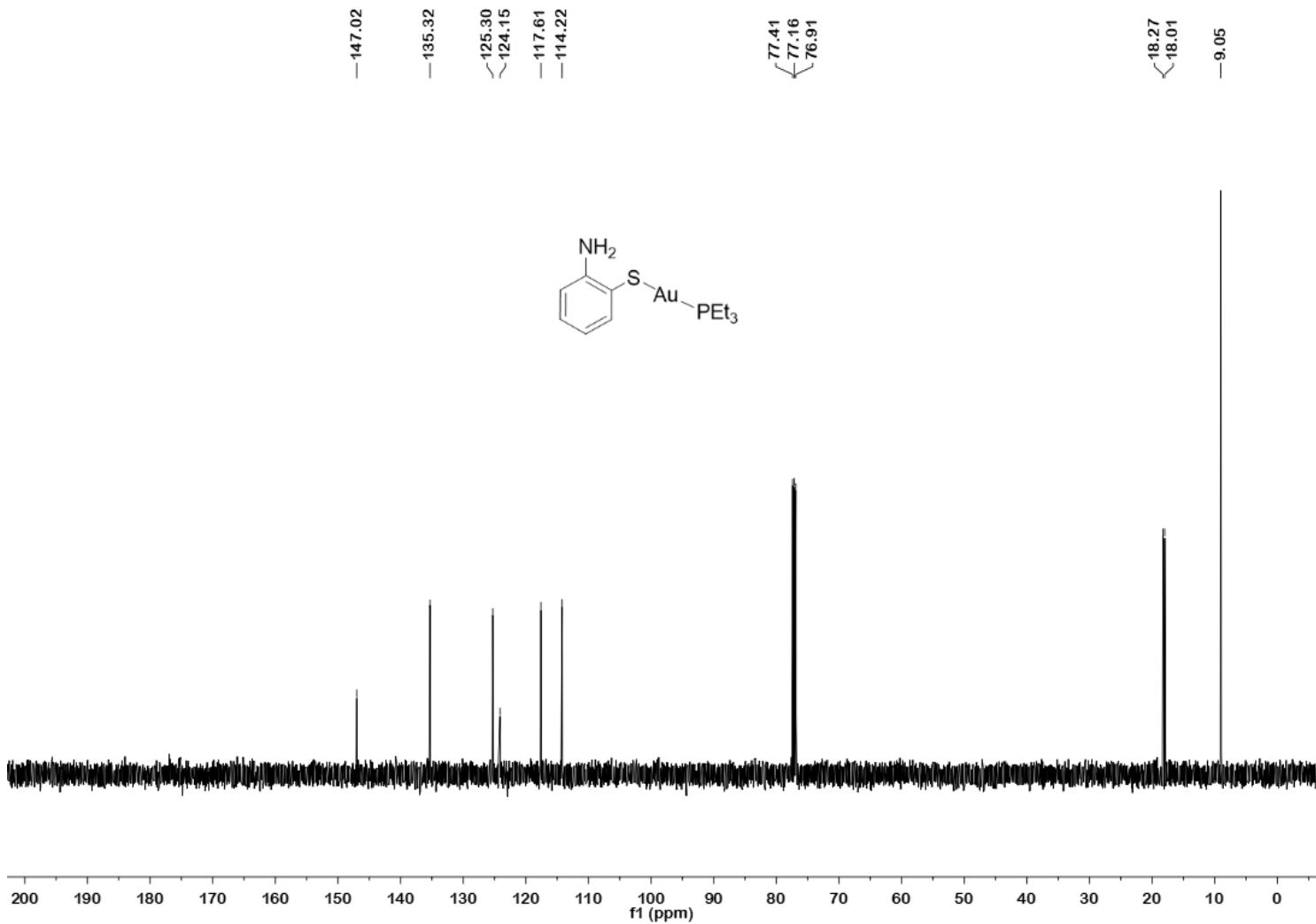


Figure S63. ^{13}C NMR spectrum of compound **16** in CDCl_3 .

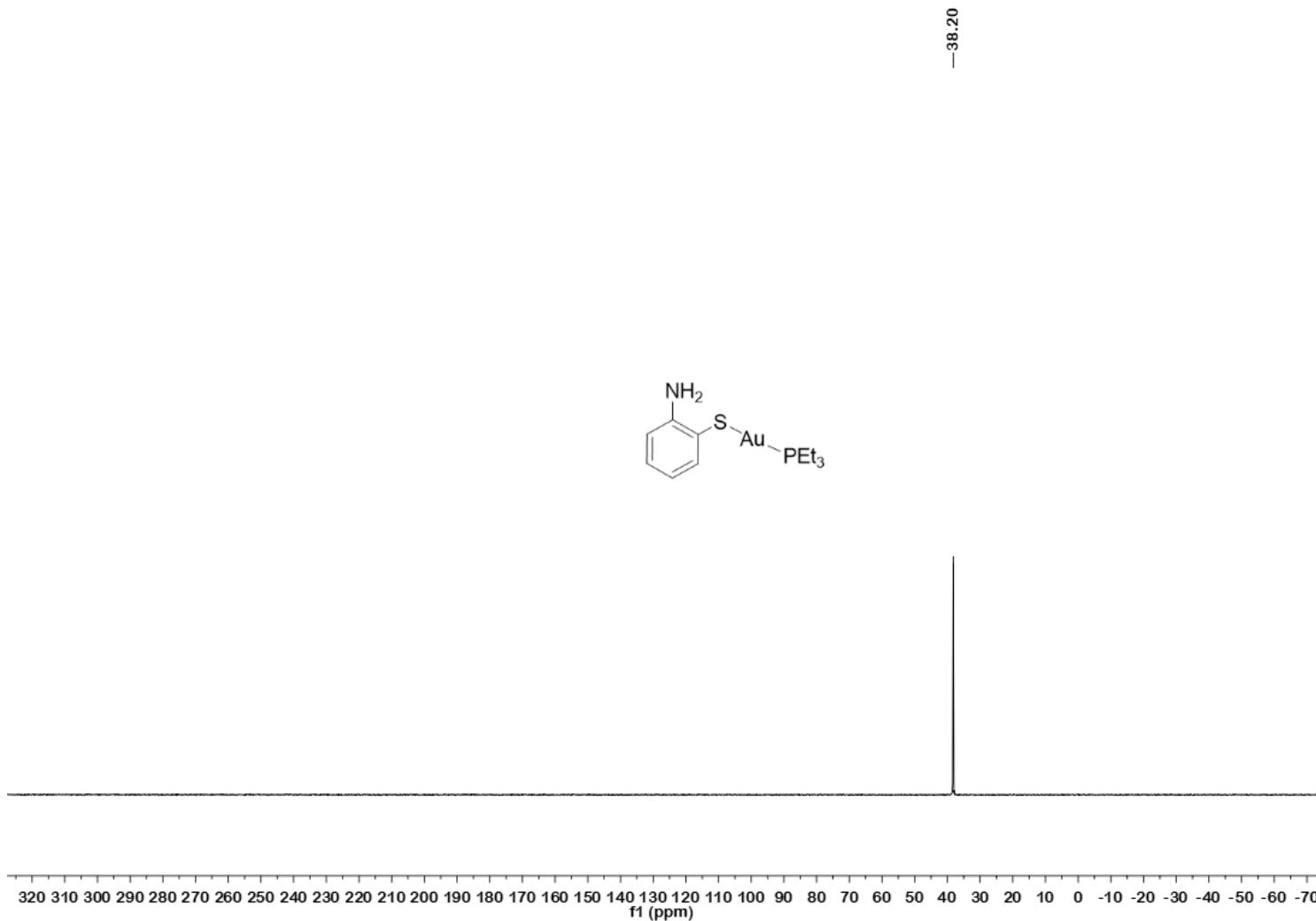


Figure S64. ^{31}P NMR spectrum of compound **16** in CDCl_3 .

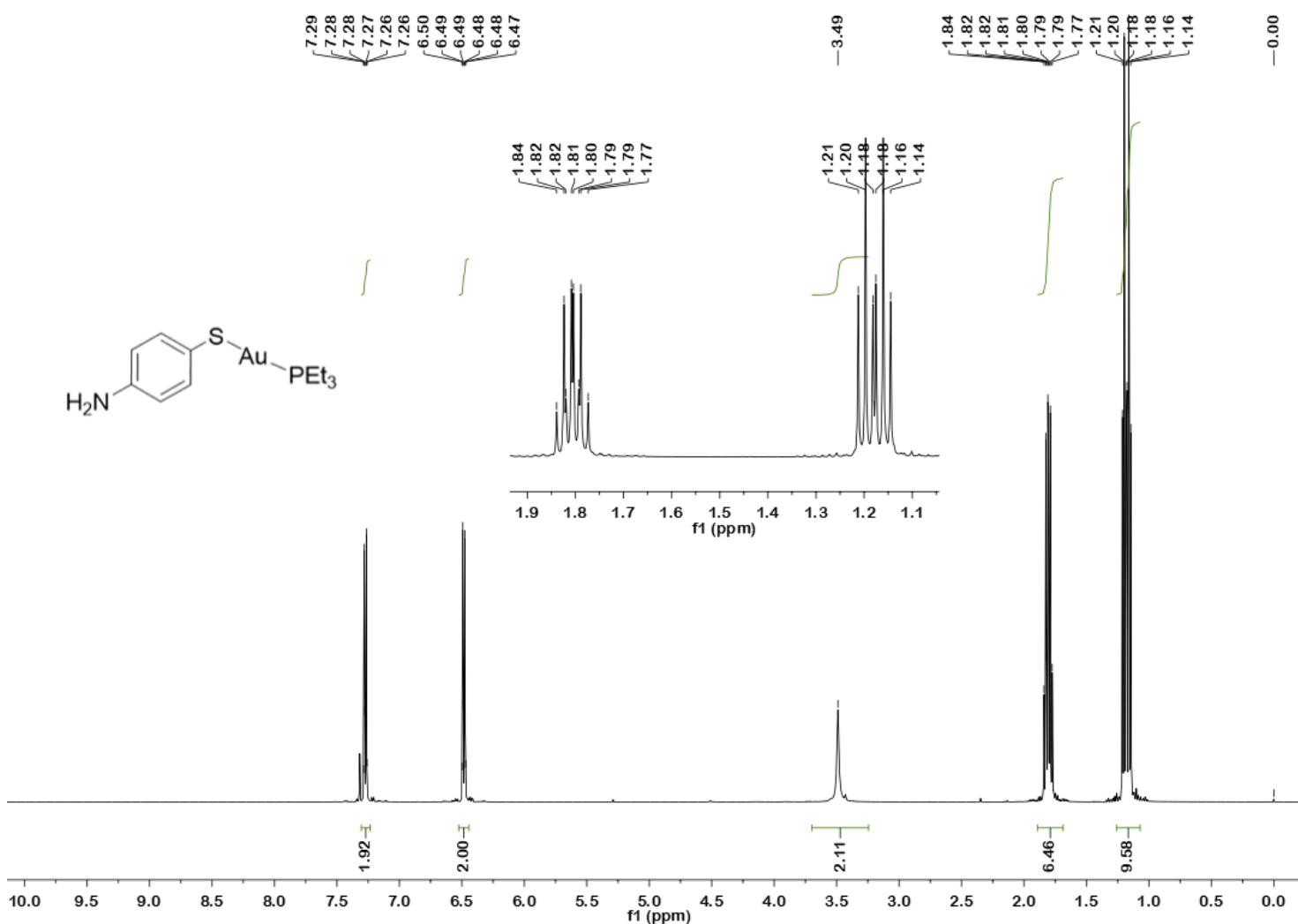


Figure S65. ^1H NMR spectrum of compound **17** in CDCl_3 .

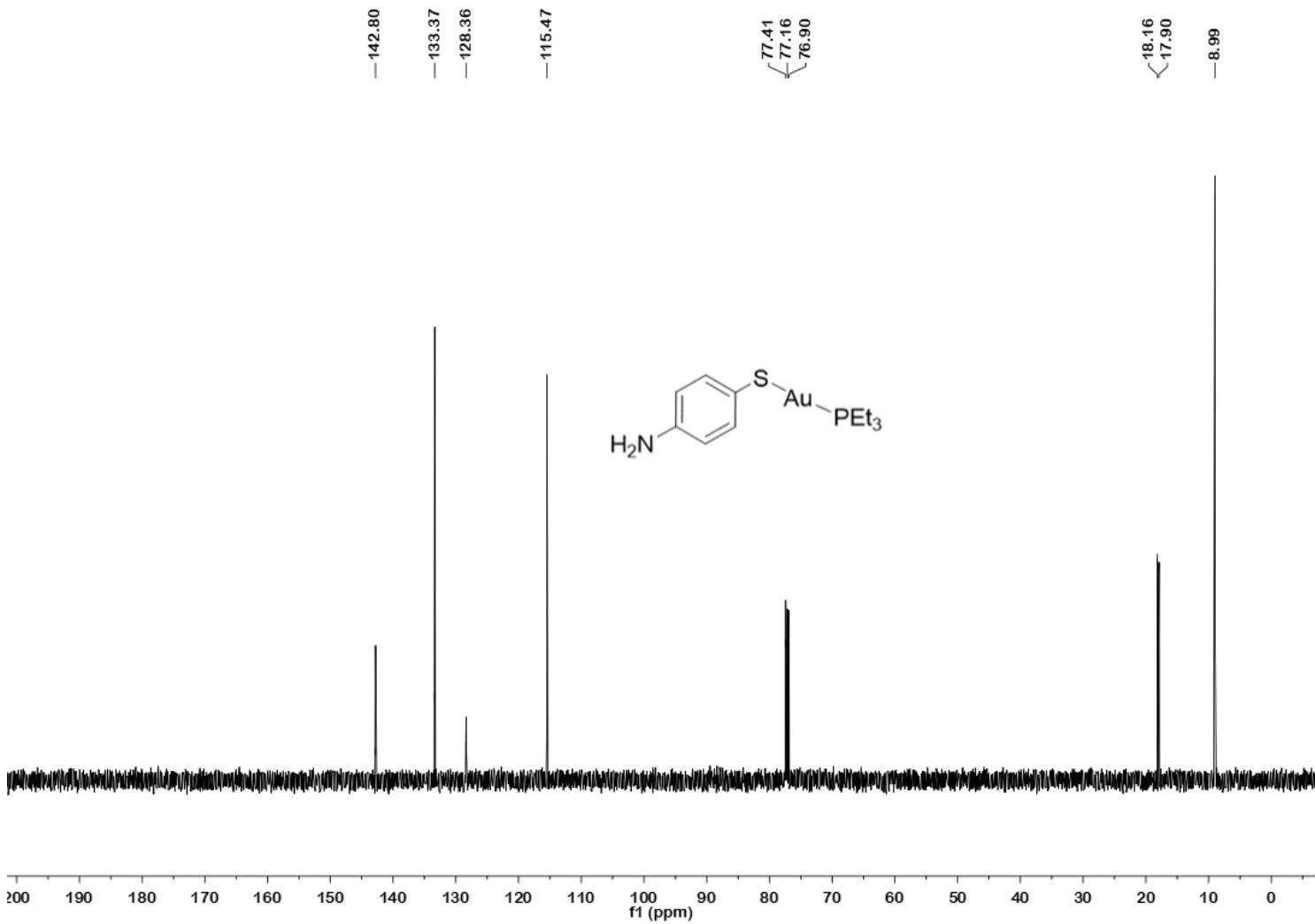


Figure S66. ^{13}C NMR spectrum of compound 17 in CDCl_3 .

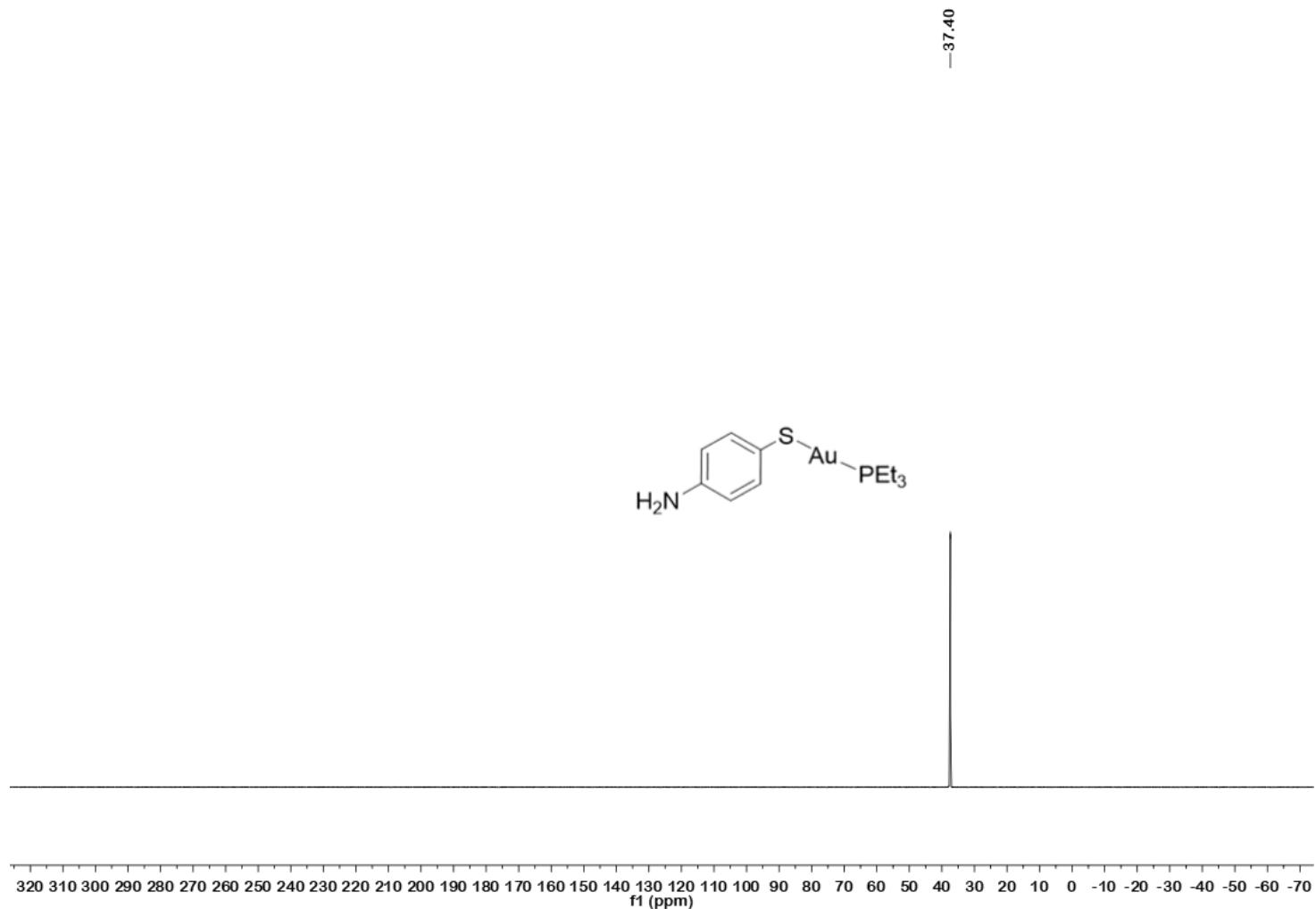


Figure S67. ^{31}P NMR spectrum of compound **17** in CDCl_3 .

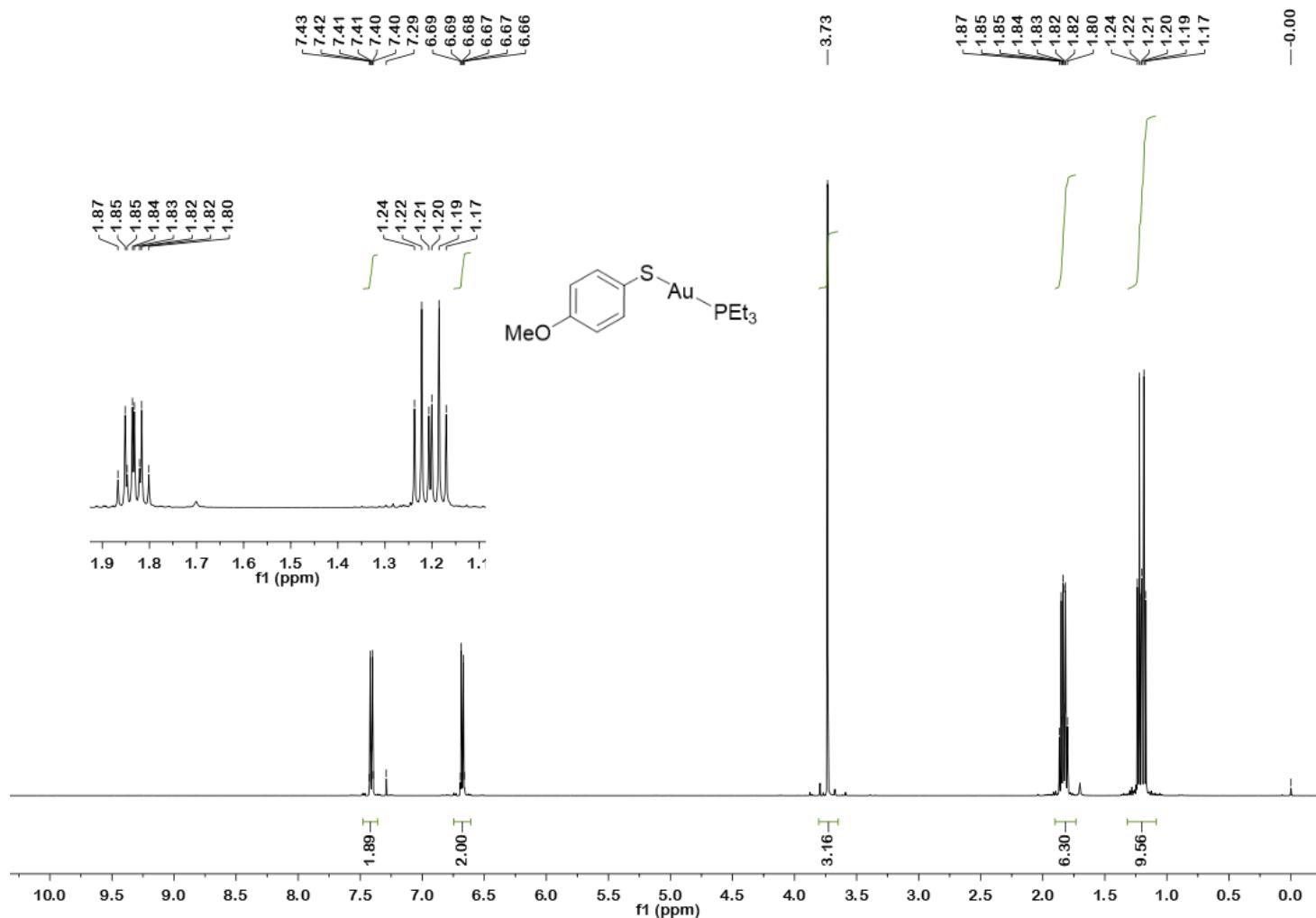


Figure S68. ^1H NMR spectrum of compound **18** in CDCl_3 .

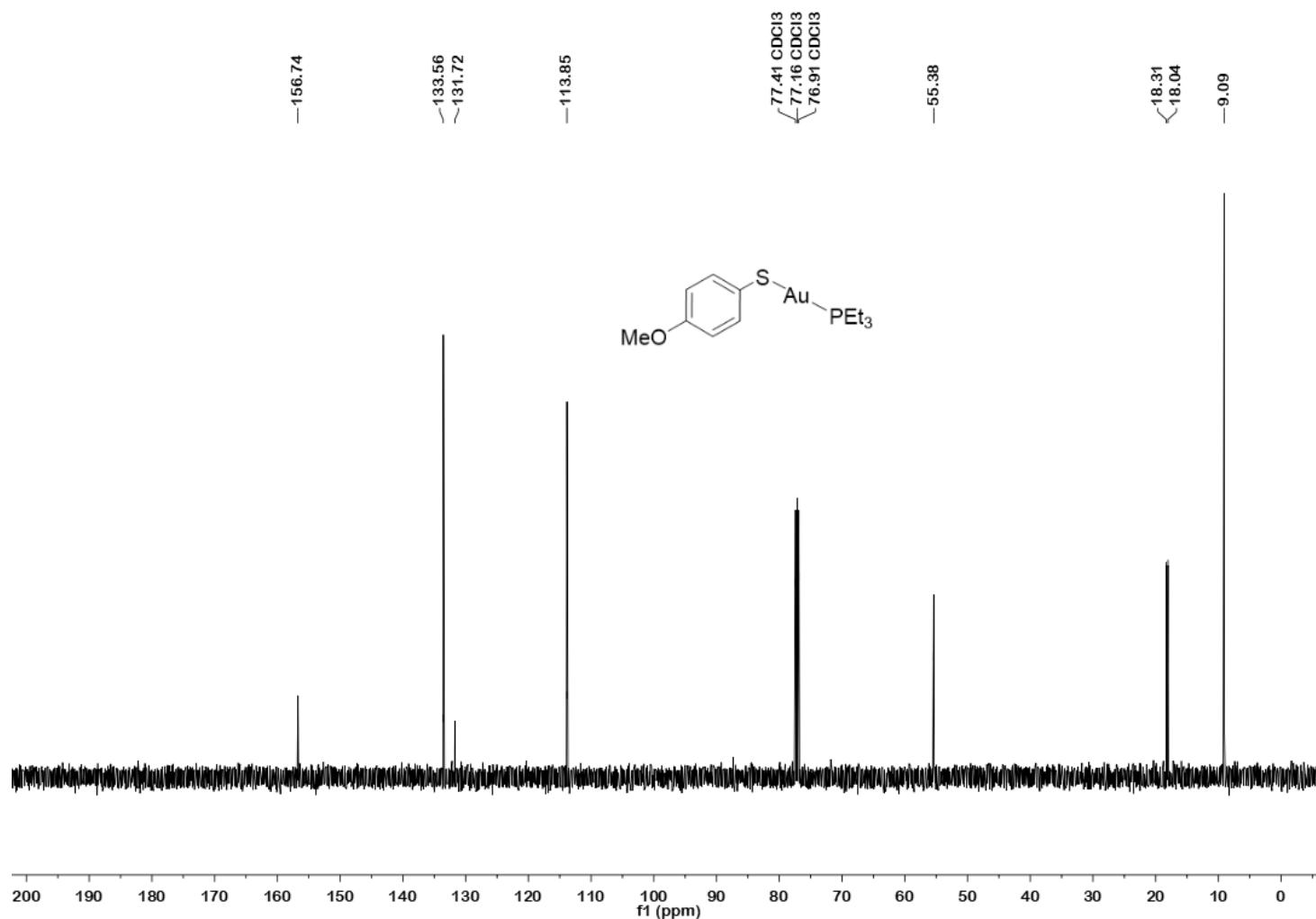


Figure S69. ^{13}C NMR spectrum of compound **18** in CDCl_3 .

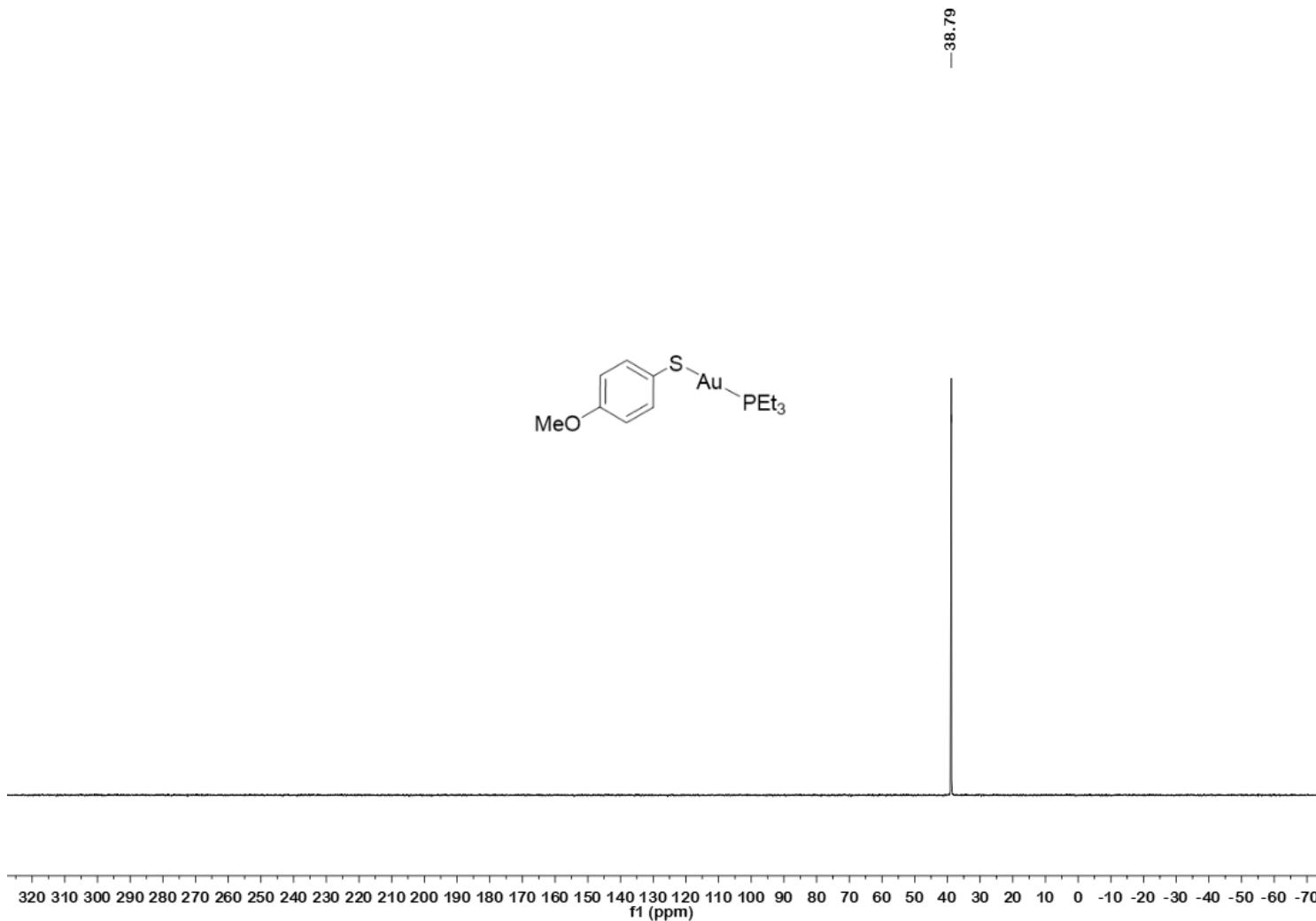


Figure S70. ^{31}P NMR spectrum of compound **18** in CDCl_3 .

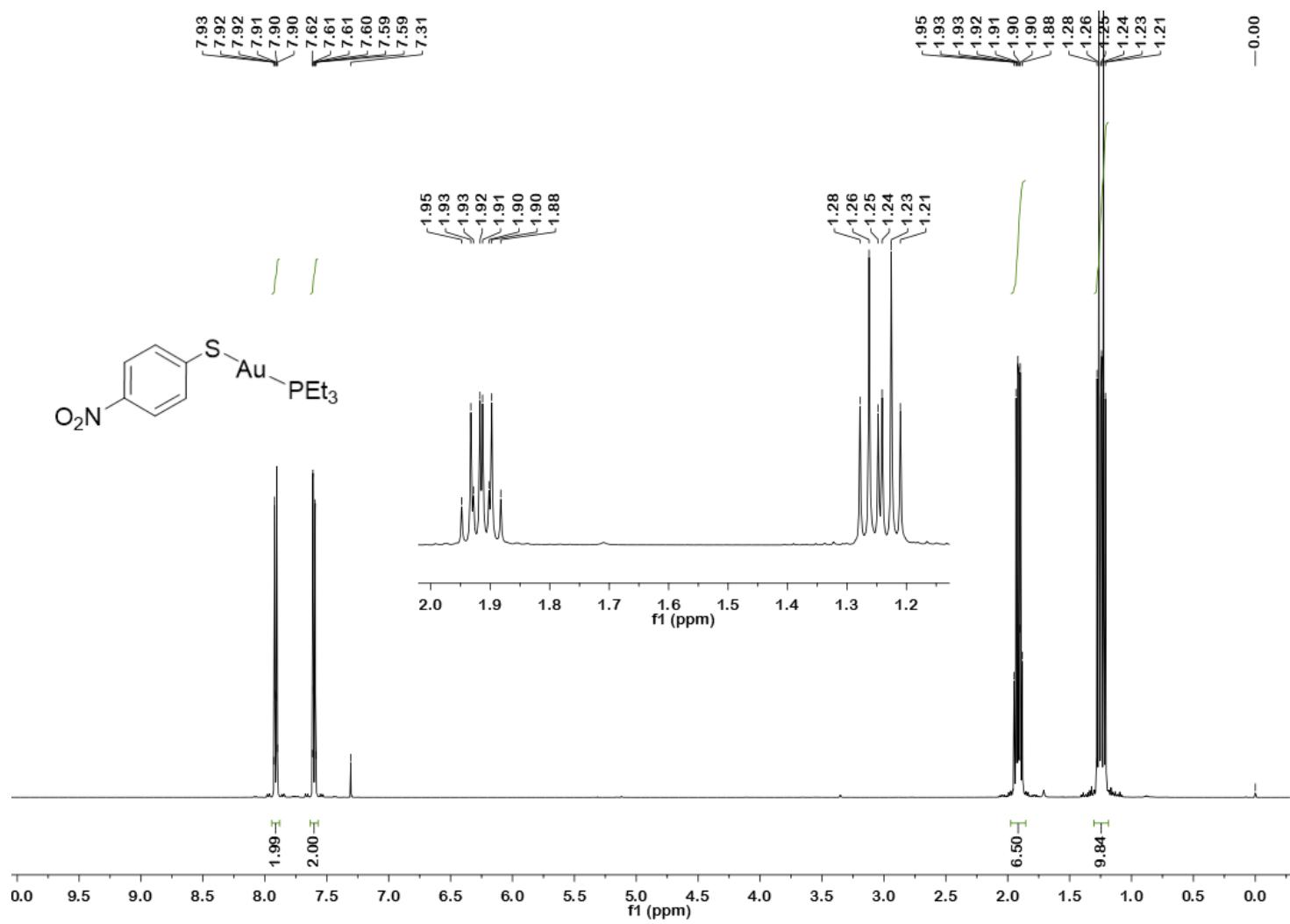


Figure S71. ^1H NMR spectrum of compound **19** in CDCl_3 .

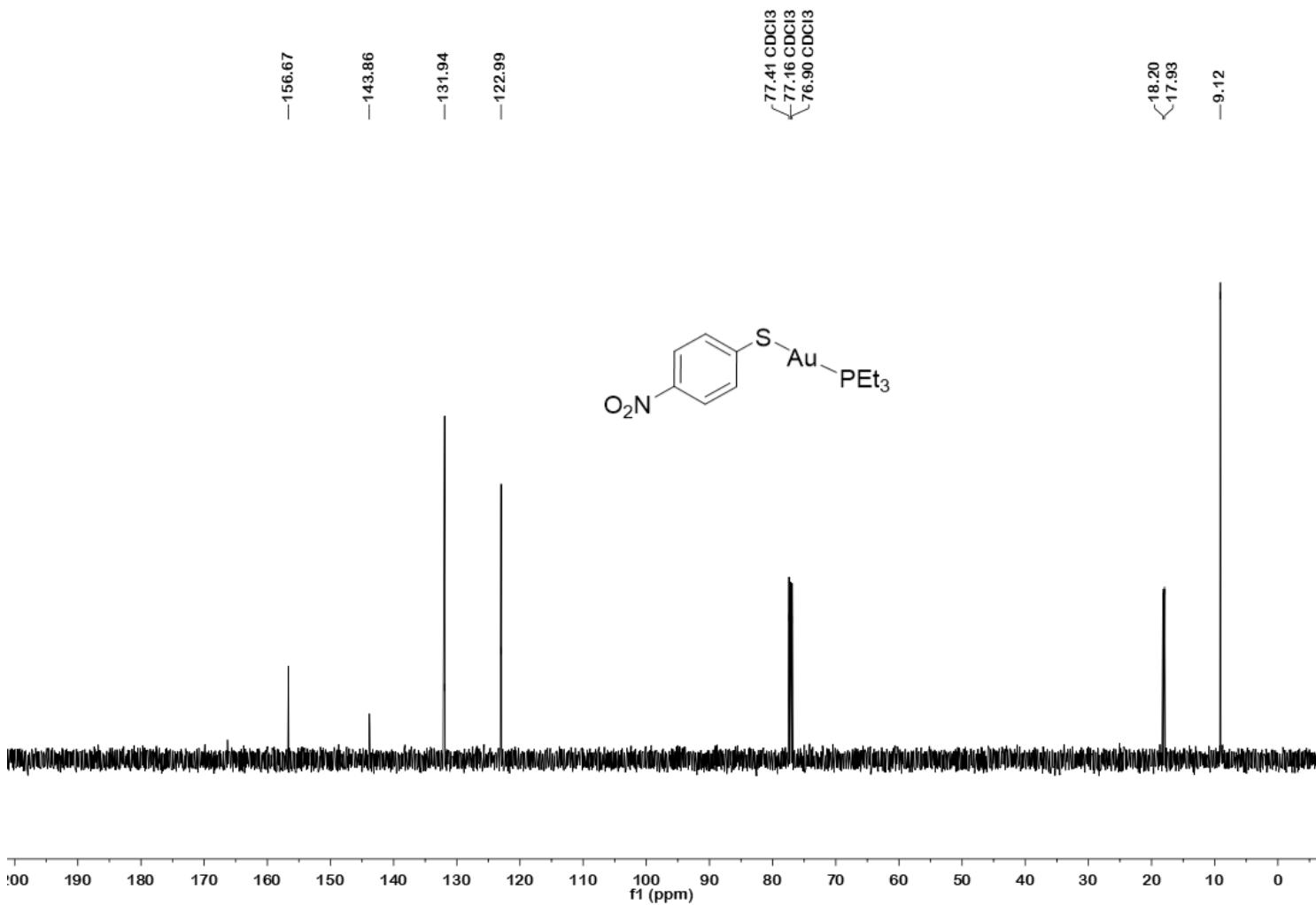


Figure S72. ^{13}C NMR spectrum of compound **19** in CDCl_3 .

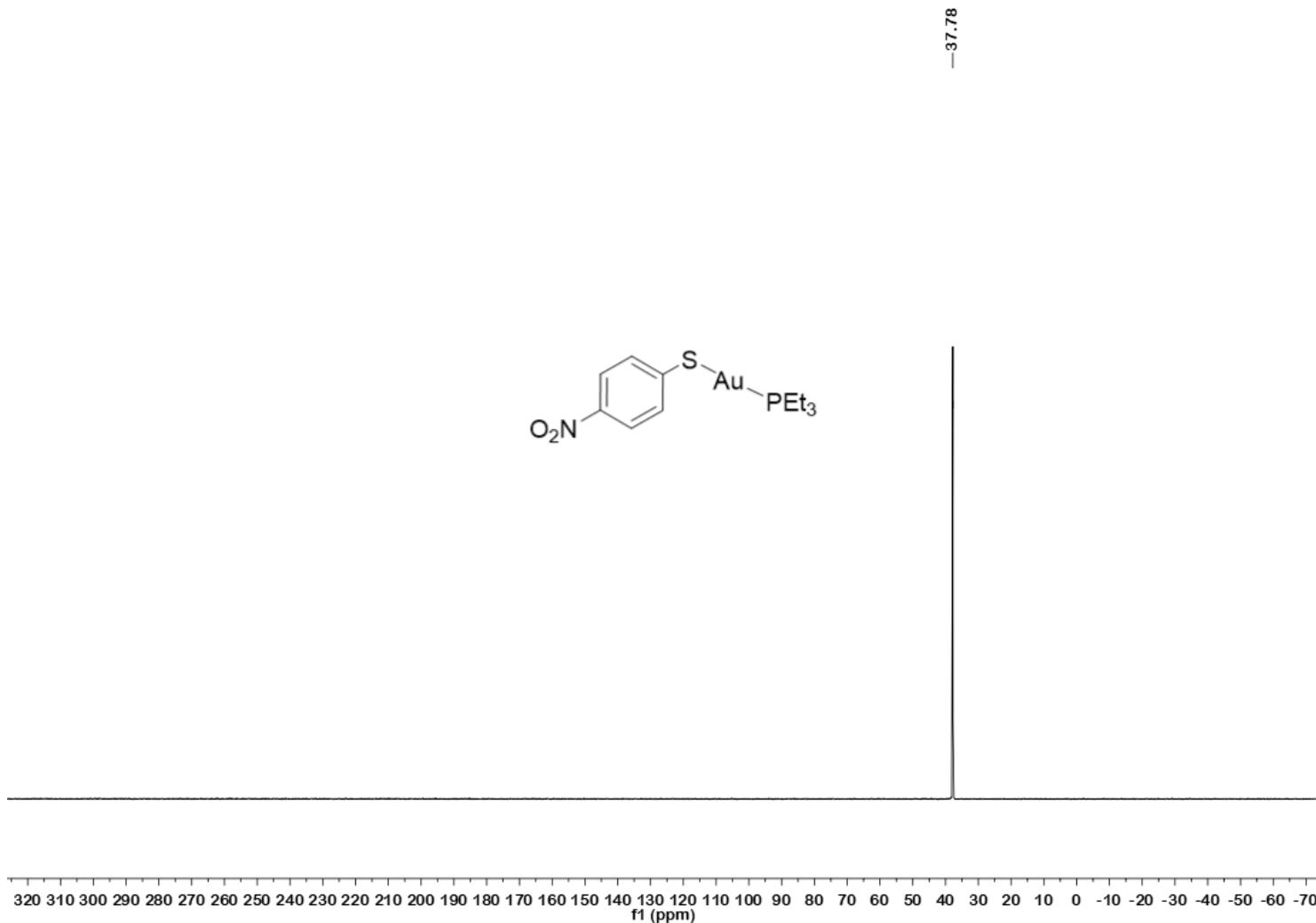


Figure S73. ^{31}P NMR spectrum of compound **19** in CDCl_3 .

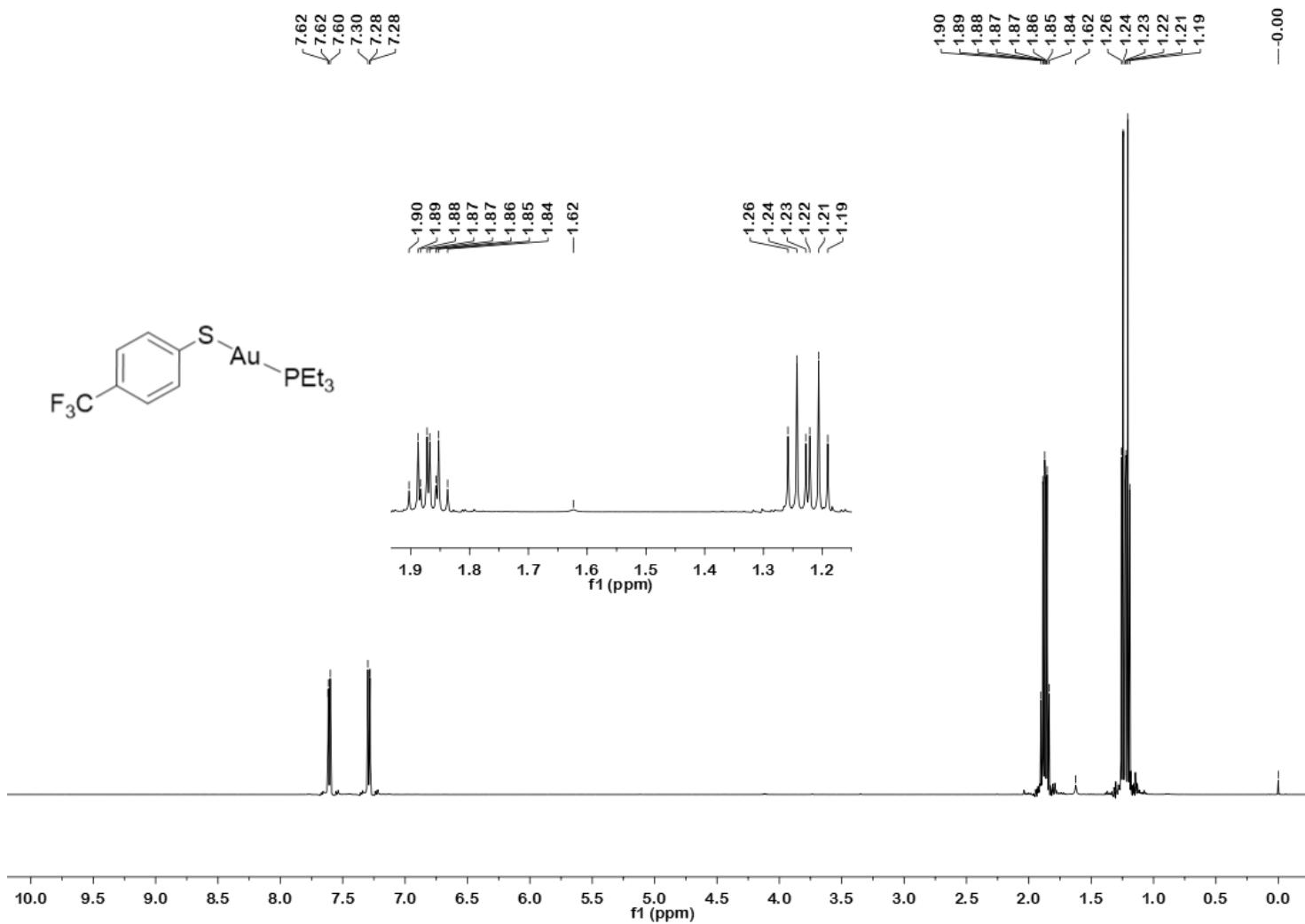
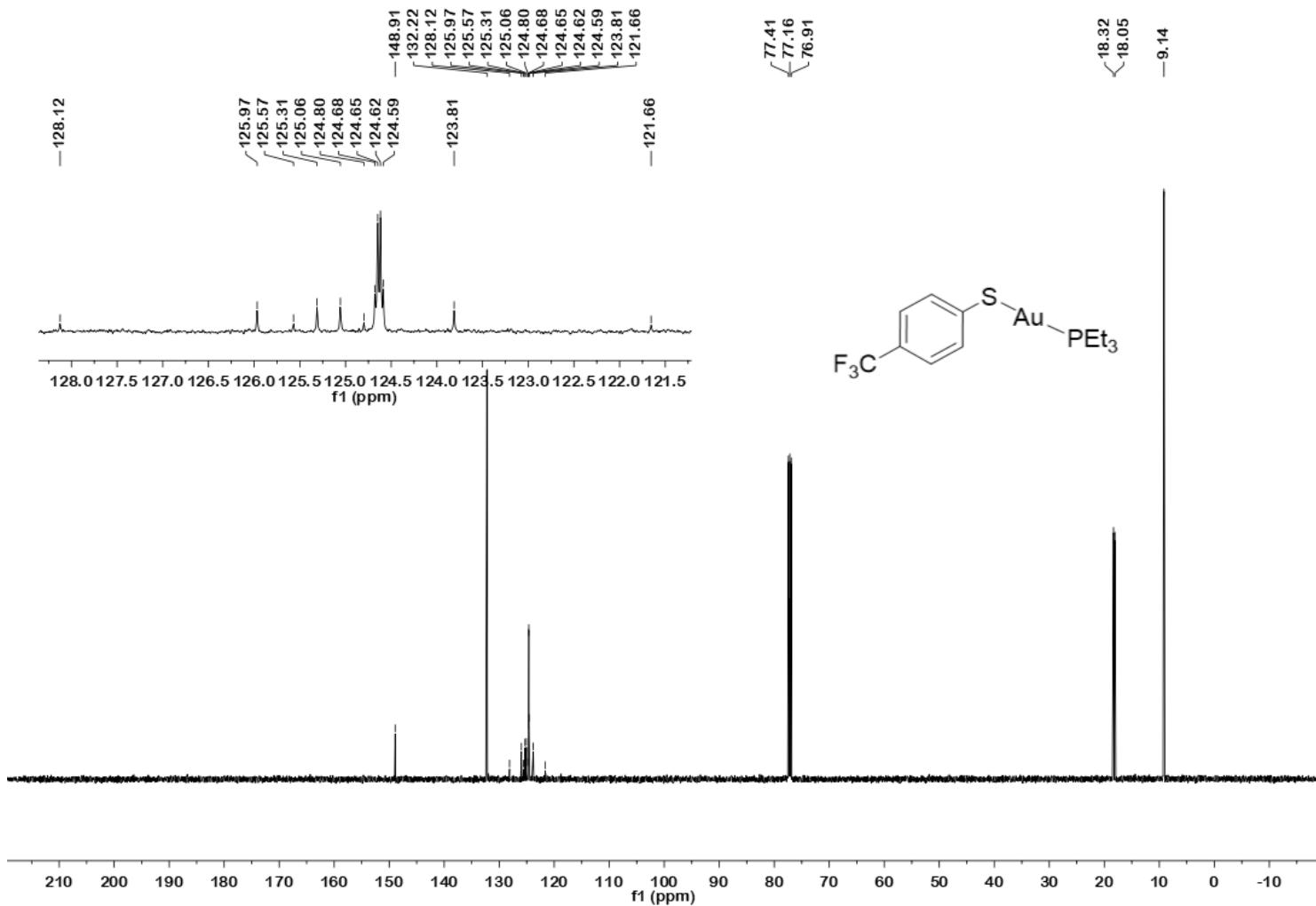


Figure S74. ^1H NMR spectrum of compound **20** in CDCl_3 .



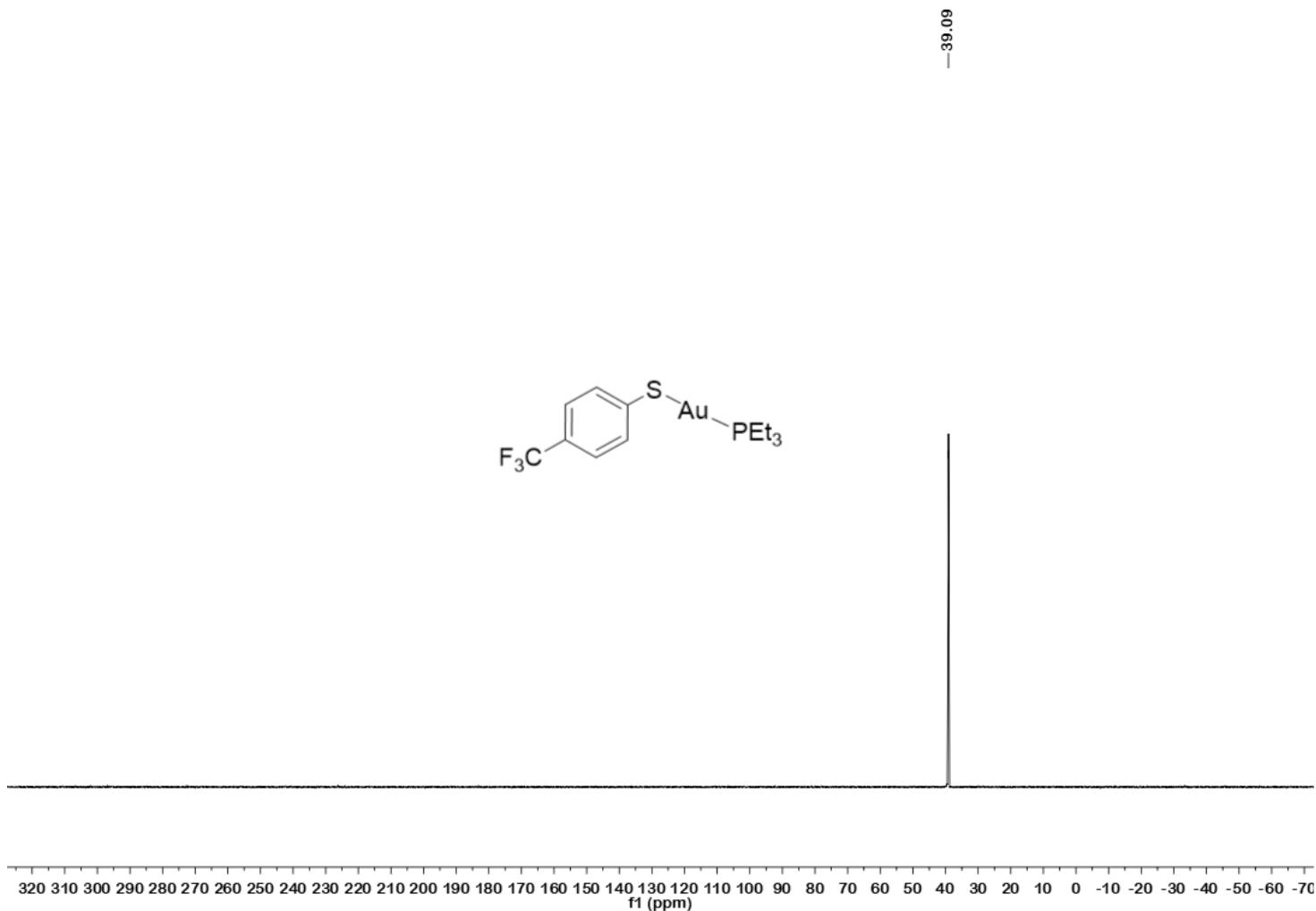


Figure S76. ^{31}P NMR spectrum of compound **20** in CDCl_3 .

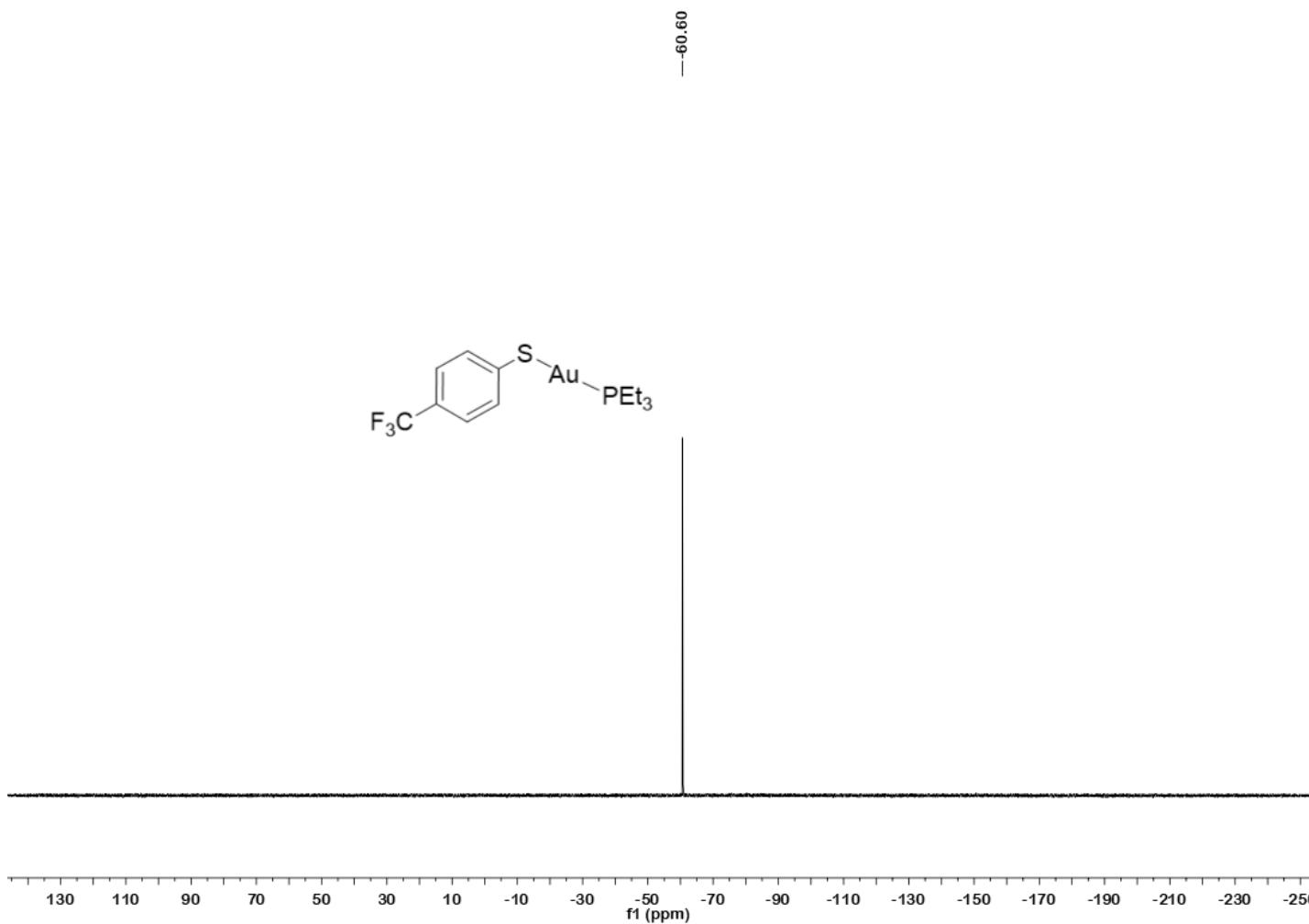


Figure S77. ^{19}F NMR spectrum of compound **20** in CDCl_3 .

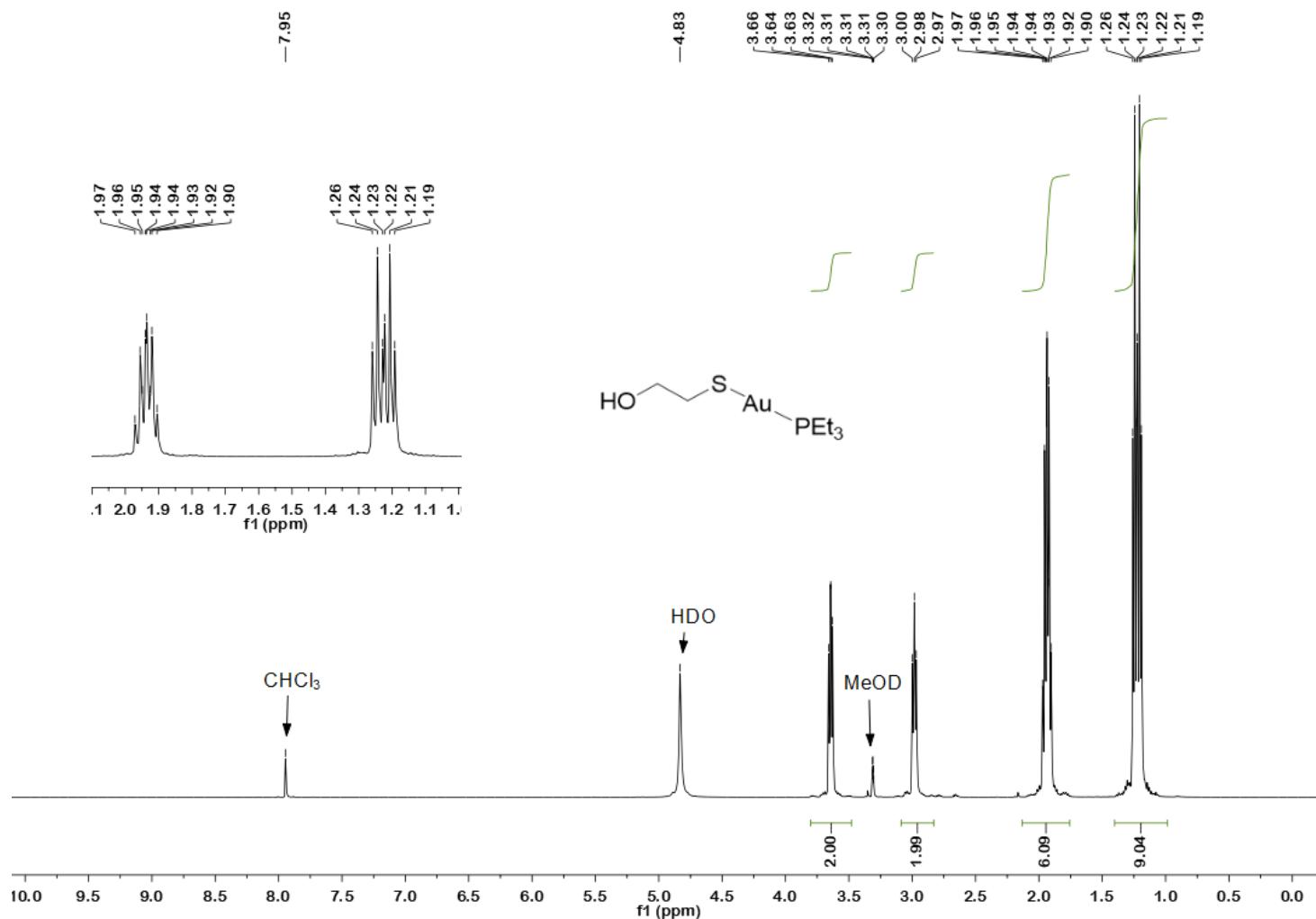


Figure S78. ^1H NMR spectrum of compound **21** in CD_3OD .

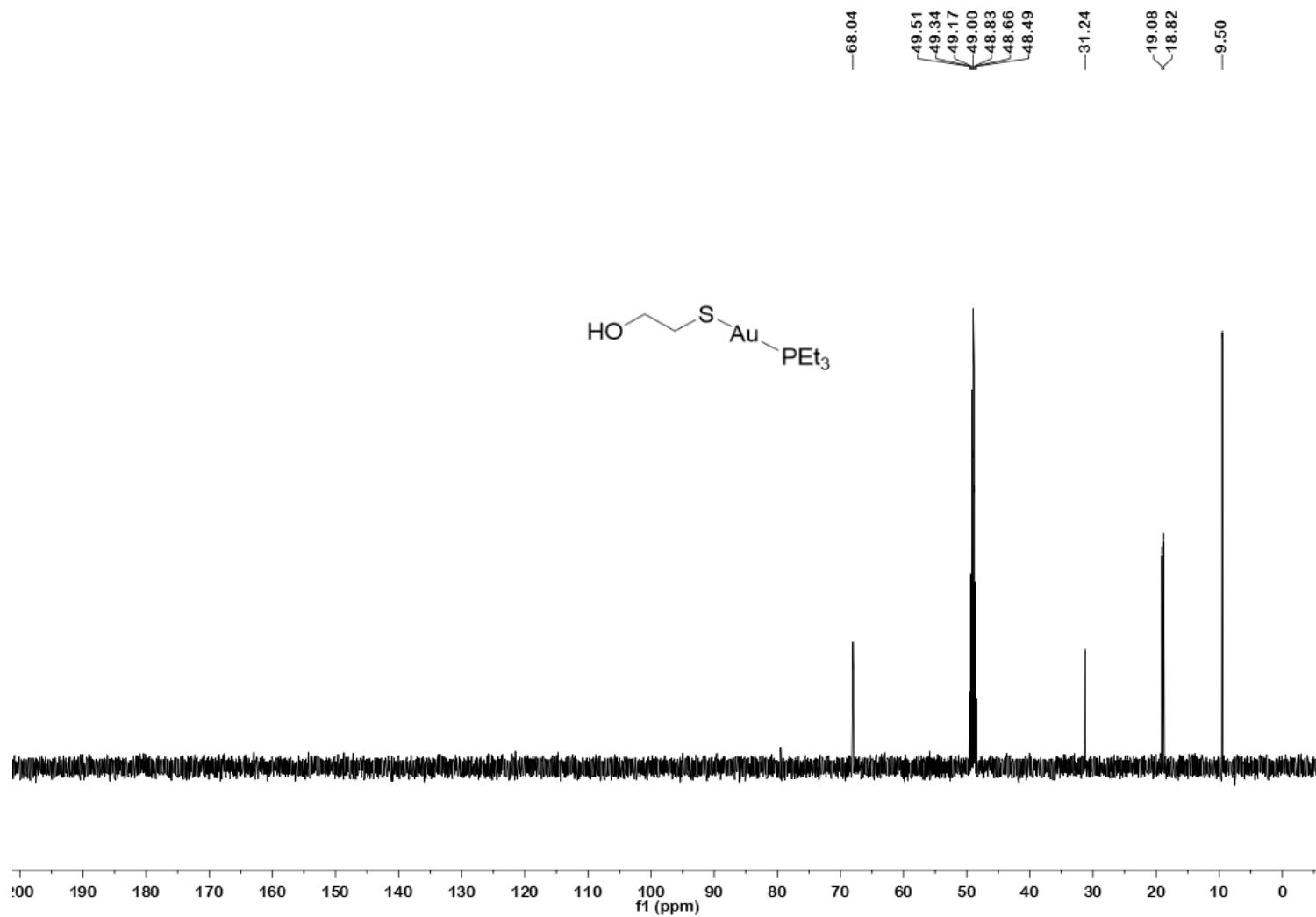


Figure S79. ^{13}C NMR spectrum of compound **21** in CD_3OD .

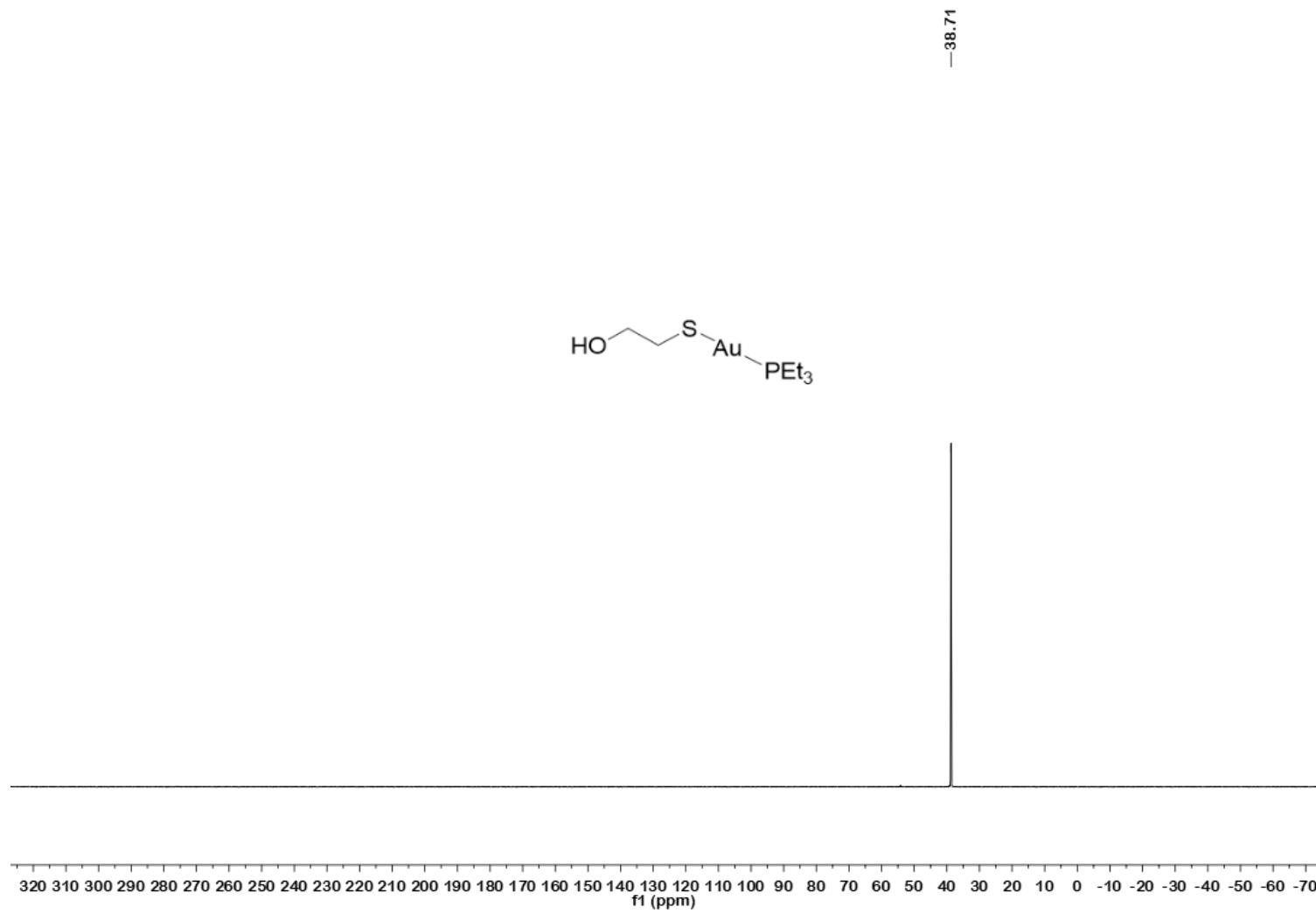


Figure S80. ^{31}P NMR spectrum of compound **21** in CD_3OD .

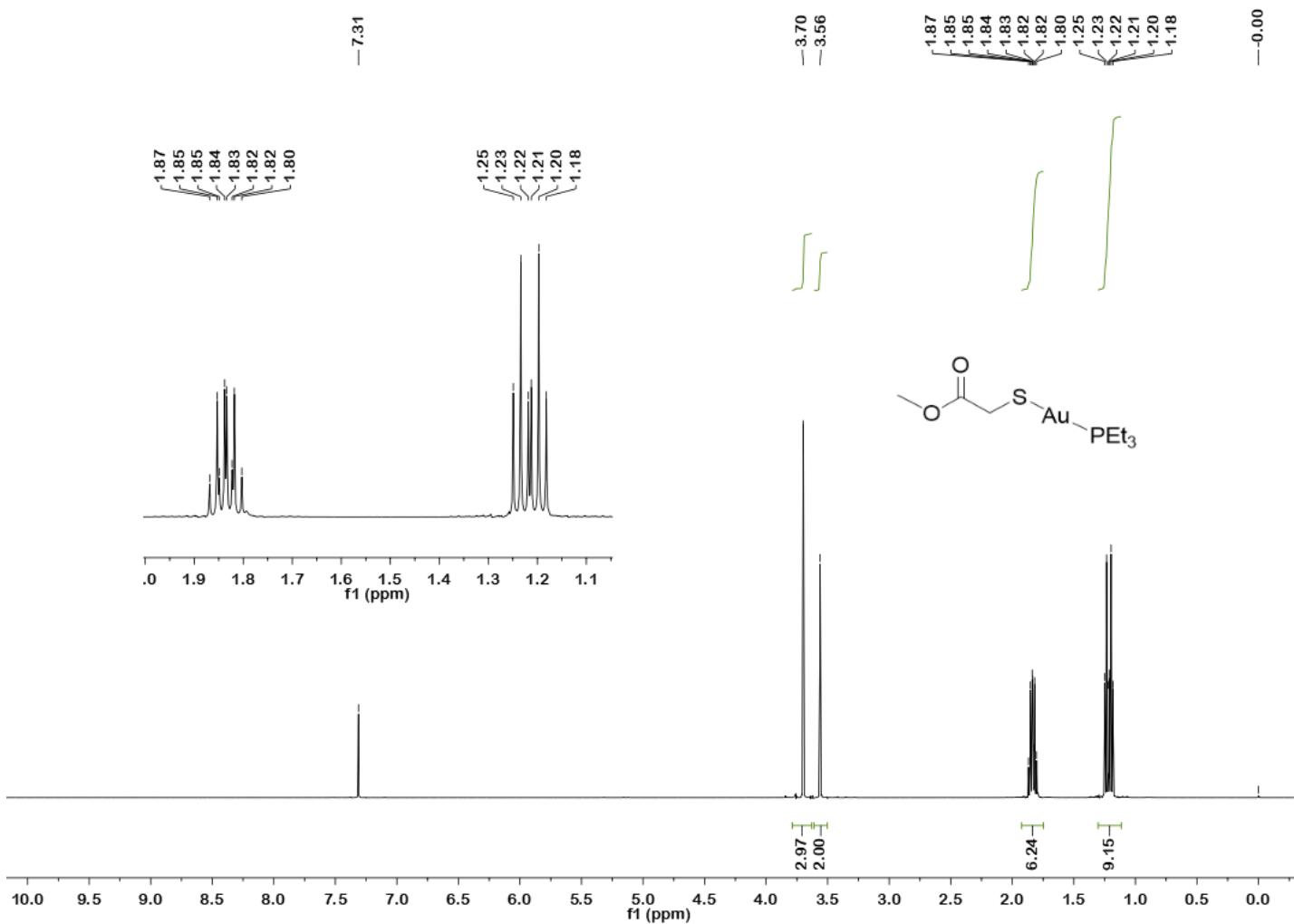


Figure S81. ^1H NMR spectrum of compound **22** in CDCl_3 .

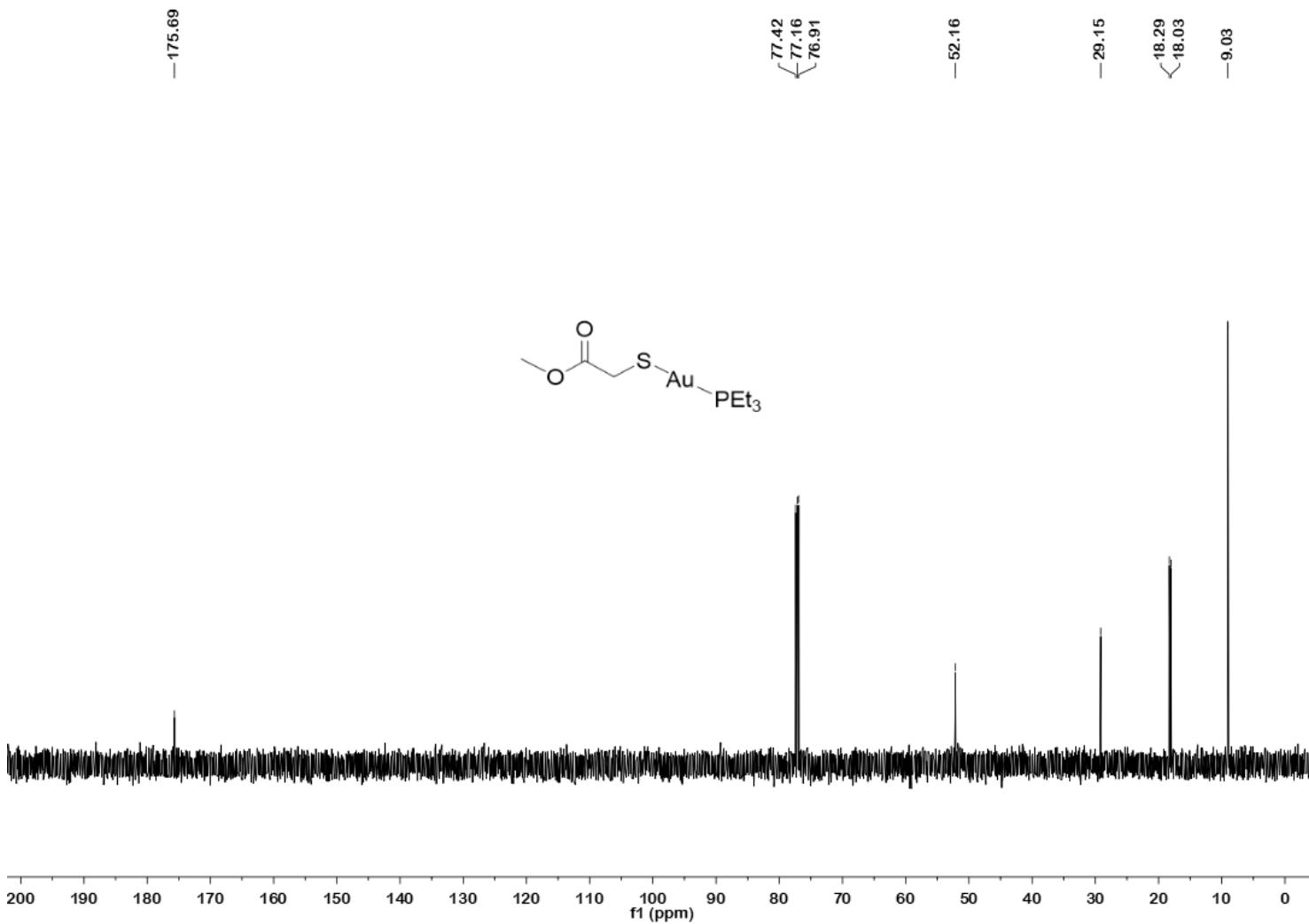


Figure S82. ^{13}C NMR spectrum of compound **22** in CDCl_3 .

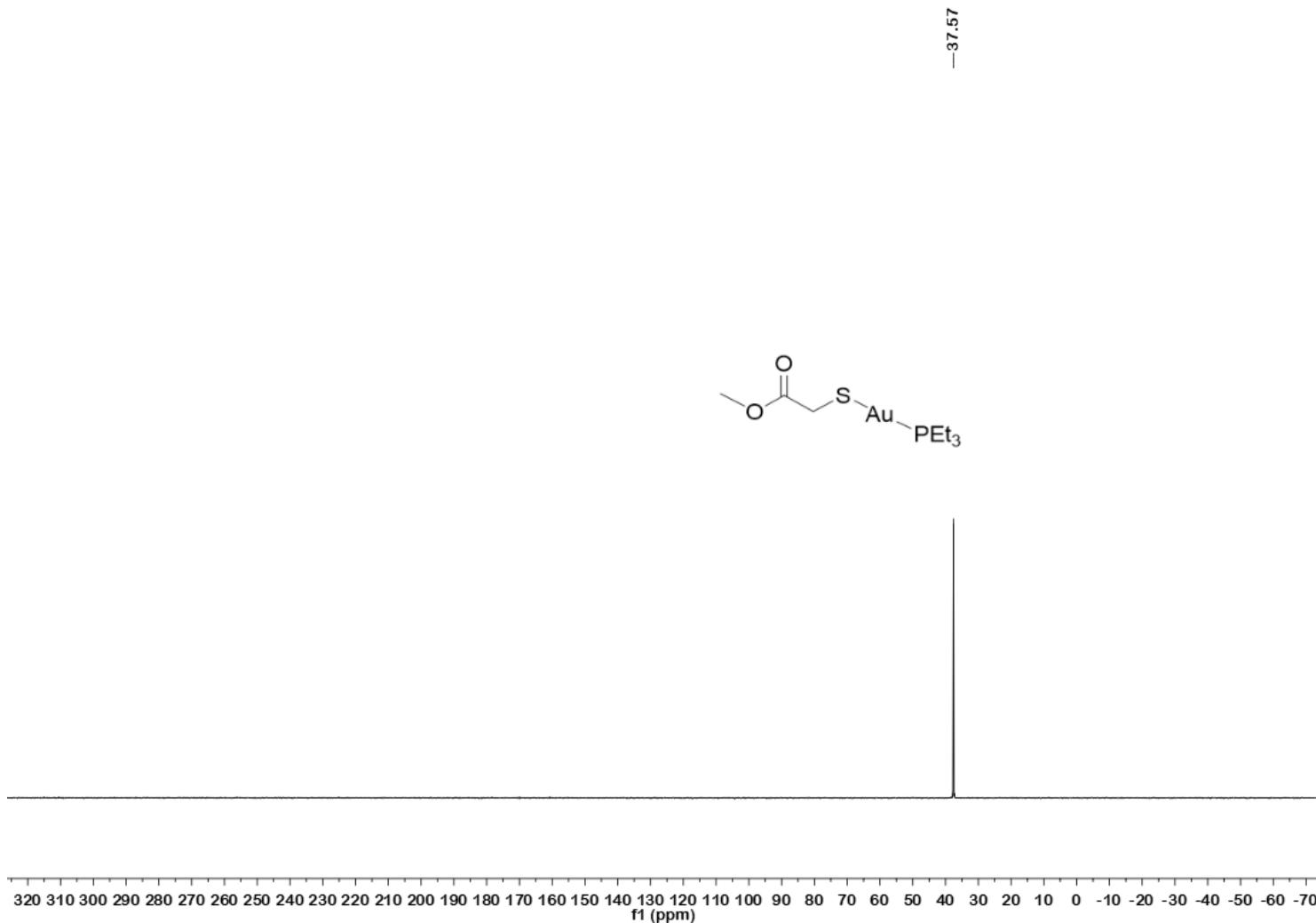


Figure S83. ^{31}P NMR spectrum of compound **22** in CDCl_3 .

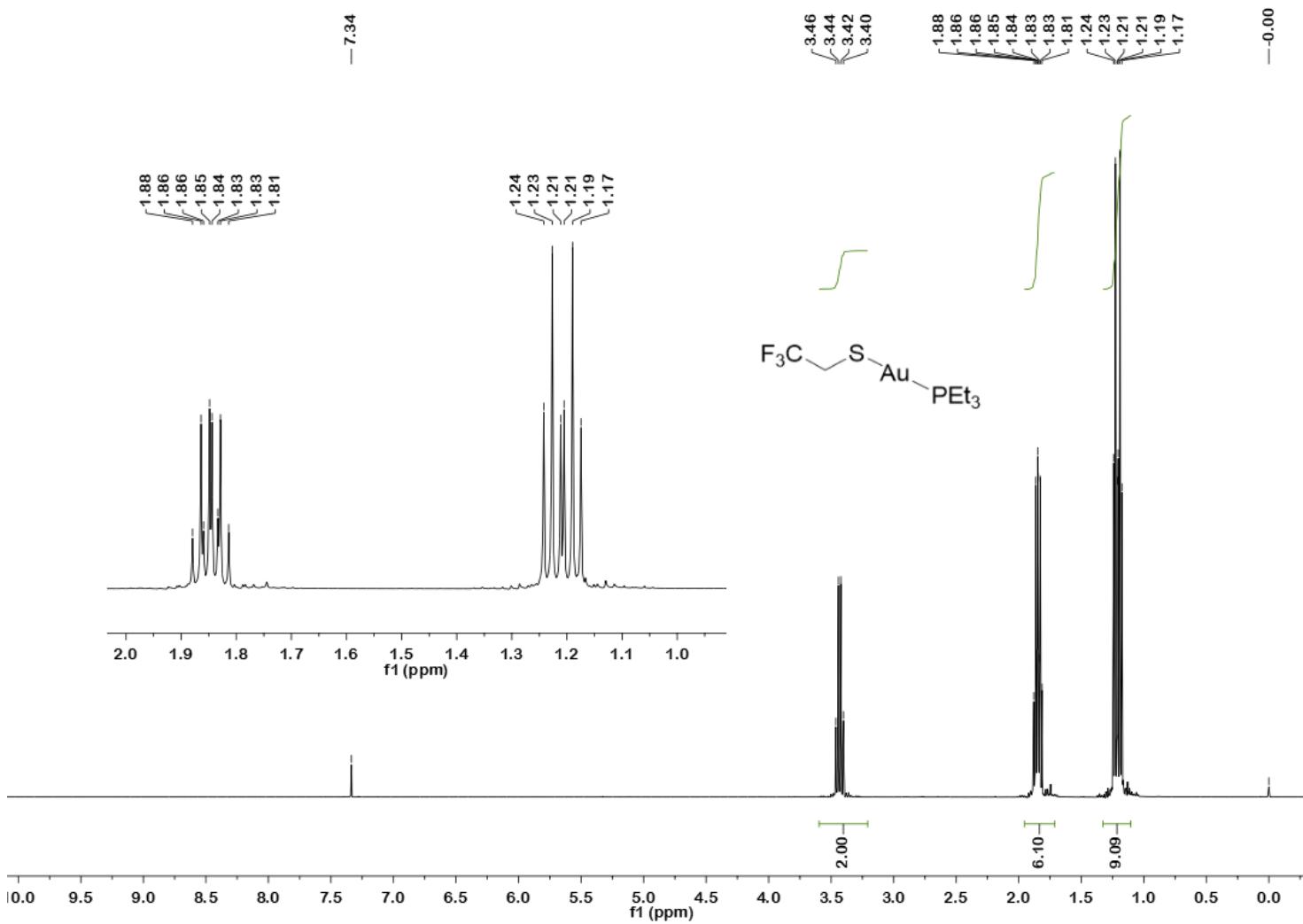


Figure S84. ^1H NMR spectrum of compound **23** in CDCl_3 .

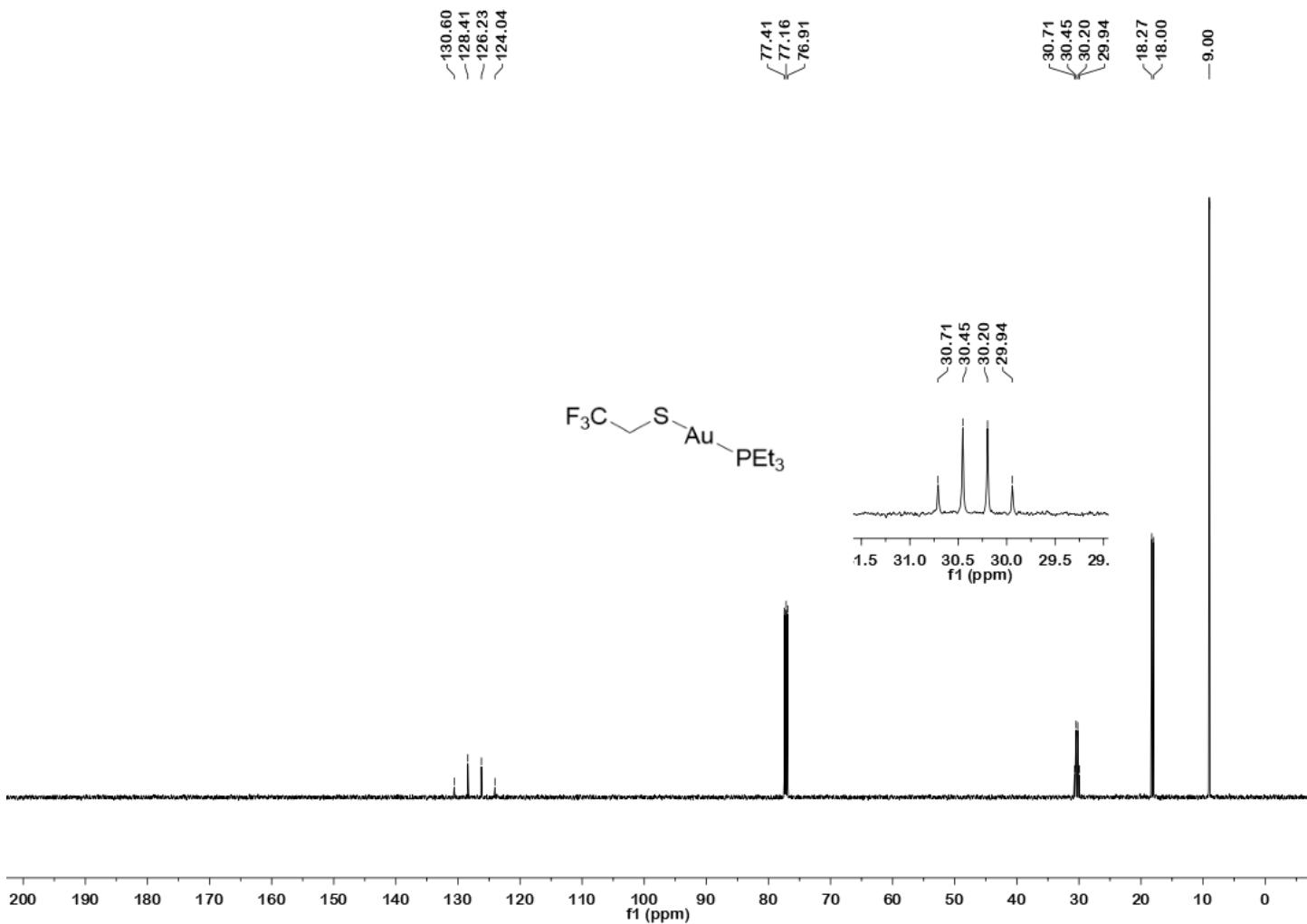


Figure S85. ^{13}C NMR spectrum of compound **23** in CDCl_3 .

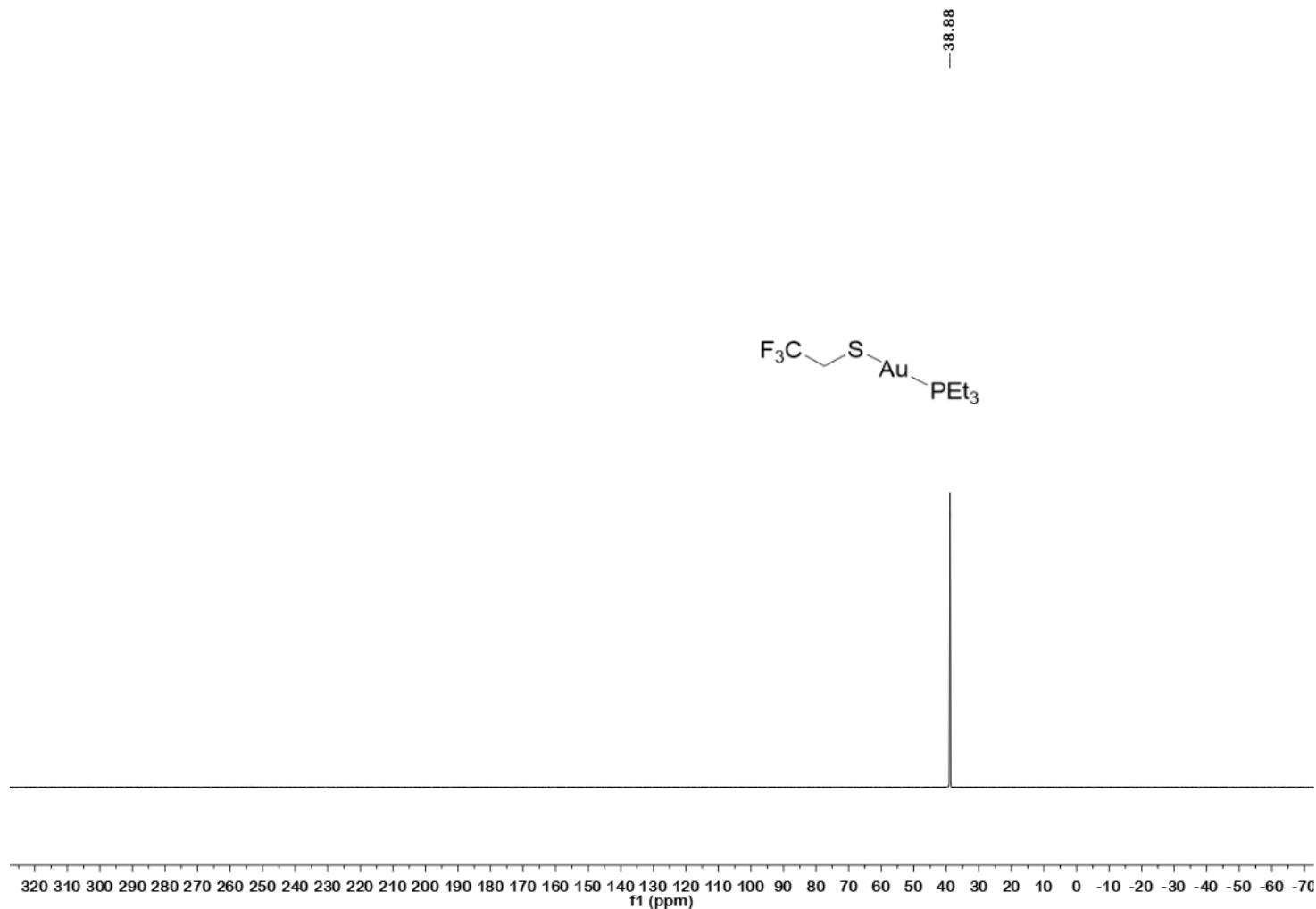


Figure S86. ${}^3\text{P}$ NMR spectrum of compound 23 in CDCl_3 .

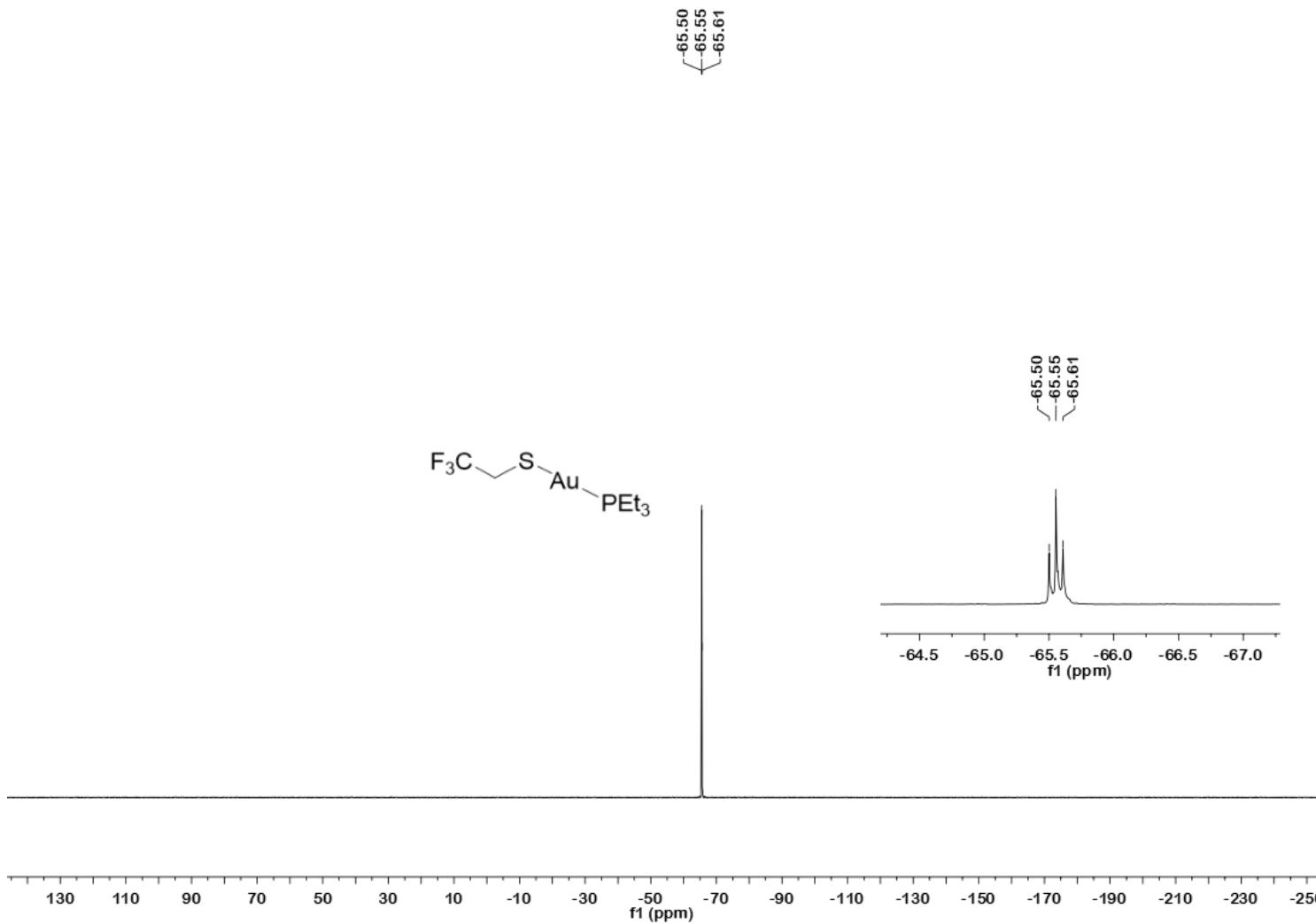


Figure S87. ^{19}F NMR spectrum of compound **23** in CDCl_3 .

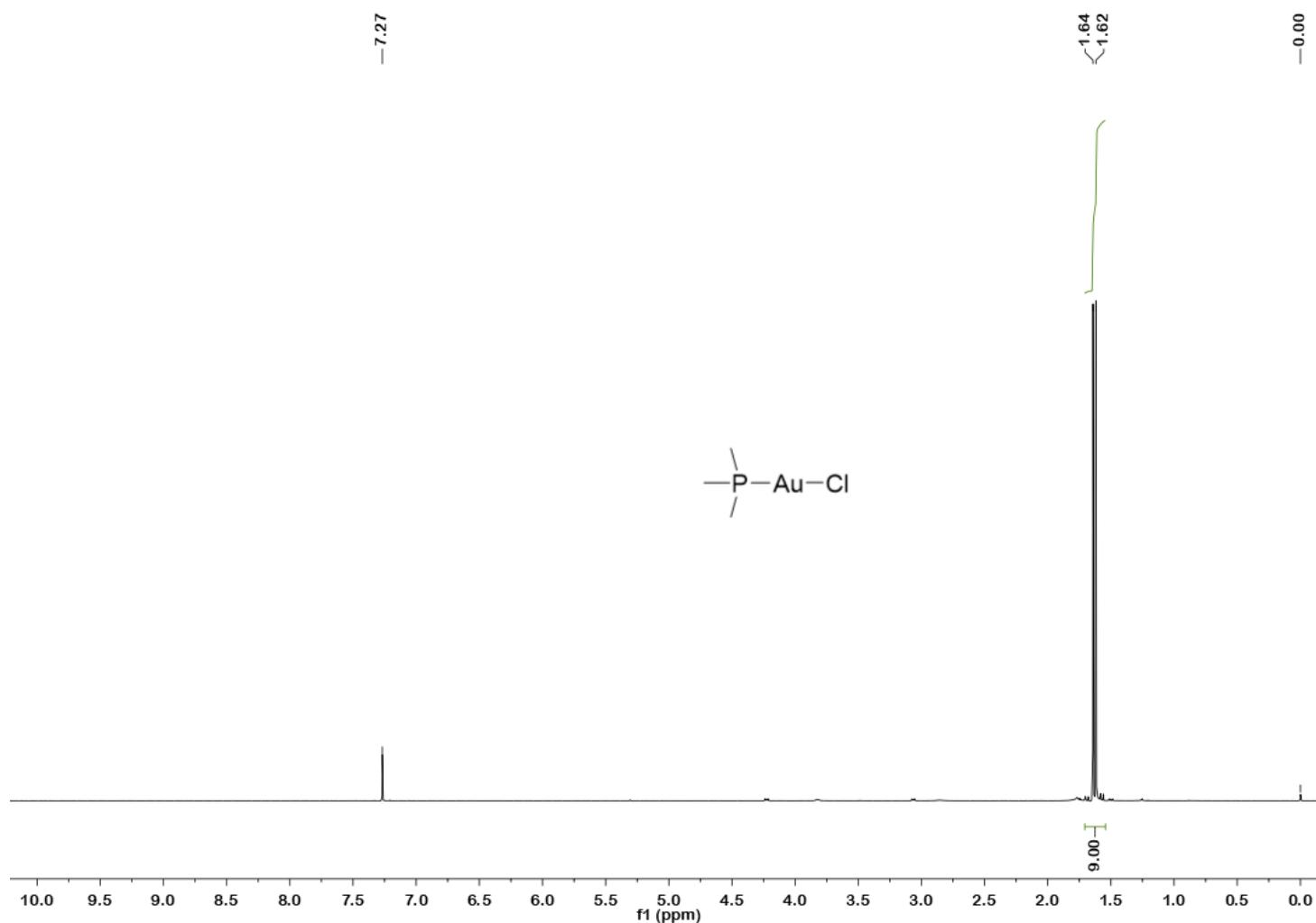


Figure S88. ^1H NMR spectrum of compound **24** in CDCl_3 .

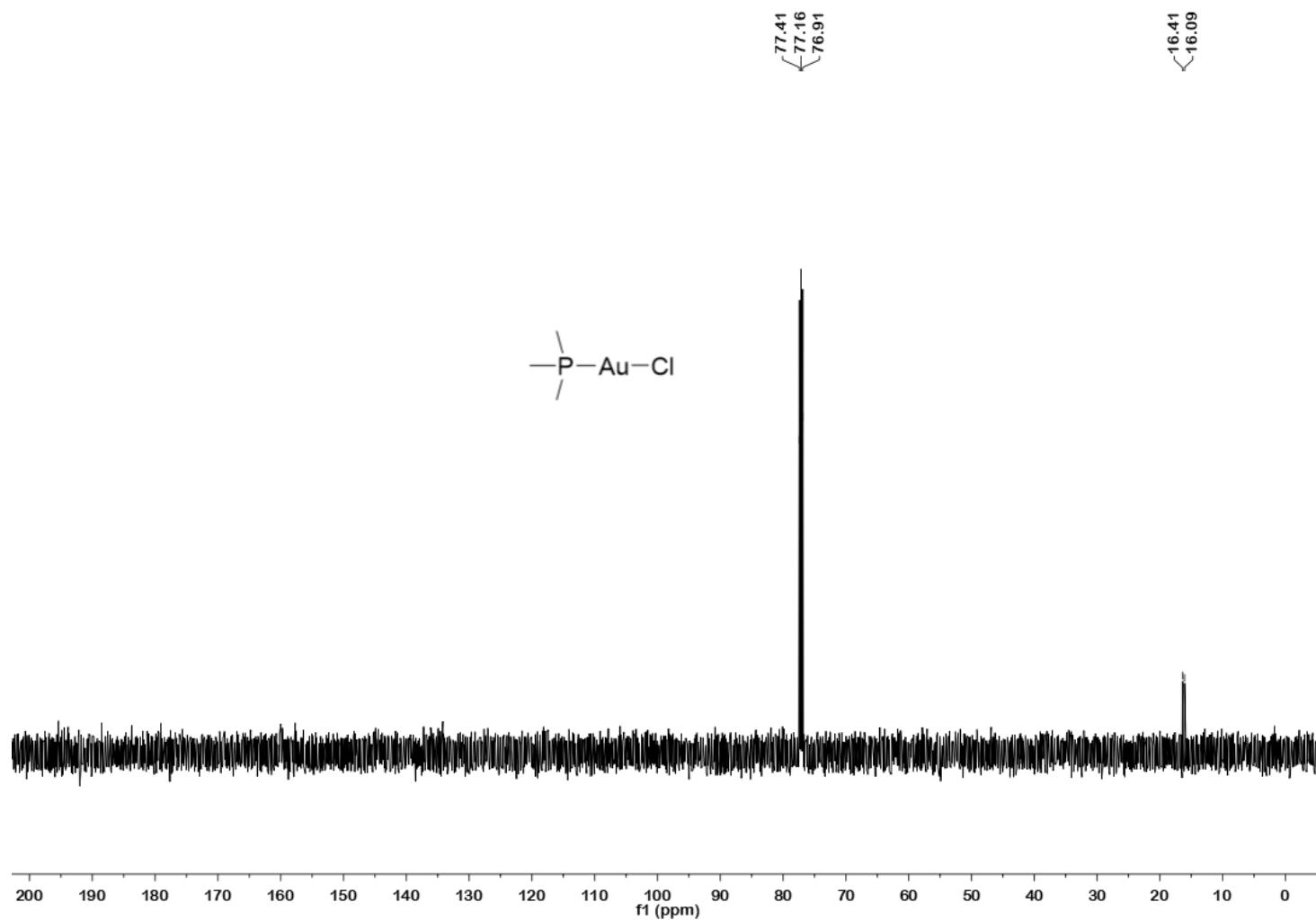


Figure S89. ^{13}C NMR spectrum of compound 24 in CDCl_3 .

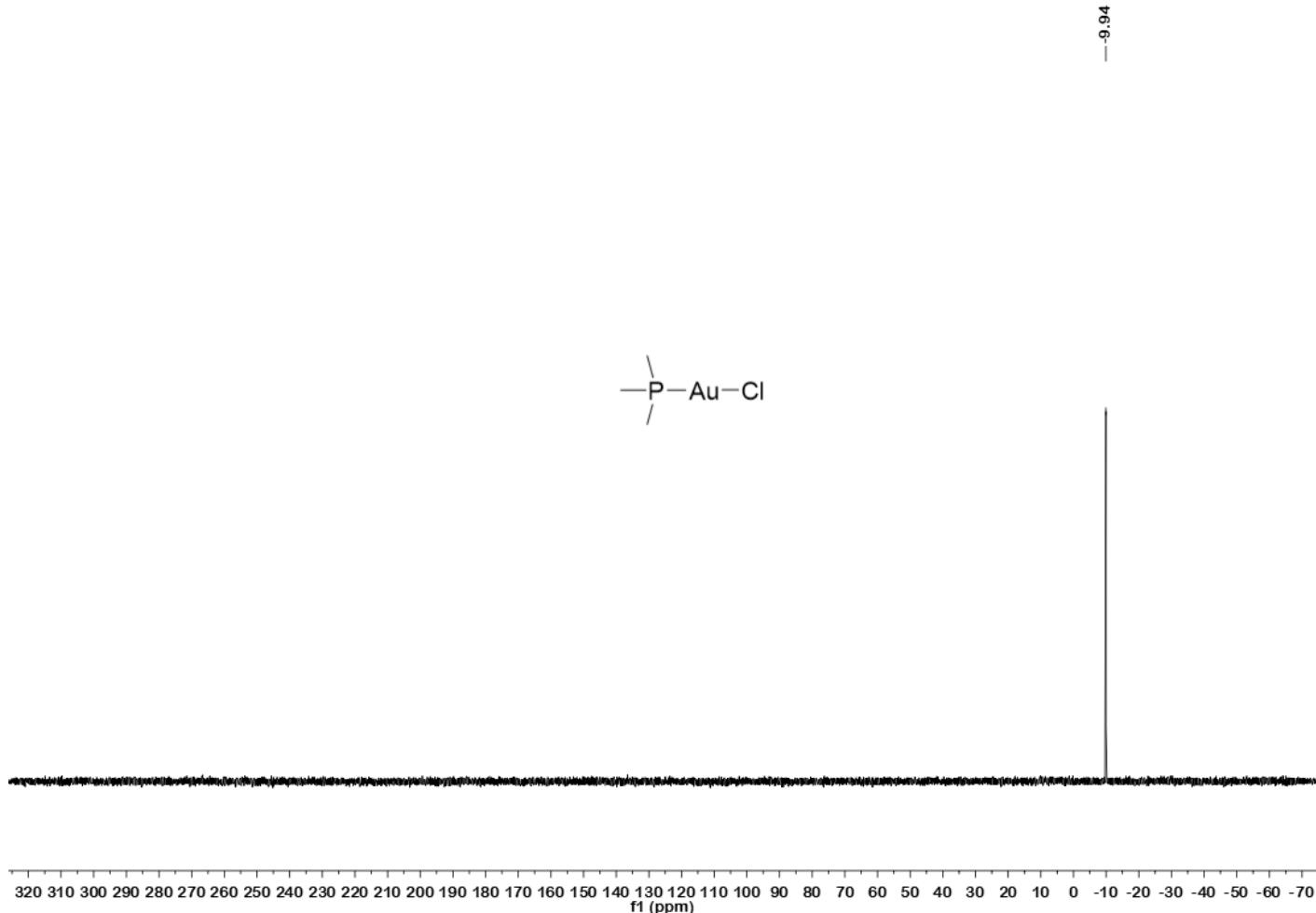


Figure S90. ^{31}P NMR spectrum of compound 24 in CDCl_3 .

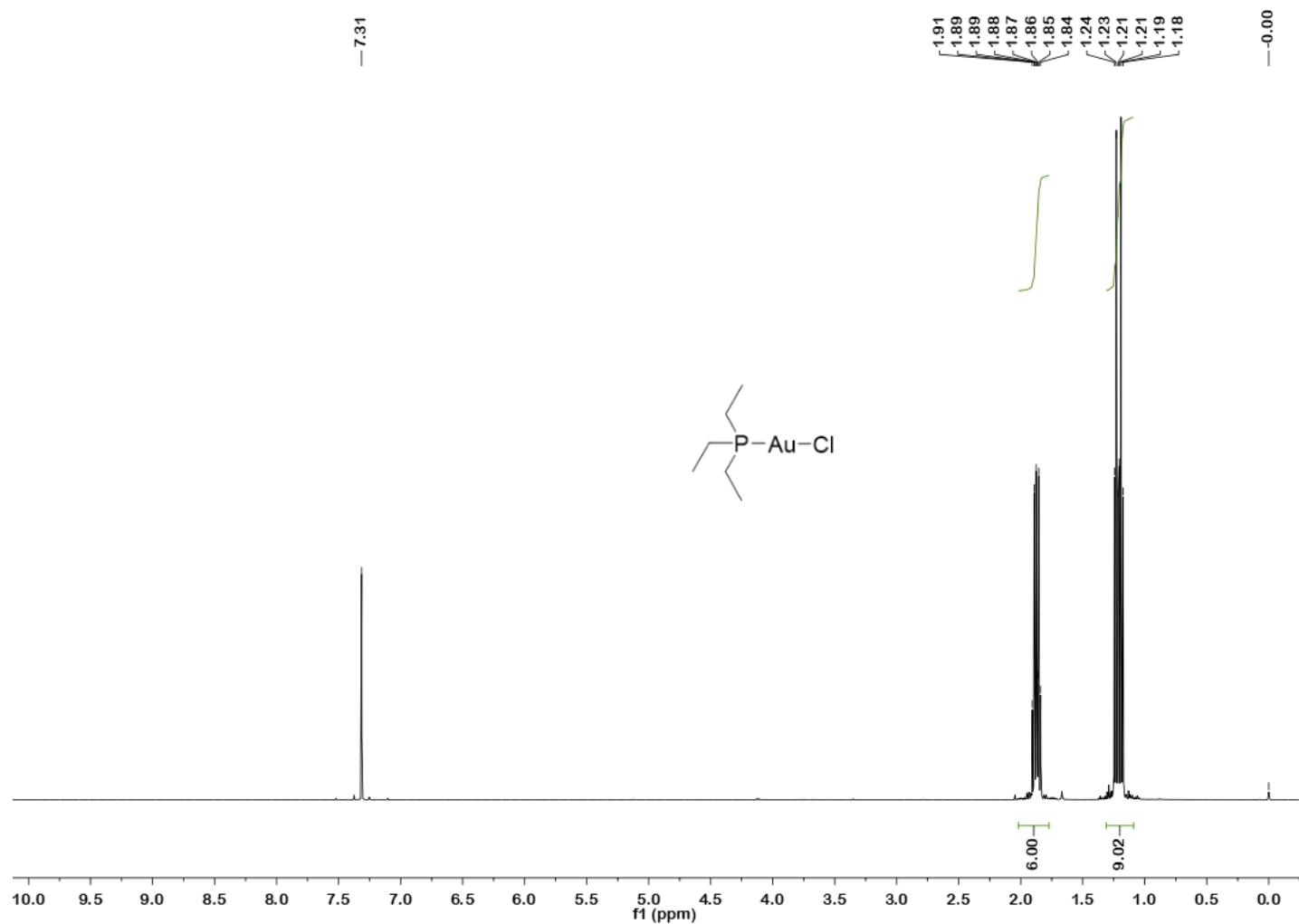


Figure S91. ^1H NMR spectrum of compound **25** in CDCl_3 .

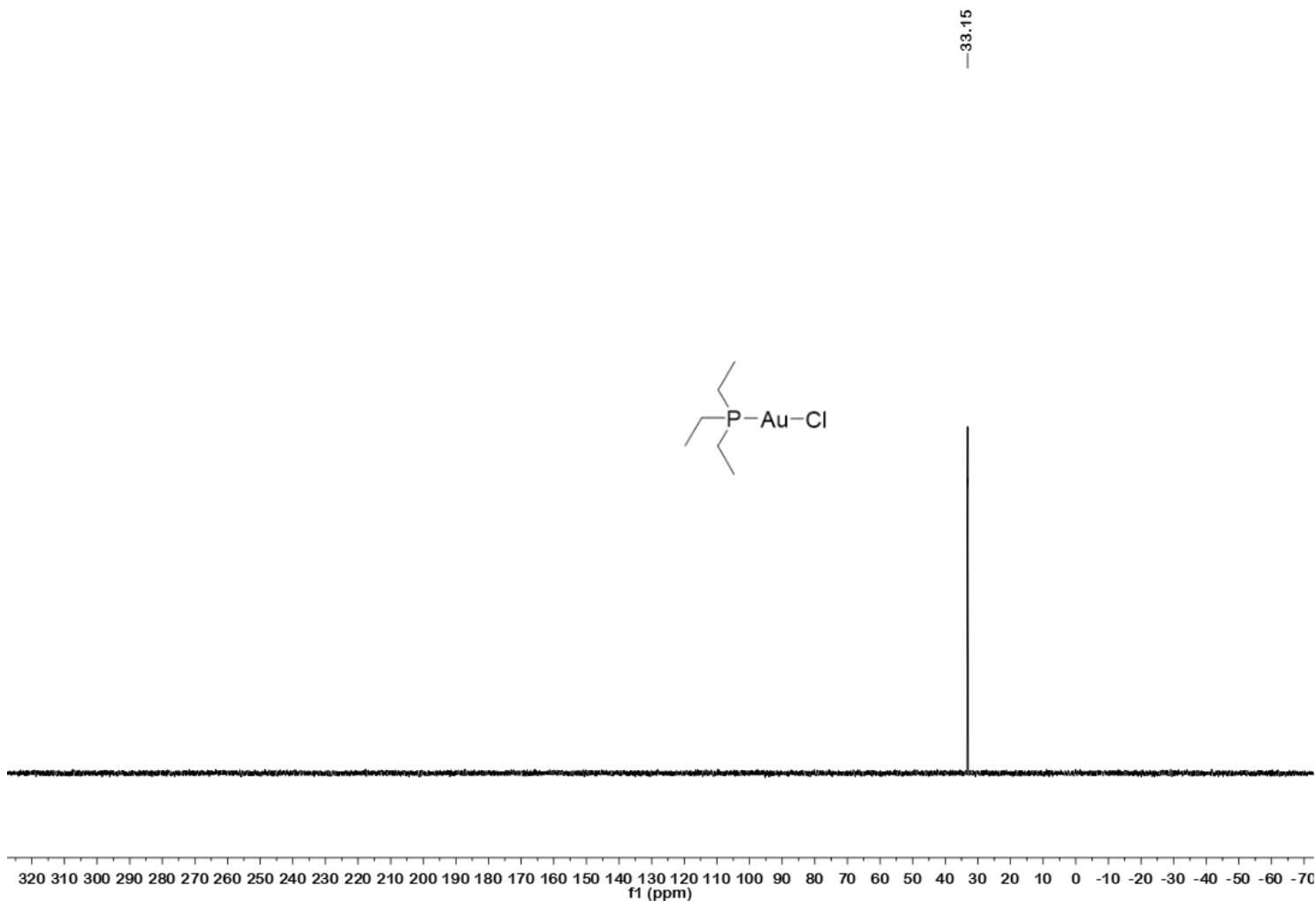


Figure S92. ^{31}P NMR spectrum of compound **25** in CDCl_3 .

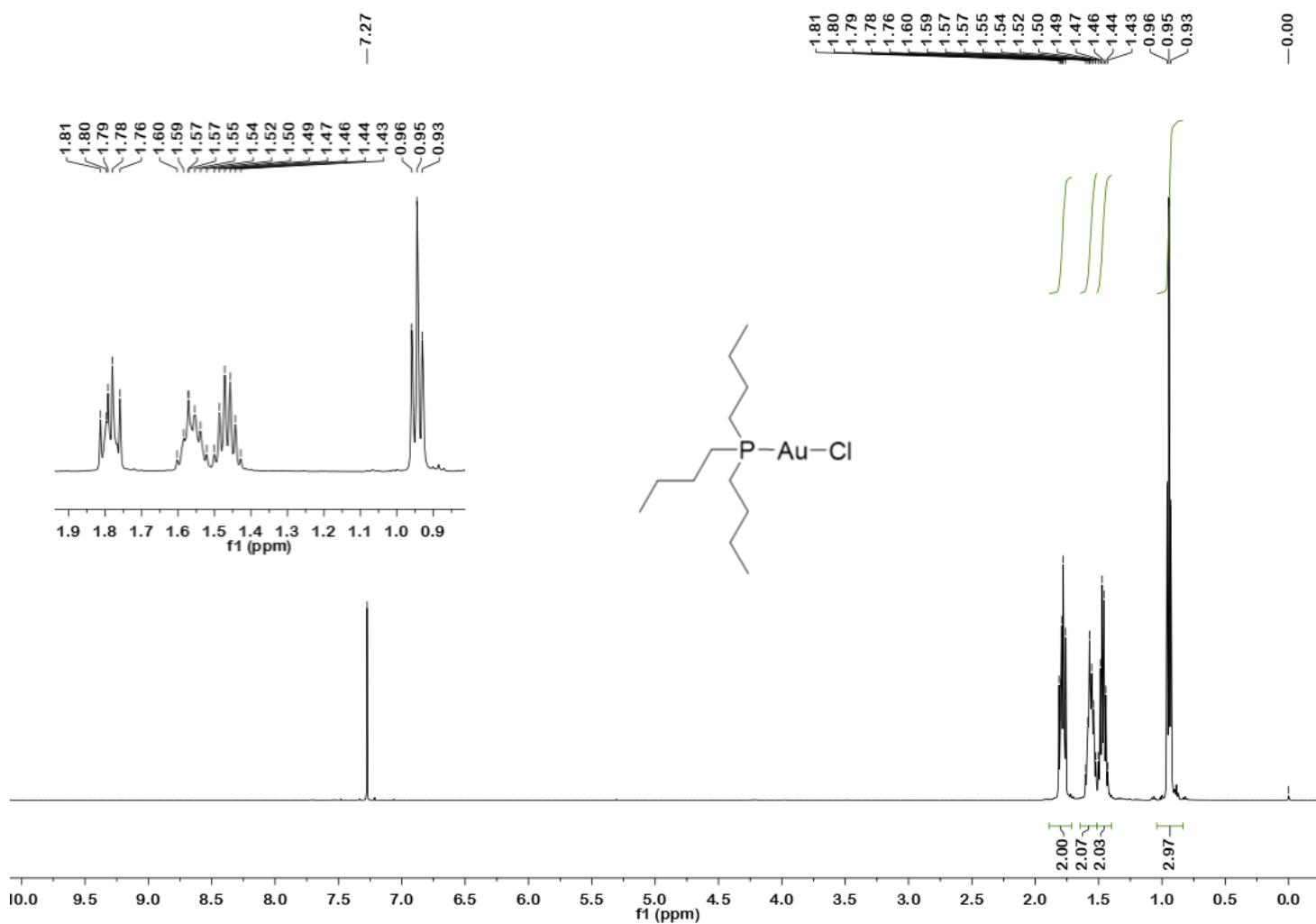


Figure S93. ^1H NMR spectrum of compound **26** in CDCl_3 .

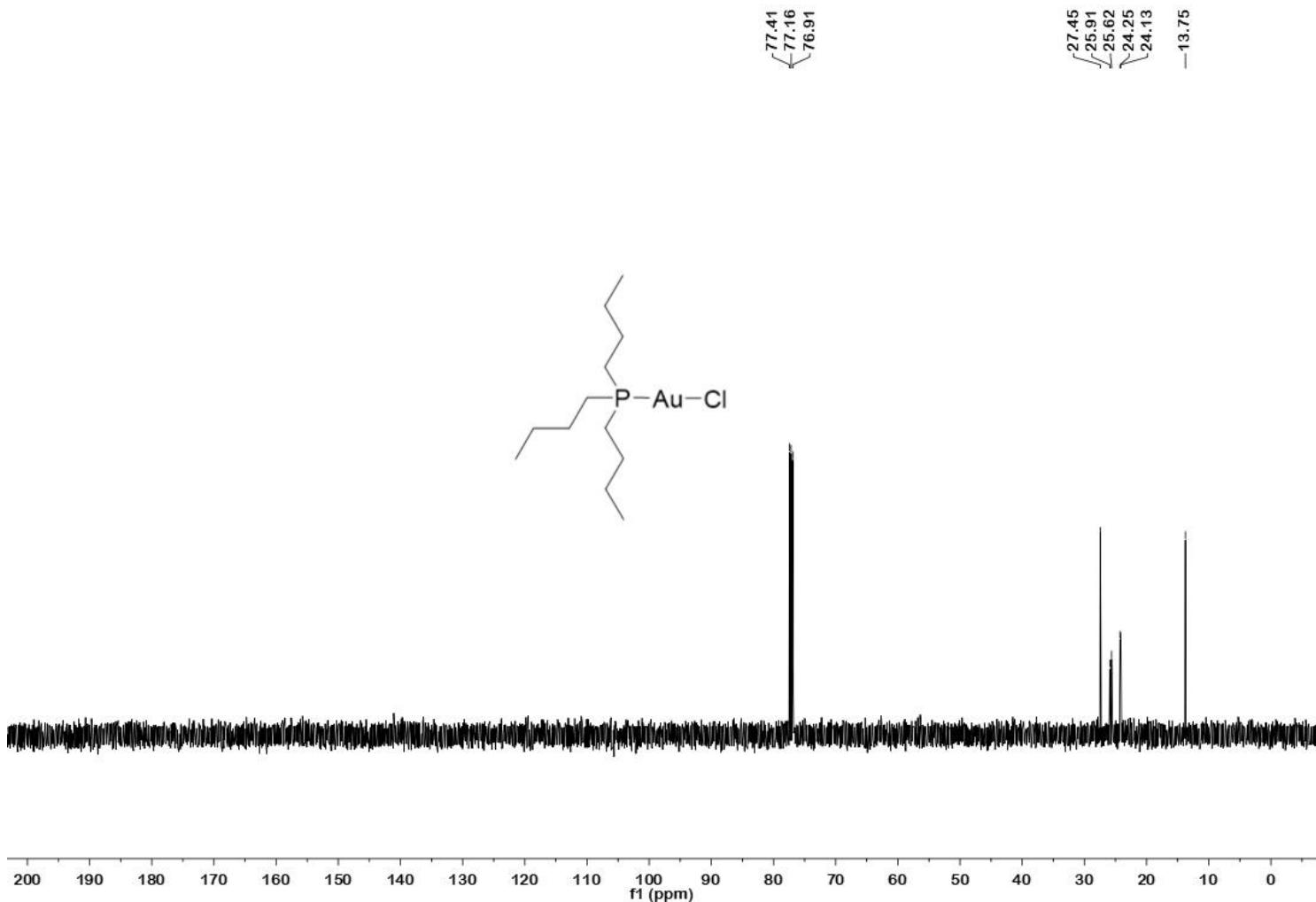


Figure S94. ^{13}C NMR spectrum of compound **26** in CDCl_3 .

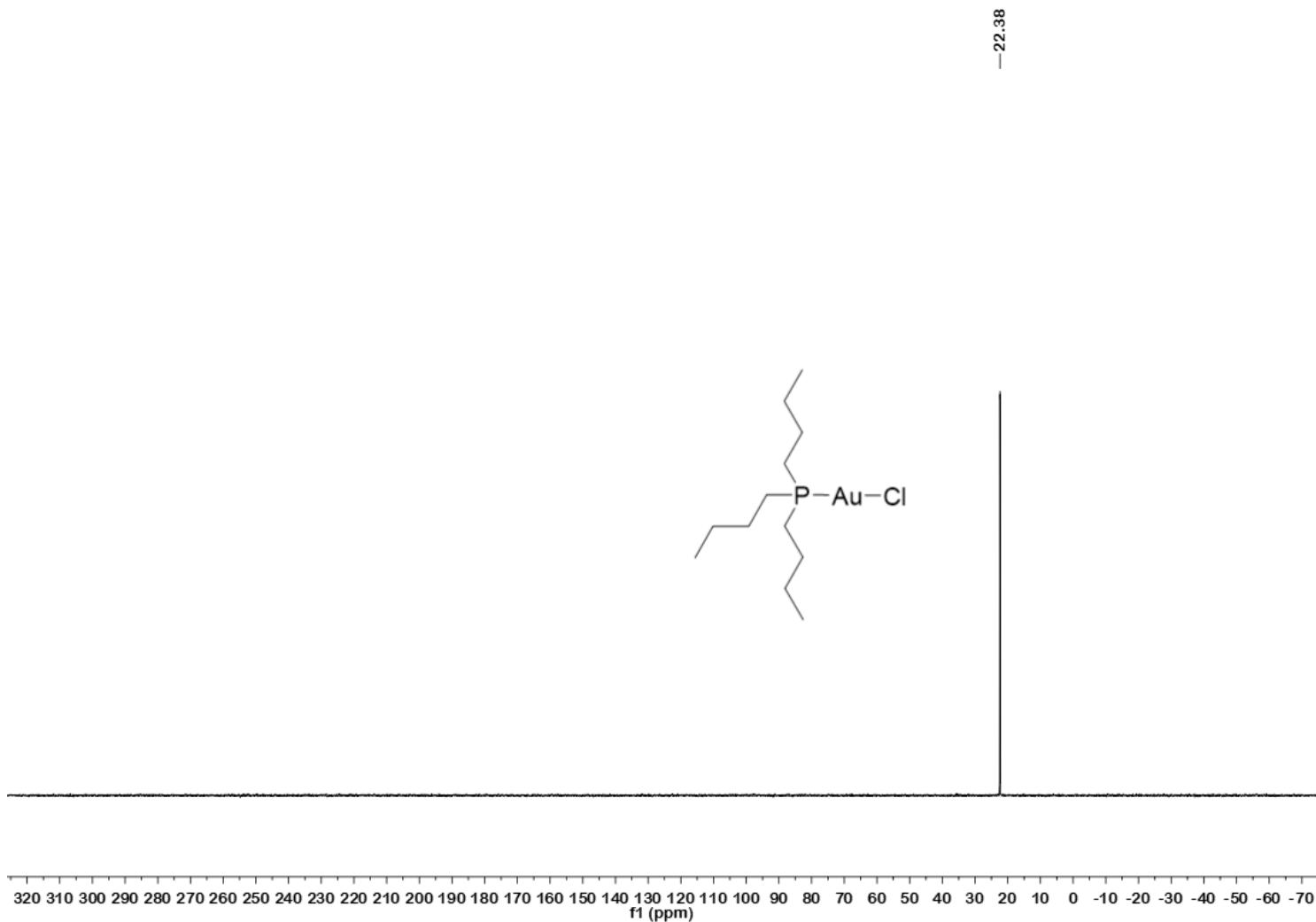


Figure S95. ^{31}P NMR spectrum of compound **26** in CDCl_3 .

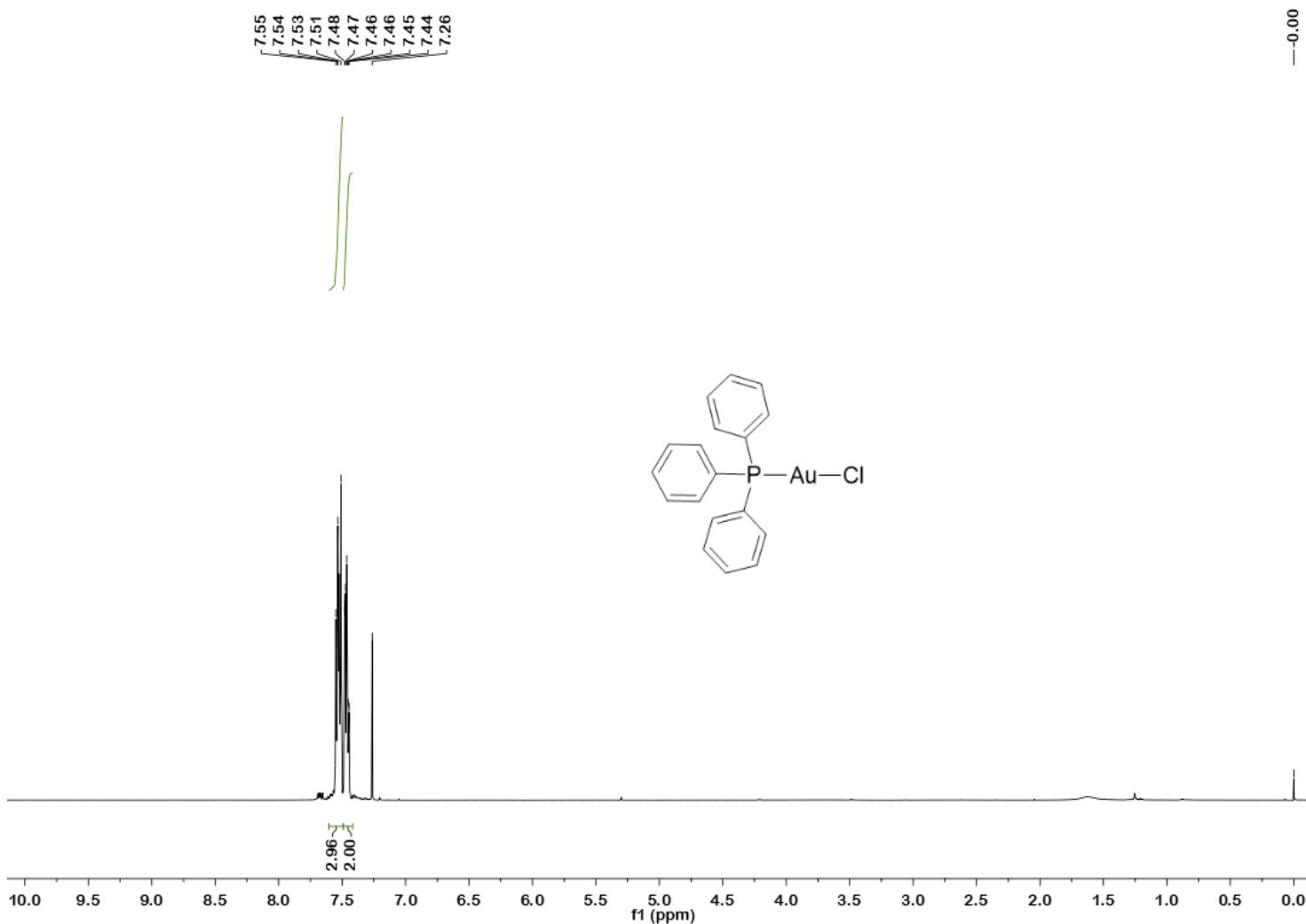


Figure S96. ^1H NMR spectrum of compound **27** in CDCl_3 .

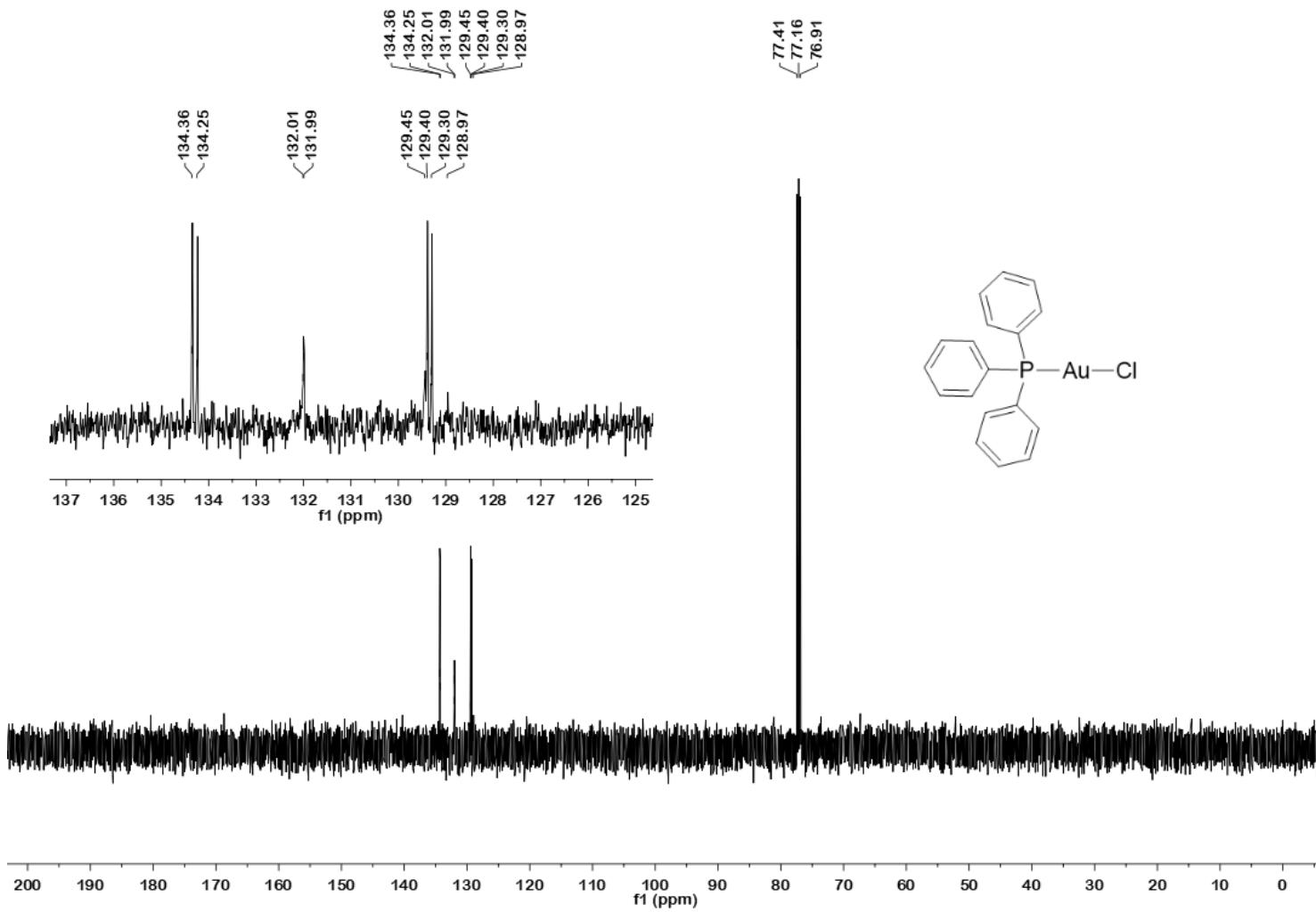


Figure S97. ^{13}C NMR spectrum of compound **27** in CDCl_3 .

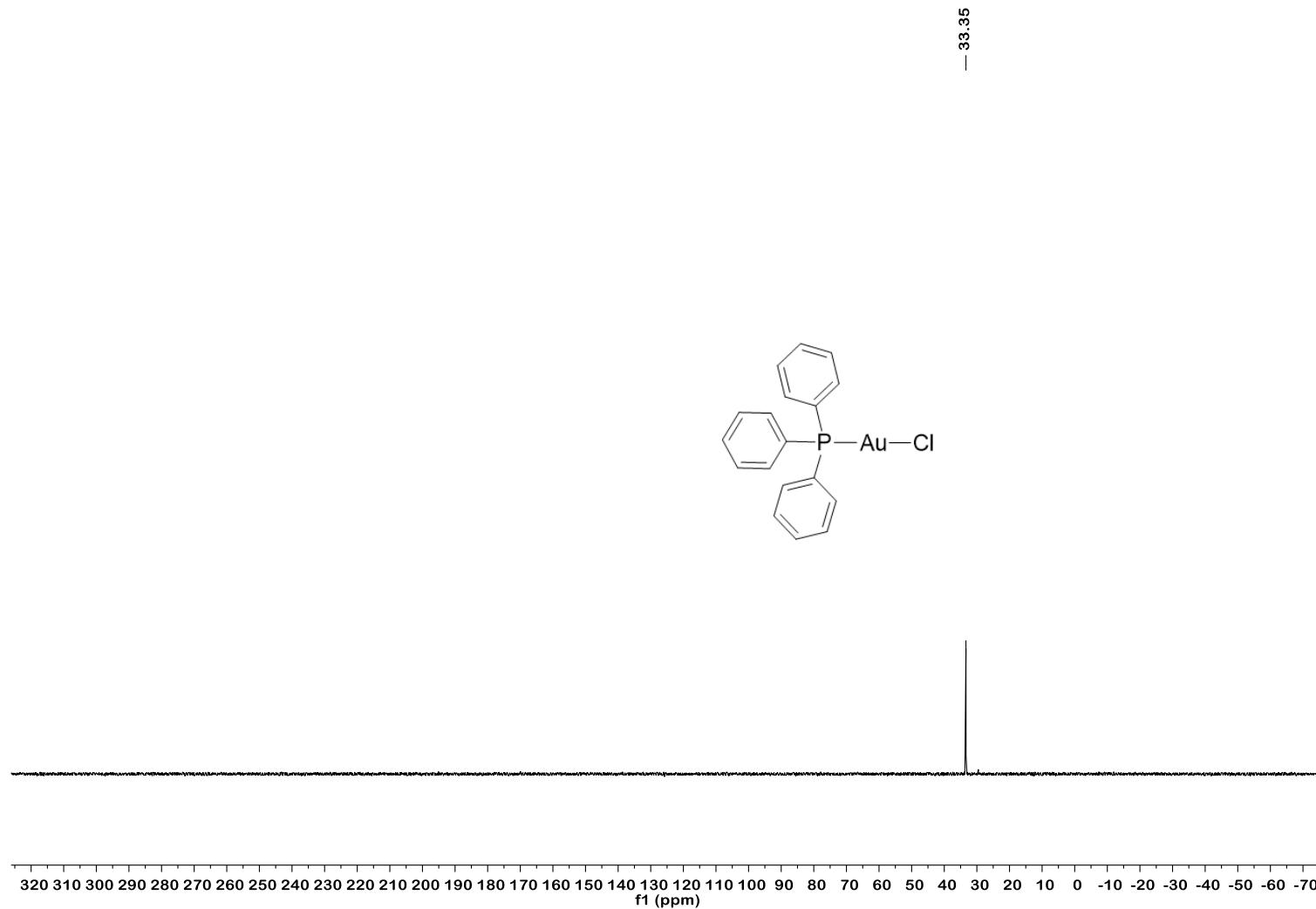


Figure S98. ^{31}P NMR spectrum of compound 27 in CDCl_3 .

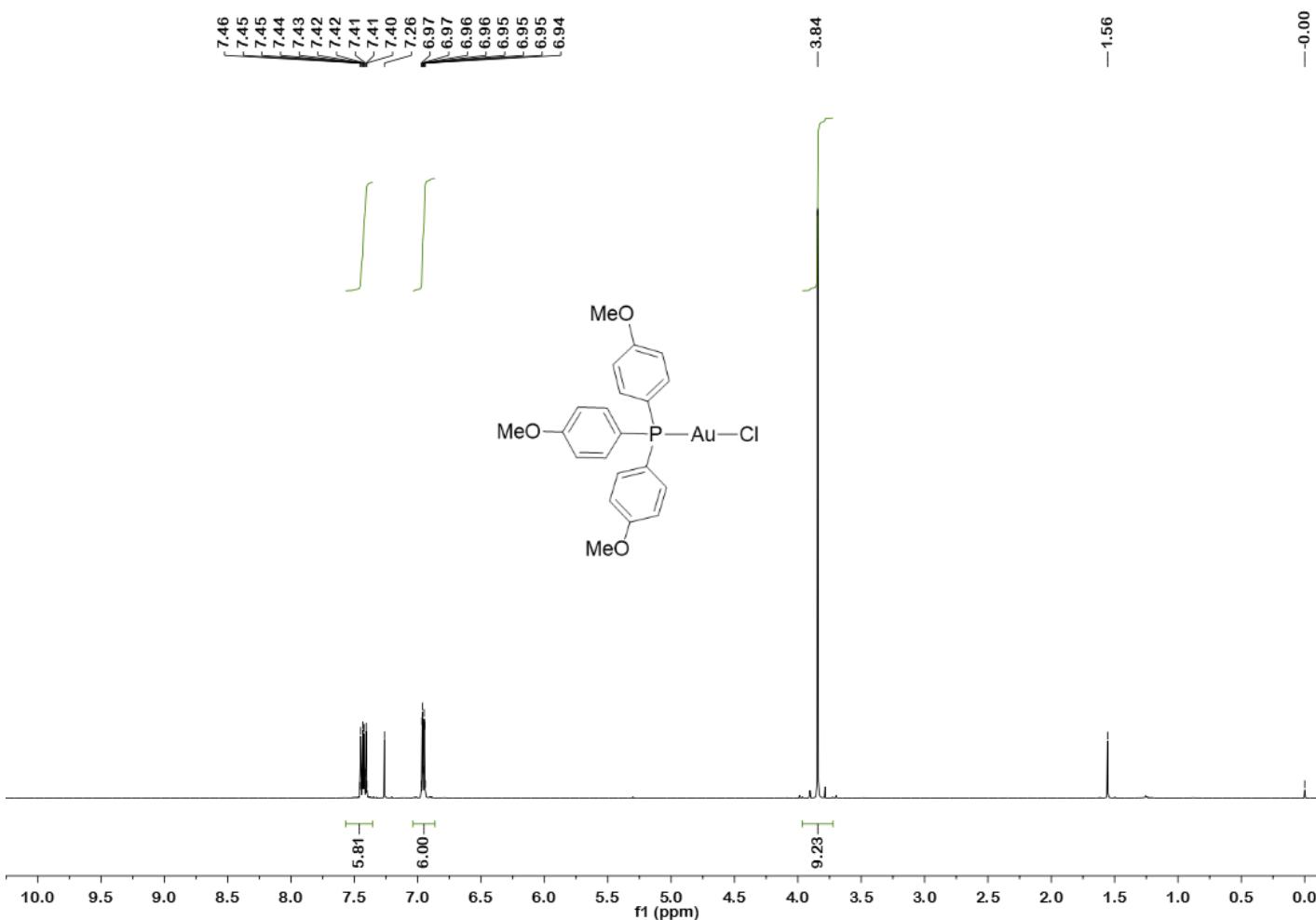


Figure S99. ^1H NMR spectrum of compound **28** in CDCl_3 .

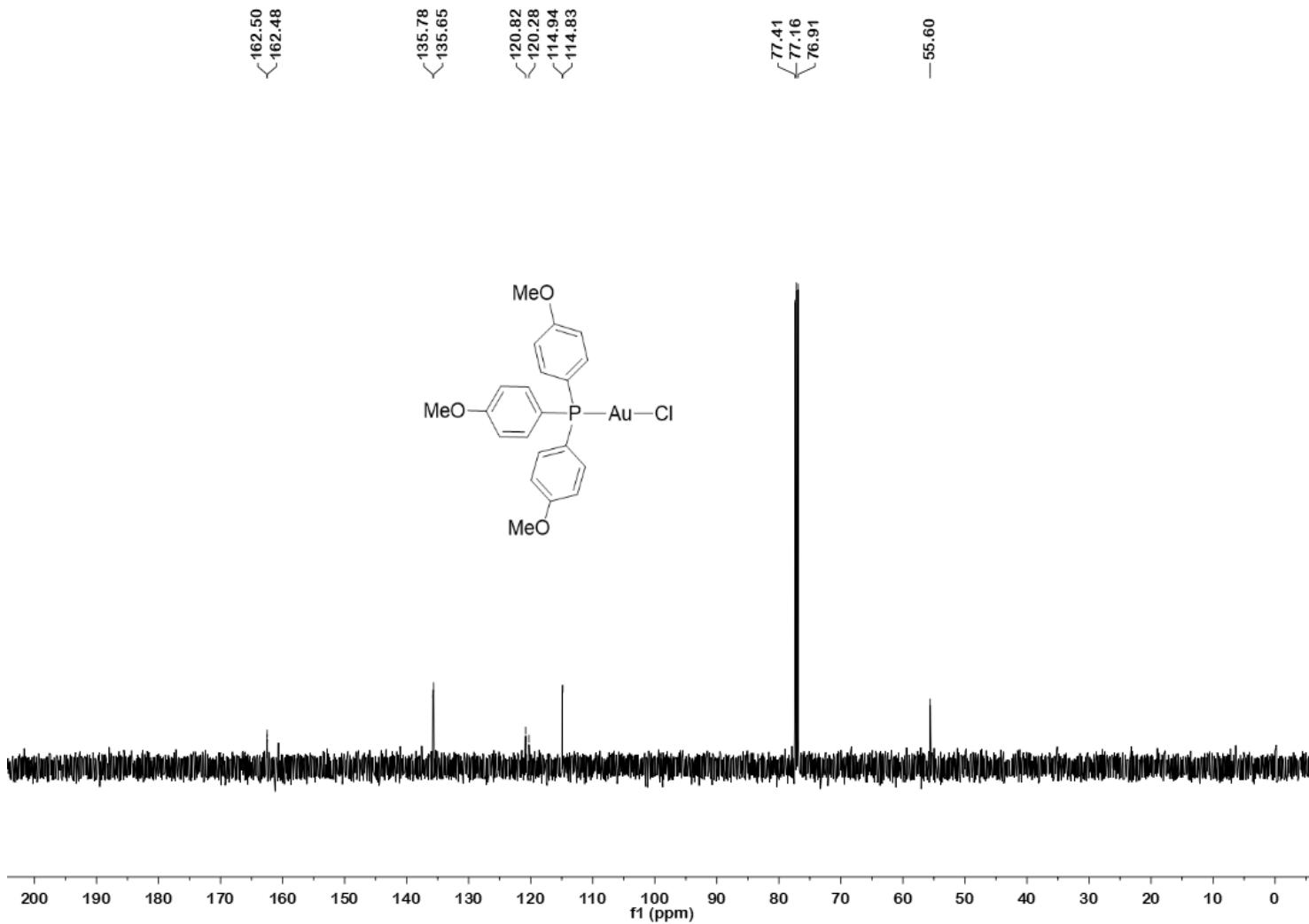


Figure S100. ^{13}C NMR spectrum of compound **28** in CDCl_3 .

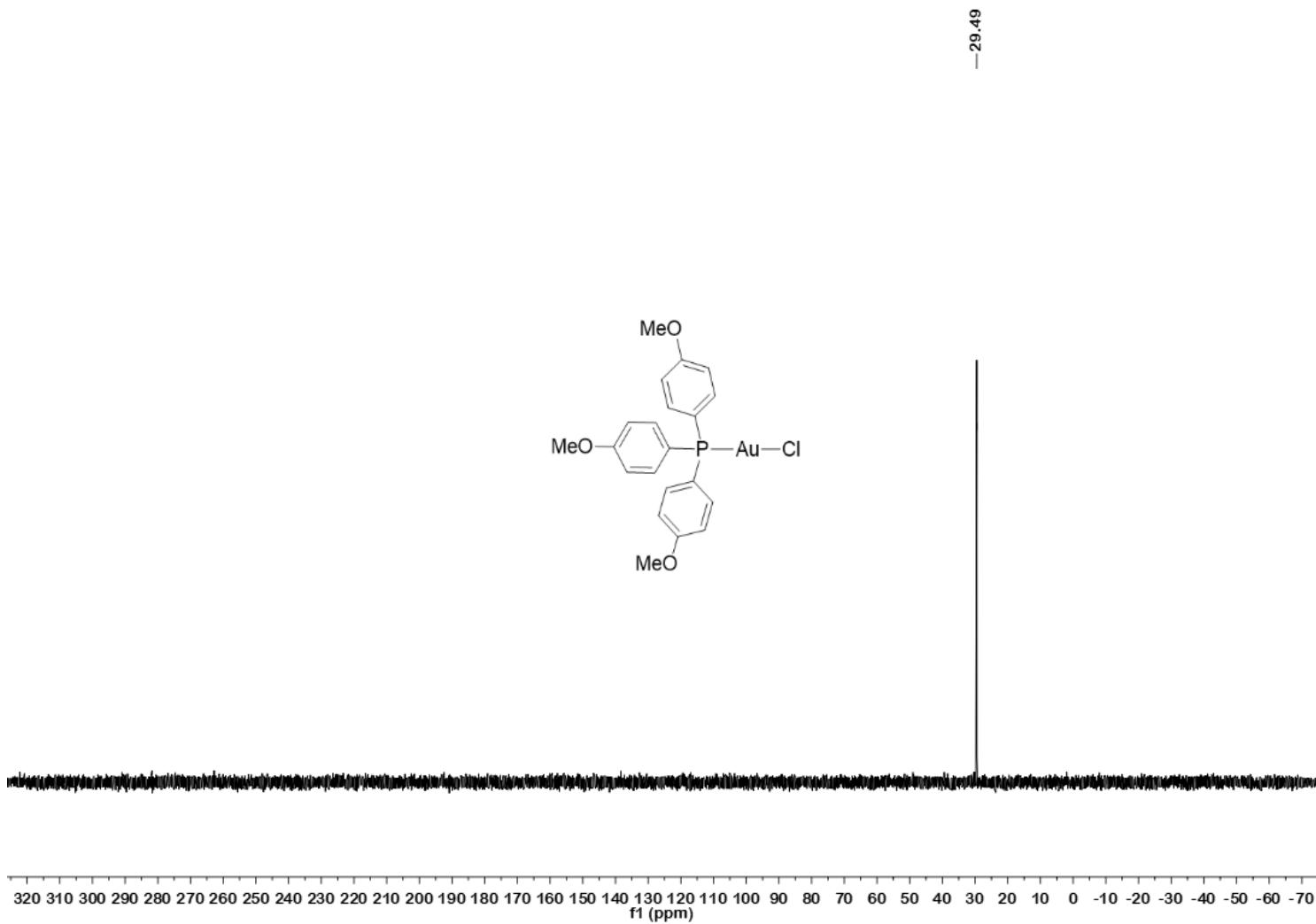


Figure S101. ^{31}P NMR spectrum of compound **28** in CDCl_3 .

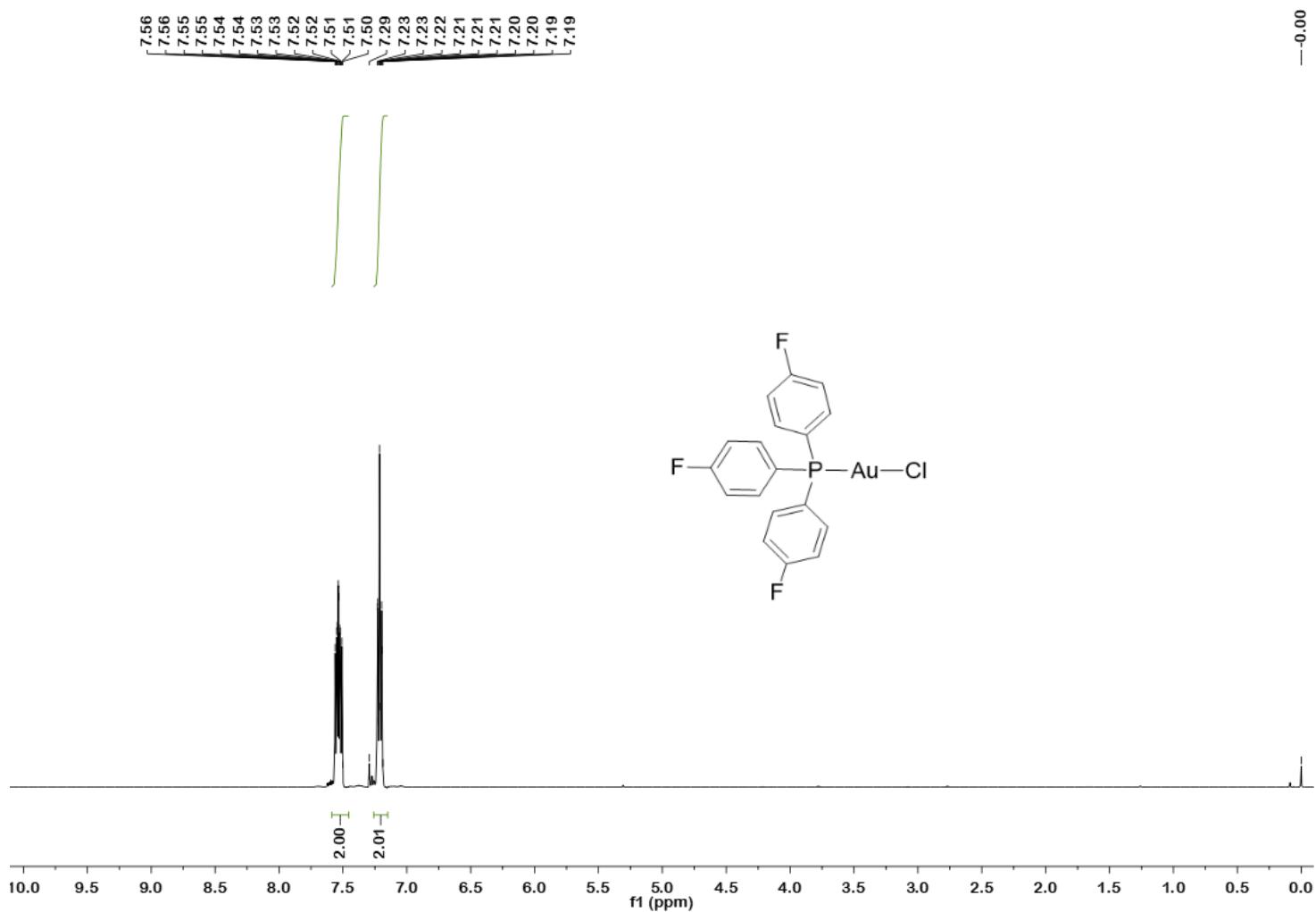


Figure S102. ^1H NMR spectrum of compound **29** in CDCl_3 .

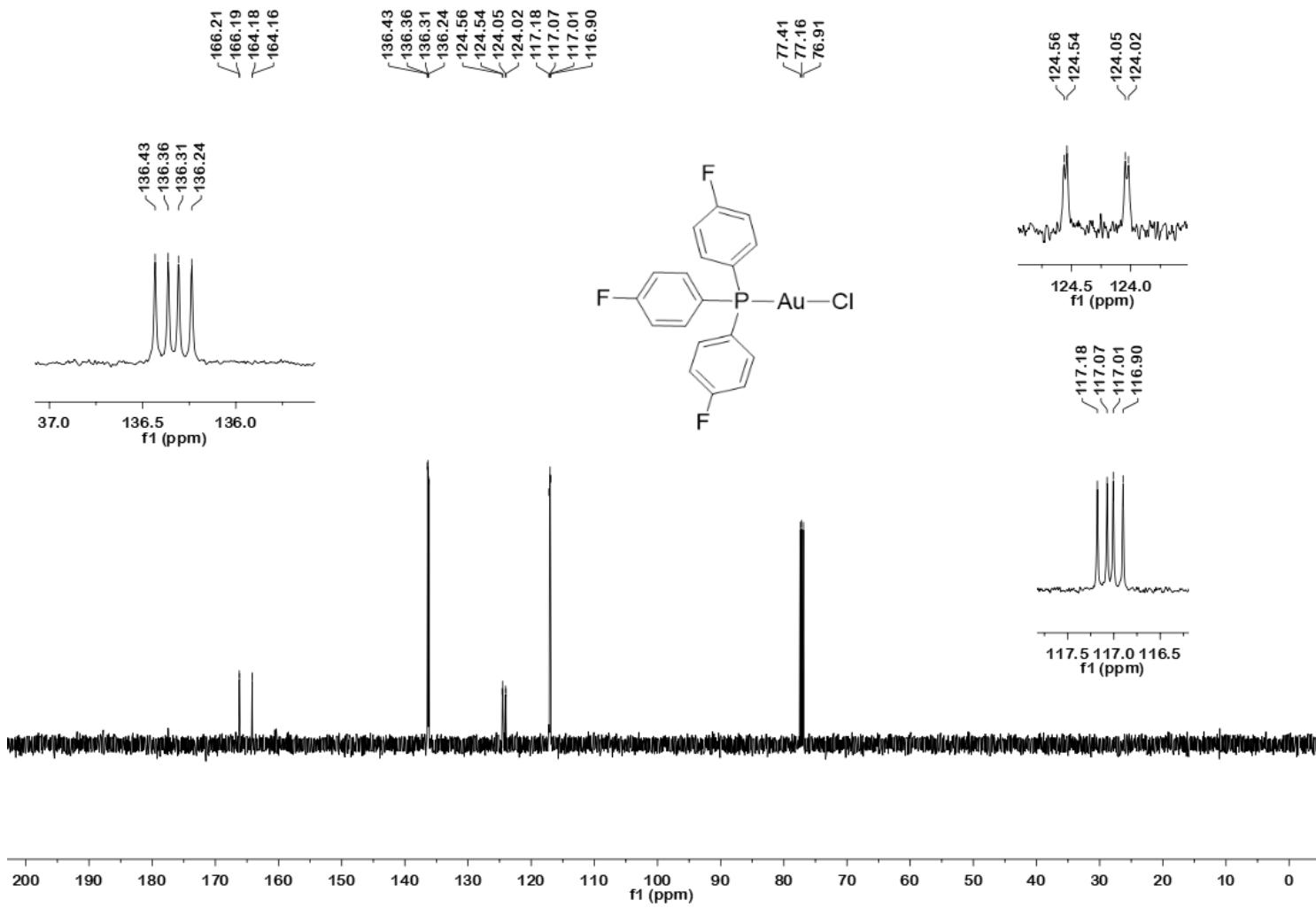


Figure S103. ^{13}C NMR spectrum of compound **29** in CDCl_3 .

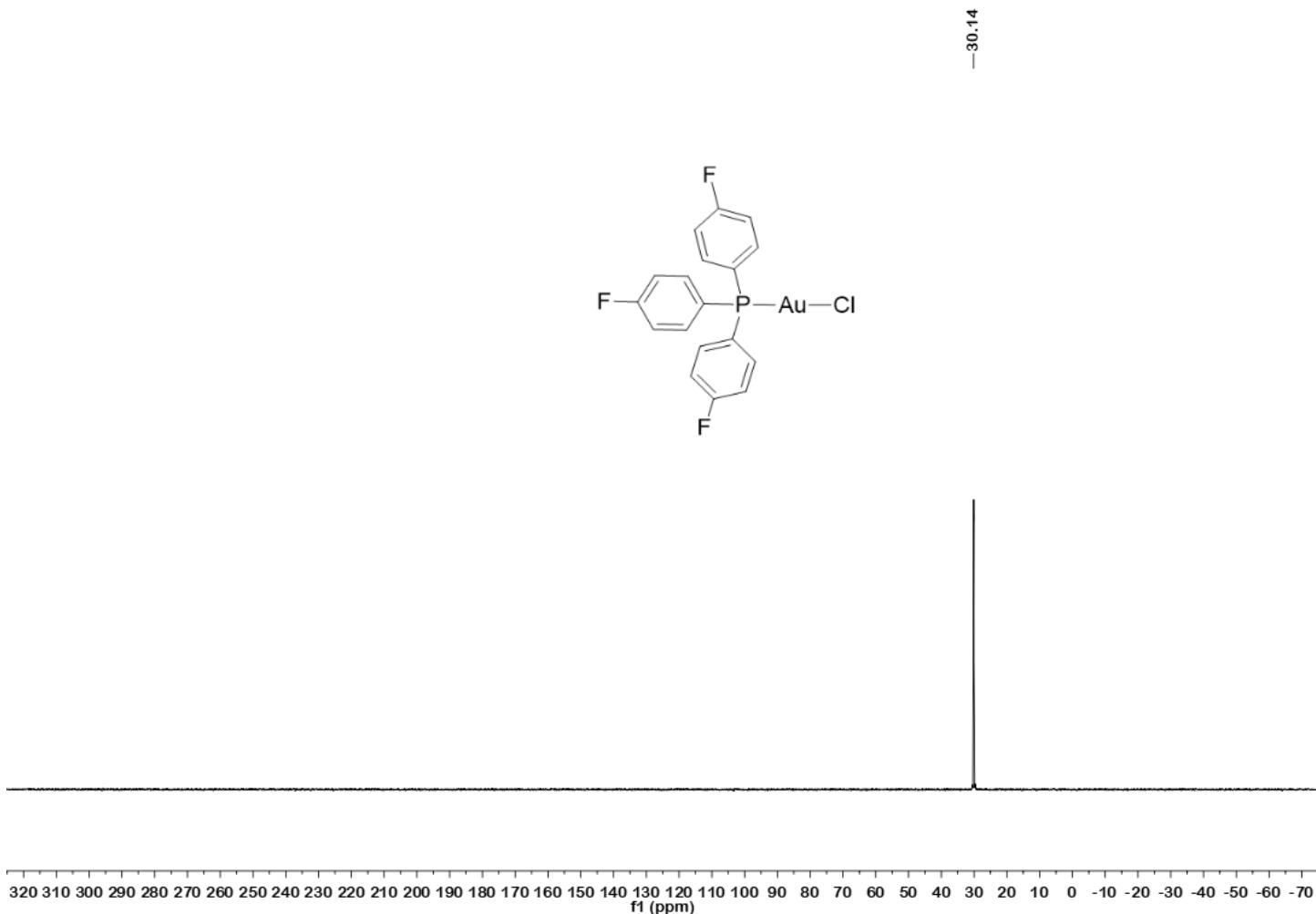


Figure S104. ^{31}P NMR spectrum of compound **29** in CDCl_3 .

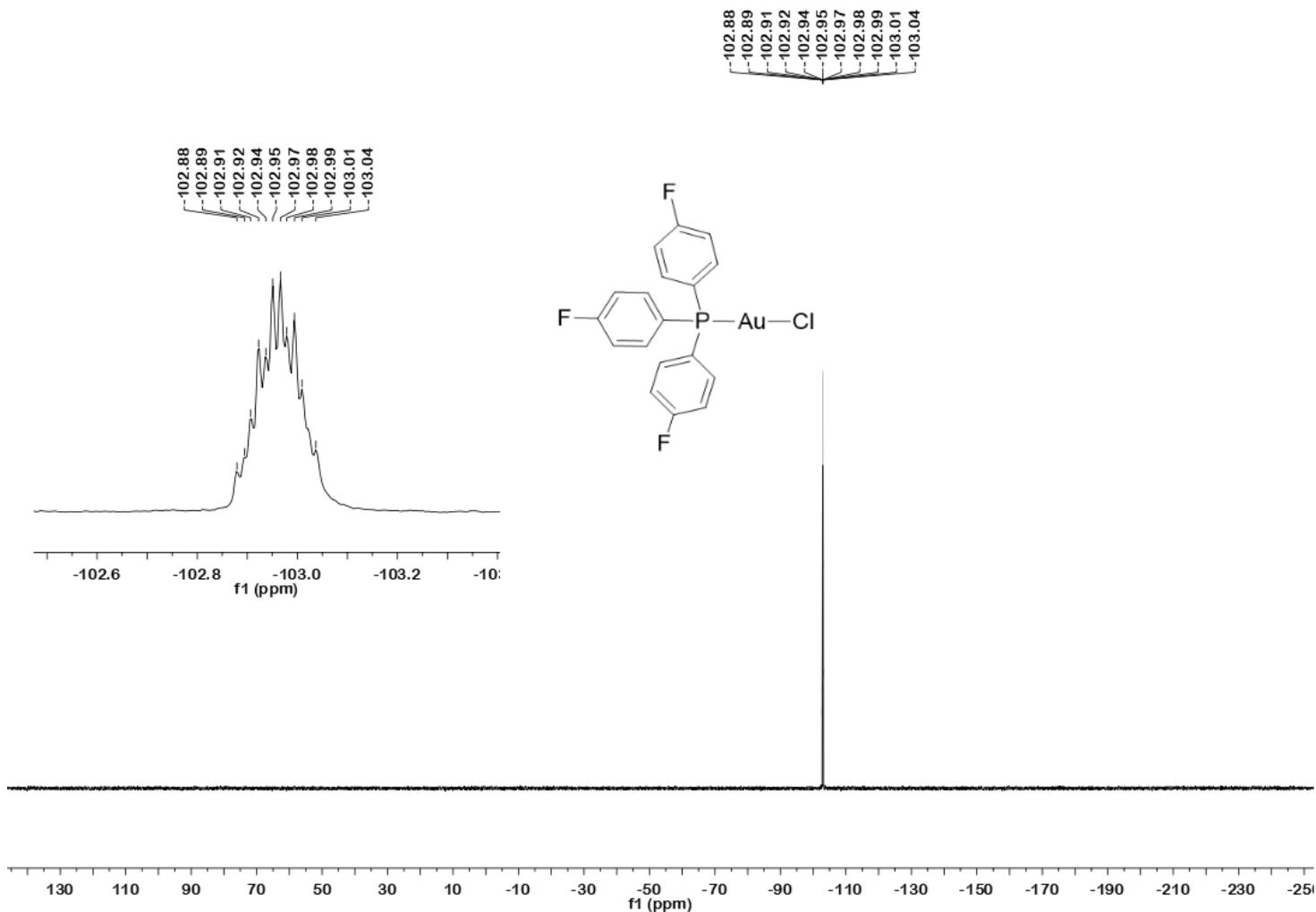


Figure S105. ^{19}F NMR spectrum of compound **29** in CDCl_3 .

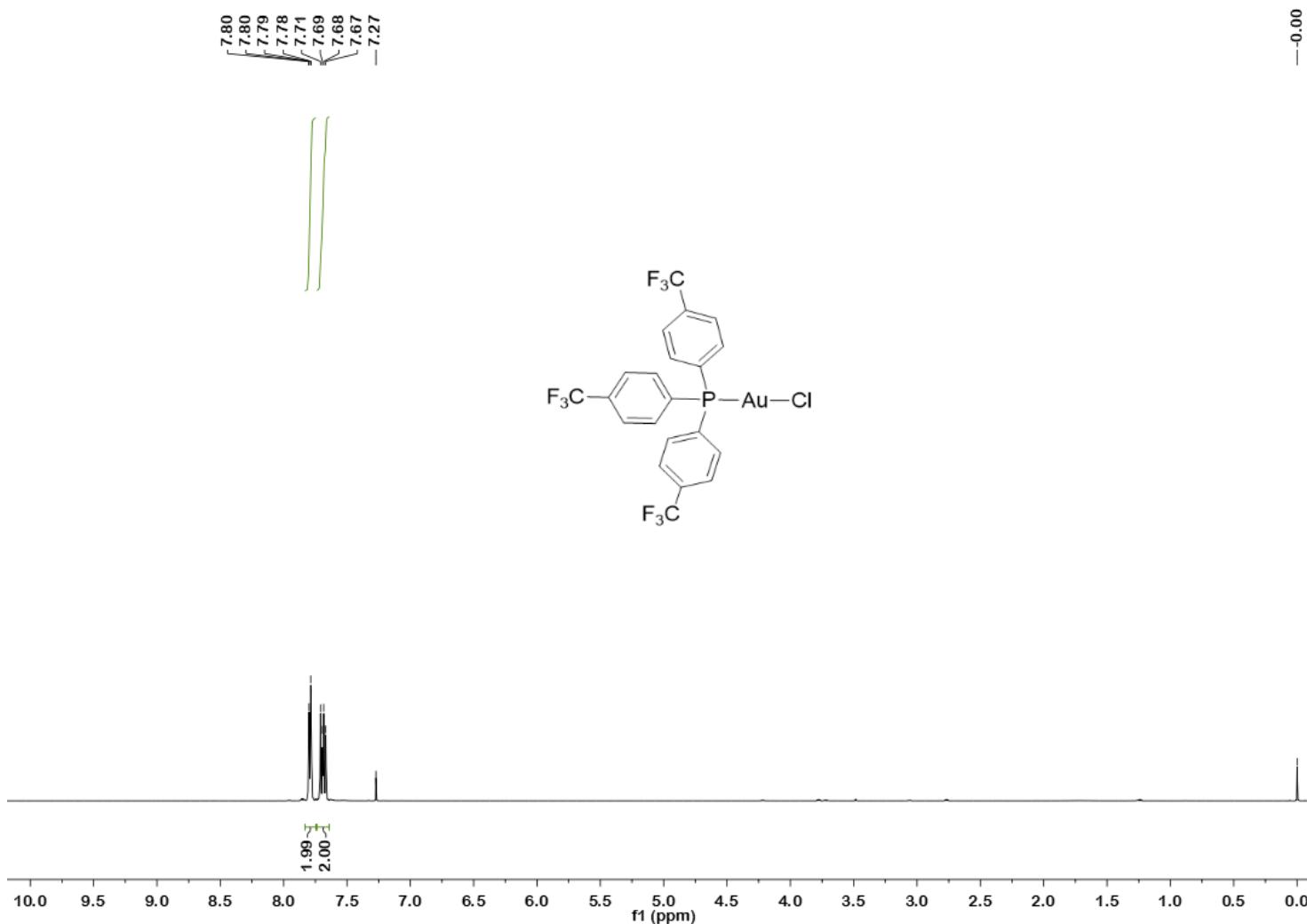


Figure S106. ^1H NMR spectrum of compound **30** in CDCl_3 .

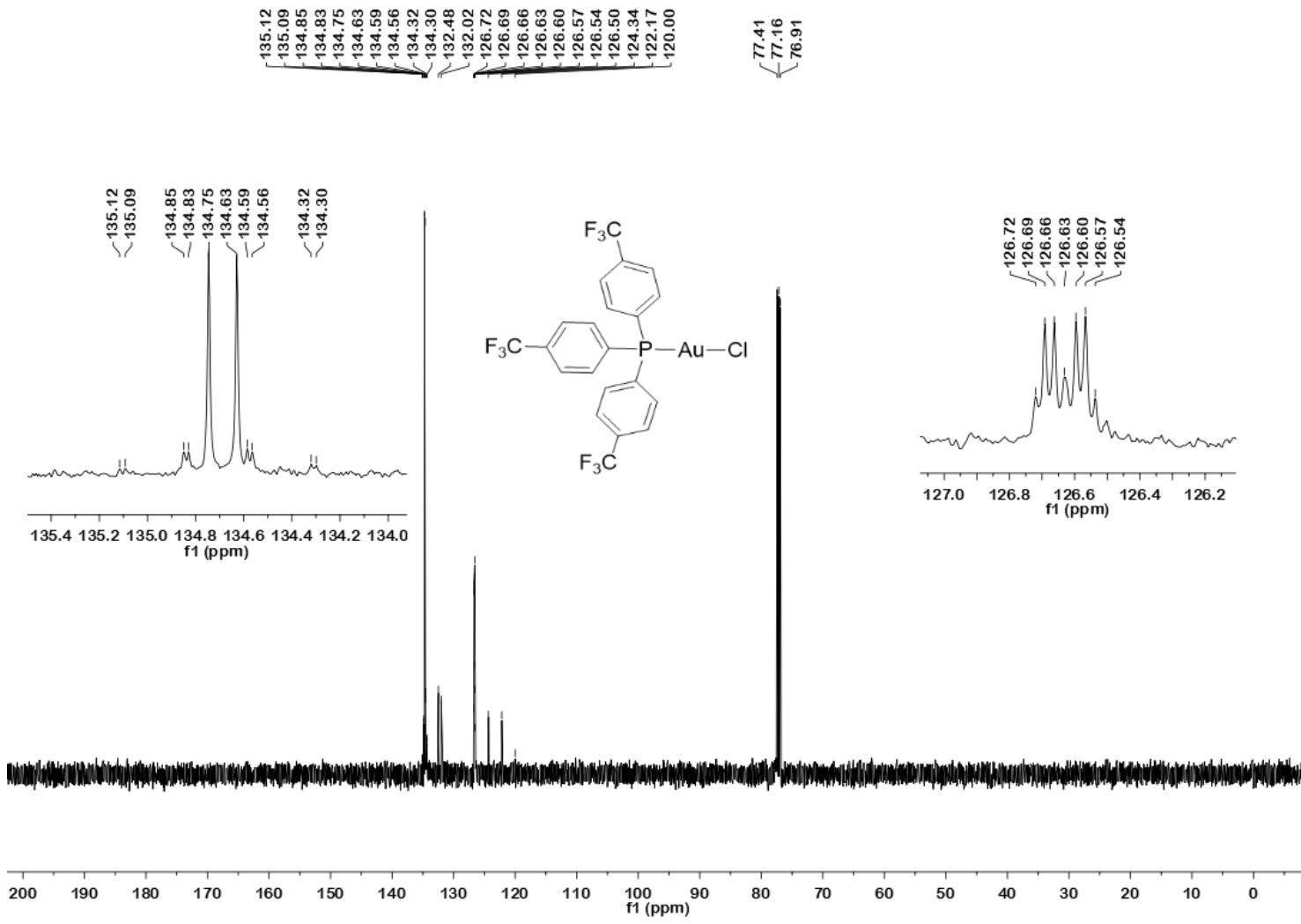


Figure S107. ^{13}C NMR spectrum of compound **30** in CDCl_3 .

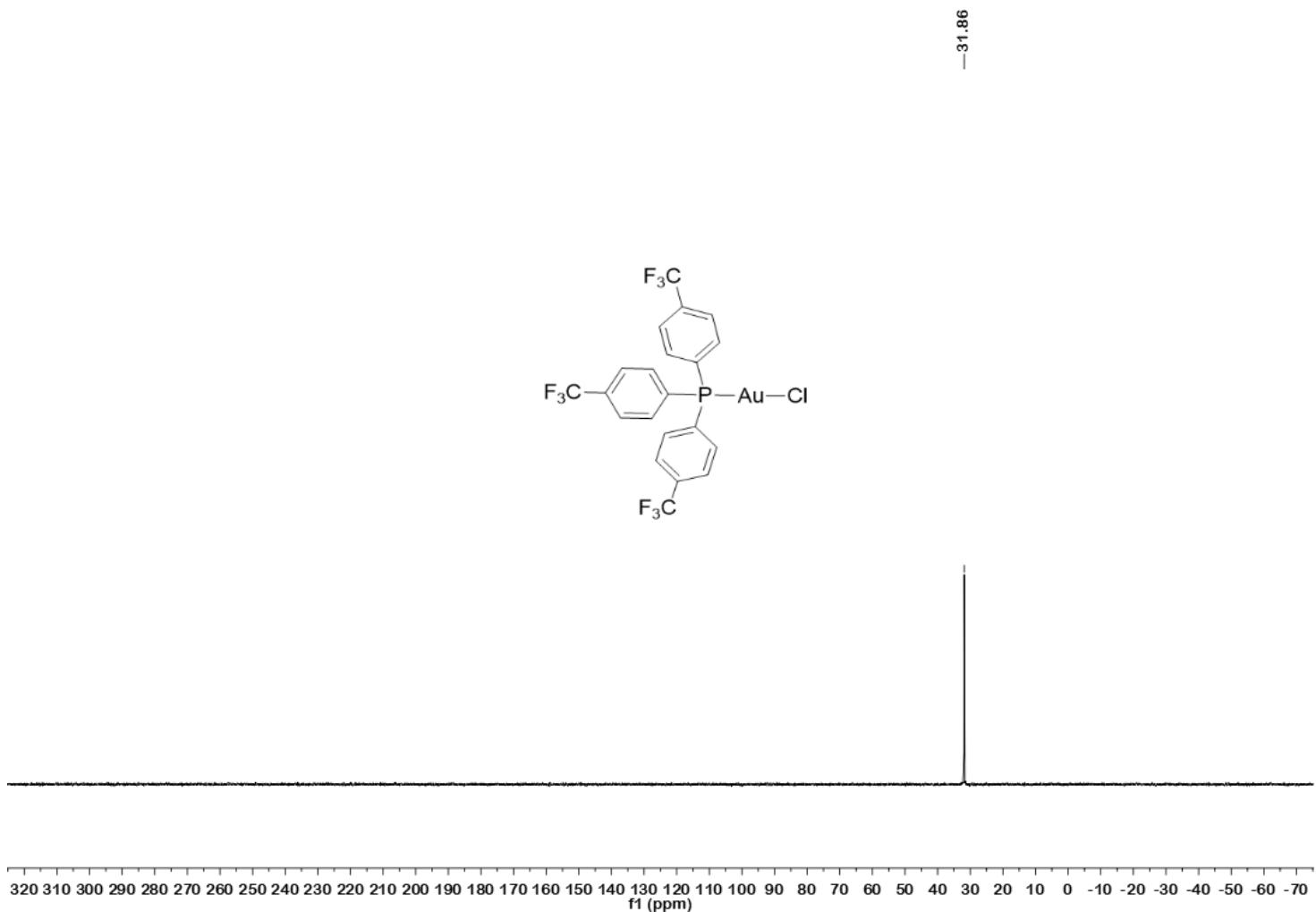


Figure S108. ^{31}P NMR spectrum of compound **30** in CDCl_3 .

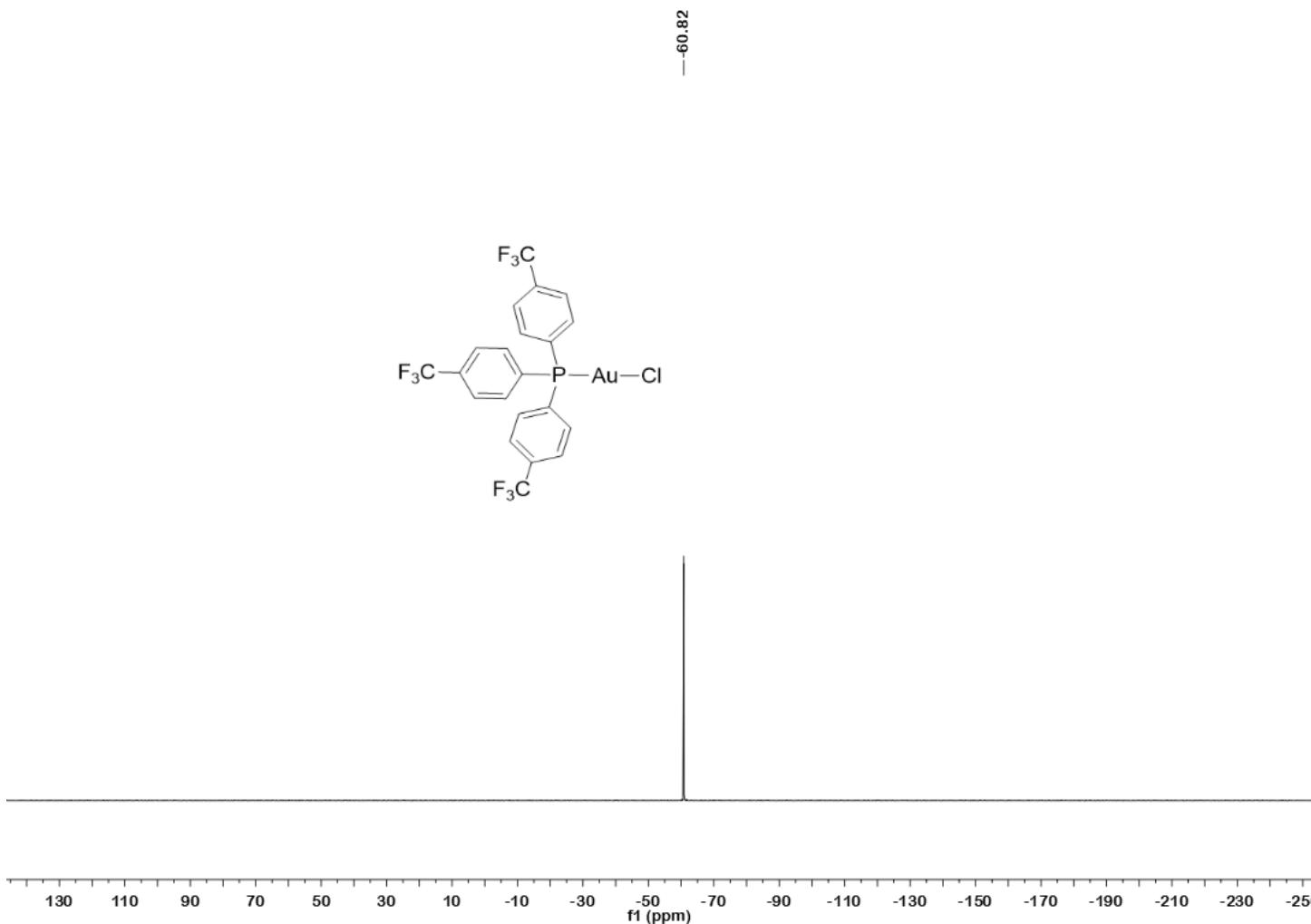


Figure S109. ^{19}F NMR spectrum of compound **30** in CDCl_3 .

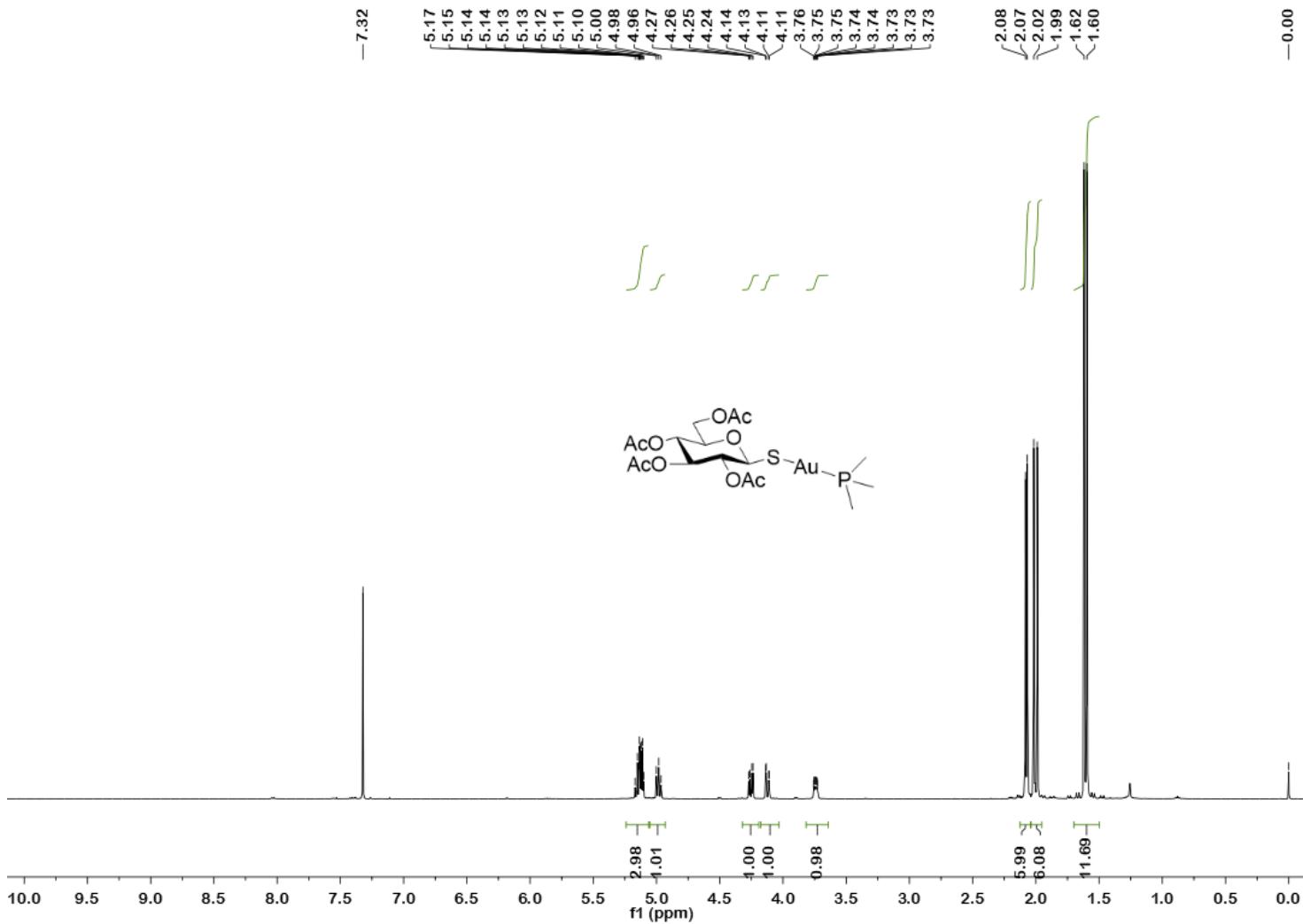


Figure S110. ^1H NMR spectrum of compound **31** in CDCl_3 .

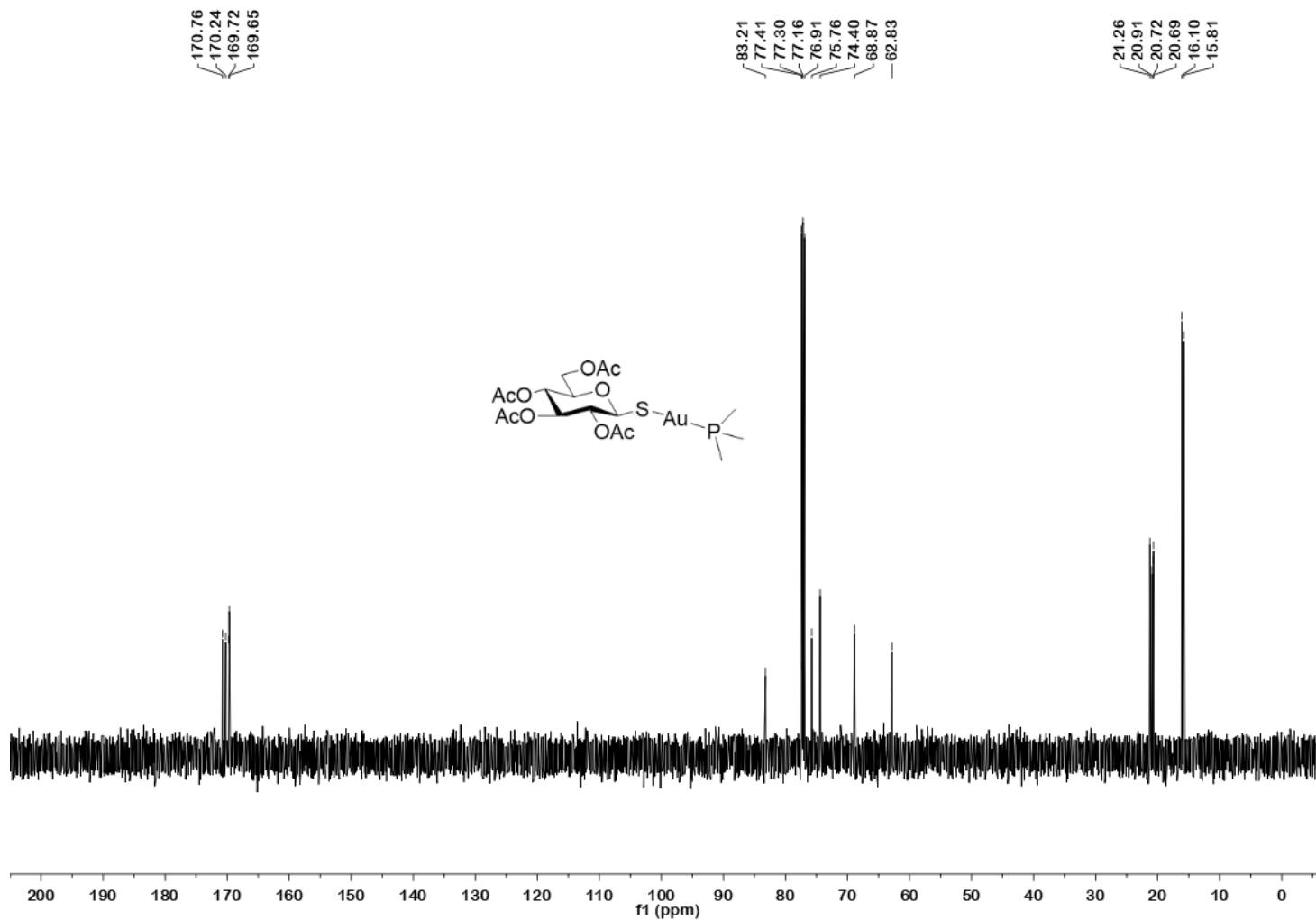


Figure S111. ^{13}C NMR spectrum of compound **31** in CDCl_3 .

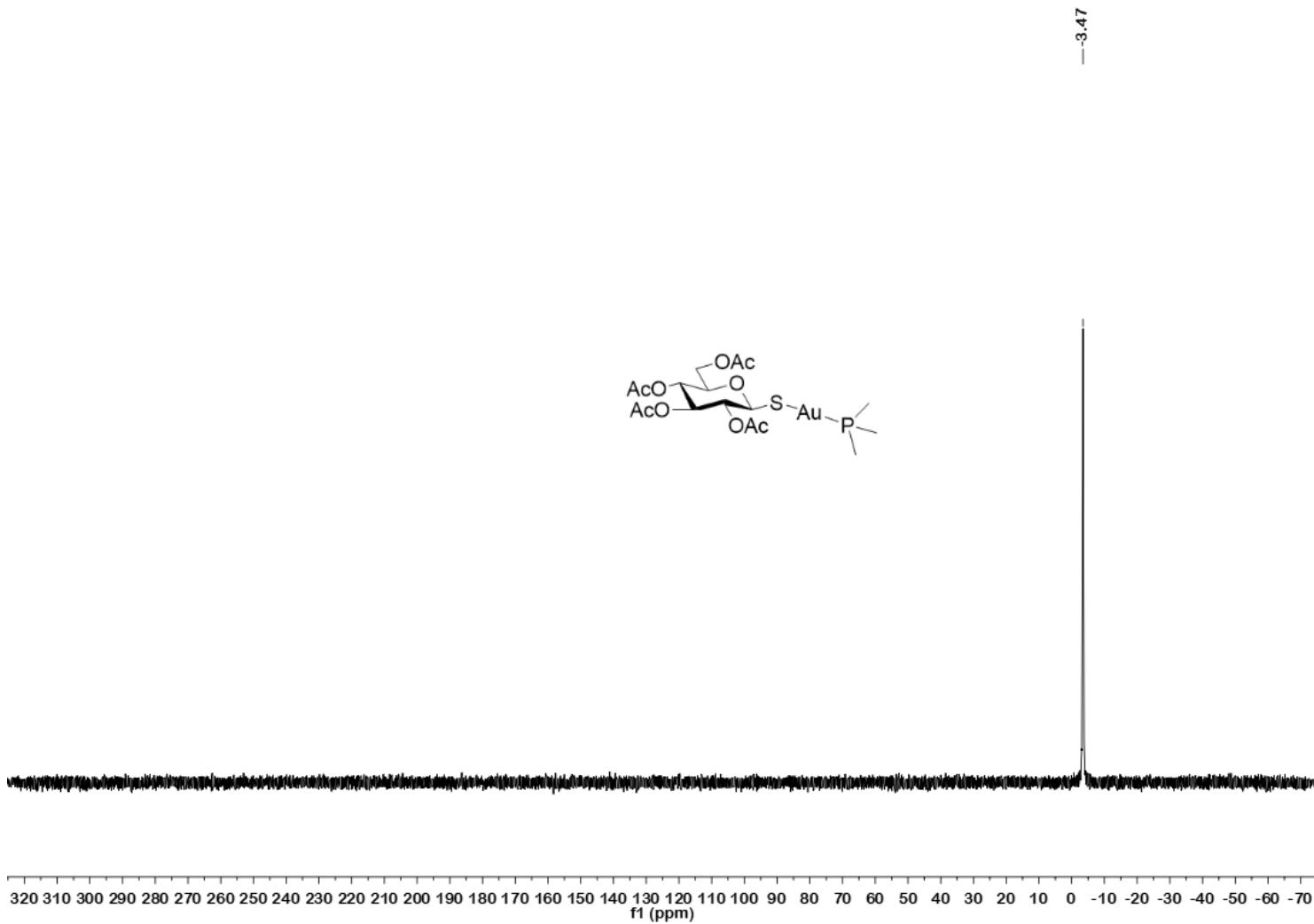
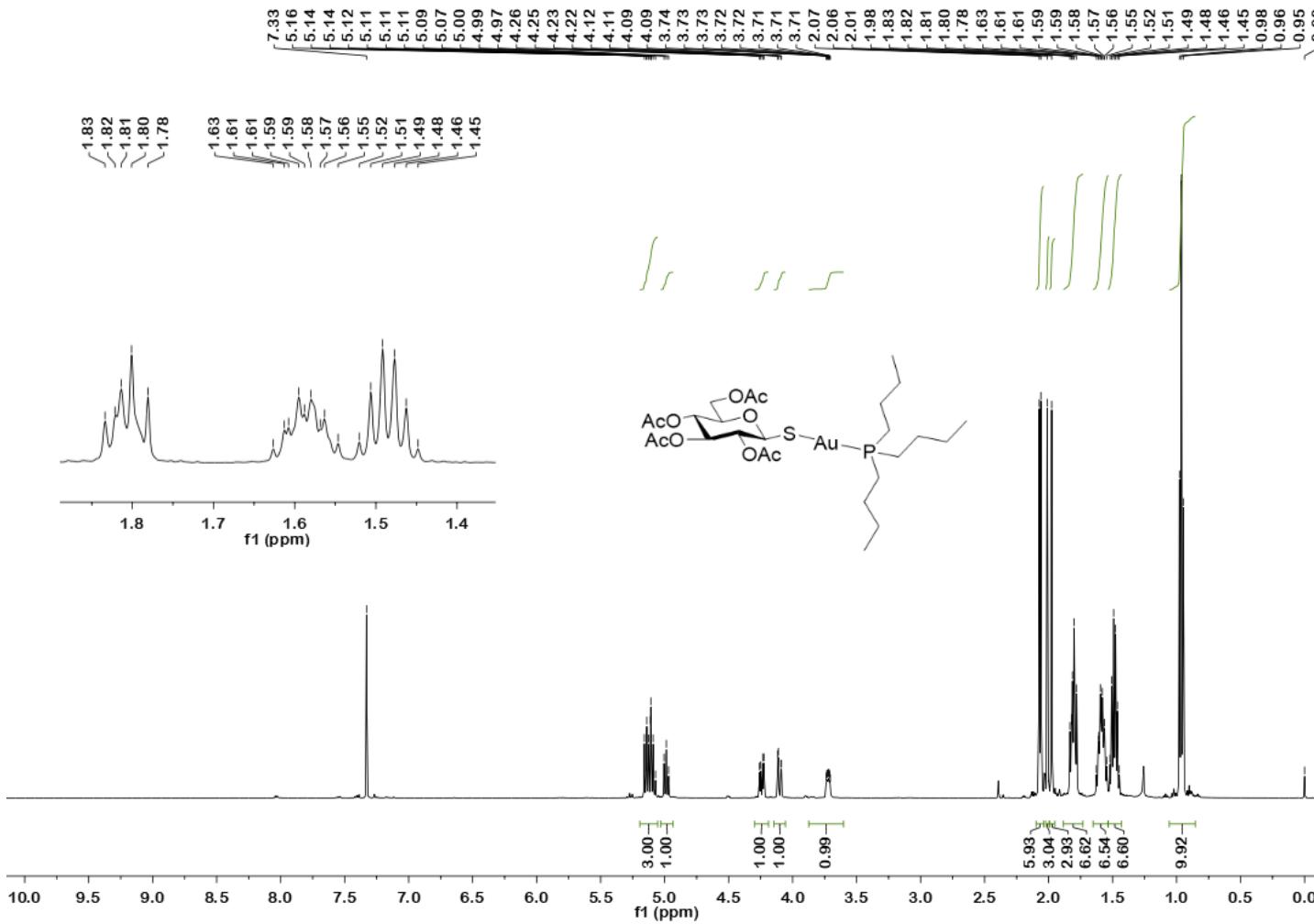


Figure S112. ^{31}P NMR spectrum of compound **31** in CDCl_3 .



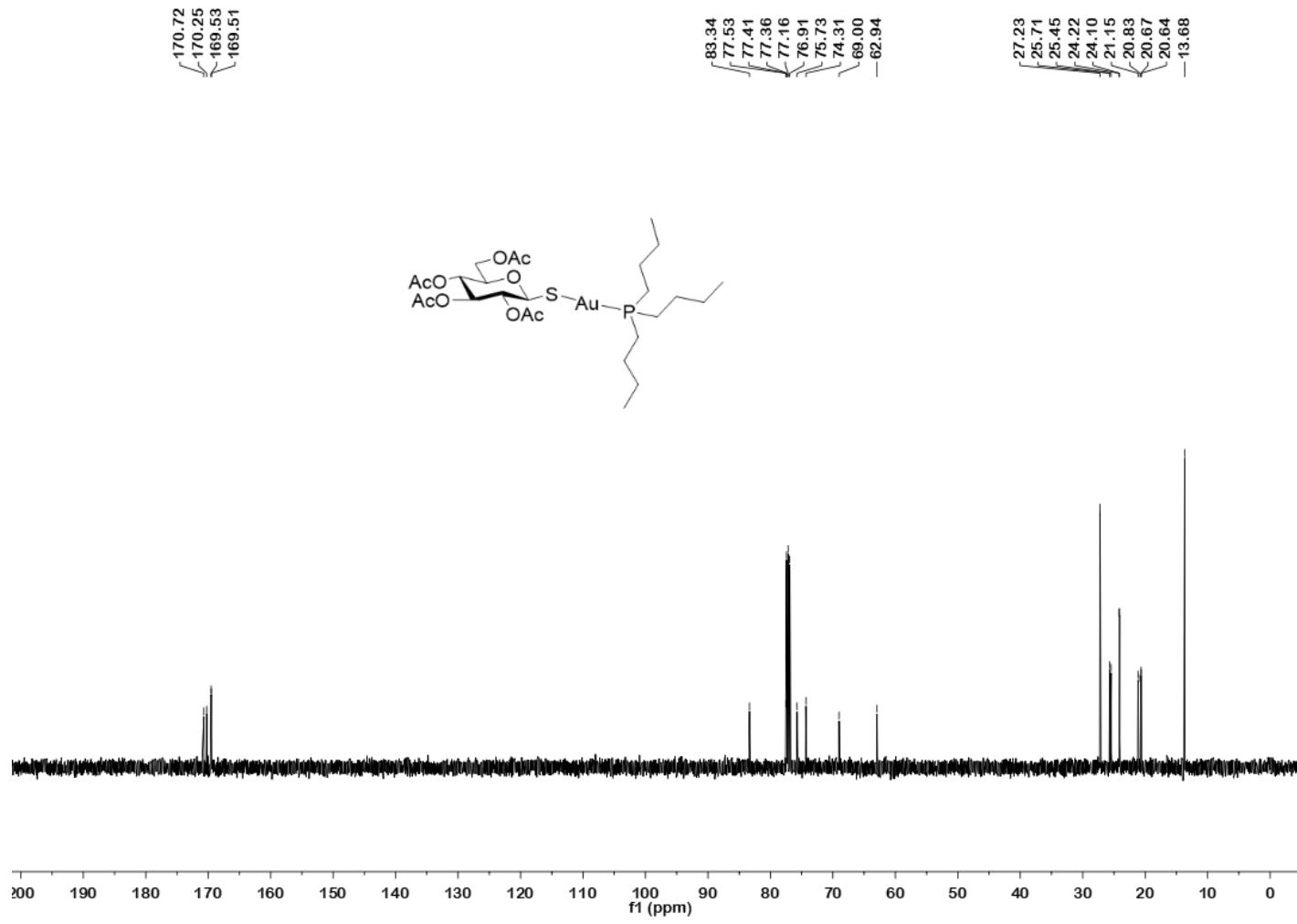


Figure S114. ^{13}C NMR spectrum of compound 32 in CDCl_3 .

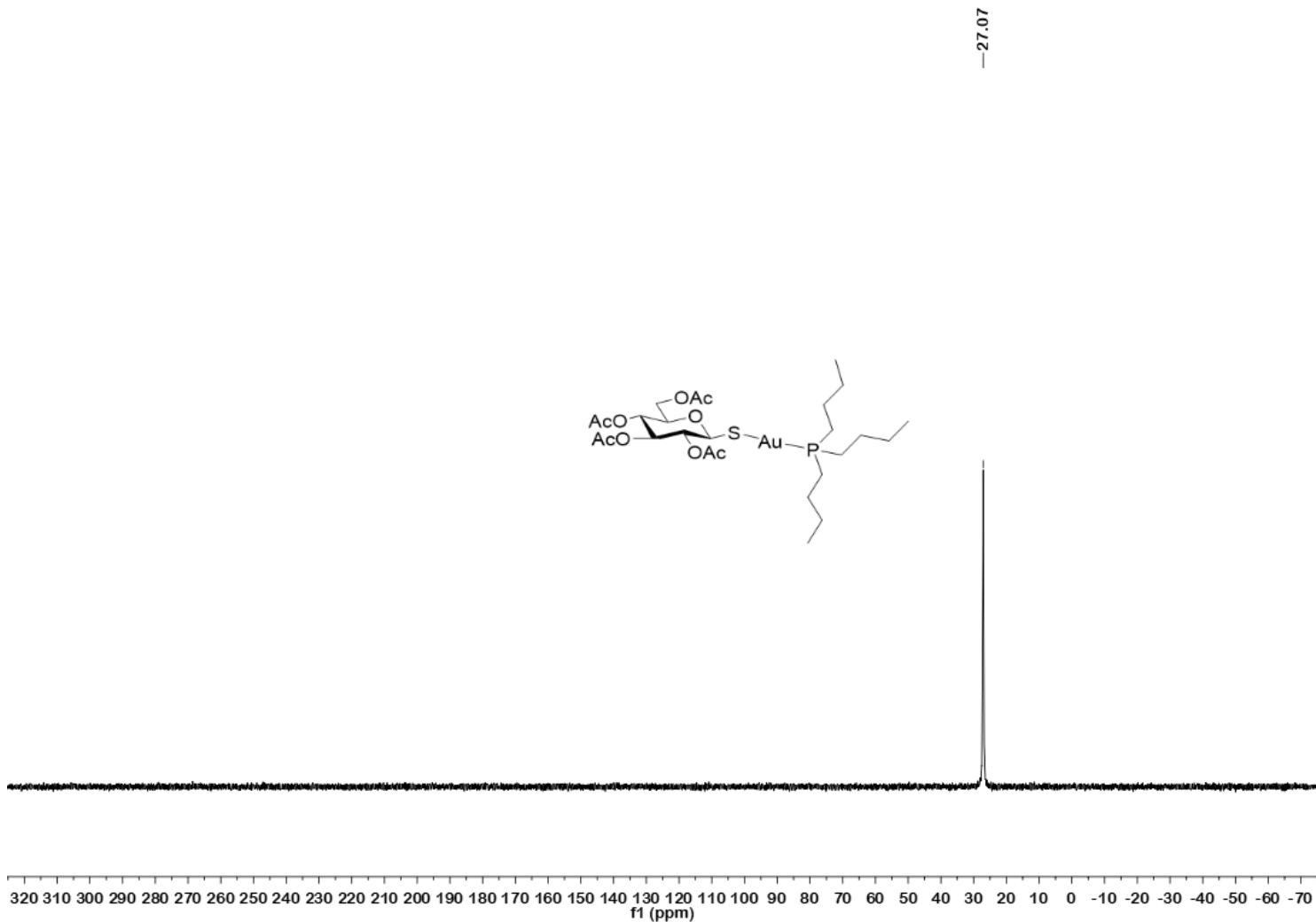


Figure S115. ^{31}P NMR spectrum of compound **32** in CDCl_3 .

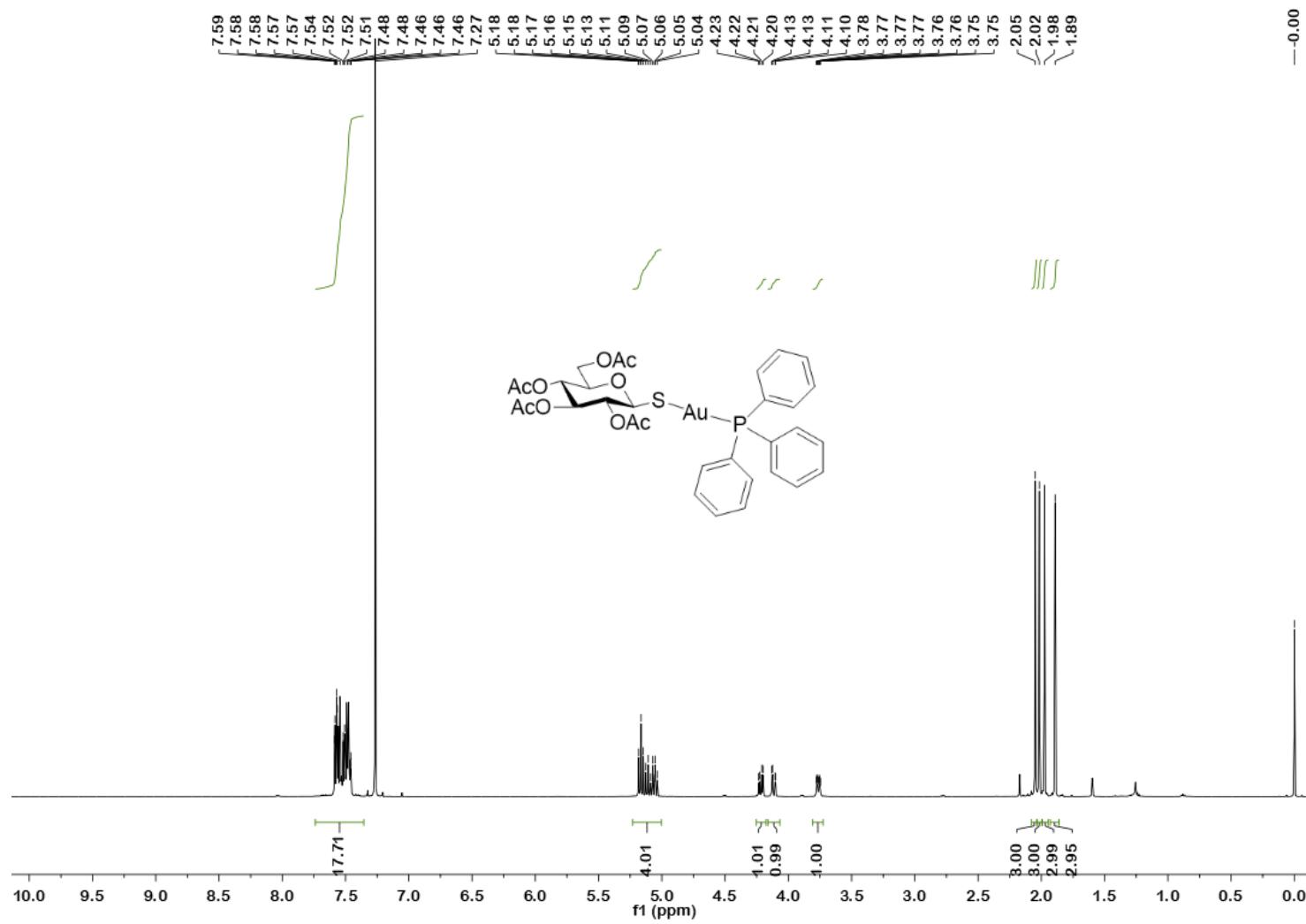


Figure S116. ^1H NMR spectrum of compound **33** in CDCl_3 .

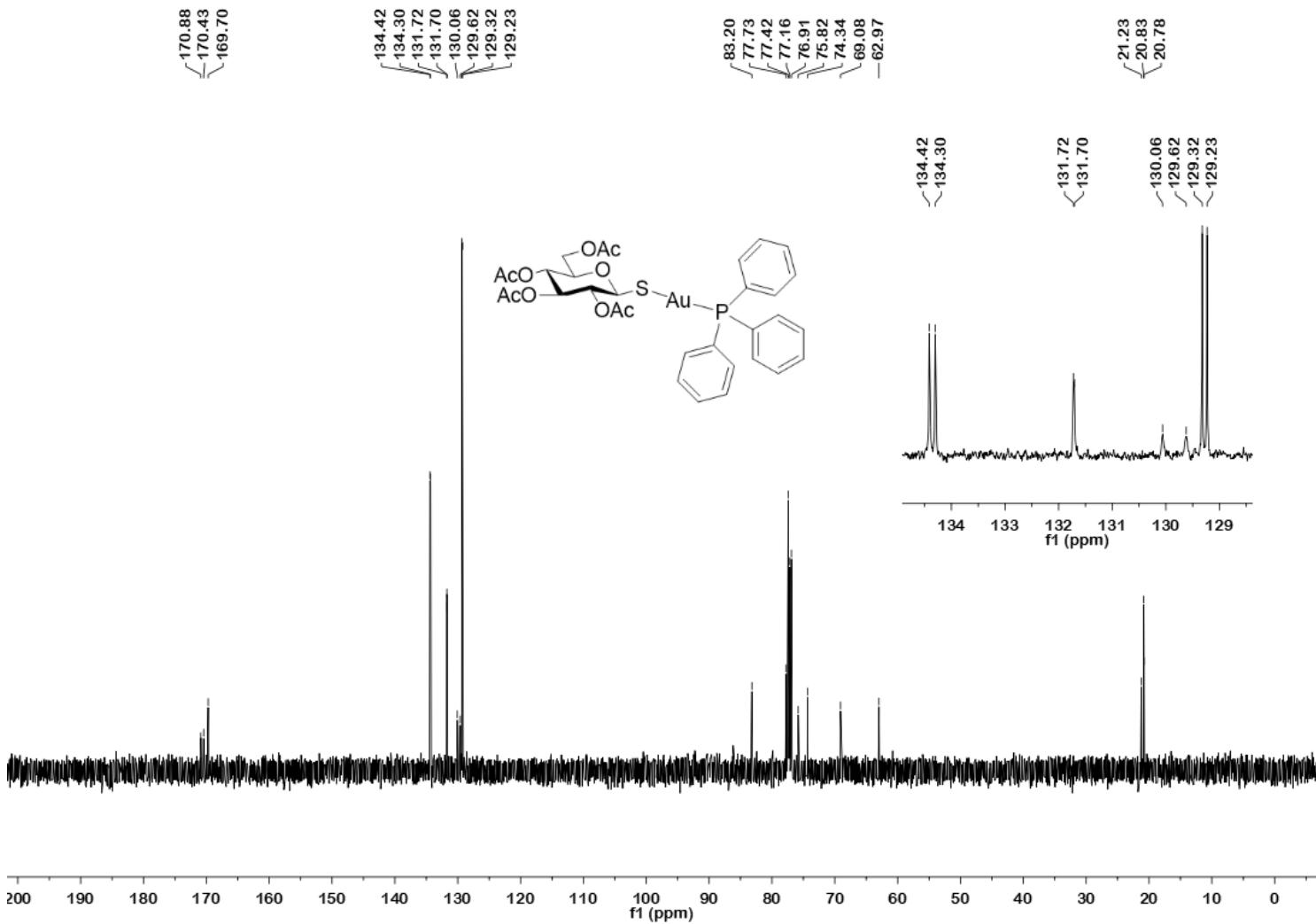


Figure S117. ^{13}C NMR spectrum of compound 33 in CDCl_3 .

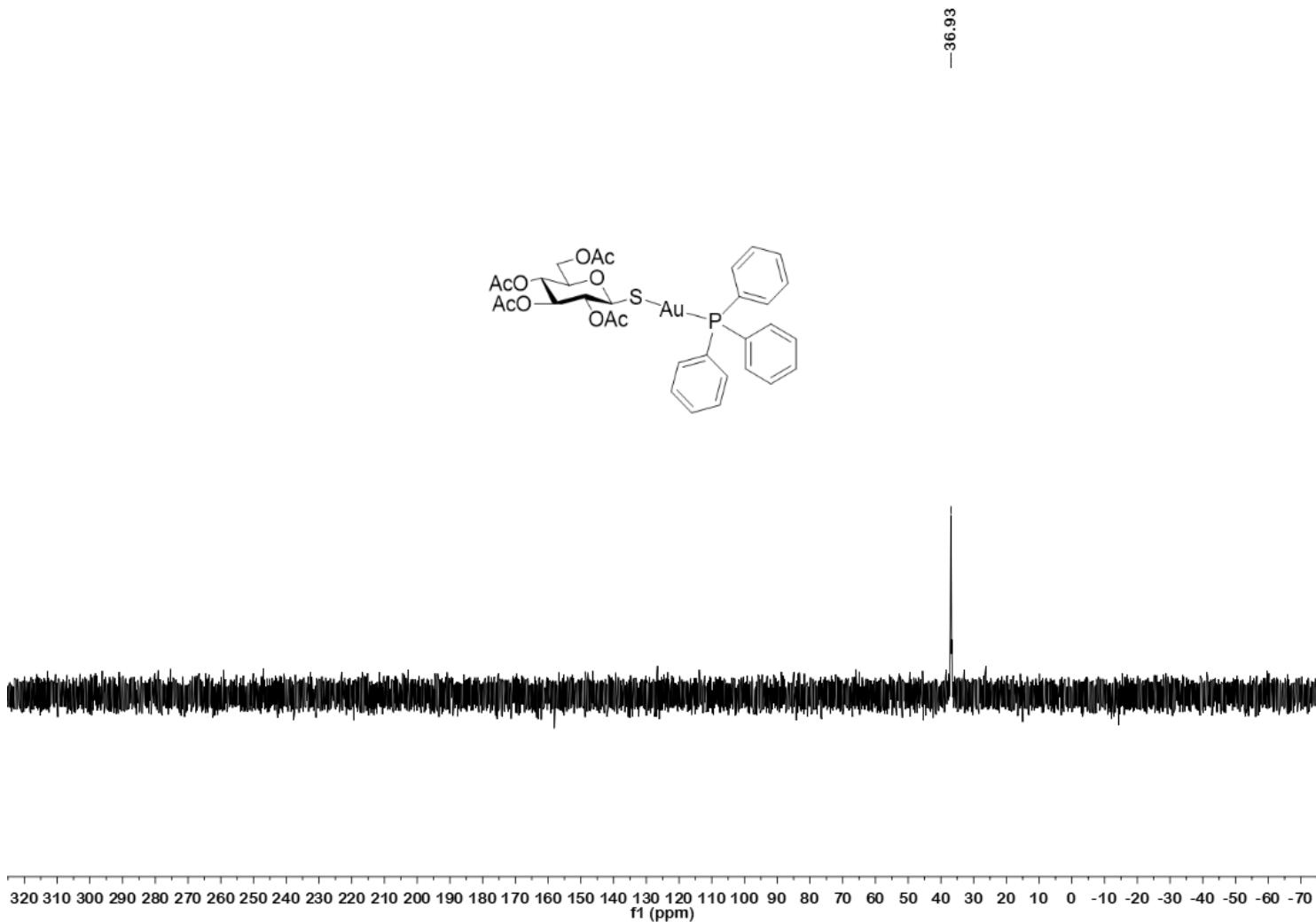
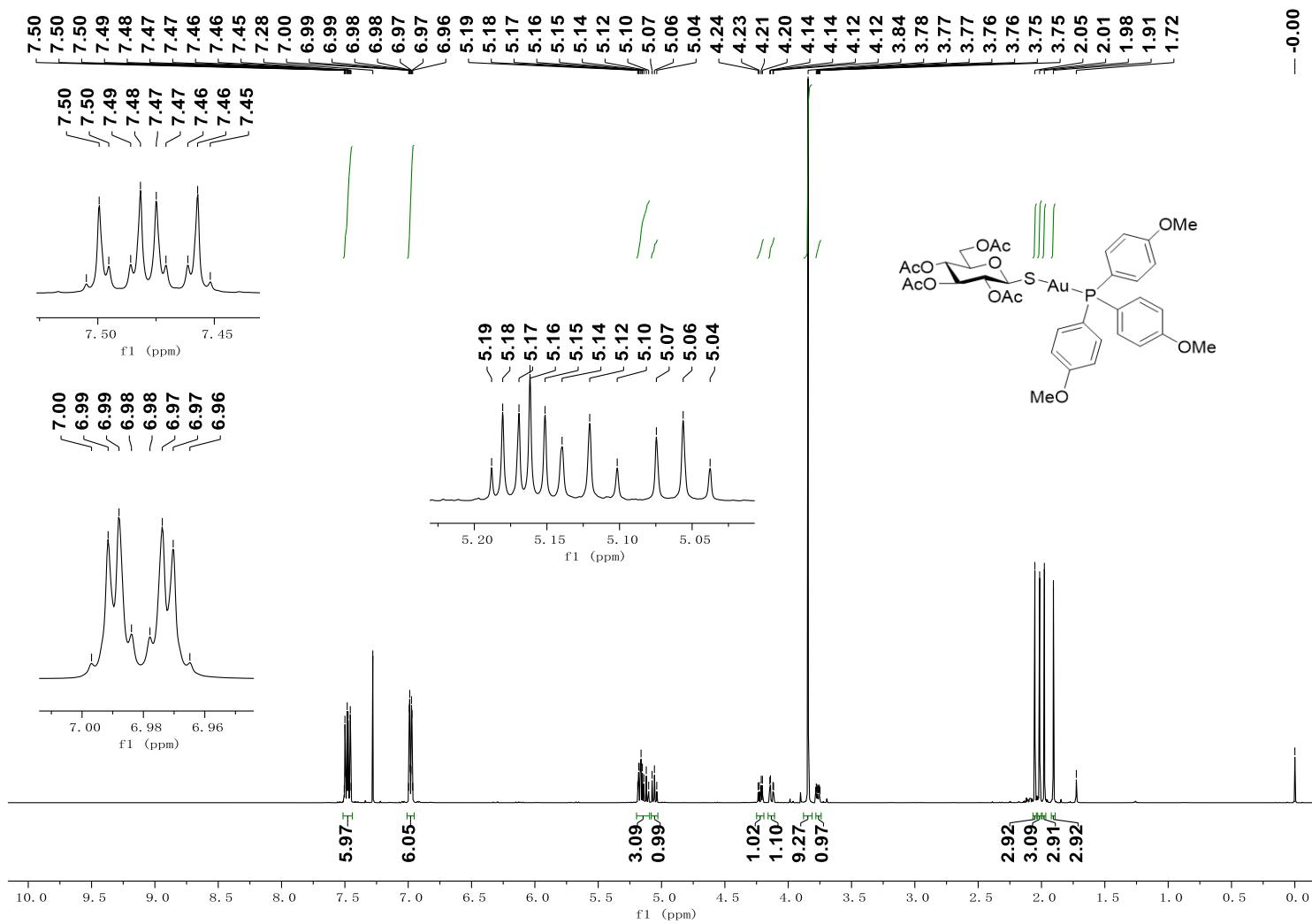


Figure S118. ^{31}P NMR spectrum of compound 33 in CDCl_3 .



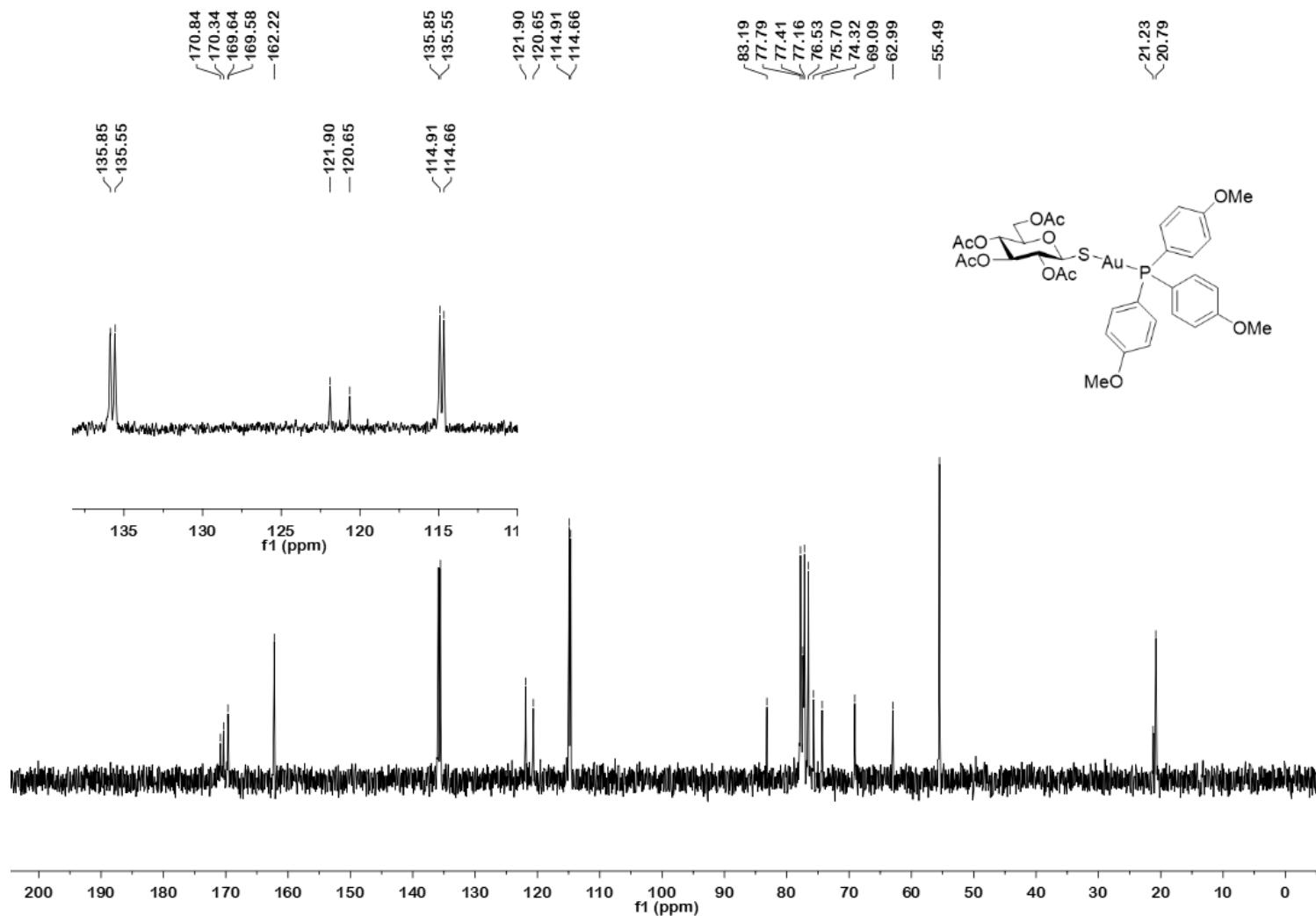


Figure S120. ^{13}C NMR spectrum of compound **34** in CDCl_3 .

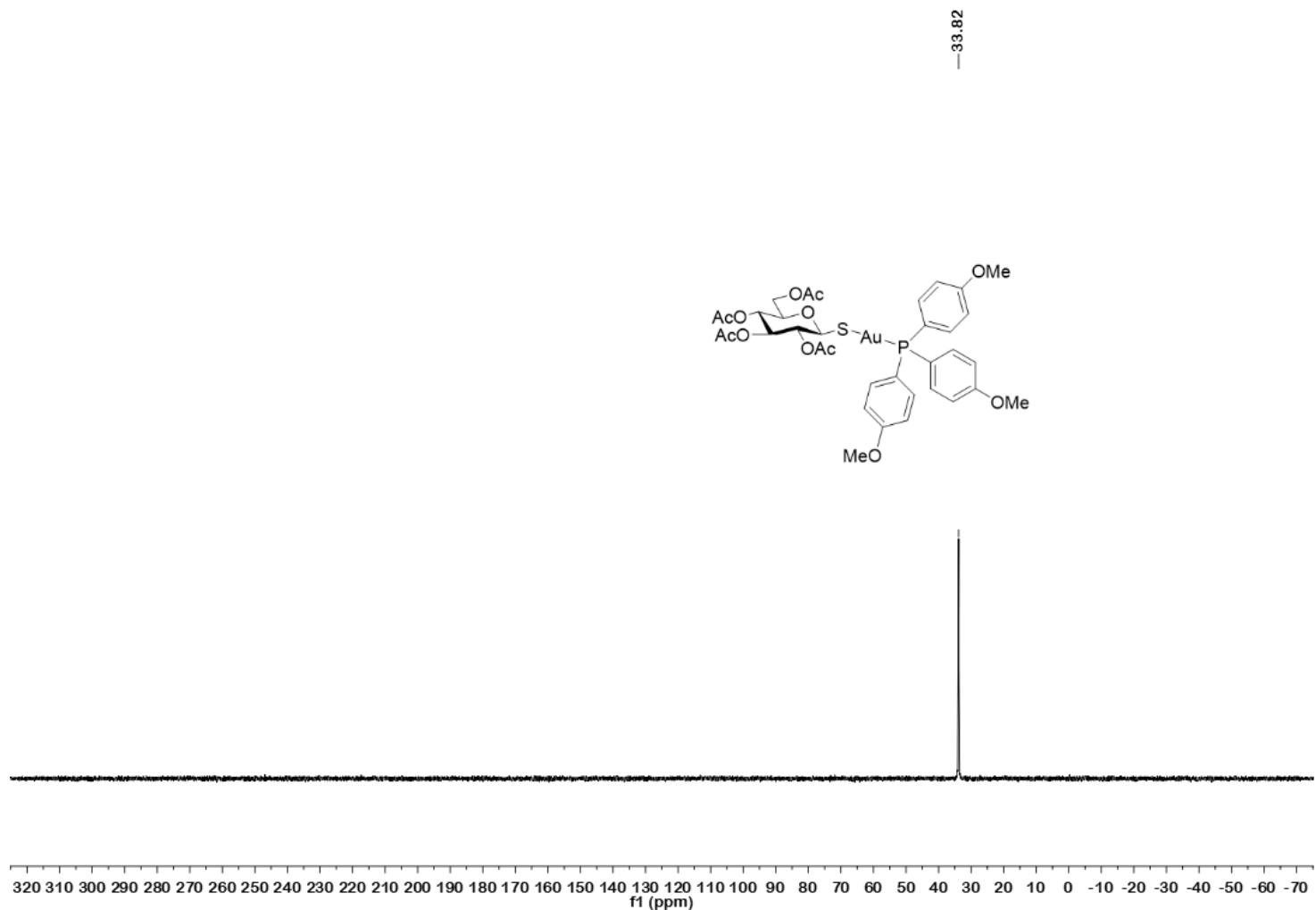


Figure S121. ^{31}P NMR spectrum of compound 34 in CDCl_3 .

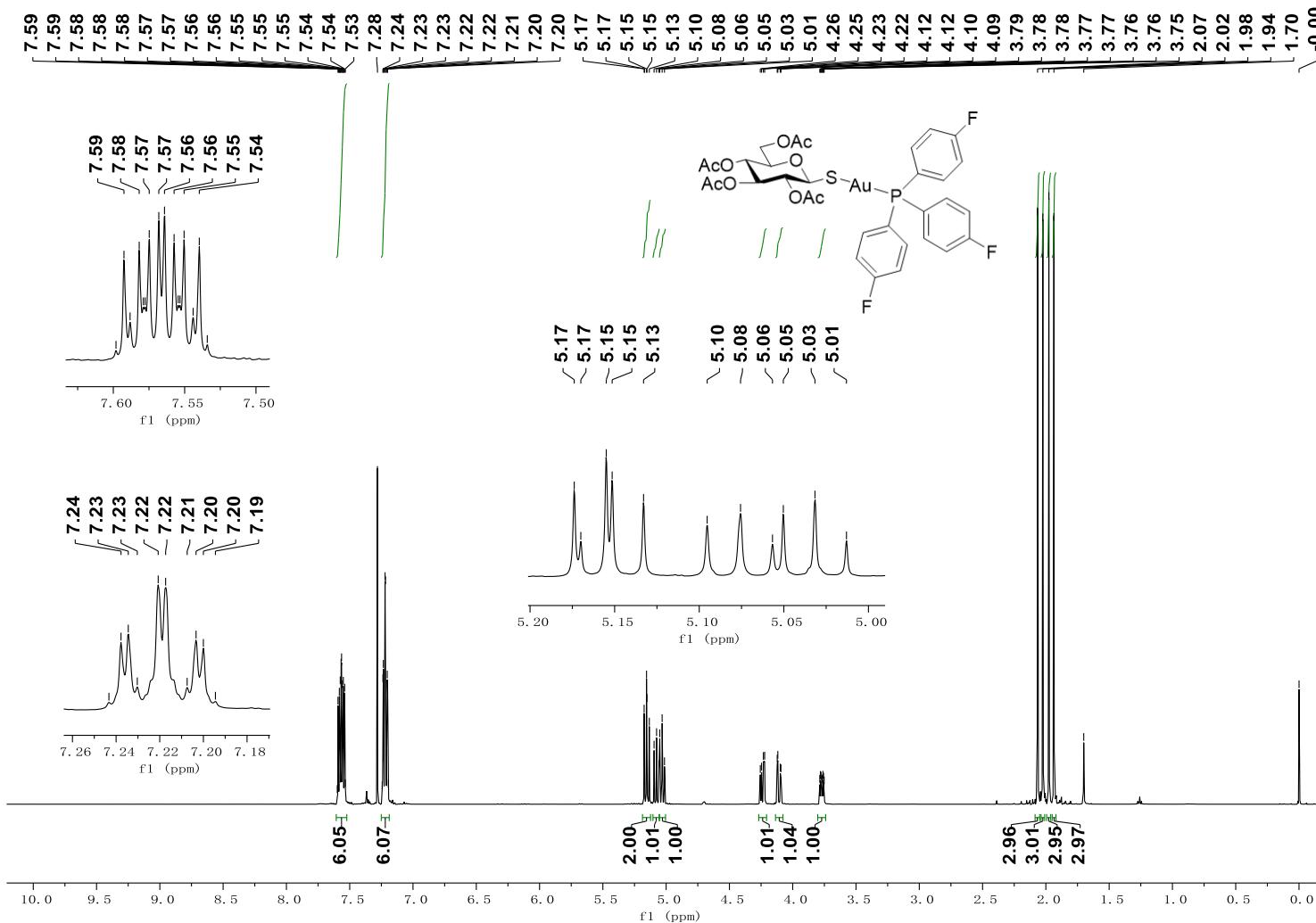
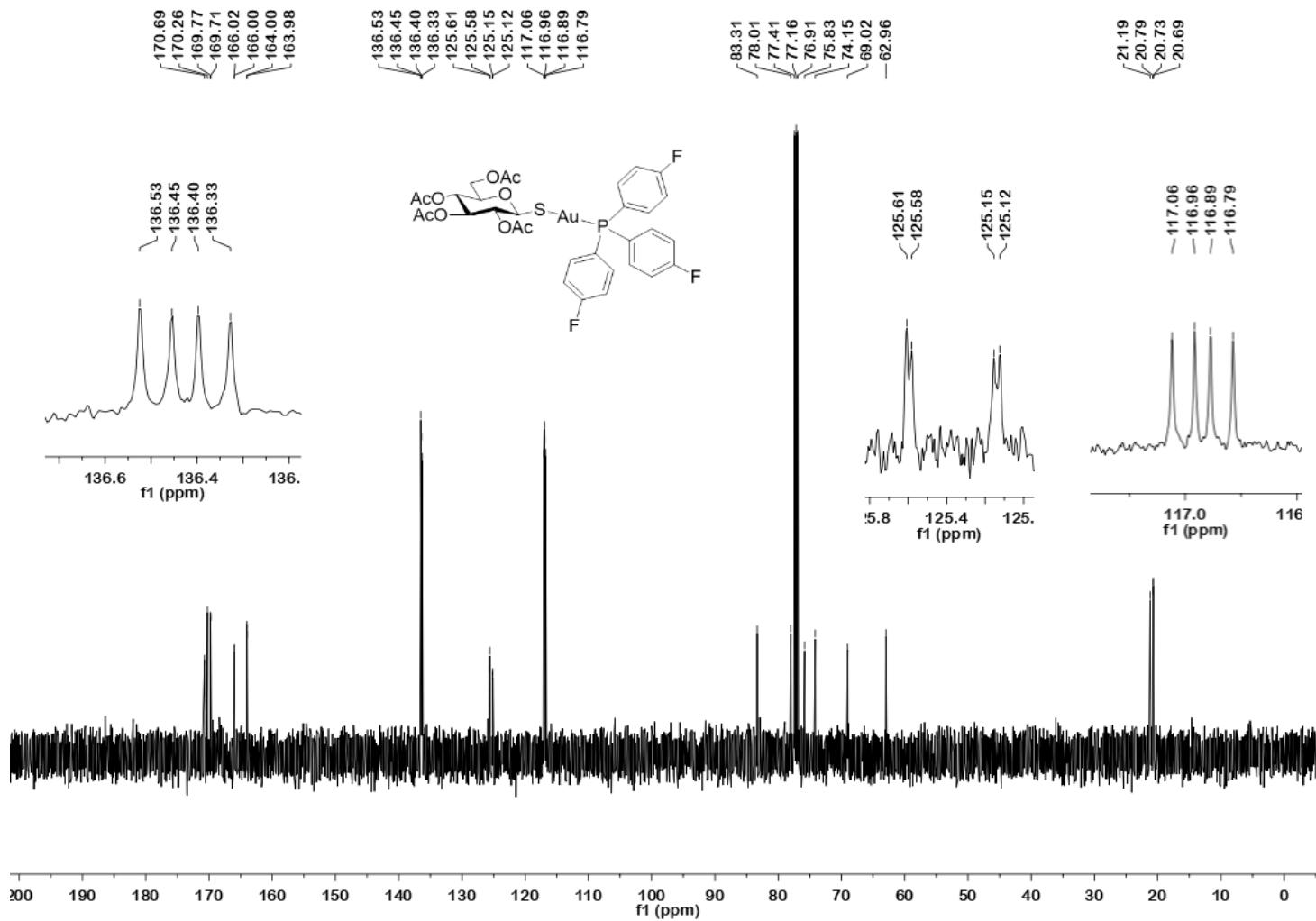


Figure S122. ^1H NMR spectrum of compound 35 in CDCl_3 .



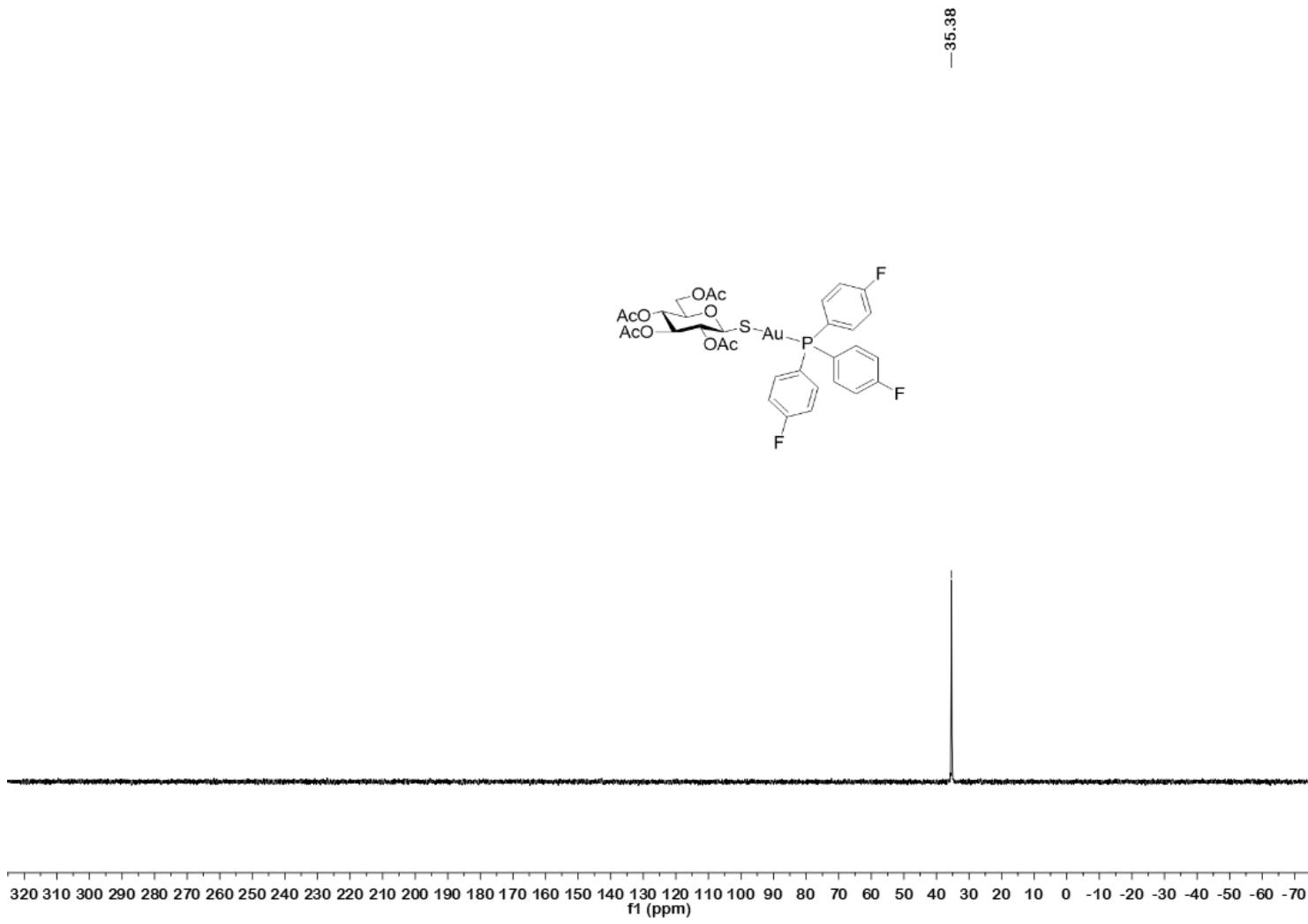


Figure S124. ^{31}P NMR spectrum of compound **35** in CDCl_3 .

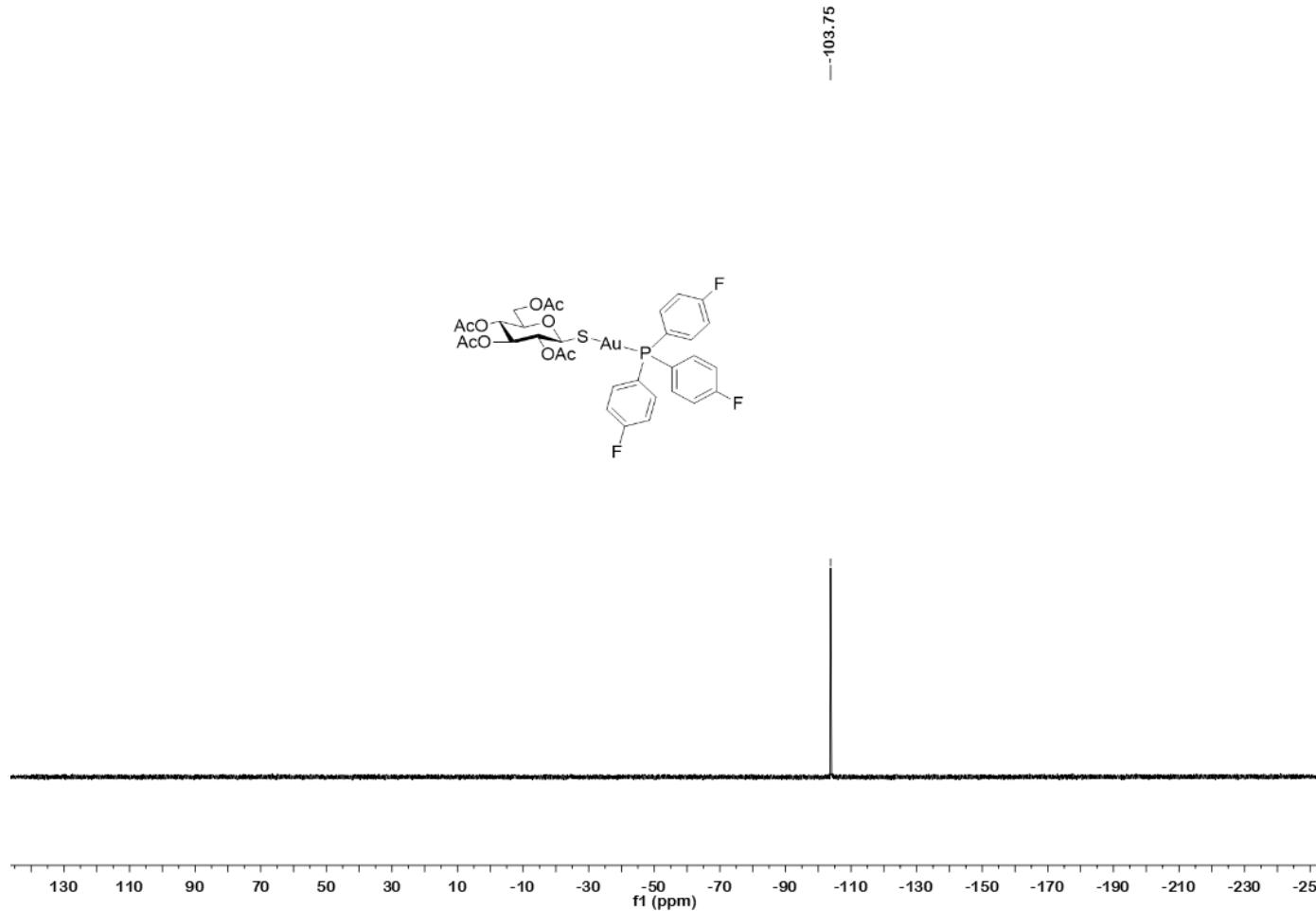


Figure S125. ^{19}F NMR spectrum of compound 35 in CDCl₃.

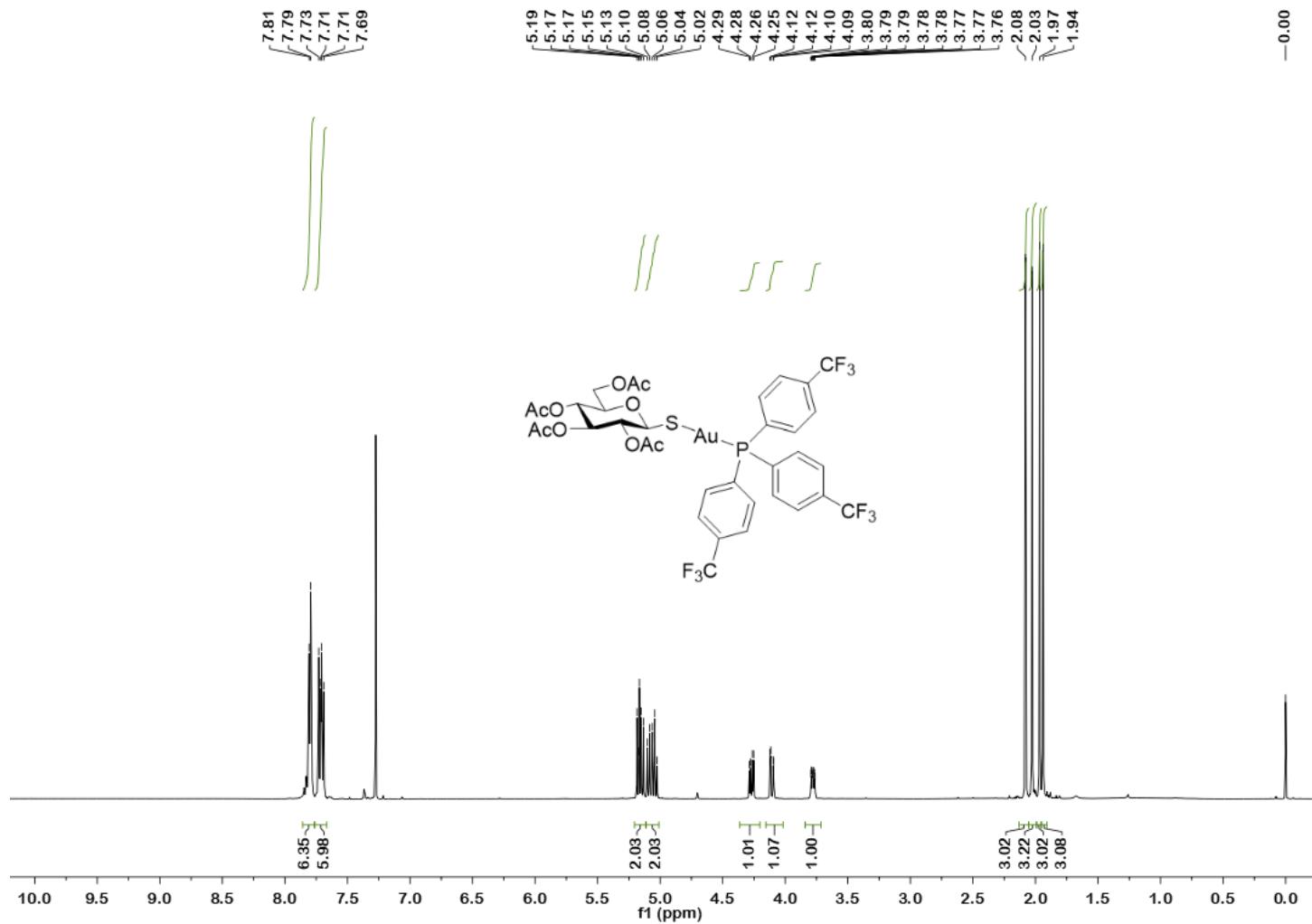


Figure S126. ^1H NMR spectrum of compound **36** in CDCl_3 .

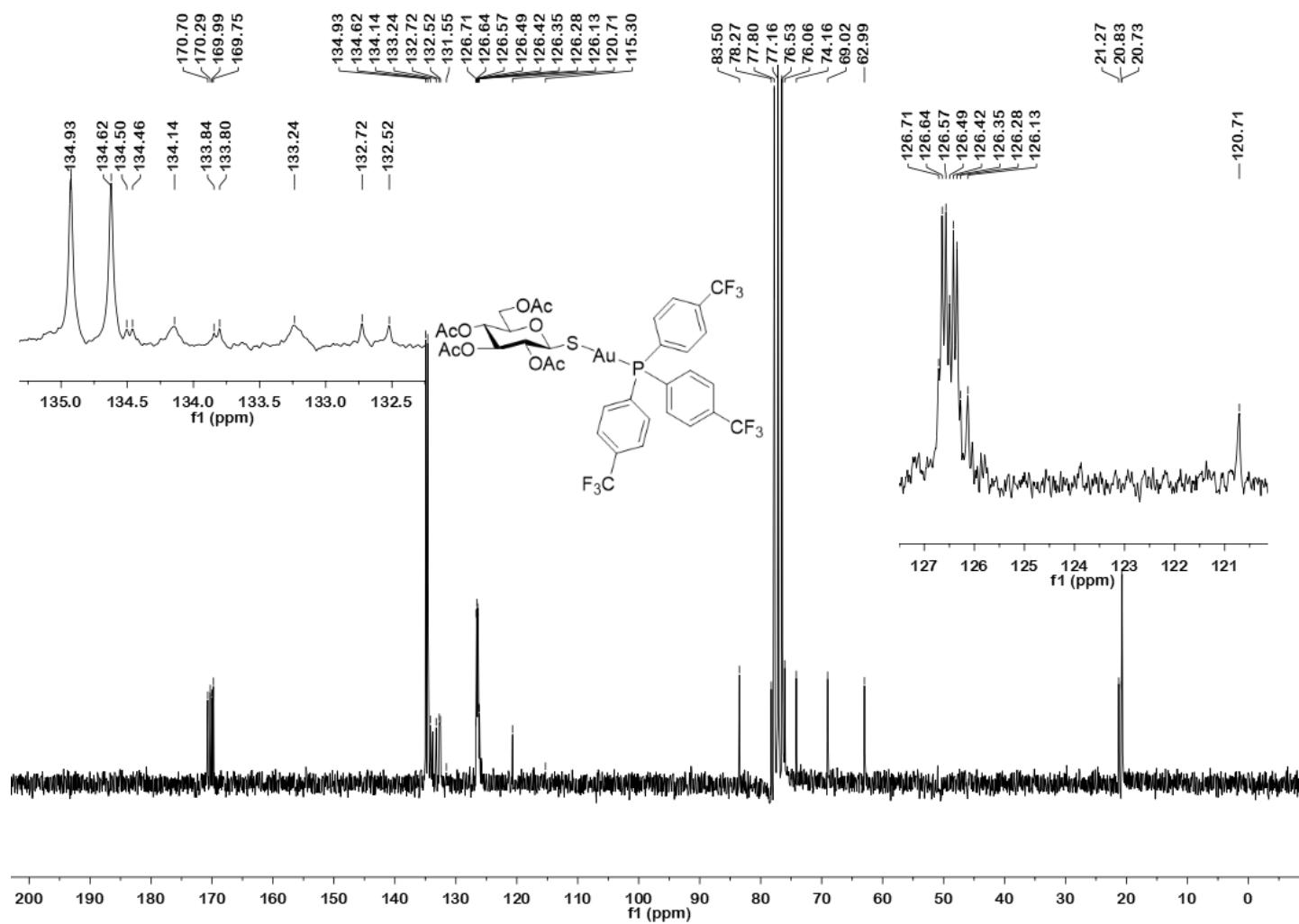


Figure S127. ^{13}C NMR spectrum of compound **36** in CDCl_3 .

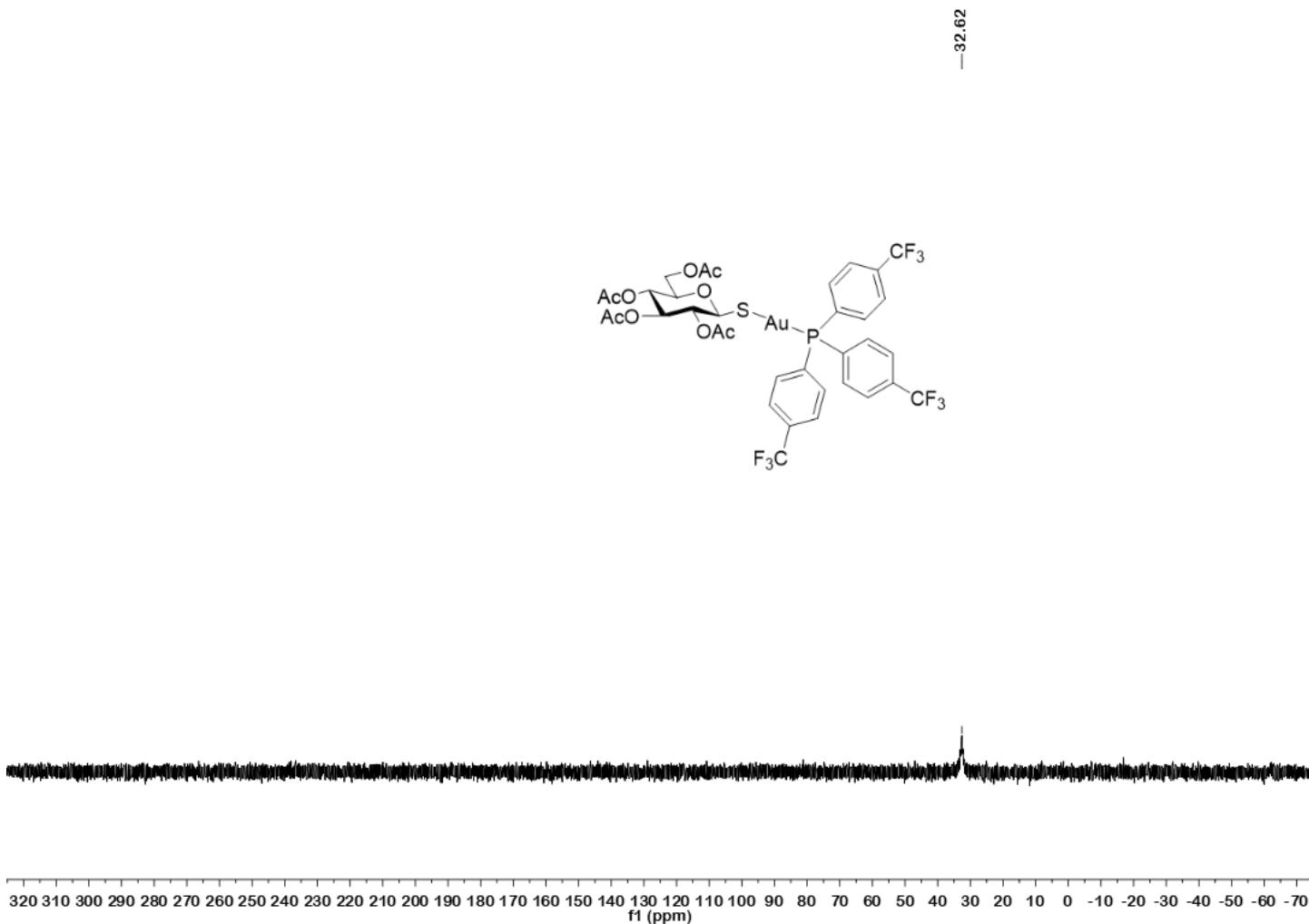


Figure S128. ^{31}P NMR spectrum of compound 36 in CDCl_3 .

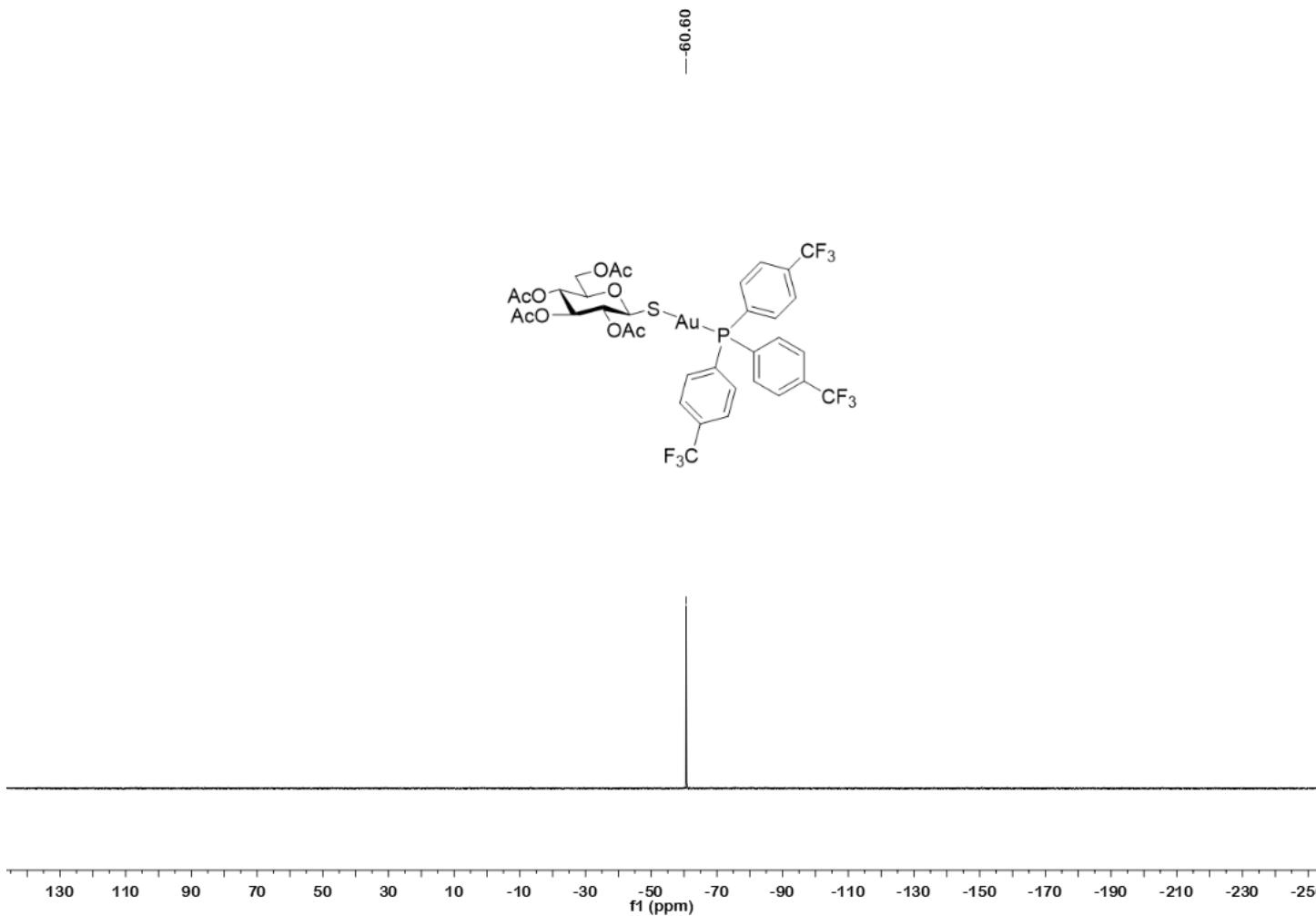


Figure S129. ^{19}F NMR spectrum of compound 36 in CDCl₃.

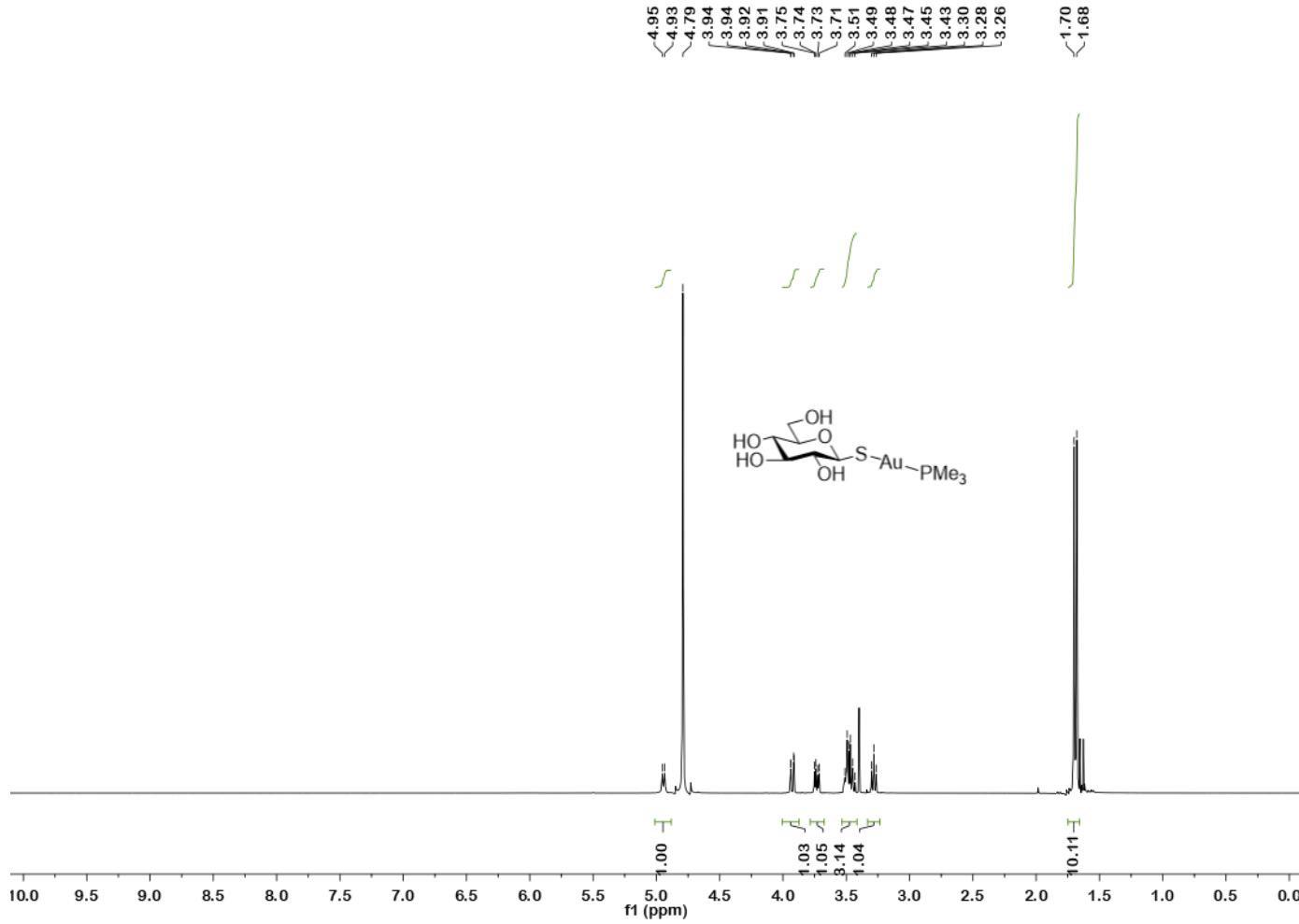


Figure S130. ^1H NMR spectrum of compound **37** in D_2O .

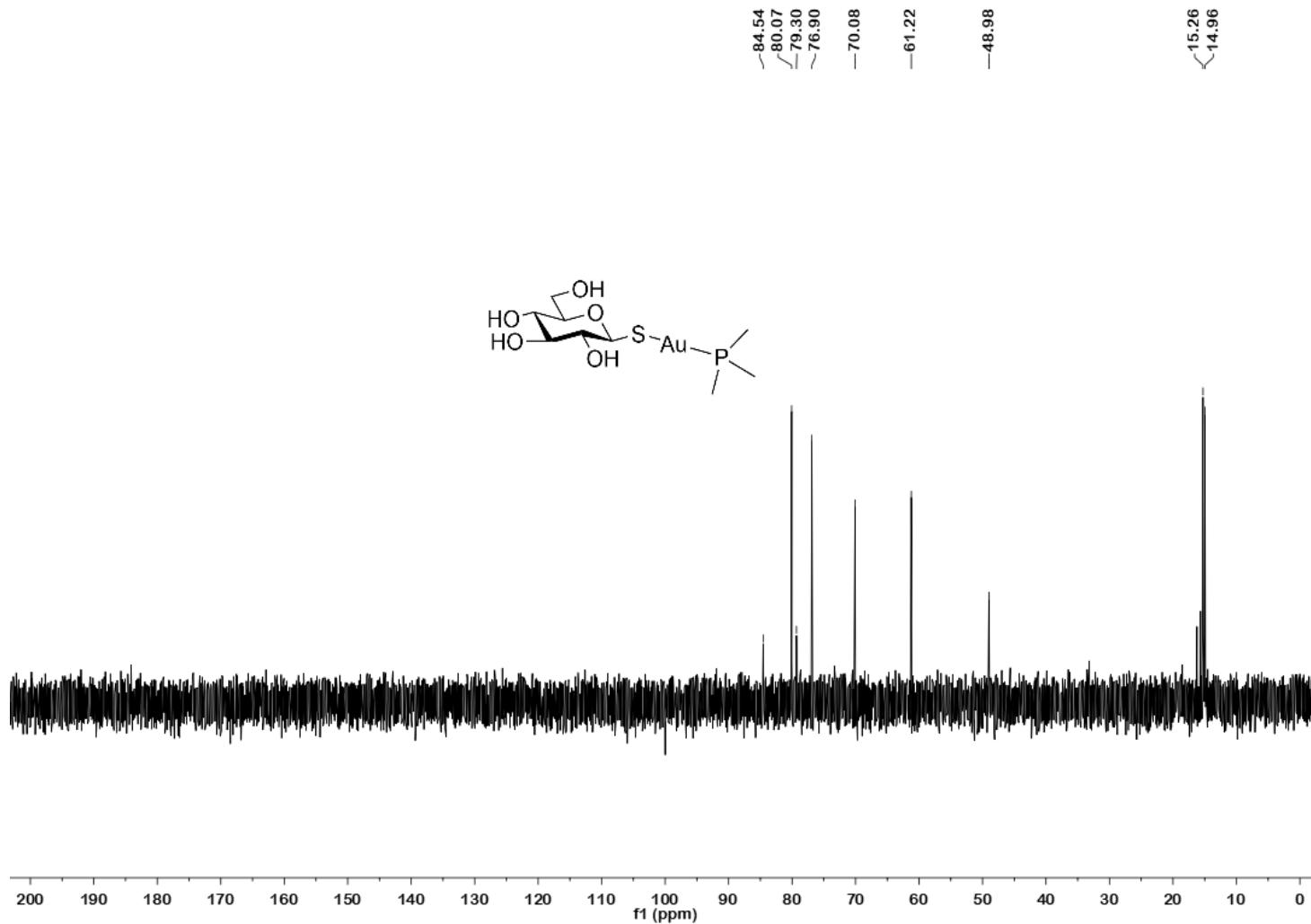


Figure S131. ^{13}C NMR spectrum of compound 37 in D_2O .

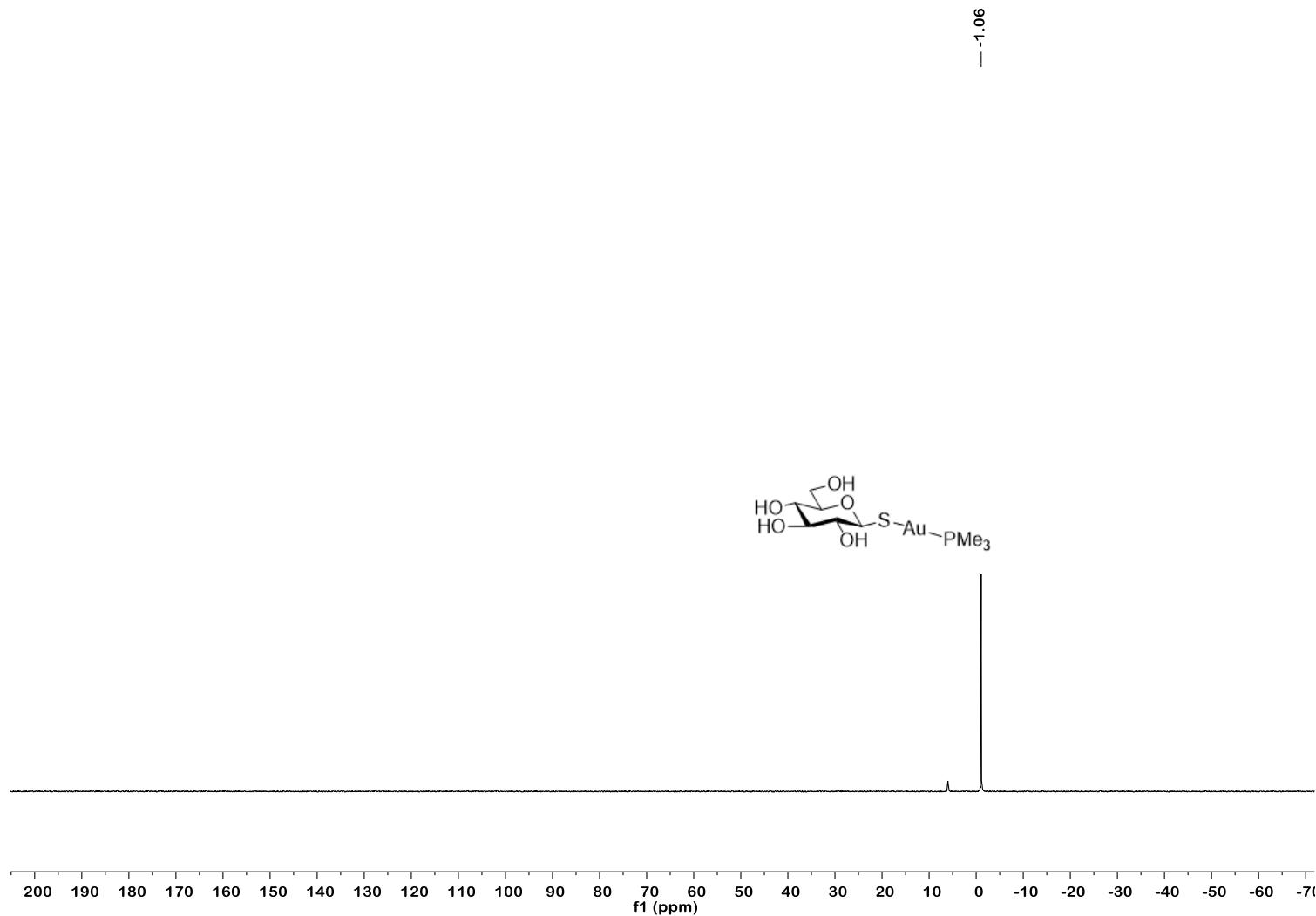


Figure S132. ^{31}P NMR spectrum of compound **37** in D_2O .

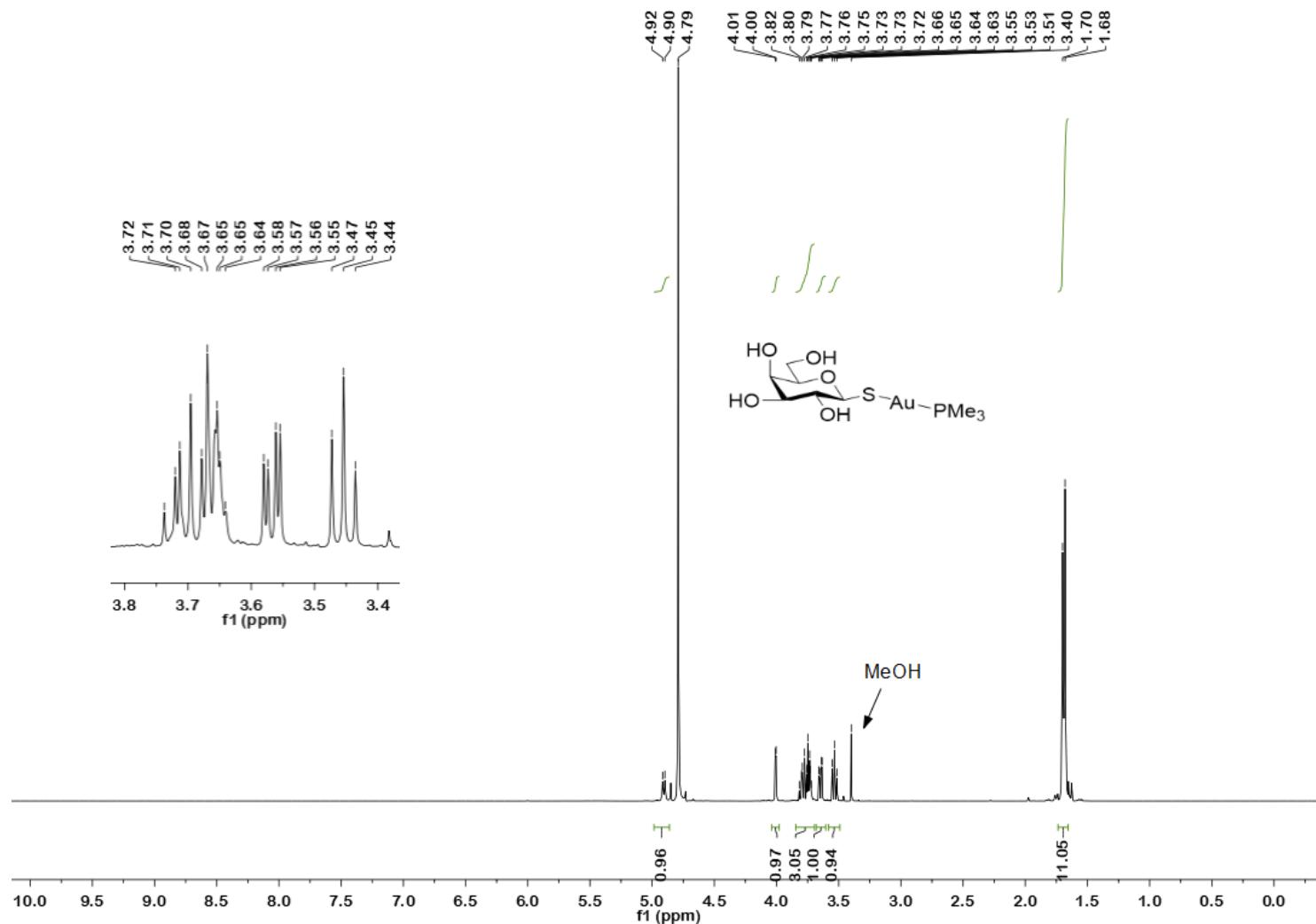


Figure S133. ^1H NMR spectrum of compound **38** in D_2O

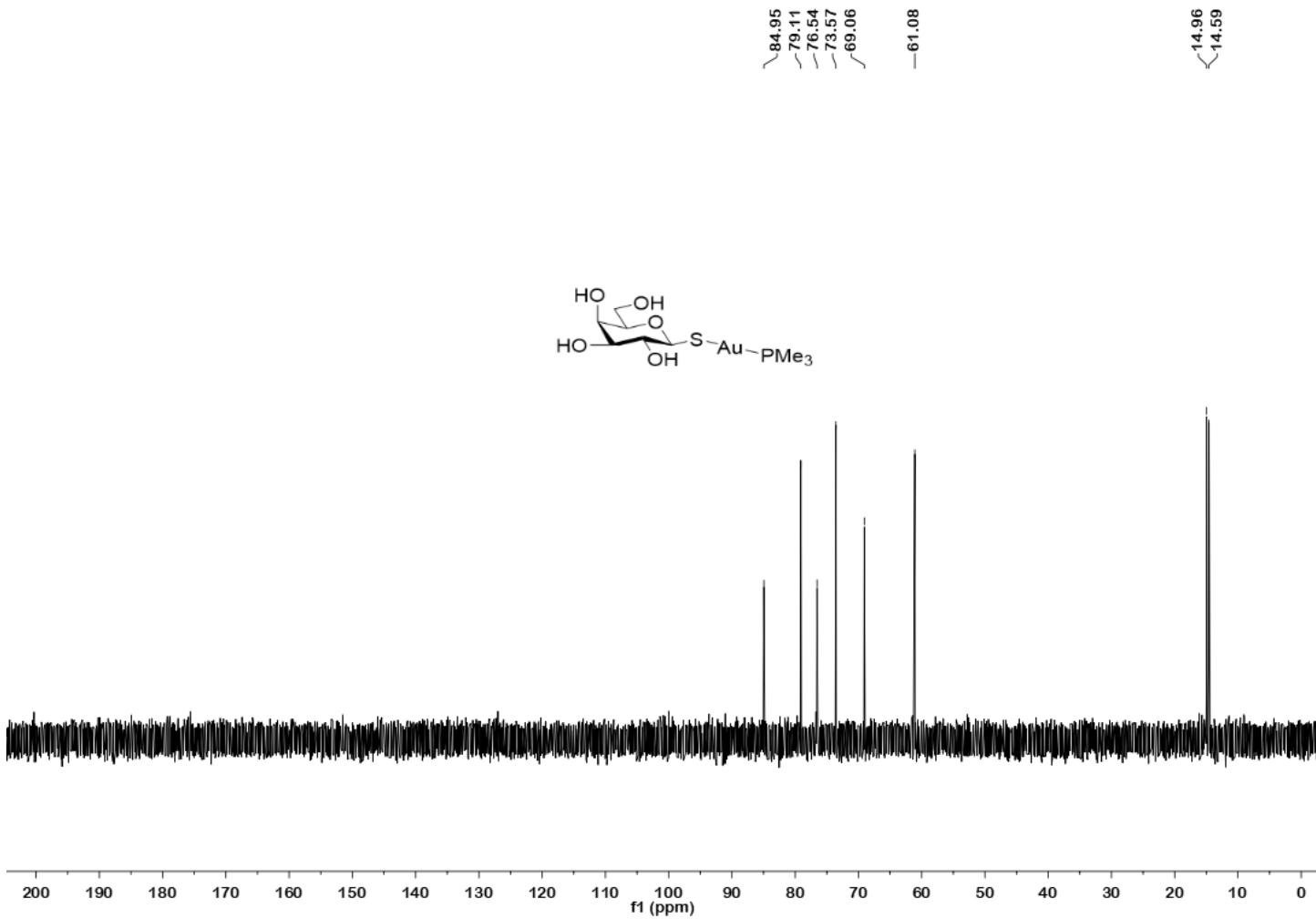


Figure S134. ^{13}C NMR spectrum of compound **38** in D_2O .

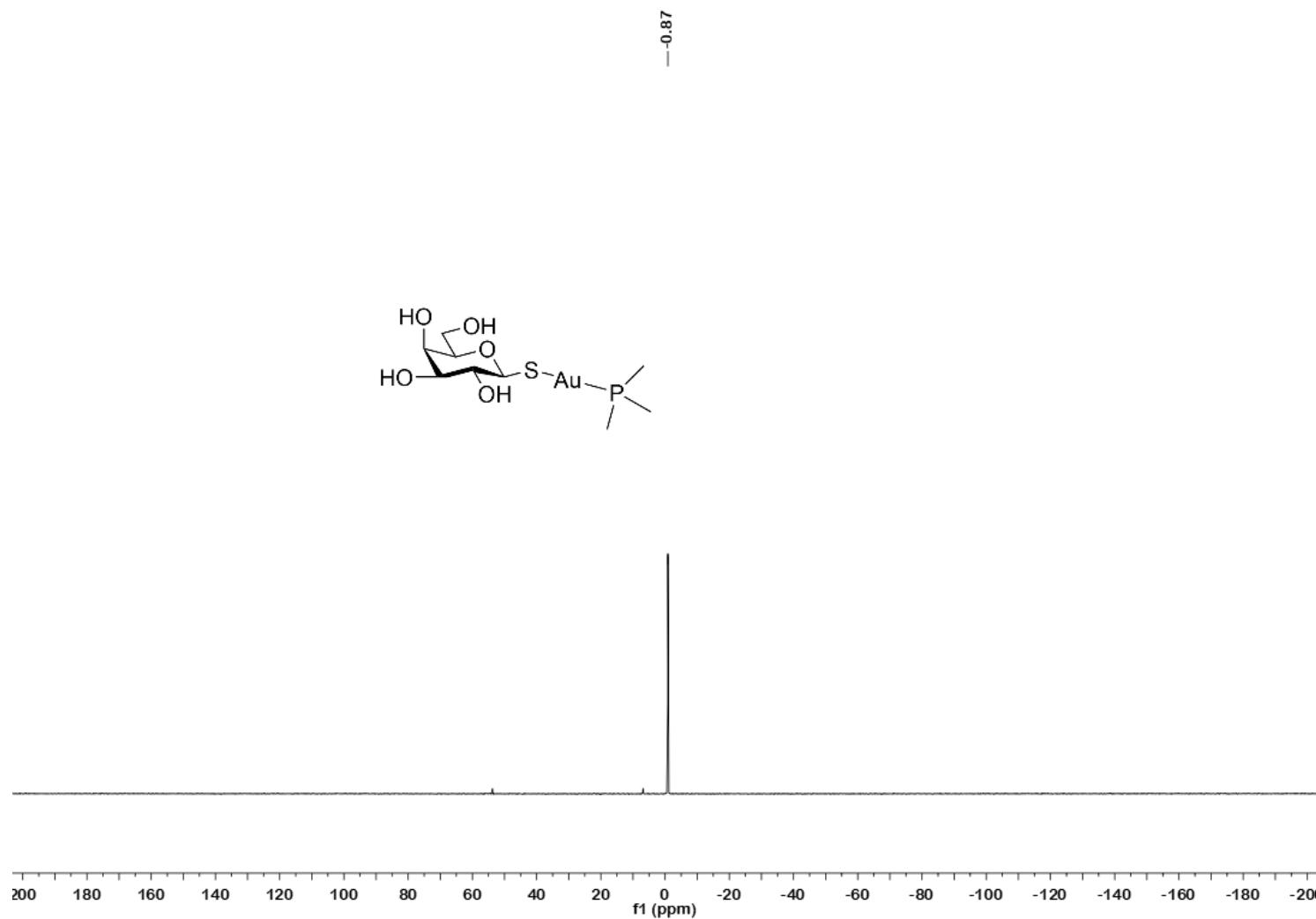


Figure S135. ^{31}P NMR spectrum of compound **38** in D_2O .

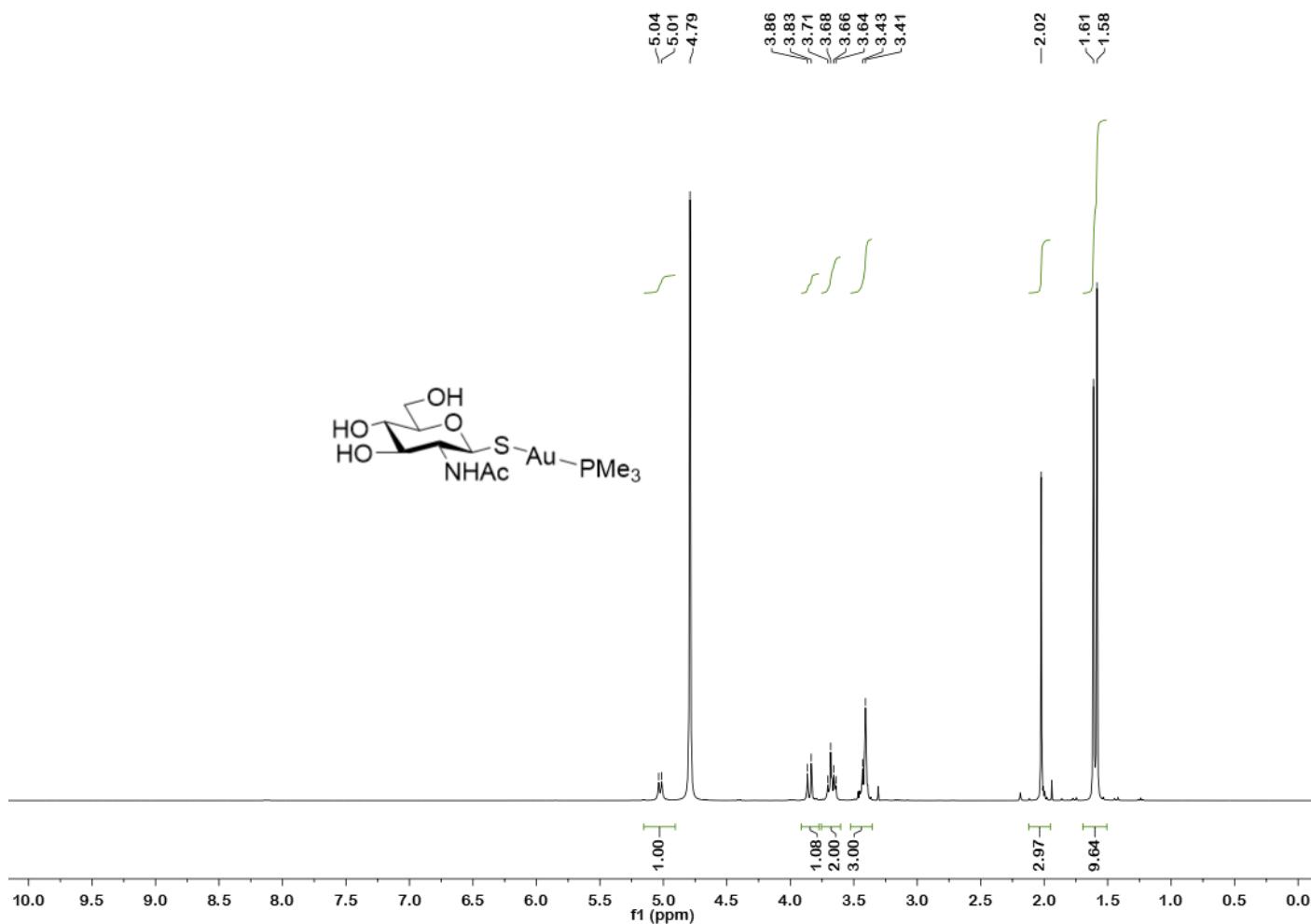


Figure S136. ^1H NMR spectrum of compound **39** in D_2O

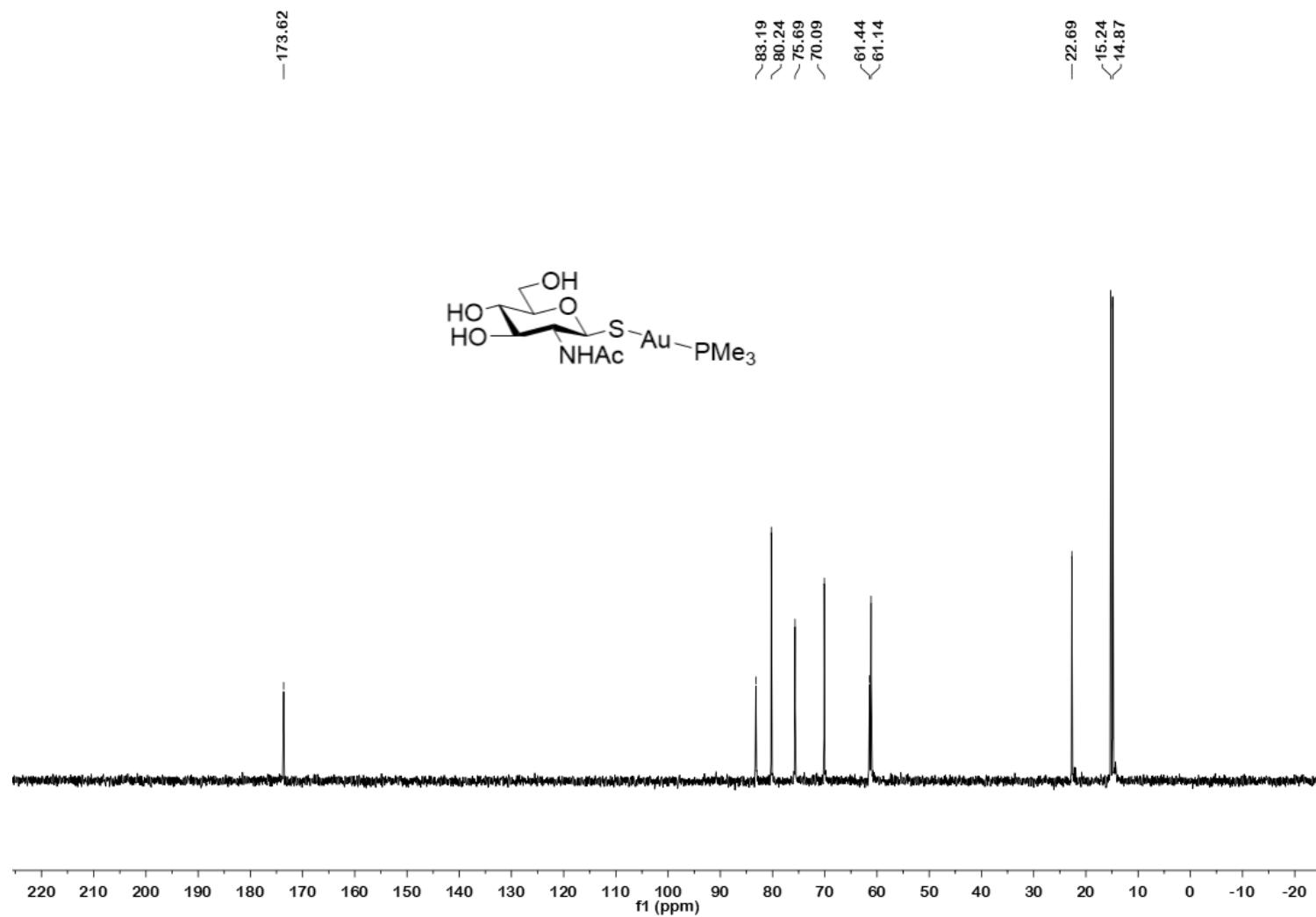


Figure S137. ^{13}C NMR spectrum of compound **39** in D_2O .

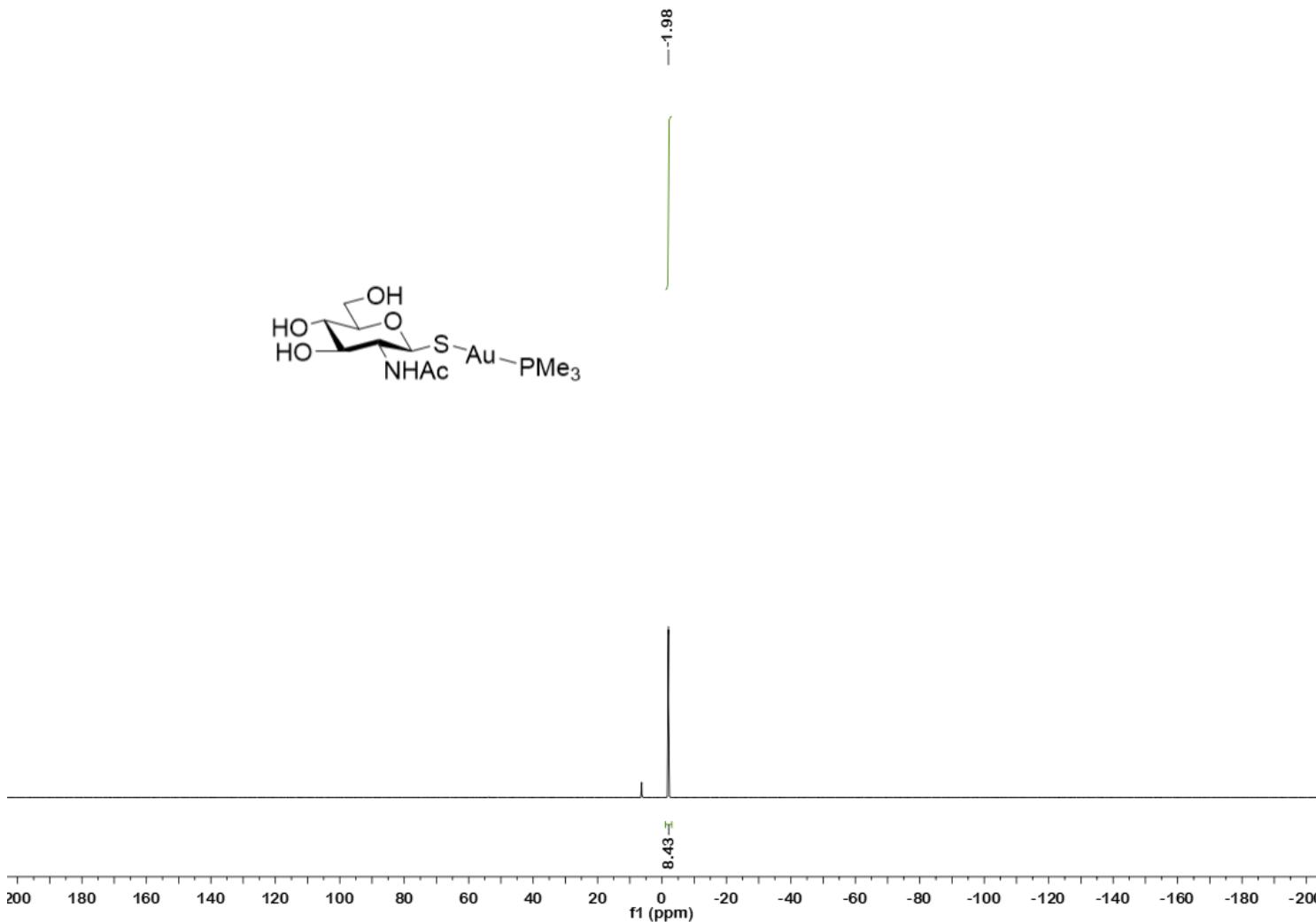


Figure S138. ^{31}P NMR spectrum of compound **39** in D_2O .

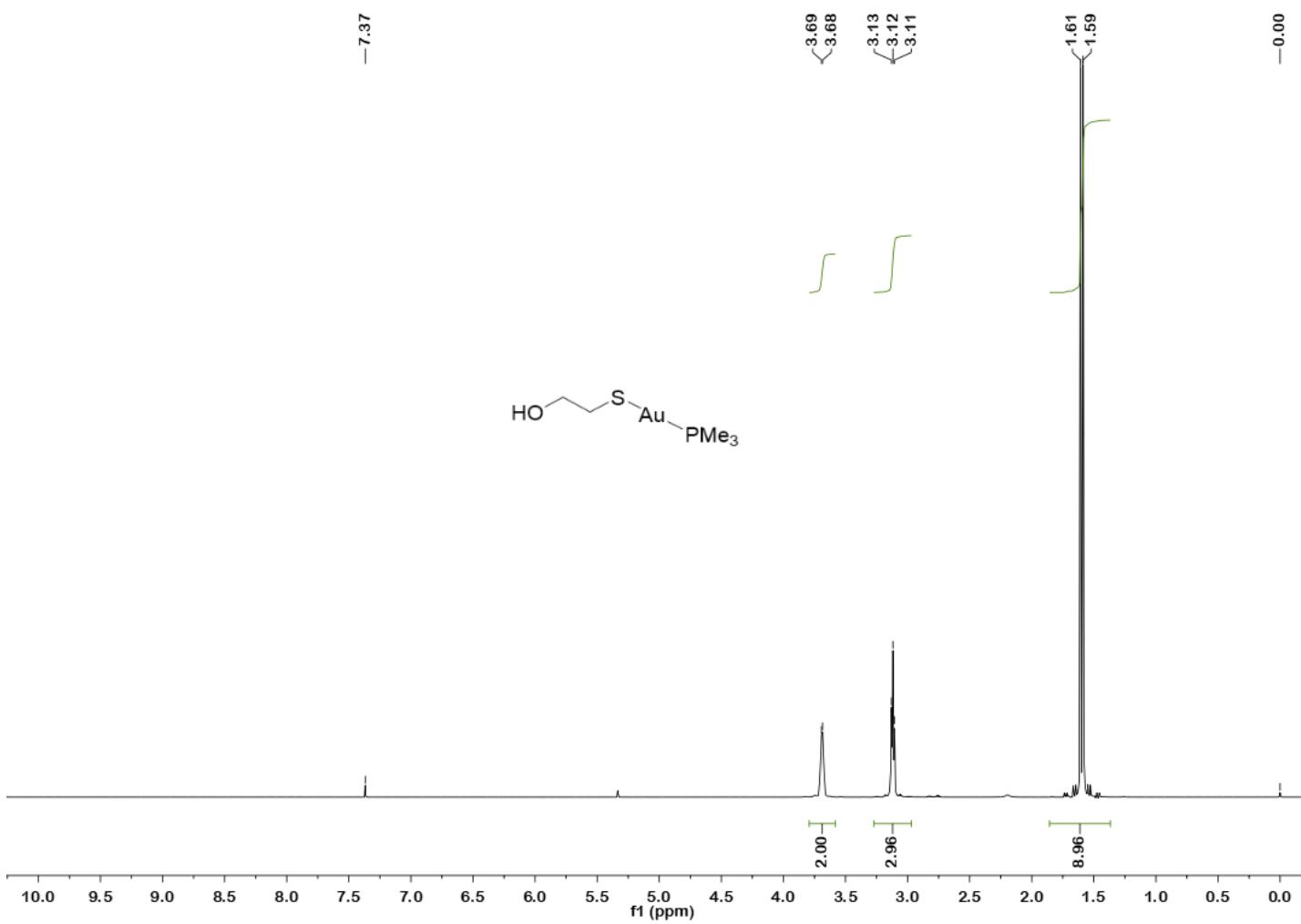


Figure S139. ¹H NMR spectrum of compound **40** in CDCl_3 .

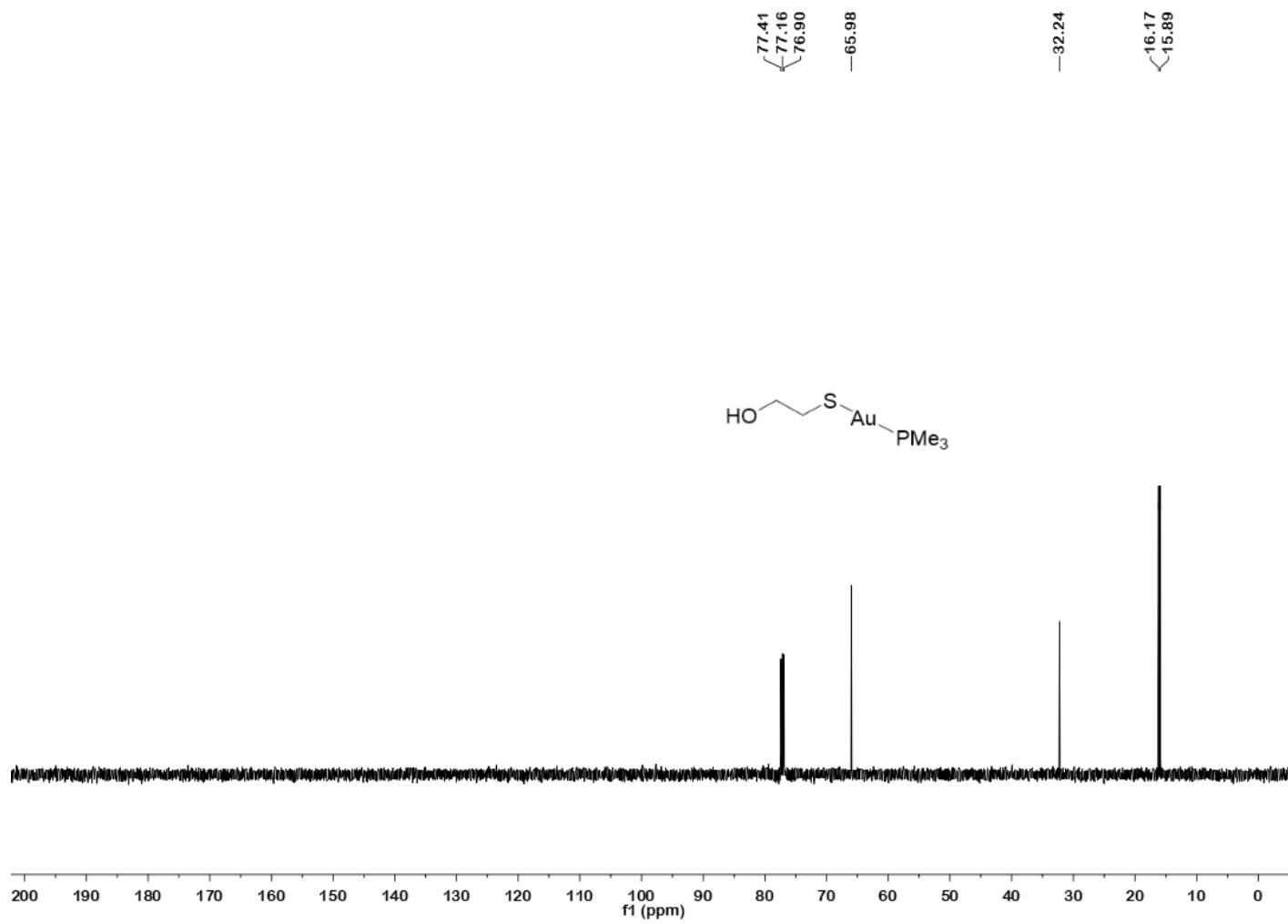


Figure S140. ¹³C NMR spectrum of compound **40** in CDCl_3 .

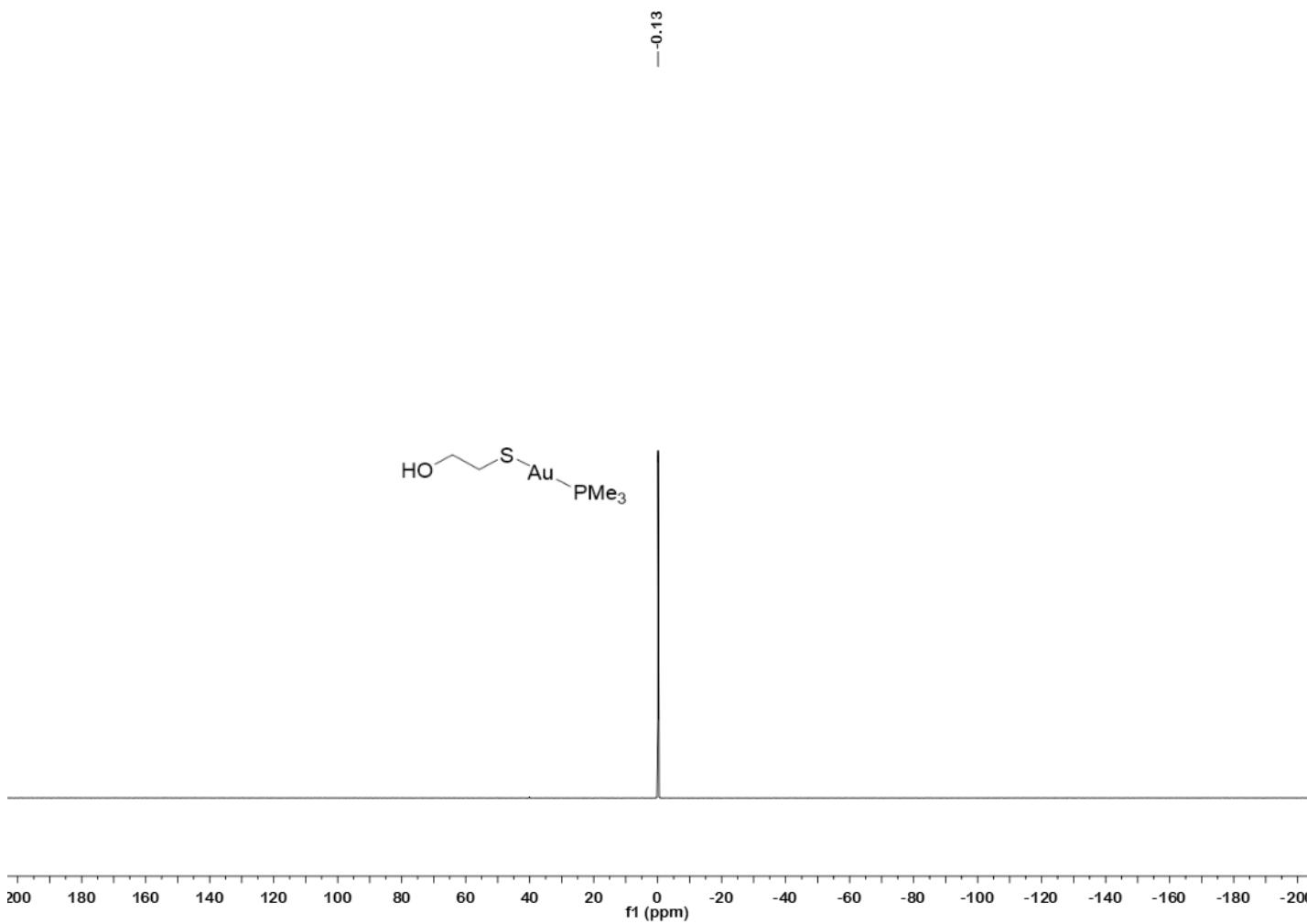
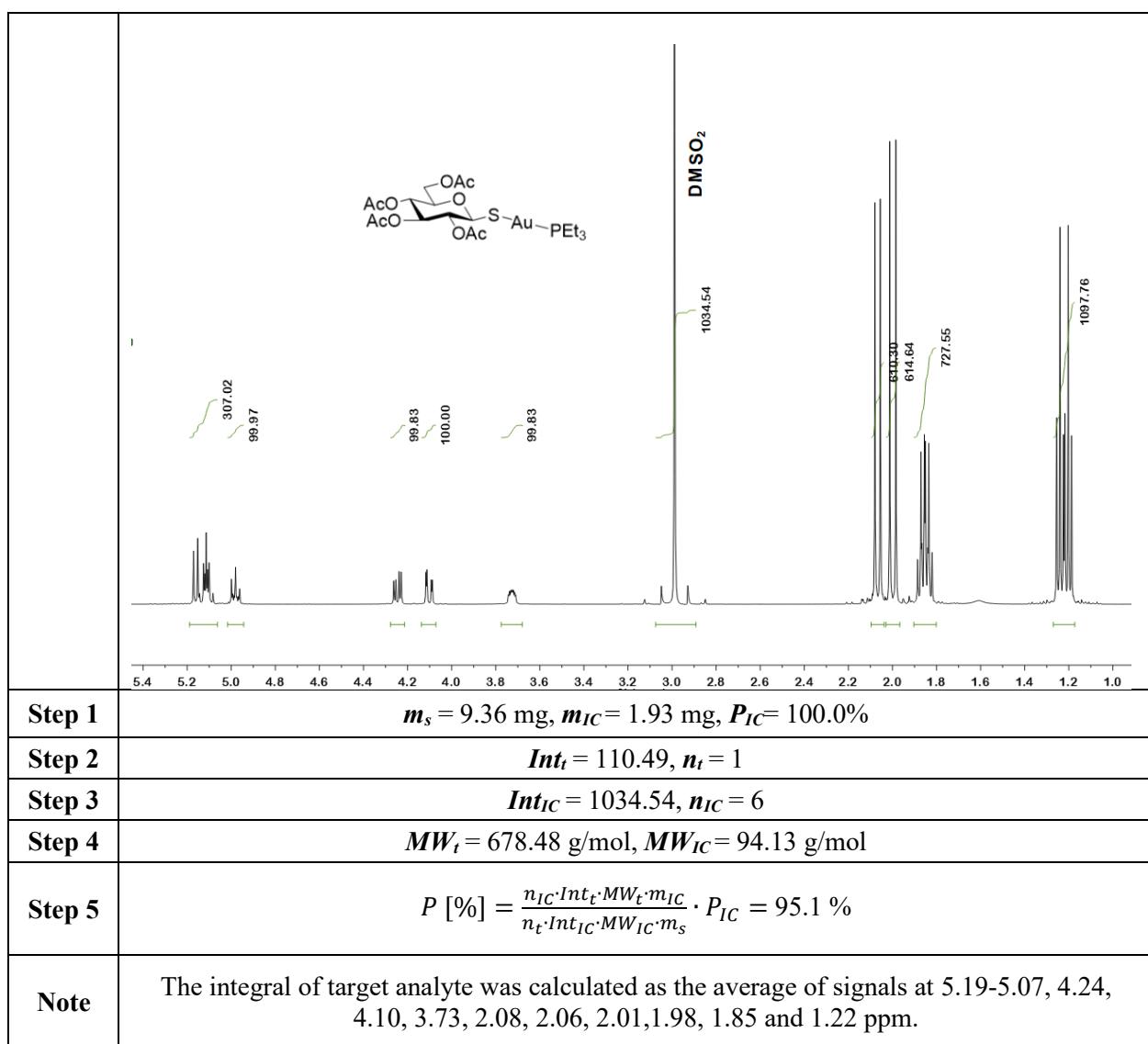


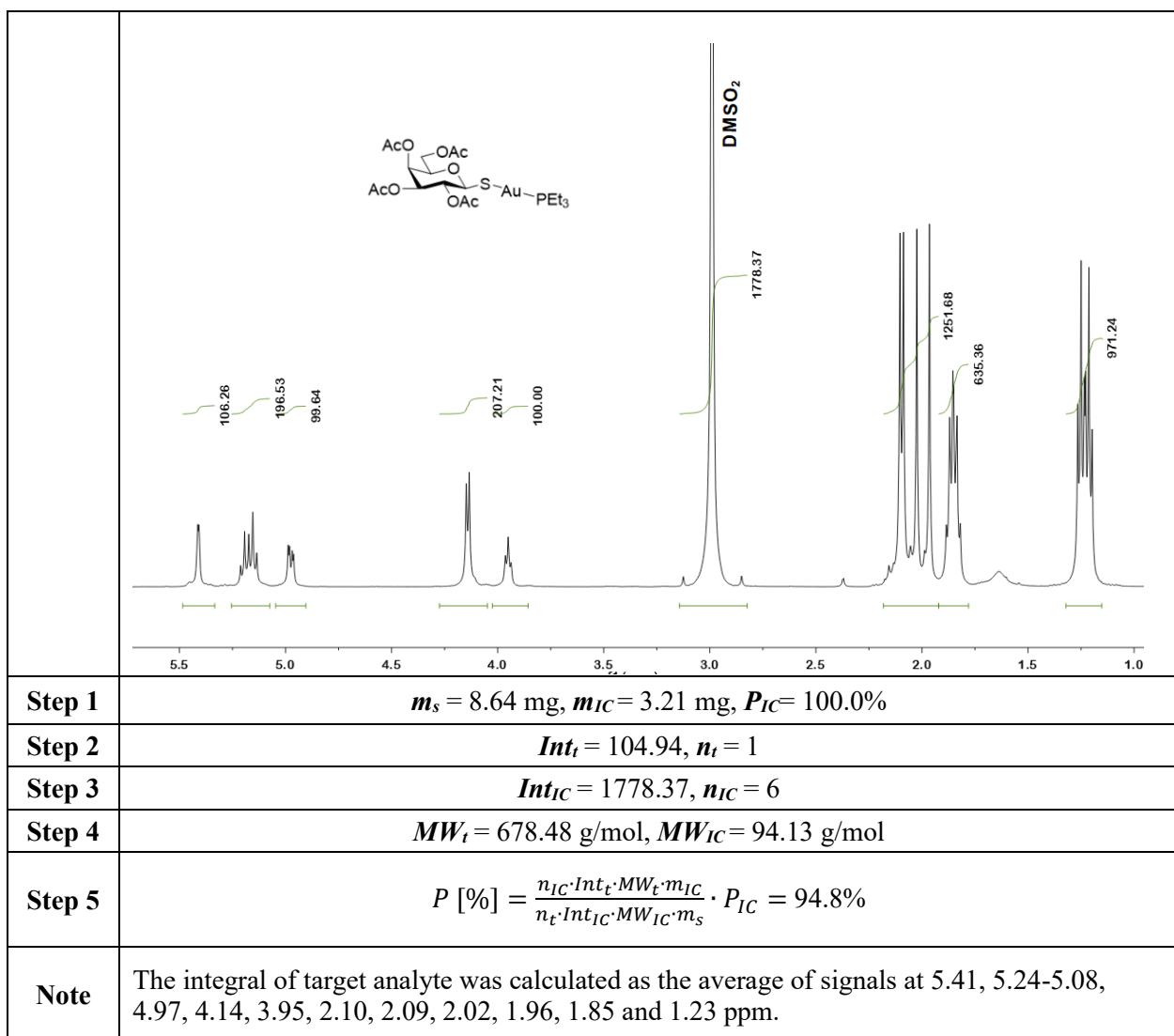
Figure S141. ^{31}P NMR spectrum of compound **40** in CDCl_3 .

Determination of purity of Compounds 1 – 40 by qHNMR.

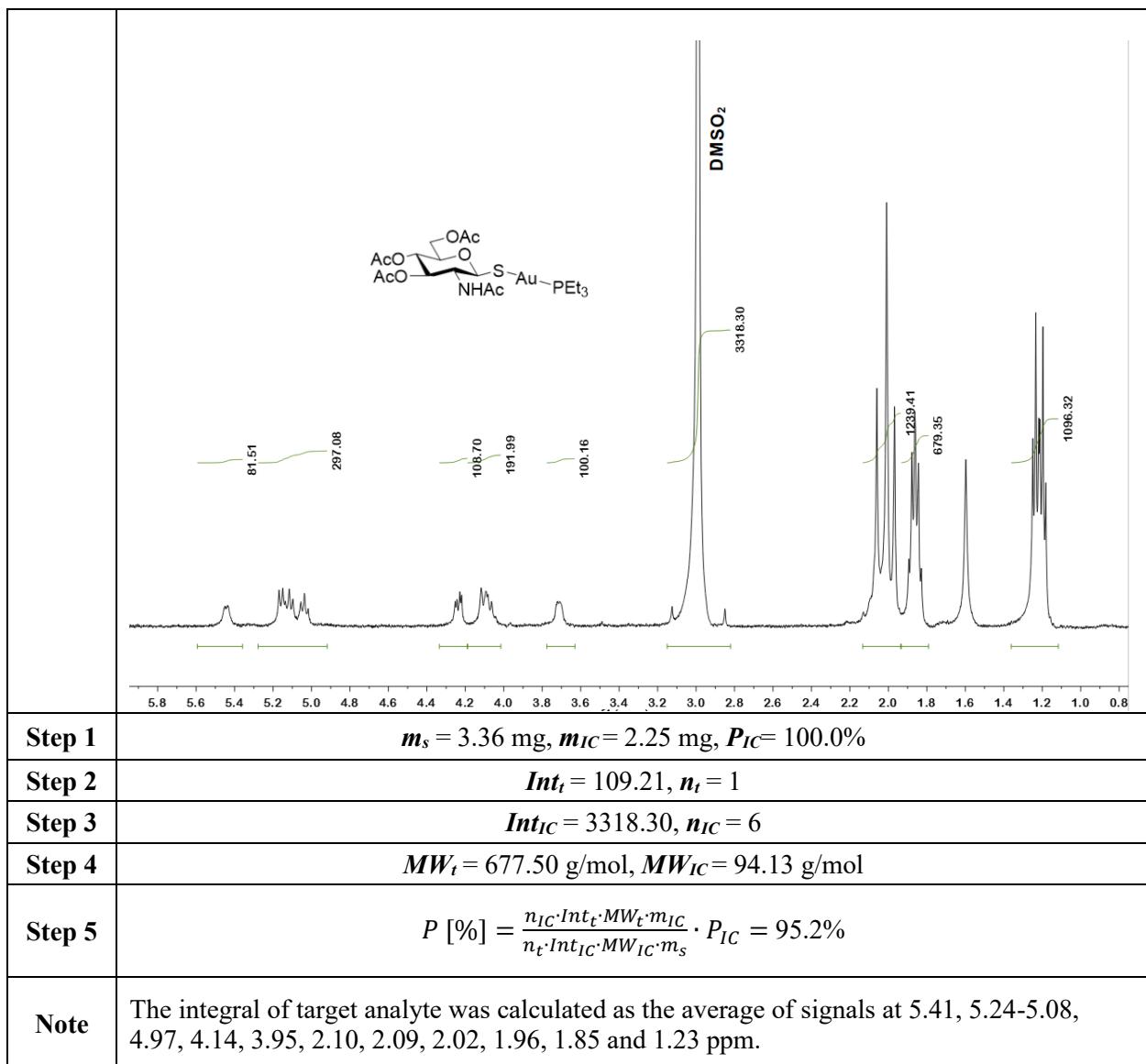
Compound **1** in CDCl_3 with the addition of dimethyl sulfone (DMSO_2 , 100.0%) as internal standard (500 MHz).



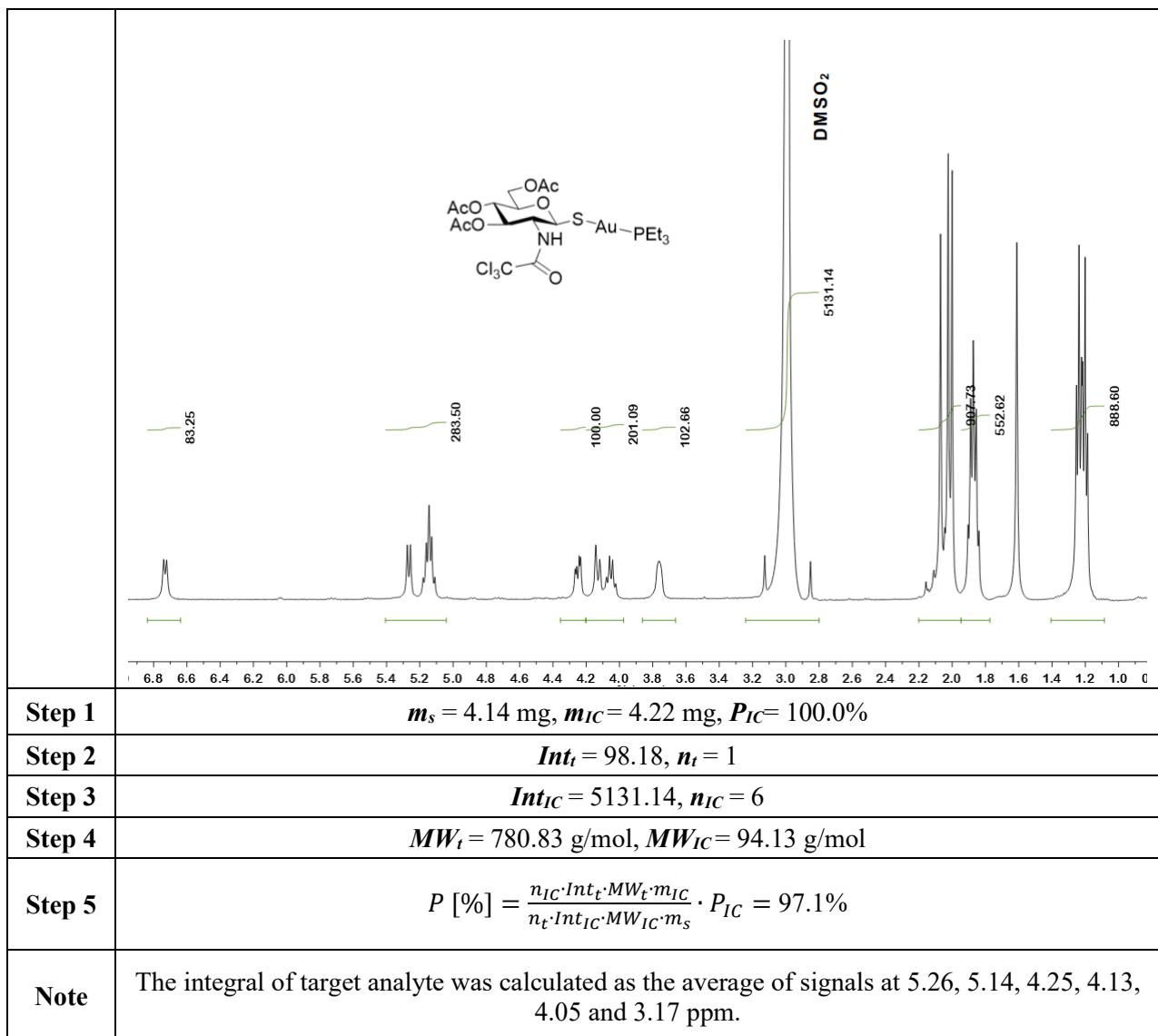
Compound **2** in CDCl_3 with the addition of dimethyl sulfone (DMSO_2 , 100.0%) as internal standard (500 MHz).



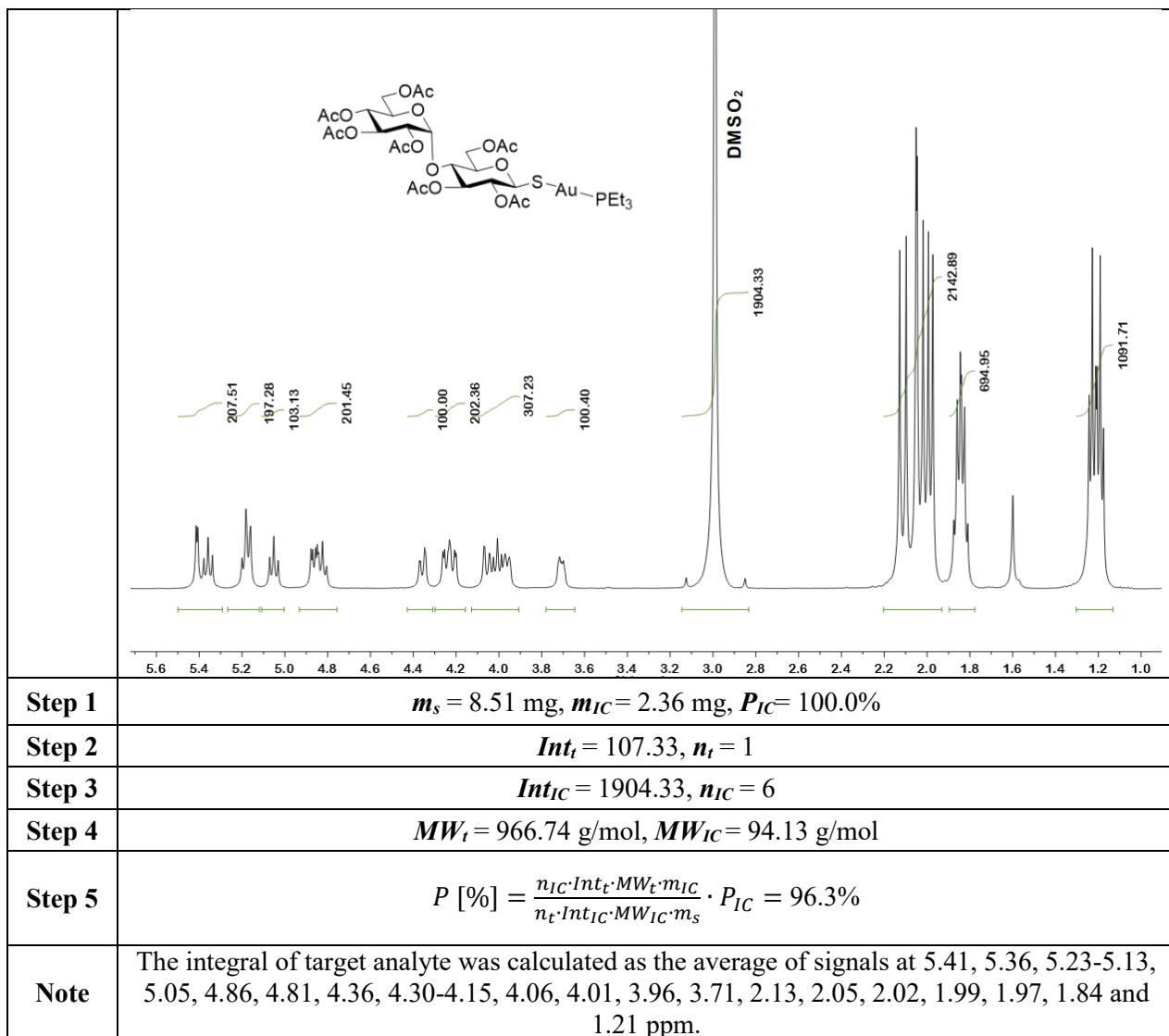
Compound **3** in CDCl_3 with the addition of dimethyl sulfone (DMSO_2 , 100.0%) as internal standard (500 MHz).



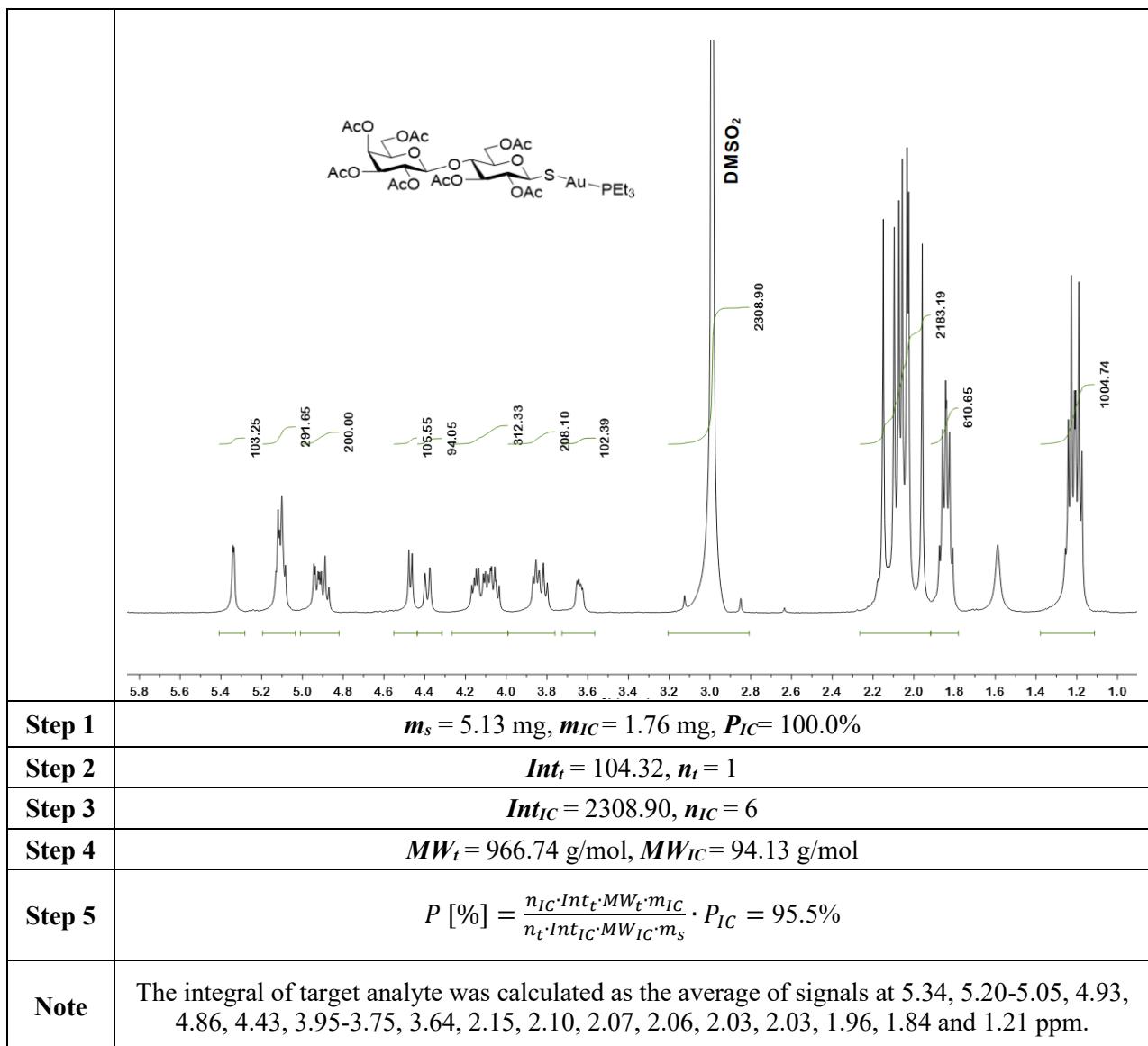
Compound **4** in CDCl_3 with the addition of dimethyl sulfone (DMSO_2 , 100.0%) as internal standard (500 MHz).



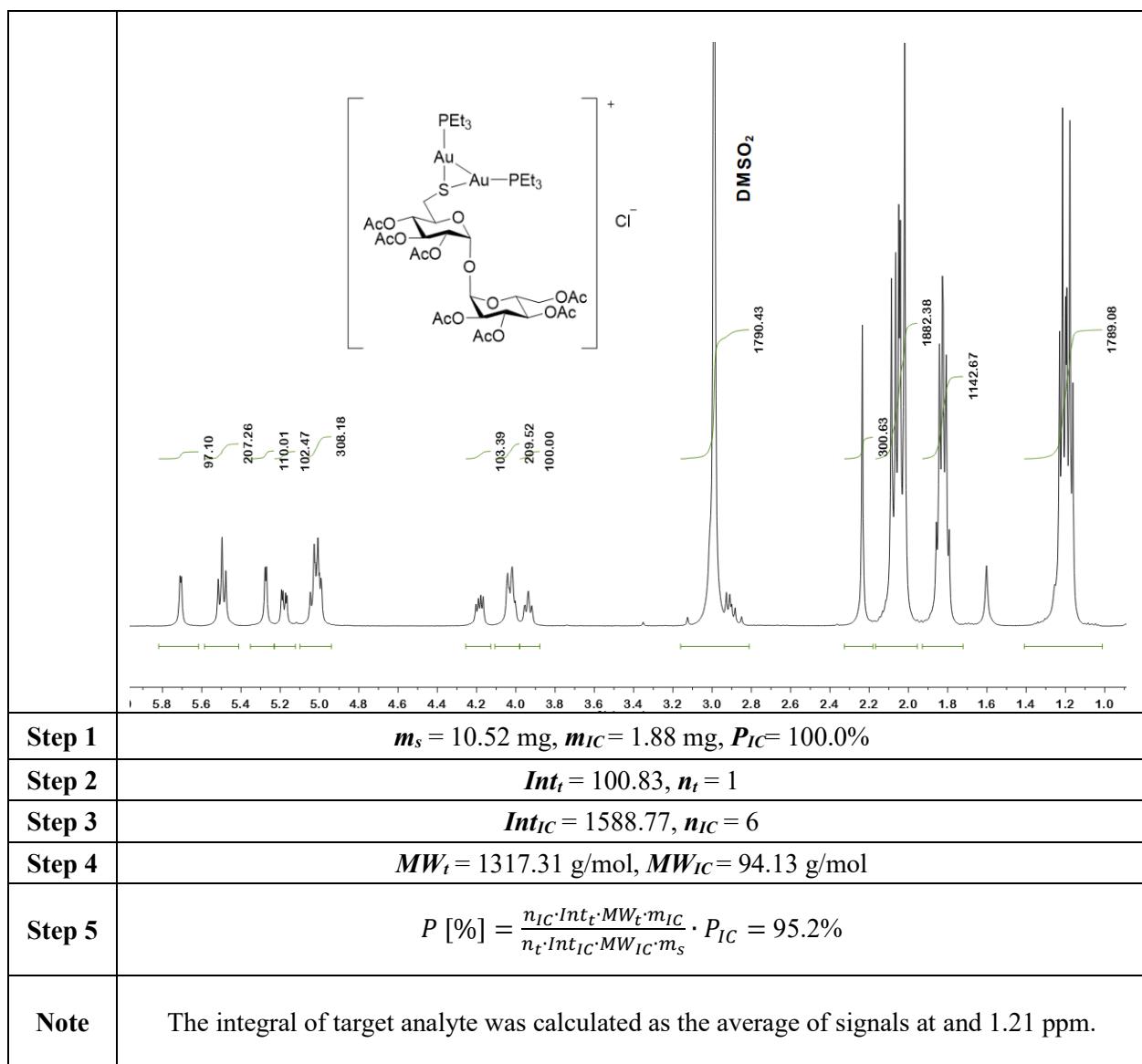
Compound **5** in CDCl_3 with the addition of dimethyl sulfone (DMSO_2 , 100.0%) as internal standard (500 MHz).



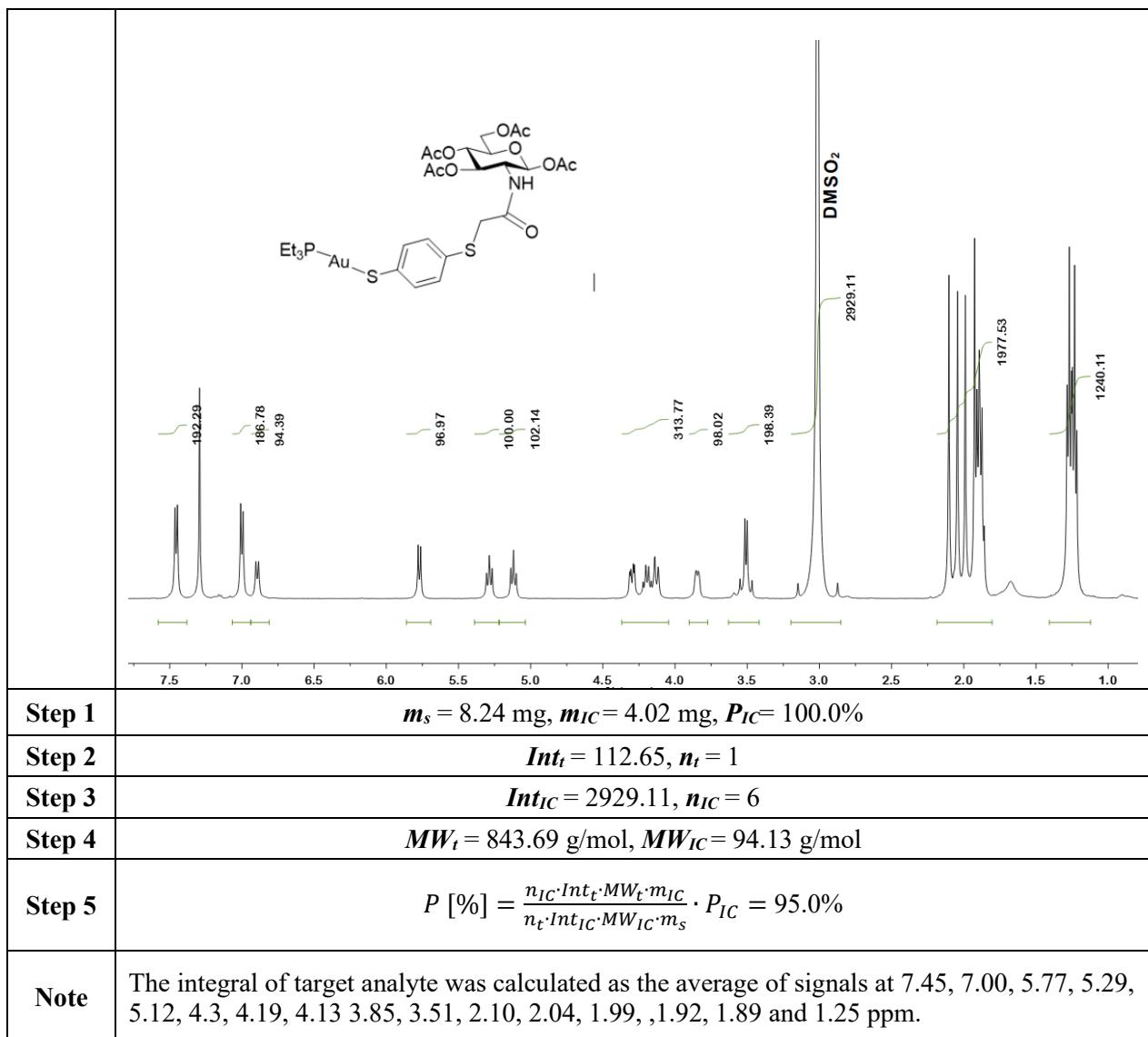
Compound **6** in CDCl_3 with the addition of dimethyl sulfone (DMSO_2 , 100.0%) as internal standard (500 MHz).



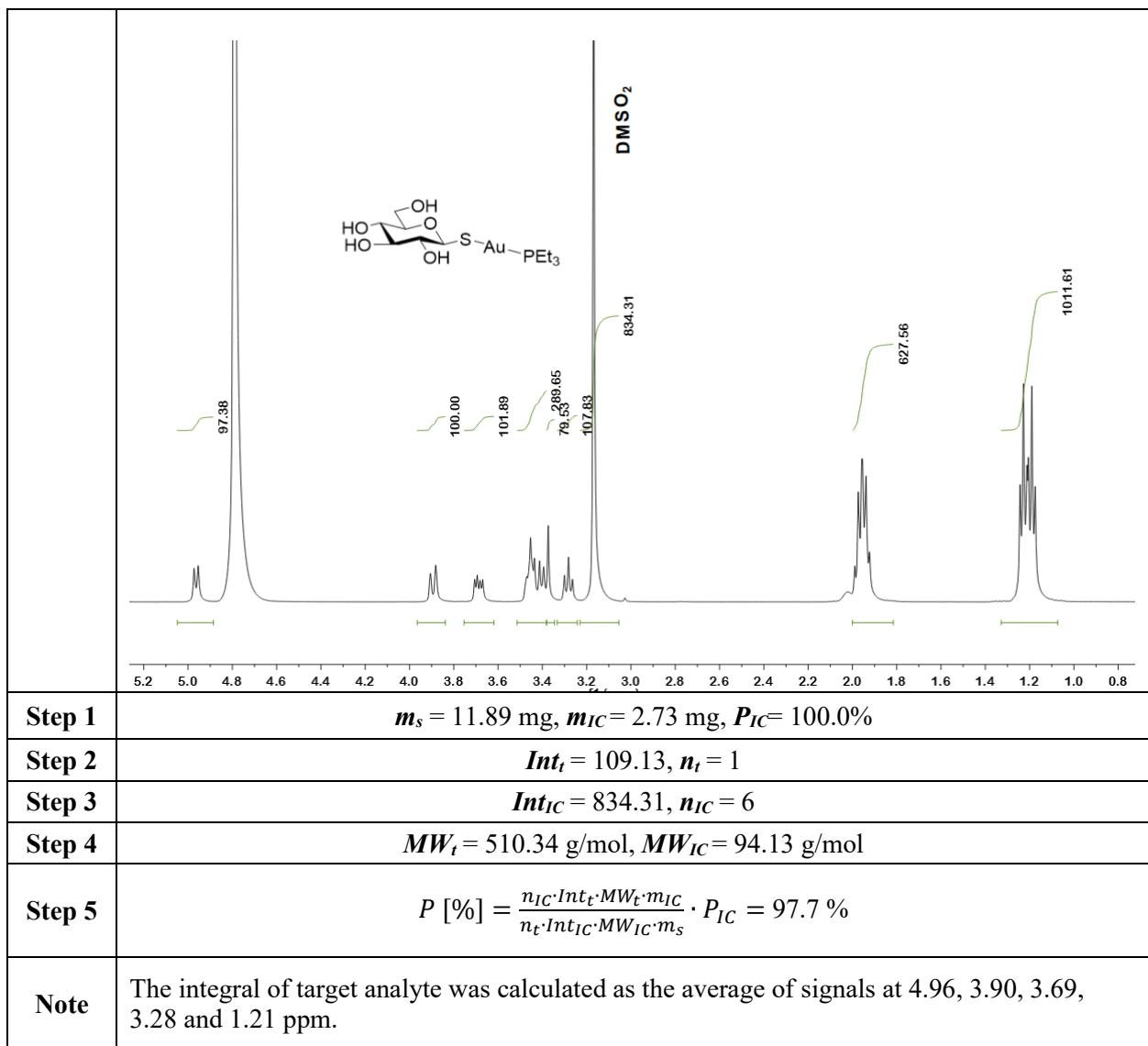
Compound **7** in CDCl_3 with the addition of dimethyl sulfone (DMSO_2 , 100.0%) as internal standard (500 MHz).



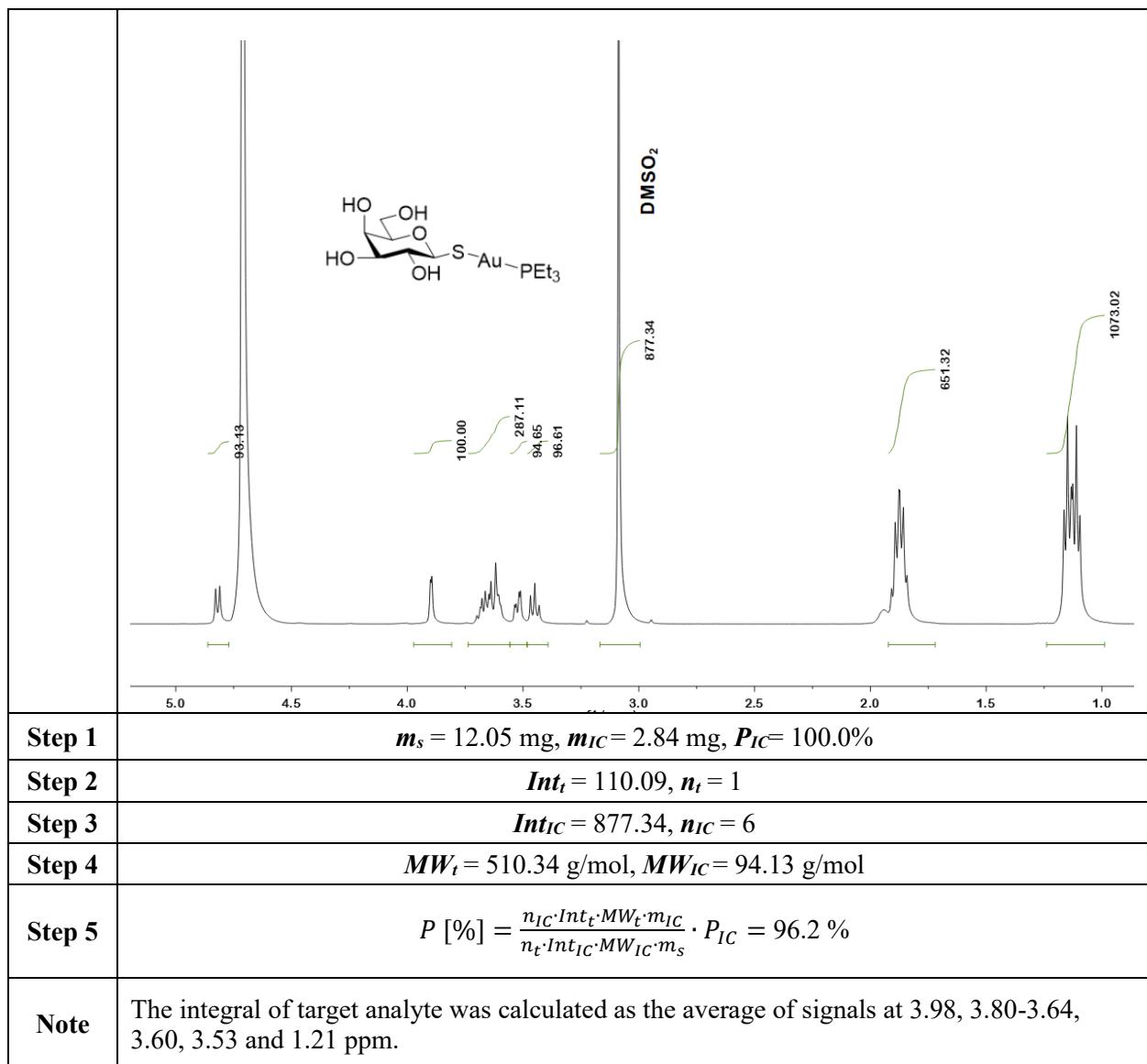
Compound **8** in CDCl_3 with the addition of dimethyl sulfone (DMSO_2 , 100.0%) as internal standard (500 MHz).



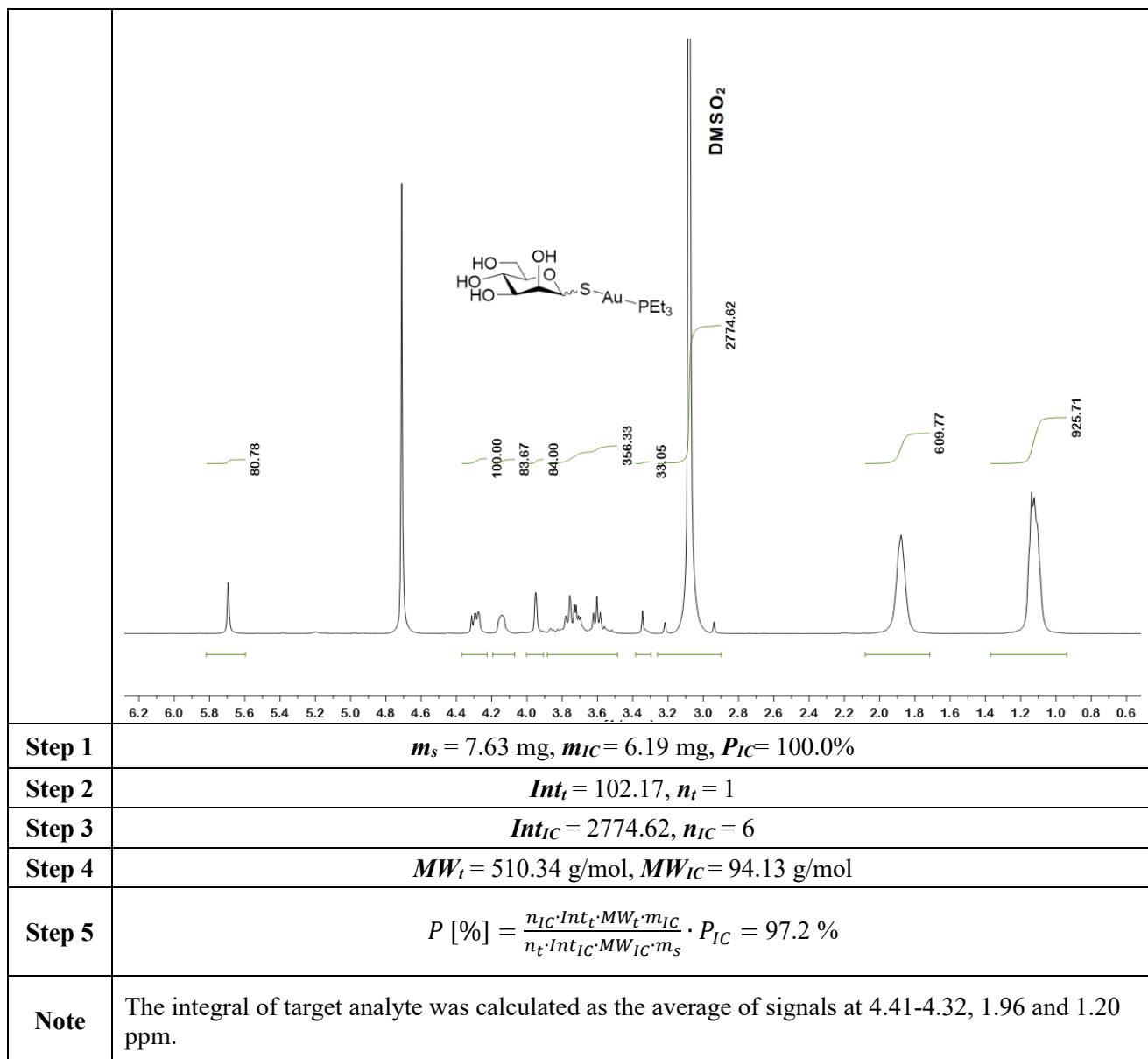
Compound **9** in D₂O with the addition of dimethyl sulfone (DMSO₂, 100.0%) as internal standard (500 MHz).



Compound **10** in D₂O with the addition of dimethyl sulfone (DMSO₂, 100.0%) as internal standard (500 MHz).



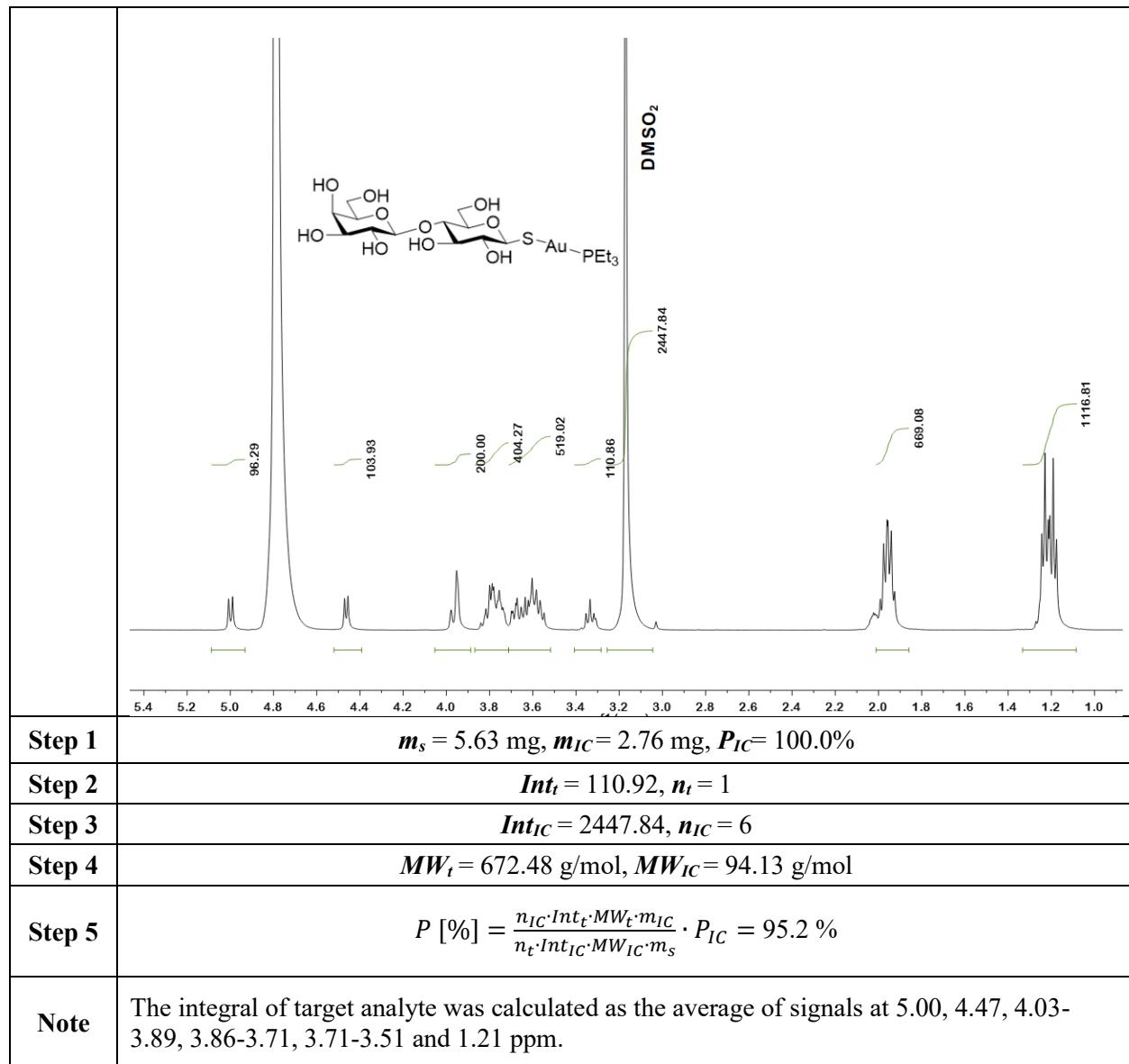
Compound **11** in D₂O with the addition of dimethyl sulfone (DMSO₂, 100.0%) as internal standard (500 MHz).



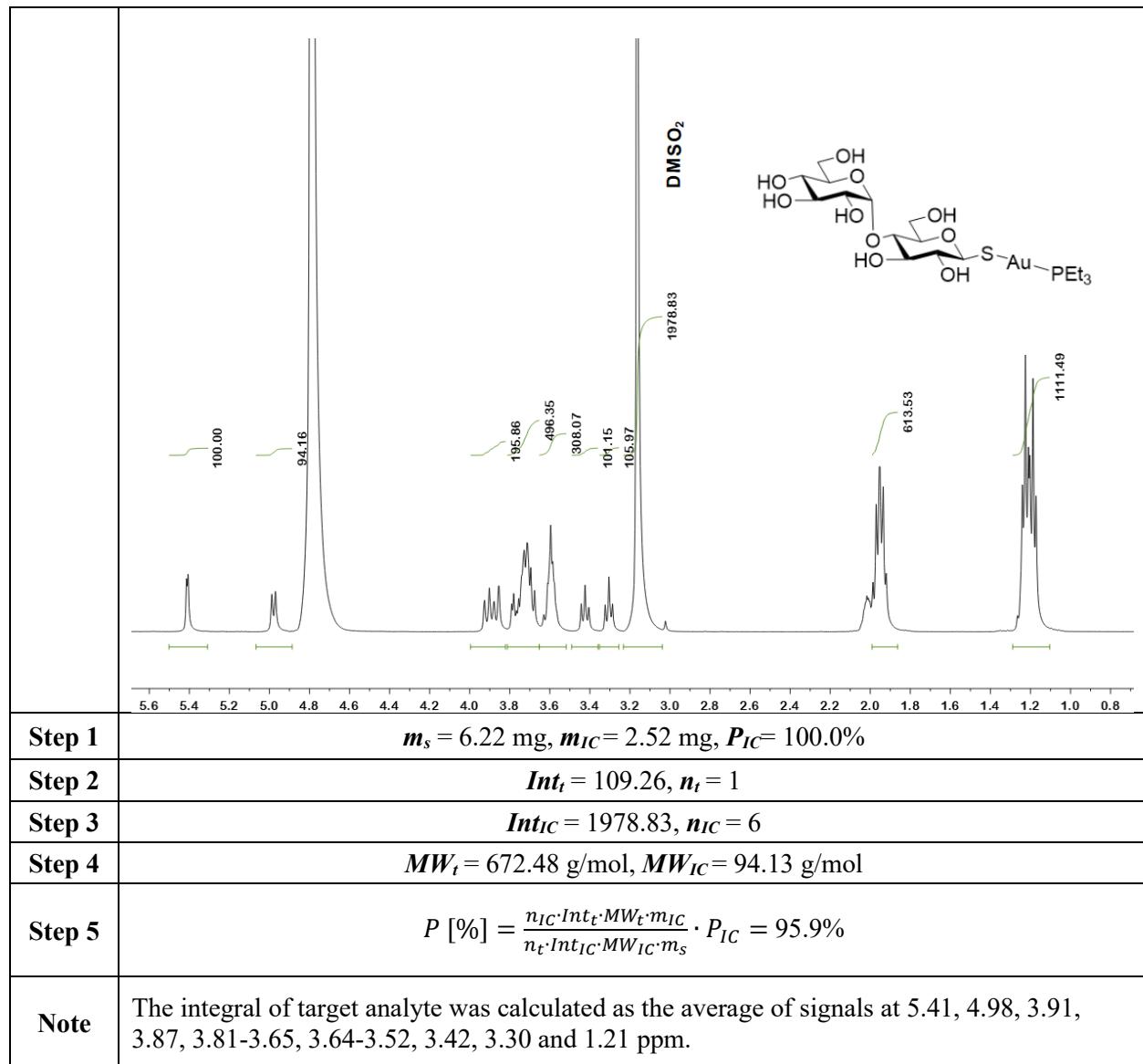
Compound **12** in D₂O with the addition of dimethyl sulfone (DMSO₂, 100.0%) as internal standard (500 MHz).

Step 1	$m_s = 6.62 \text{ mg}$, $m_{IC} = 3.32 \text{ mg}$, $P_{IC} = 100.0\%$
Step 2	$Int_t = 107.32$, $n_t = 1$
Step 3	$Int_{IC} = 1952.91$, $n_{IC} = 6$
Step 4	$MW_t = 551.39 \text{ g/mol}$, $MW_{IC} = 94.13 \text{ g/mol}$
Step 5	$P [\%] = \frac{n_{IC} \cdot Int_t \cdot MW_t \cdot m_{IC}}{n_t \cdot Int_{IC} \cdot MW_{IC} \cdot m_s} \cdot P_{IC} = 96.9 \%$
Note	The integral of target analyte was calculated as the average of signals at 5.07, 3.90, 3.78, 3.70, 3.53-3.35 and 1.20 ppm.

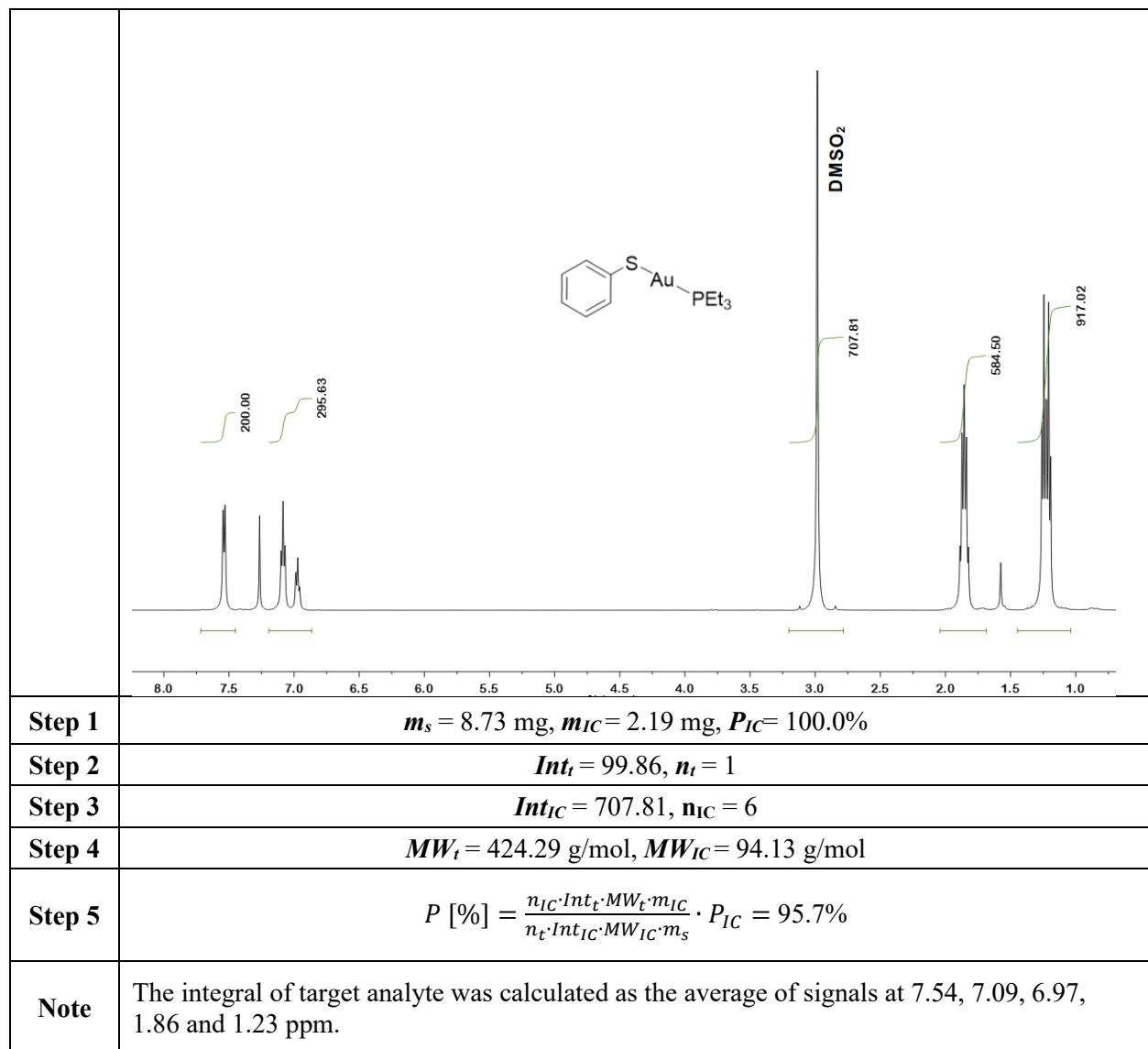
Compound **13** in D₂O with the addition of dimethyl sulfone (DMSO₂, 100.0%) as internal standard (500 MHz).



Compound **14** in D₂O with the addition of dimethyl sulfone (DMSO₂, 100.0%) as internal standard (500 MHz).



Compound **15** in CDCl_3 with the addition of dimethyl sulfone (DMSO_2 , 100.0%) as internal standard (500 MHz).



Compound **16** in CDCl_3 with the addition of dimethyl sulfone (DMSO_2 , 100.0%) as internal standard (500 MHz).

	<p>The figure shows the ^1H NMR spectrum of compound 16. The chemical structure of the compound is shown above the spectrum, featuring a phenyl ring with an amino group (NH_2) at position 4 and a gold atom coordinated to a thiomphenyl group (S-Au-PEt_3). The spectrum displays several aromatic peaks in the 7-8 ppm range, aliphatic peaks in the 1-4 ppm range, and a sharp peak for the internal standard DMSO_2 at 3.0 ppm. Peak labels include: 101.24, 100.00, 102.66, 98.78, 170.00, 2122.26, 632.64, and 977.34.</p>
Step 1	$m_s = 4.79 \text{ mg}$, $m_{IC} = 3.45 \text{ mg}$, $P_{IC} = 100.0\%$
Step 2	$Int_t = 103.53$, $n_t = 1$
Step 3	$Int_{IC} = 2122.26$, $n_{IC} = 6$
Step 4	$MW_t = 439.31 \text{ g/mol}$, $MW_{IC} = 94.13 \text{ g/mol}$
Step 5	$P [\%] = \frac{n_{IC} \cdot Int_t \cdot MW_t \cdot m_{IC}}{n_t \cdot Int_{IC} \cdot MW_{IC} \cdot m_s} \cdot P_{IC} = 98.4 \%$
Note	The integral of target analyte was calculated as the average of signals at 7.52, 6.87, 6.67, 6.56 and 1.82 ppm

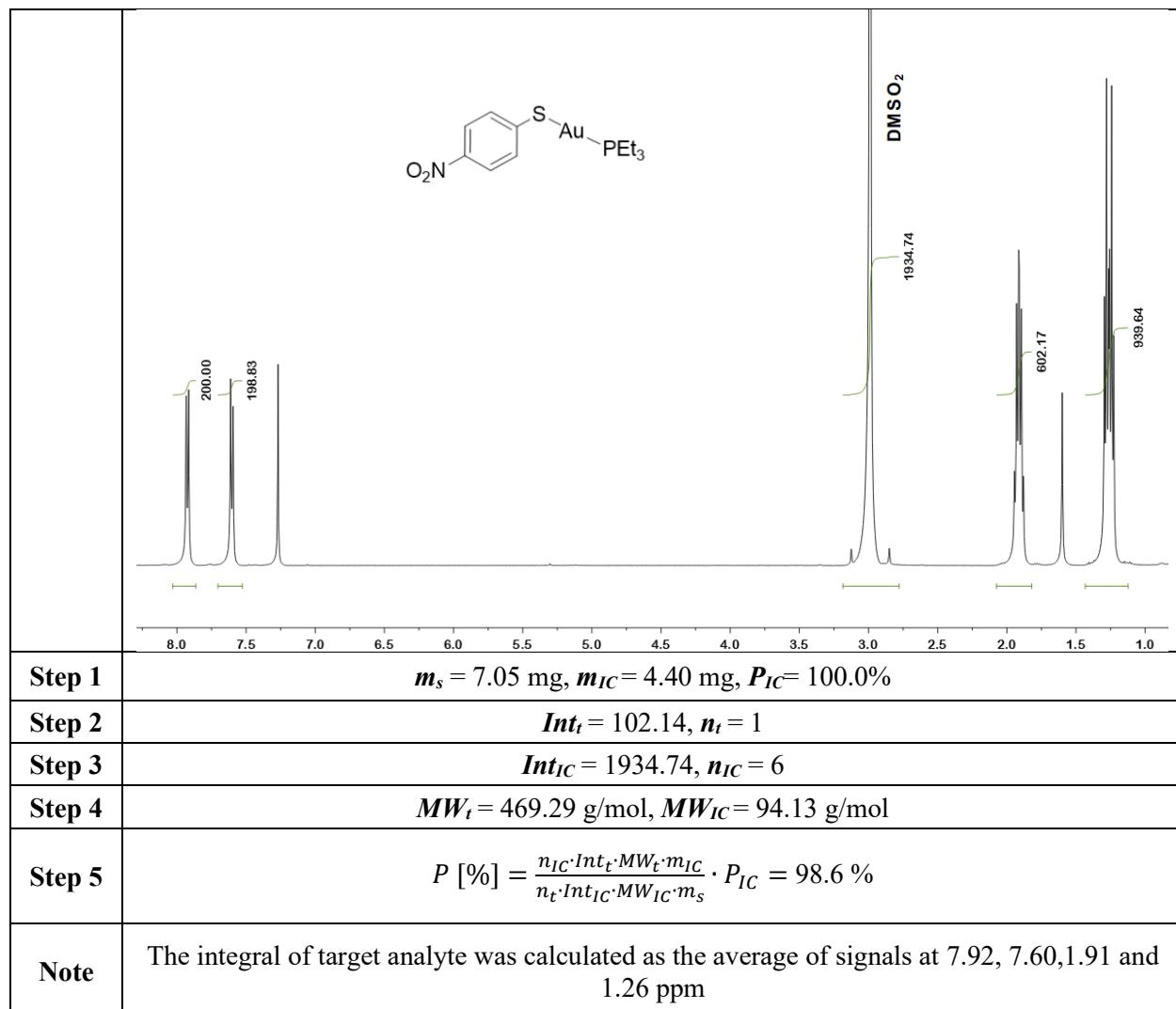
Compound **17** in CDCl_3 with the addition of dimethyl sulfone (DMSO_2 , 100.0%) as internal standard (500 MHz).

	<p style="text-align: center;">$\text{H}_2\text{N}-\text{C}_6\text{H}_4-\text{S}-\text{Au}-\text{P}(\text{Et})_3$</p>
Step 1	$m_s = 4.96 \text{ mg}$, $m_{IC} = 4.08 \text{ mg}$, $P_{IC} = 100.0\%$
Step 2	$Int_t = 102.82$, $n_t = 1$
Step 3	$Int_{IC} = 2414.86$, $n_{IC} = 6$
Step 4	$MW_t = 439.31 \text{ g/mol}$, $MW_{IC} = 94.13 \text{ g/mol}$
Step 5	$P [\%] = \frac{n_{IC} \cdot Int_t \cdot MW_t \cdot m_{IC}}{n_t \cdot Int_{IC} \cdot MW_{IC} \cdot m_s} \cdot P_{IC} = 98.08 \%$
Note	The integral of target analyte was calculated as the average of signals at 6.50, 1.83 and 1.21 ppm.

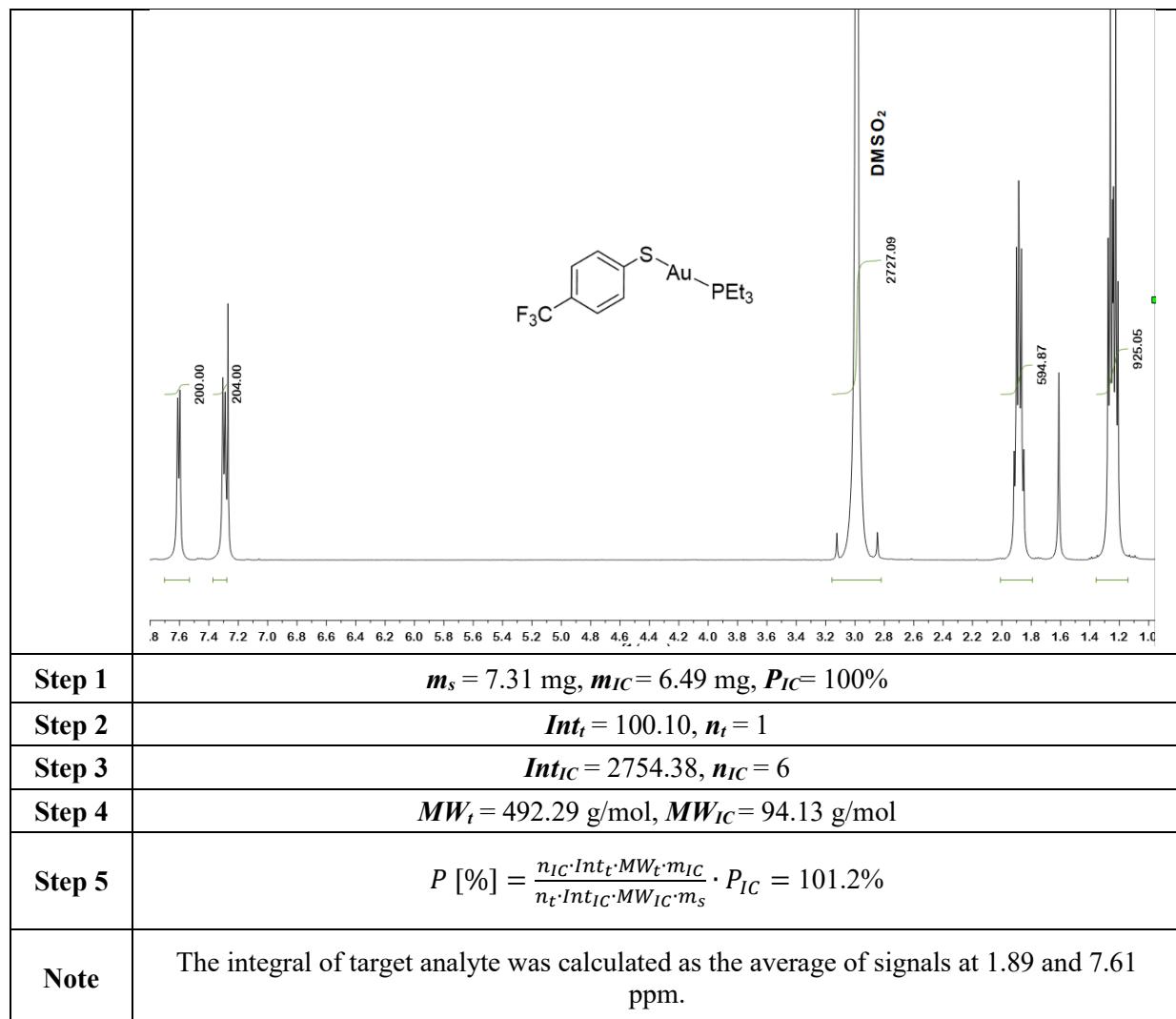
Compound **18** in CDCl_3 with the addition of dimethyl sulfone (DMSO_2 , 100.0%) as internal standard (500 MHz).

	<p>The figure shows the ^1H NMR spectrum of compound 18. The chemical structure of the compound is shown above the spectrum. The spectrum displays several peaks corresponding to the protons in the molecule. Key peaks are labeled with their chemical shifts (δ) in ppm: 7.41, 6.69, 3.74, 1.84, and 1.22. A sharp peak at approximately 2.5 ppm is labeled DMSO_2. Other labeled peaks include 200.00, 198.71, 303.91, 2060.75, 596.32, and 916.94. Green integration curves are overlaid on the peaks, and green horizontal bars below the x-axis indicate the integration ranges for specific signals.</p>
Step 1	$m_s = 6.65 \text{ mg}, m_{IC} = 4.66 \text{ mg}, P_{IC} = 100.0\%$
Step 2	$Int_t = 100.70, n_t = 1$
Step 3	$Int_{IC} = 1030.38, n_{IC} = 6$
Step 4	$MW_t = 454.32 \text{ g/mol}, MW_{IC} = 94.13 \text{ g/mol}$
Step 5	$P [\%] = \frac{n_{IC} \cdot Int_t \cdot MW_t \cdot m_{IC}}{n_t \cdot Int_{IC} \cdot MW_{IC} \cdot m_s} \cdot P_{IC} = 99.2 \%$
Note	The integral of target analyte was calculated as the average of signals at 7.41, 6.69, 3.74, 1.84 and 1.22 ppm

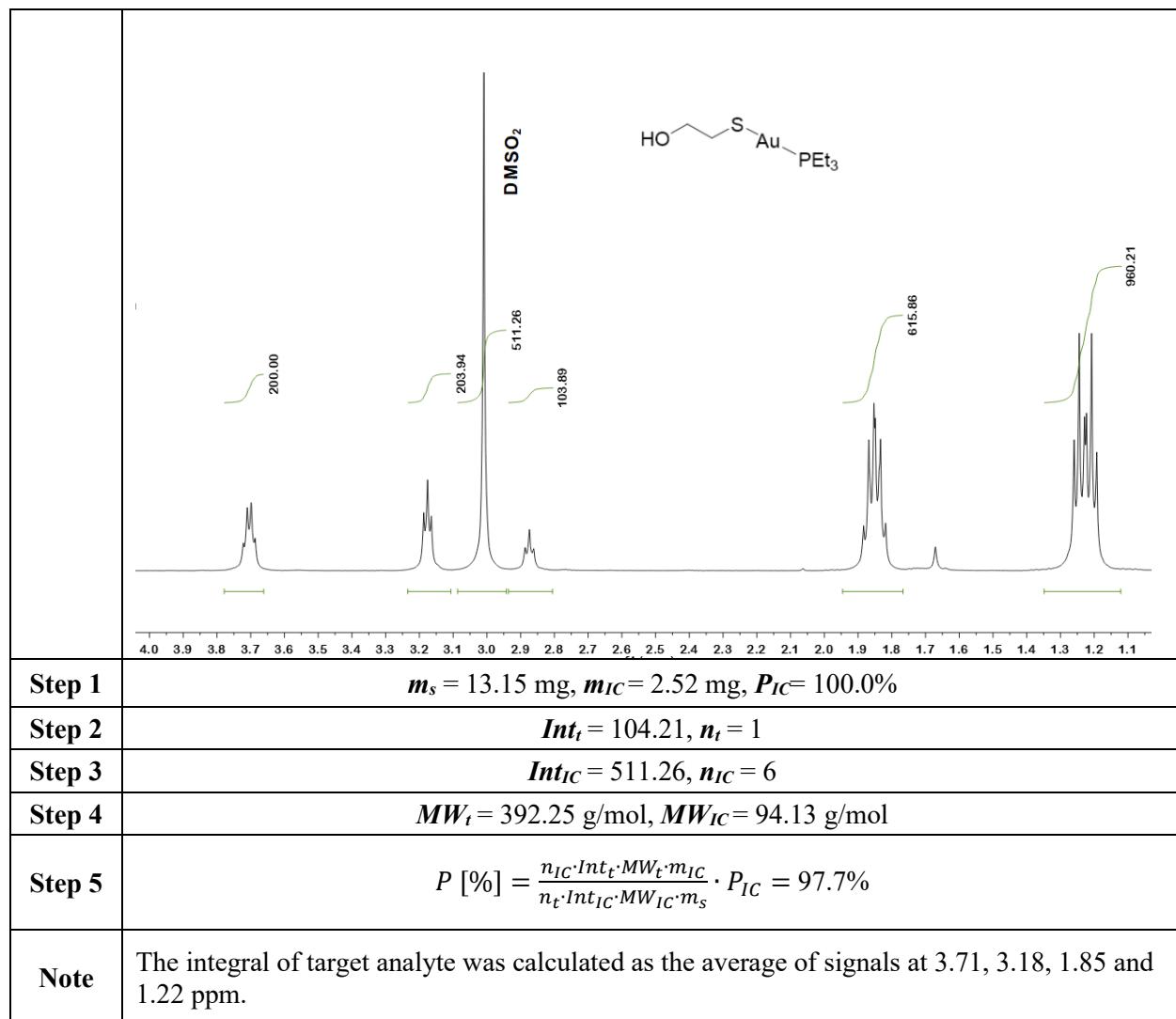
Compound **19** in CDCl_3 with the addition of dimethyl sulfone (DMSO_2 , 100.0%) as internal standard (500 MHz).



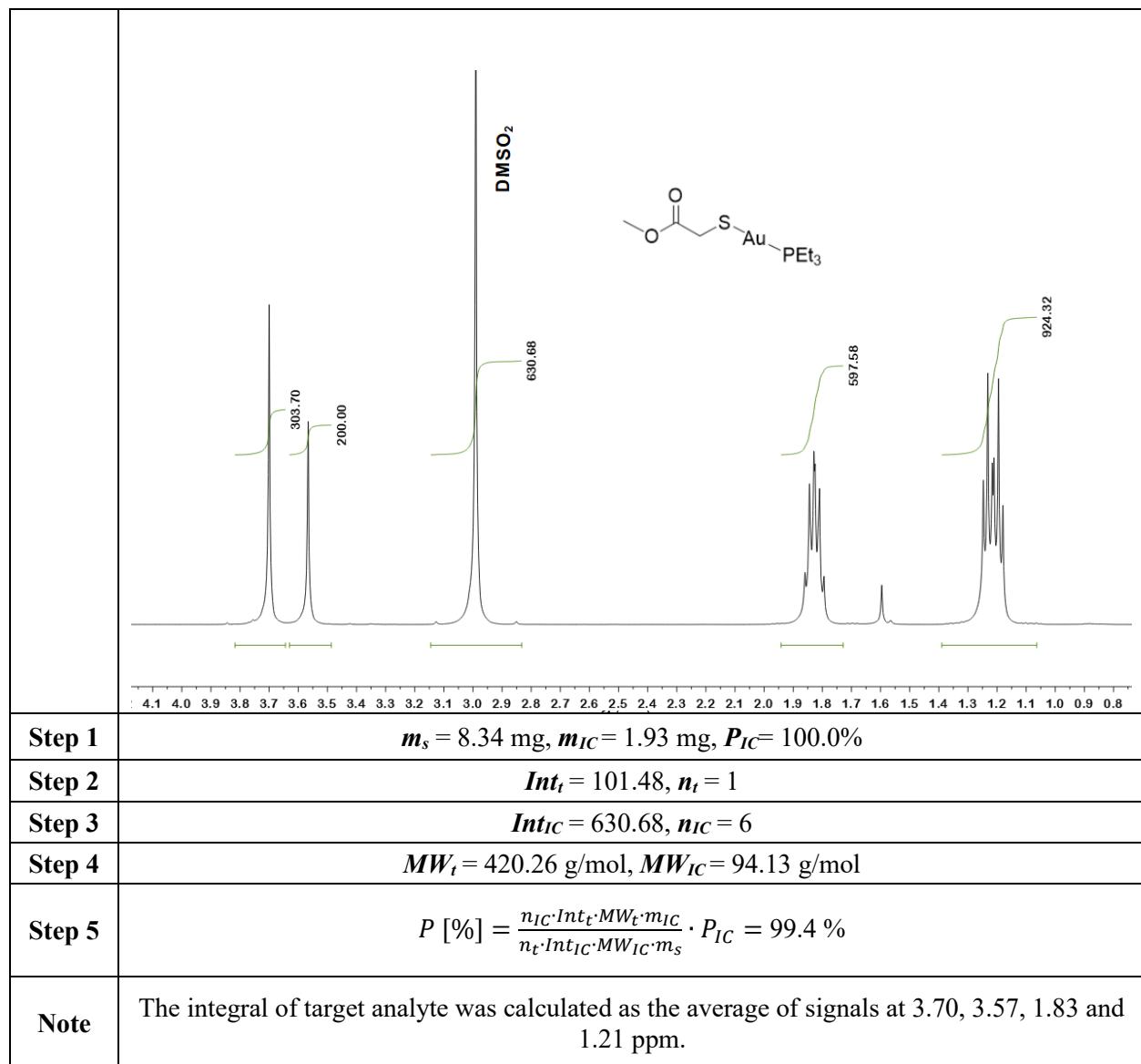
Compound **20** in CDCl_3 with the addition of dimethyl sulfone (DMSO_2 , 100.0%) as internal standard (500 MHz).



Compound **21** in CDCl_3 with the addition of dimethyl sulfone (DMSO_2 , 100.0%) as internal standard (500 MHz).



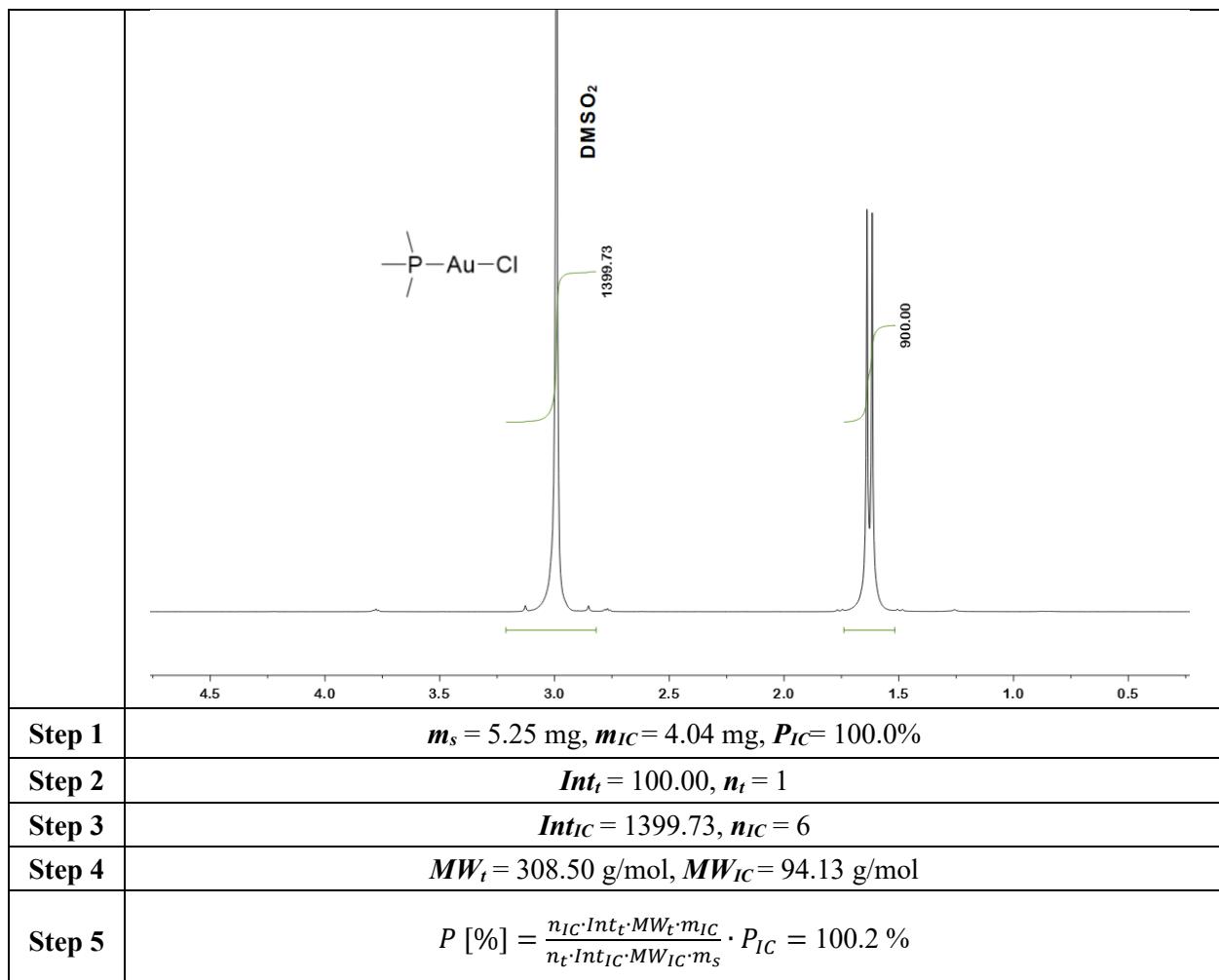
Compound **22** in CDCl_3 with the addition of dimethyl sulfone (DMSO_2 , 100.0%) as internal standard (500 MHz).



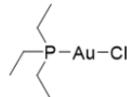
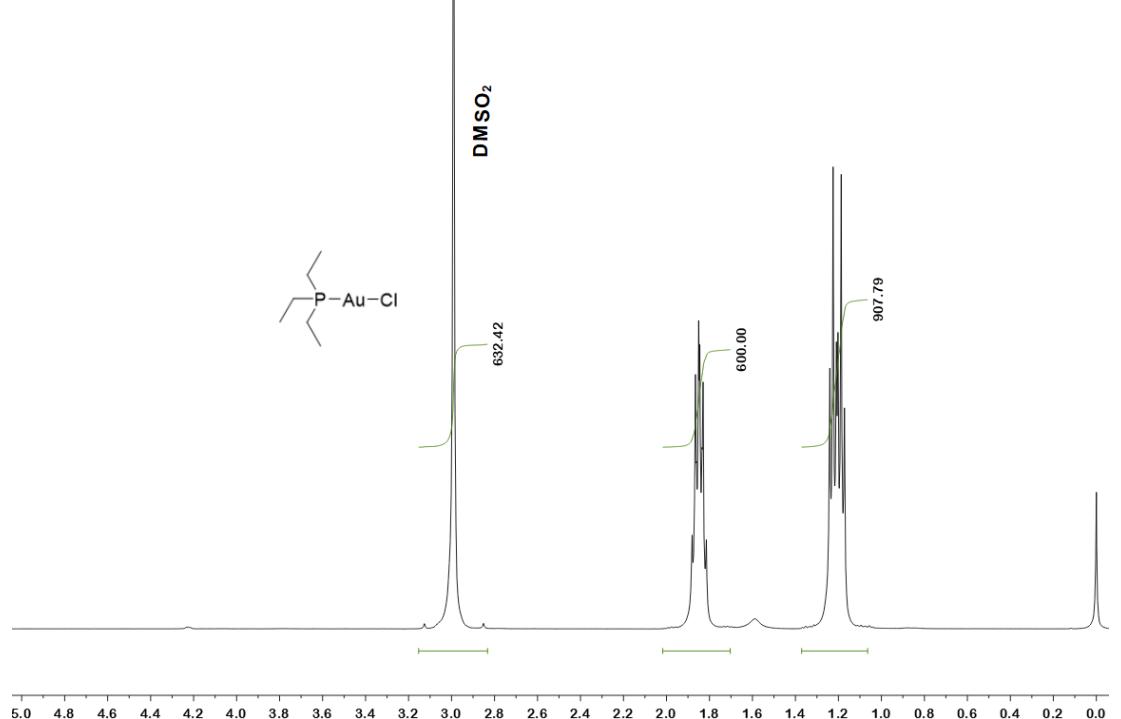
Compound **23** in CDCl_3 with the addition of dimethyl sulfone (DMSO_2 , 100.0%) as internal standard (500 MHz).

Step 1	$m_s = 7.77 \text{ mg}$, $m_{IC} = 4.60 \text{ mg}$, $P_{IC} = 100.0\%$
Step 2	$Int_t = 100.61$, $n_t = 1$
Step 3	$Int_{IC} = 1605.53$, $n_{IC} = 6$
Step 4	$MW_t = 430.22 \text{ g/mol}$, $MW_{IC} = 94.13 \text{ g/mol}$
Step 5	$P [\%] = \frac{n_{IC} \cdot Int_t \cdot MW_t \cdot m_{IC}}{n_t \cdot Int_{IC} \cdot MW_{IC} \cdot m_s} \cdot P_{IC} = 101.7\%$
Note	The integral of target analyte was calculated as the average of signals at 3.43 and 1.83 ppm.

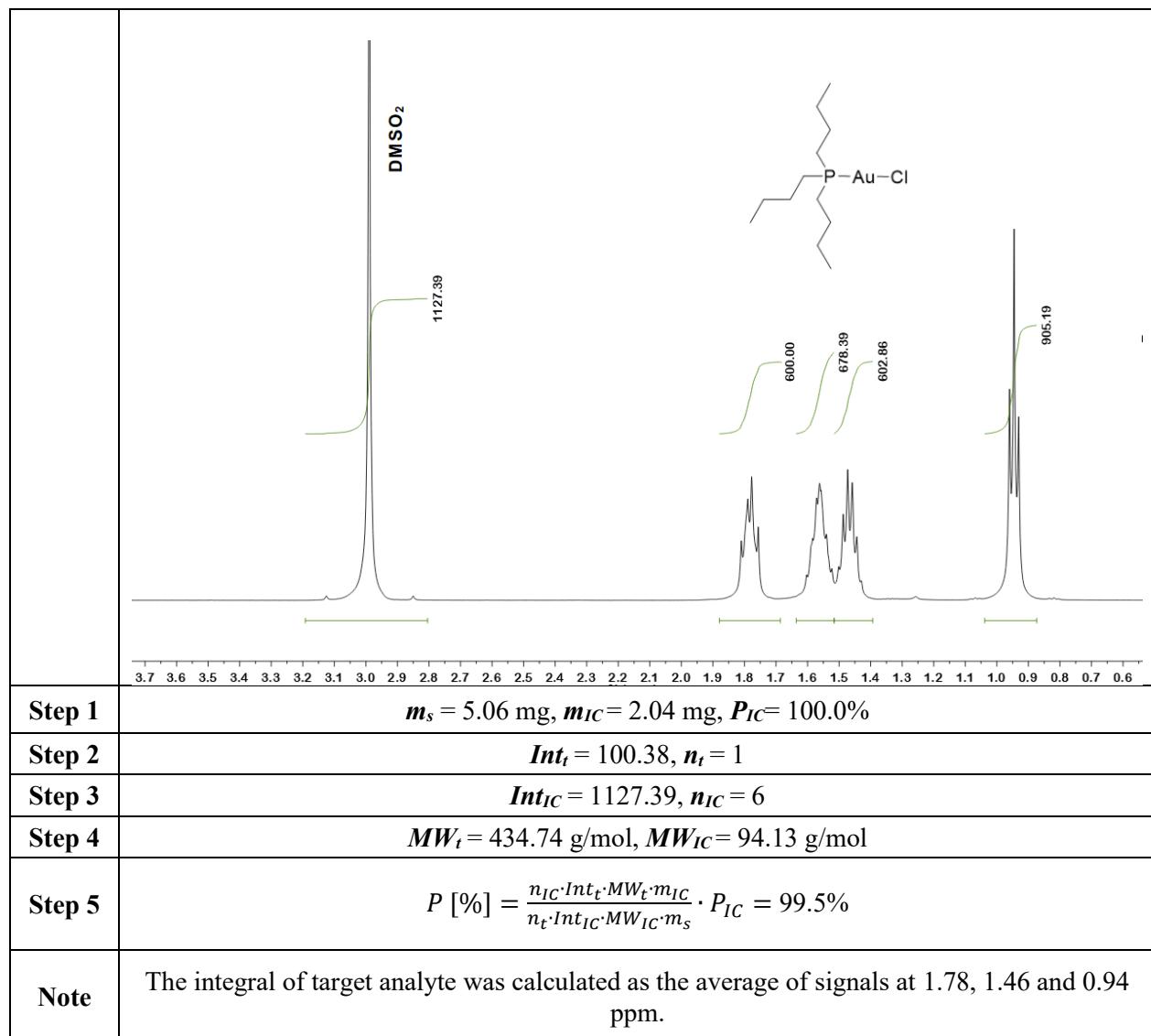
Compound **24** in CDCl_3 with the addition of dimethyl sulfone (DMSO_2 , 100.0%) as internal standard (500 MHz).



Compound **25** in CDCl_3 with the addition of dimethyl sulfone (DMSO_2 , 100.0%) as internal standard (500 MHz).

	 <p>DMSO₂</p> 
Step 1	$m_s = 7.06 \text{ mg}$, $m_{IC} = 1.93 \text{ mg}$, $P_{IC} = 100.0\%$
Step 2	$Int_t = 100.52$, $n_t = 1$
Step 3	$Int_{IC} = 632.42$, $n_{IC} = 6$
Step 4	$MW_t = 350.58 \text{ g/mol}$, $MW_{IC} = 94.13 \text{ g/mol}$
Step 5	$P [\%] = \frac{n_{IC} \cdot Int_t \cdot MW_t \cdot m_{IC}}{n_t \cdot Int_{IC} \cdot MW_{IC} \cdot m_s} \cdot P_{IC} = 97.1\%$
Note	The integral of target analyte was calculated as the average of signals 1.85, and 1.21 ppm.

Compound **26** in CDCl_3 with the addition of dimethyl sulfone (DMSO_2 , 100.0%) as internal standard (500 MHz).



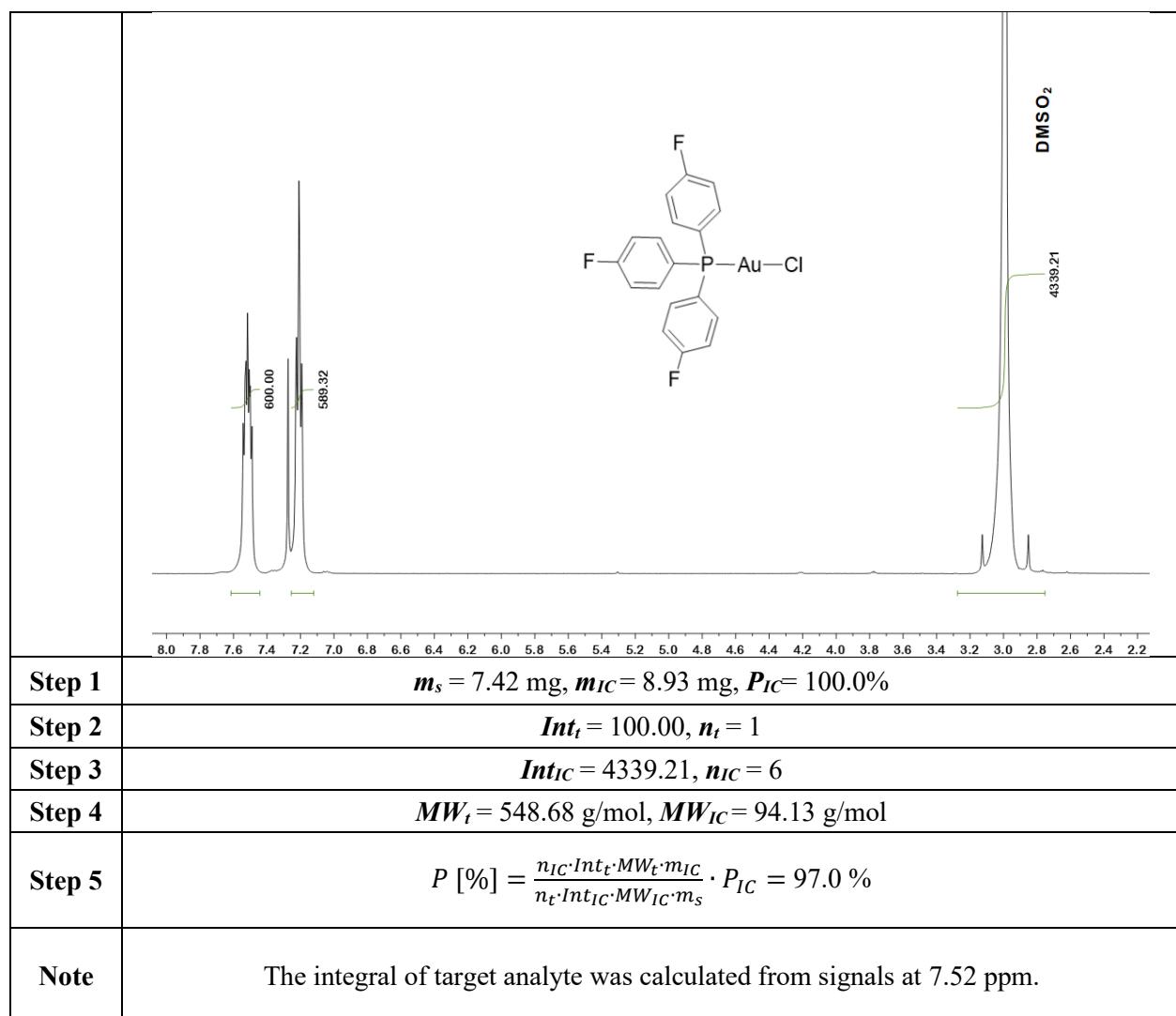
Compound **27** in CDCl_3 with the addition of dimethyl sulfone (DMSO_2 , 100.0%) as internal standard (500 MHz).

Step 1	$m_s = 6.59 \text{ mg}$, $m_{IC} = 2.86 \text{ mg}$, $P_{IC} = 100.0\%$
Step 2	$Int_t = 100.00$, $n_t = 1$
Step 3	$Int_{IC} = 1388.95$, $n_{IC} = 6$
Step 4	$MW_t = 494.71 \text{ g/mol}$, $MW_{IC} = 94.13 \text{ g/mol}$
Step 5	$P [\%] = \frac{n_{IC} \cdot Int_t \cdot MW_t \cdot m_{IC}}{n_t \cdot Int_{IC} \cdot MW_{IC} \cdot m_s} \cdot P_{IC} = 98.5\%$
Note	The integral of target analyte was calculated as the average of signals 7.65-7.40 ppm.

Compound **28** in CDCl_3 with the addition of dimethyl sulfone (DMSO_2 , 100.0%) as internal standard (500 MHz).

Step 1	$m_s = 5.44 \text{ mg}$, $m_{IC} = 7.97 \text{ mg}$, $P_{IC} = 100.0\%$
Step 2	$Int_t = 100.68$, $n_t = 1$
Step 3	$Int_{IC} = 5592.83$, $n_{IC} = 6$
Step 4	$MW_t = 584.79 \text{ g/mol}$, $MW_{IC} = 94.13 \text{ g/mol}$
Step 5	$P [\%] = \frac{n_{IC} \cdot Int_t \cdot MW_t \cdot m_{IC}}{n_t \cdot Int_{IC} \cdot MW_{IC} \cdot m_s} \cdot P_{IC} = 98.3 \%$
Note	The integral of target analyte was calculated as the average of signals at 7.43, 6.96 and 3.85 ppm.

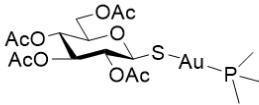
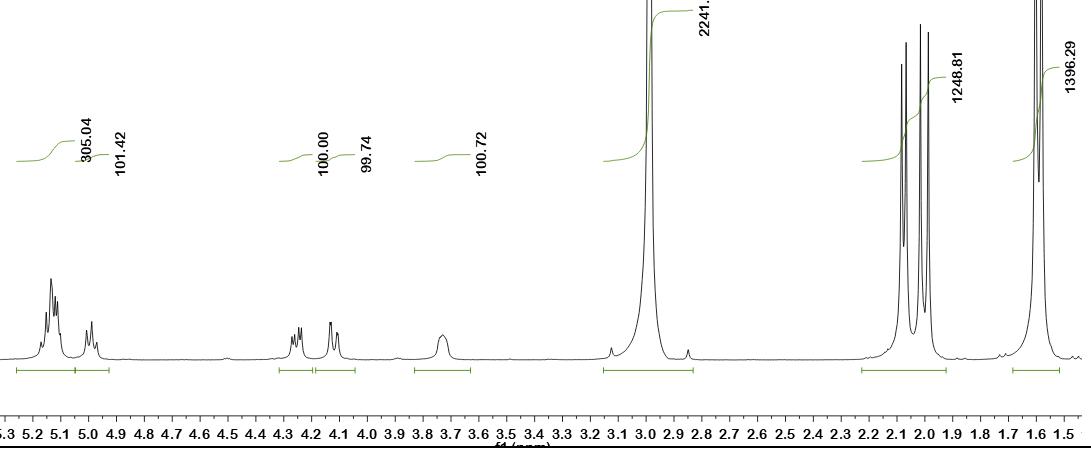
Compound **29** in CDCl_3 with the addition of dimethyl sulfone (DMSO_2 , 100.0%) as internal standard (500 MHz).



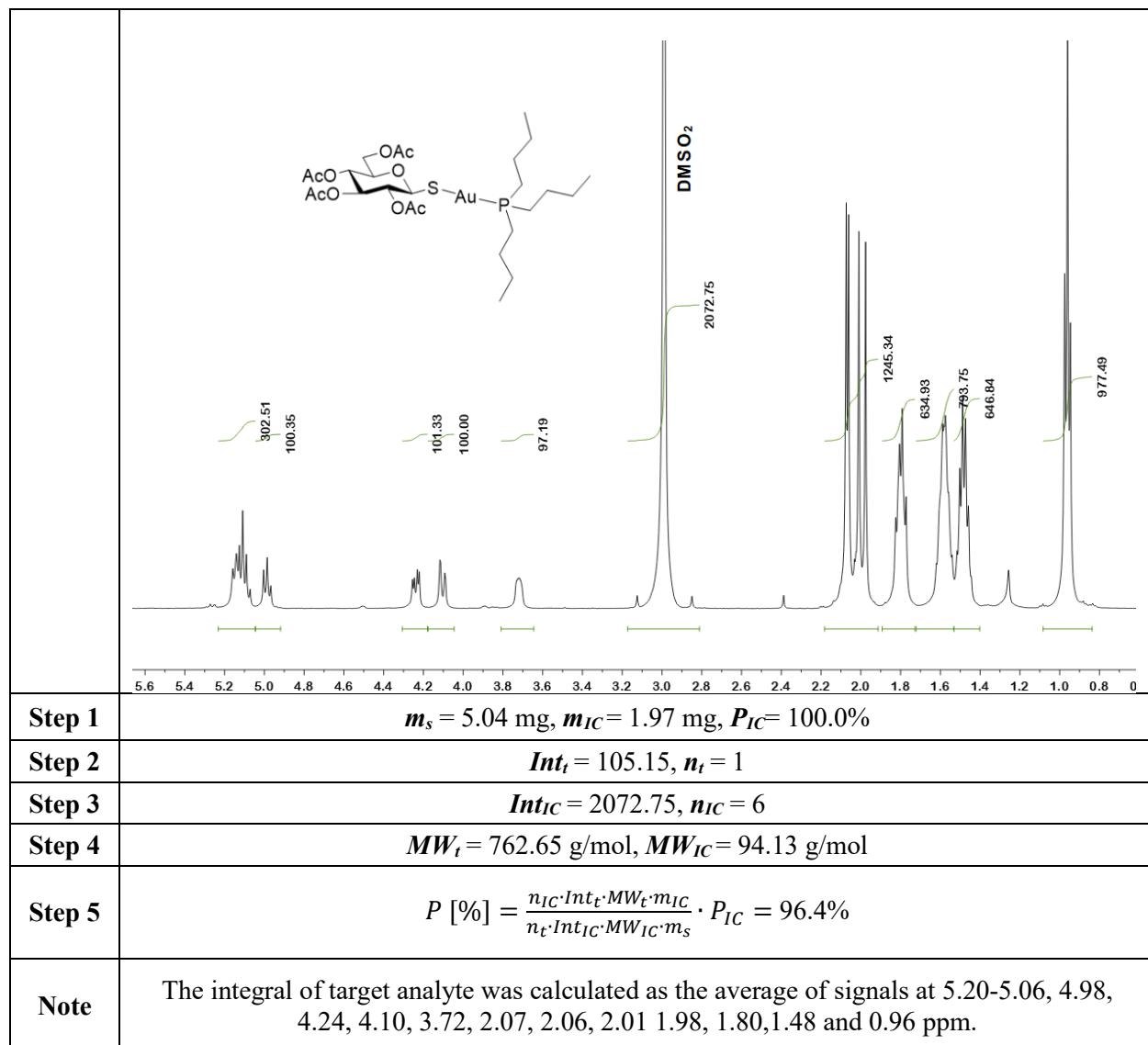
Compound **30** in CDCl_3 with the addition of dimethyl sulfone (DMSO_2 , 100.0%) as internal standard (500 MHz).

	<p style="text-align: center;"> F_3C $\text{F}_3\text{C}-\text{C}_6\text{H}_3-\text{P}(\text{Au}-\text{Cl})-\text{C}_6\text{H}_3-\text{C}_6\text{F}_{12}$ </p>
Step 1	$m_s = 5.39 \text{ mg}$, $m_{IC} = 3.31 \text{ mg}$, $P_{IC} = 100.0\%$
Step 2	$Int_t = 99.20$, $n_t = 1$
Step 3	$Int_{IC} = 2693.83$, $n_{IC} = 6$
Step 4	$MW_t = 698.70 \text{ g/mol}$, $MW_{IC} = 94.13 \text{ g/mol}$
Step 5	$P [\%] = \frac{n_{IC} \cdot Int_t \cdot MW_t \cdot m_{IC}}{n_t \cdot Int_{IC} \cdot MW_{IC} \cdot m_s} \cdot P_{IC} = 100.7 \%$
Note	The integral of target analyte was calculated as the average of signals at 7.79 and 7.67 ppm.

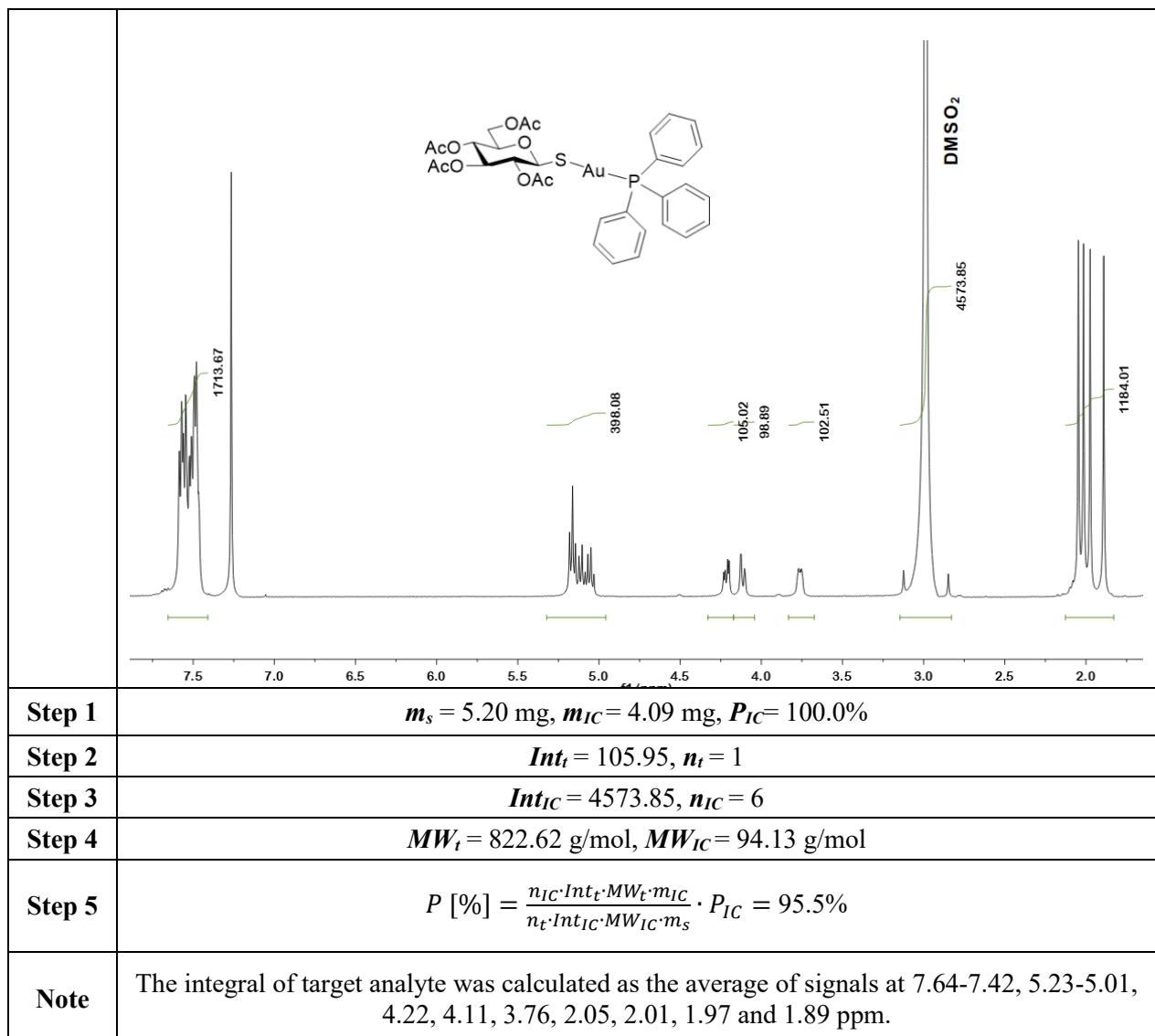
Compound **31** in CDCl_3 with the addition of dimethyl sulfone (DMSO_2 , 100.0%) as internal standard (500 MHz).

	 
Step 1	$m_s = 5.59 \text{ mg}$, $m_{IC} = 2.85 \text{ mg}$, $P_{IC} = 100.0\%$
Step 2	$Int_t = 102.93$, $n_t = 1$
Step 3	$Int_{IC} = 2241.35$ $n_{IC} = 6$
Step 4	$MW_t = 636.40 \text{ g/mol}$, $MW_{IC} = 94.13 \text{ g/mol}$
Step 5	$P [\%] = \frac{n_{IC} \cdot Int_t \cdot MW_t \cdot m_{IC}}{n_t \cdot Int_{IC} \cdot MW_{IC} \cdot m_s} \cdot P_{IC} = 95.0\%$
Note	The integral of target analyte was calculated as the average of signals at 5.20-5.06, 4.99, 4.26, 4.12, 3.73, 2.08, 2.07, 2.01 and 1.99 ppm.

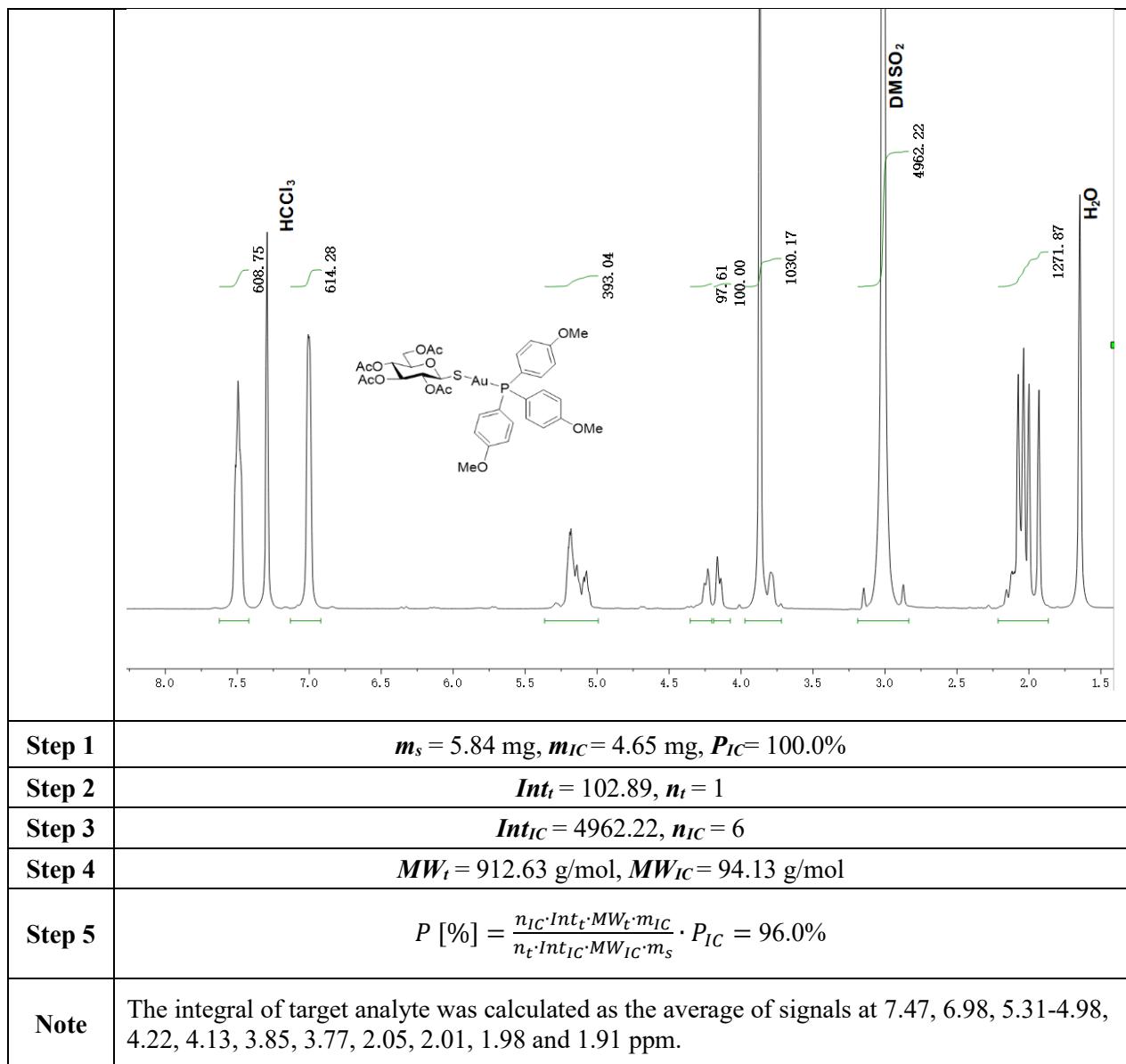
Compound **32** in CDCl_3 with the addition of dimethyl sulfone (DMSO_2 , 100.0%) as internal standard (500 MHz).



Compound **33** in CDCl_3 with the addition of dimethyl sulfone (DMSO_2 , 100.0%) as internal standard (500 MHz).



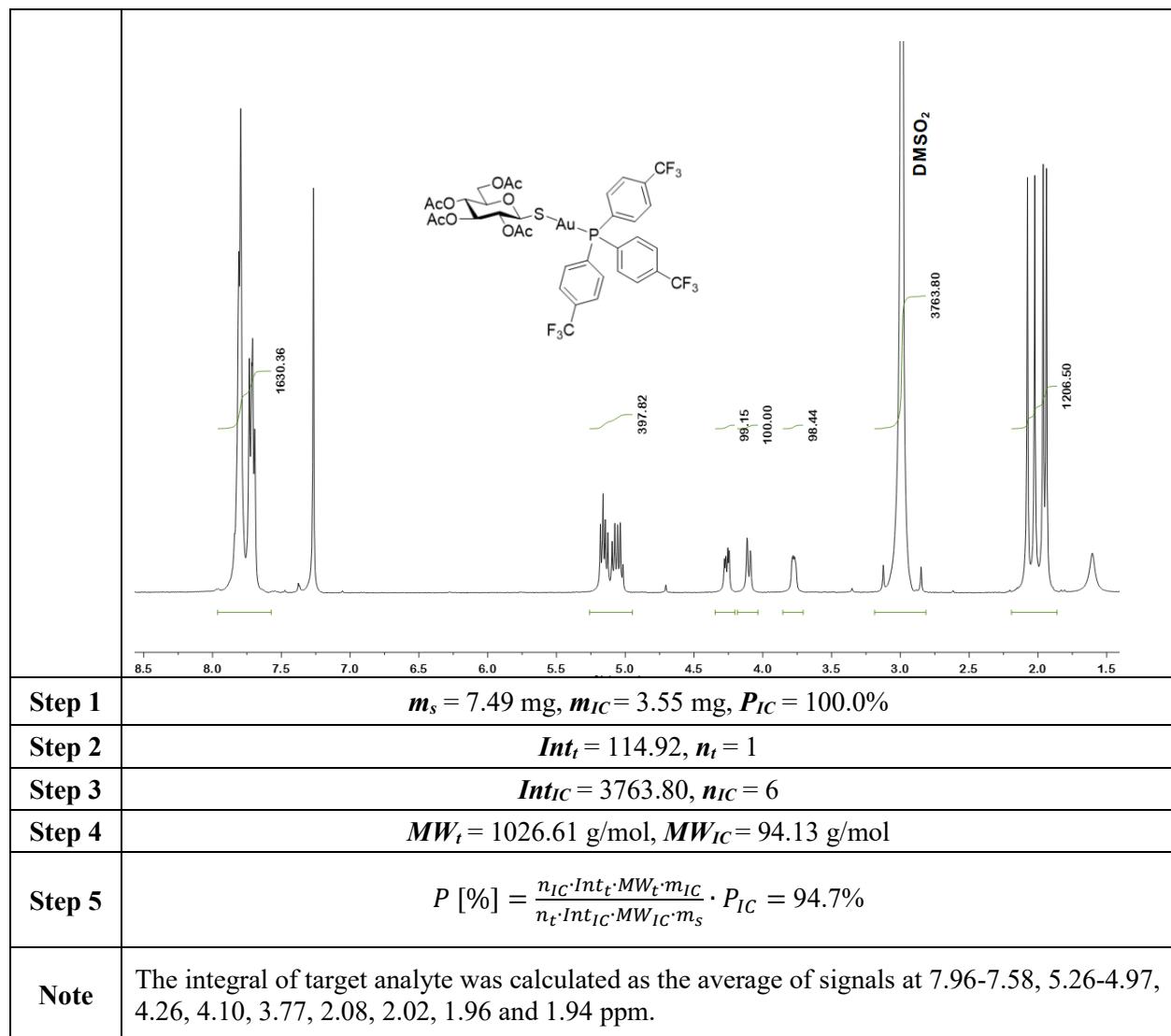
Compound **34** in CDCl_3 with the addition of dimethyl sulfone (DMSO_2 , 100.0%) as internal standard (500 MHz).



Compound **35** in CDCl_3 with the addition of dimethyl sulfone (DMSO_2 , 100.0%) as internal standard (500 MHz).

	<p style="text-align: center;"> </p>
Step 1	$m_s = 5.81 \text{ mg}$, $m_{IC} = 2.42 \text{ mg}$, $P_{IC} = 100.0\%$
Step 2	$Int_t = 100.86$, $n_t = 1$
Step 3	$Int_{IC} = 2463.66$, $n_{IC} = 6$
Step 4	$MW_t = 876.59 \text{ g/mol}$, $MW_{IC} = 94.13 \text{ g/mol}$
Step 5	$P [\%] = \frac{n_{IC} \cdot Int_t \cdot MW_t \cdot m_{IC}}{n_t \cdot Int_{IC} \cdot MW_{IC} \cdot m_s} \cdot P_{IC} = 95.3\%$
Note	The integral of target analyte was calculated as the average of signals at 7.56, 5.32-4.96, 4.24, 4.11, 3.77, 2.07, 1.98, and 1.94 ppm.

Compound **36** in CDCl_3 with the addition of dimethyl sulfone (DMSO_2 , 100.0%) as internal standard (500 MHz).



Compound **37** in D₂O with the addition of dimethyl sulfone (DMSO₂, 100.0%) as internal standard (500 MHz).

Step 1	$m_s = 10.01 \text{ mg}$, $m_{IC} = 2.81 \text{ mg}$, $P_{IC} = 100.0\%$
Step 2	$Int_t = 112.77$, $n_t = 1$
Step 3	$Int_{IC} = 994.63$, $n_{IC} = 6$
Step 4	$MW_t = 468.25 \text{ g/mol}$, $MW_{IC} = 94.13 \text{ g/mol}$
Step 5	$P [\%] = \frac{n_{IC} \cdot Int_t \cdot MW_t \cdot m_{IC}}{n_t \cdot Int_{IC} \cdot MW_{IC} \cdot m_s} \cdot P_{IC} = 95.0\%$
Note	The integral of target analyte was calculated as the average of signals 3.89, 3.69, 3.53-3.34 and 1.62 ppm.

Compound **38** in D₂O with the addition of dimethyl sulfone (DMSO₂, 100.0%) as internal standard (500 MHz).

	<p><chem>CC(C)(C)S(=O)(=O)c1ccc(O)c(O)c1</chem></p>
Step 1	$m_s = 6.37 \text{ mg}$, $m_{IC} = 3.22 \text{ mg}$, $P_{IC} = 100.0\%$
Step 2	$Int_t = 101.79$, $n_t = 1$
Step 3	$Int_{IC} = 1593.91$, $n_{IC} = 6$
Step 4	$MW_t = 468.25 \text{ g/mol}$, $MW_{IC} = 94.13 \text{ g/mol}$
Step 5	$P [\%] = \frac{n_{IC} \cdot Int_t \cdot MW_t \cdot m_{IC}}{n_t \cdot Int_{IC} \cdot MW_{IC} \cdot m_s} \cdot P_{IC} = 96.4\%$
Note	The integral of target analyte was calculated as the average of signals 3.97, 3.82-3.64, 3.60, 3.49 and 1.61 ppm.

Compound **39** in D₂O with the addition of dimethyl sulfone (DMSO₂, 100.0%) as internal standard (500 MHz).

Step 1	$m_s = 11.47 \text{ mg}$, $m_{IC} = 4.25 \text{ mg}$, $P_{IC} = 100.0\%$
Step 2	$Int_t = 104.78$, $n_t = 1$
Step 3	$Int_{IC} = 1330.45$, $n_{IC} = 6$
Step 4	$MW_t = 509.31 \text{ g/mol}$, $MW_{IC} = 94.13 \text{ g/mol}$
Step 5	$P [\%] = \frac{n_{IC} \cdot Int_t \cdot MW_t \cdot m_{IC}}{n_t \cdot Int_{IC} \cdot MW_{IC} \cdot m_s} \cdot P_{IC} = 94.7 \%$
Note	The integral of target analyte was calculated as the average of signals 3.91, 3.81-3.65, 2.08 and 1.63 ppm.

Compound **40** in D₂O with the addition of dimethyl sulfone (DMSO₂, 100.0%) as internal standard (500 MHz).

	<p>DMSO₂</p> <p><chem>CC(C)(C)P(=O)(=O)SCCO</chem></p>
Step 1	$m_s = 7.05 \text{ mg}$, $m_{IC} = 4.08 \text{ mg}$, $P_{IC} = 100.0\%$
Step 2	$Int_t = 101.80$, $n_t = 1$
Step 3	$Int_{IC} = 1316.75$, $n_{IC} = 6$
Step 4	$MW_t = 350.17 \text{ g/mol}$, $MW_{IC} = 94.13 \text{ g/mol}$
Step 5	$P [\%] = \frac{n_{IC} \cdot Int_t \cdot MW_t \cdot m_{IC}}{n_t \cdot Int_{IC} \cdot MW_{IC} \cdot m_s} \cdot P_{IC} = 99.9 \%$
Note:	The integral of target analyte was calculated as the average of signals at 3.68, 3.13 and 1.56 ppm.

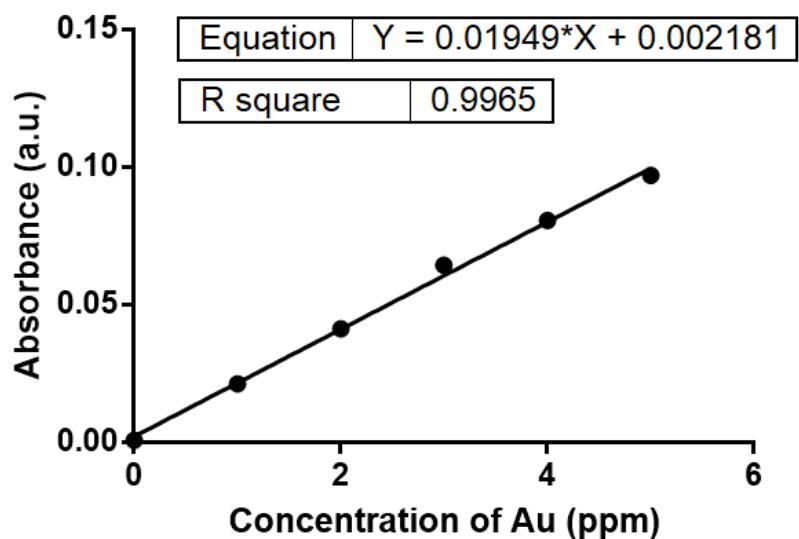


Figure S142. Calibration Curve A of compound **40** in *n*-octanol-saturated water.

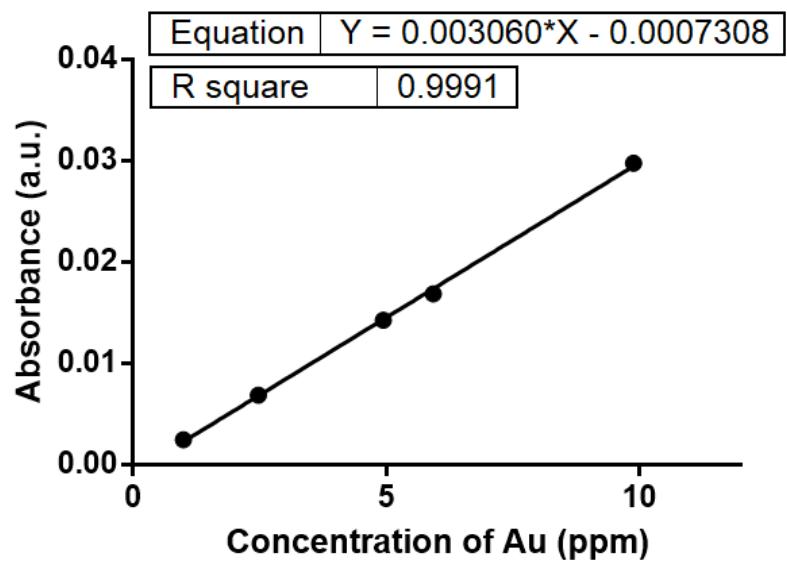


Figure S143. Calibration Curve B of compound **40** in water-saturated *n*-octanol.

Table S1. MIC/MBC ($\mu\text{g}/\text{mL}$)^a of Group 1 analogs having varying thio sugar structures.

	<i>A. baumannii</i>	<i>P. aeruginosa</i>	<i>E. cloacae</i>	<i>K. pneumoniae</i>	<i>S. aureus</i>	<i>E. faecium</i>	<i>E. coli</i>	Log P
	NCTC 13420	NCTC 13437	NCTC 13405	ATCC 700603	JE2 (USA300)	ATCC 700221	ATCC 25922	
1	32 (32)	256 (256)	128 (128)	256 (256)	0.03 (0.06)	0.12/0.06 (0.25)	16 (16)	0.56
2	32 (64)	256 (256)	256 (256)	>256 (>256)	0.03 (0.06)	0.12/0.25 (0.25)	16 (16)	0.38
3	16 (32)	128 (128)	32 (64)	128 (128)	0.03/0.06 (0.06)	0.06 (0.25)	8 (8)	0.26
4 ^f	64 (128)	>256	>256	>256	0.008 (0.01)	0.06 (0.12)	16 (16)	1.01
5 ^f	32 (32)	128 (128)	64 (64)	>256	0.008 (0.06)	0.03/0.06 (0.12)	16 (16)	0.45
6 ^g	64 (128)	128 (128)	256 (256)	>256	0.03 (0.12)	0.12 (0.25)	32 (32)	1.04
7 ^c	16 (16)	>256	32 (32)	>256	0.004/0.008 (0.008)	0.06 (0.12)	32 (32)	-0.28
8 ^f	64/16 (16)	>256	128/256	>256	0.03/0.008 (0.12)	0.03/0.06 (0.25)	128/>256 (128)	-0.36
9	32/8 (128)	256 (256)	64 (64)	256 (256)	0.03/0.06 (0.25)	0.06 (1)	16 (16)	-0.81
10	16 (16)	128 (128)	32 (32)	128 (128)	0.01 (0.06)	0.06/0.12 (0.25)	8 (8)	-0.73
11	16 (32)	256 (256)	64 (128)	256 (256)	0.01/0.03 (0.06)	0.03/0.06 (0.25)	8 (8)	-0.59
12	8/16 (64)	256 (256)	64 (128)	256 (256)	0.008 (0.03)	0.03/0.06 (0.25)	4/2 (4)	-0.89
13	16/32 (128)	256 (256)	128 (128)	256 (256)	0.01/0.03 (0.06)	0.06 (0.25)	16 (16)	-1.81
14	16 (16)	256 (256)	128 (128)	>256 (>256)	0.03/0.06 (0.06)	0.06/0.12 (0.25)	8 (8)	-1.69

^aAssays were repeated twice. Only one value is presented unless both data are shown.

Lowest precipitation concentration at ^b4, ^c8, ^d16, ^e32 ^f64, ^g128 $\mu\text{g}/\text{mL}$.

Table S2. MIC/MBC ($\mu\text{g}/\text{mL}$)^a of Group 2 analogs having an aromatic or aliphatic thiol ligand.

	<i>A. baumannii</i>	<i>P. aeruginosa</i>	<i>E. cloacae</i>	<i>K. pneumoniae</i>	<i>S. aureus</i>	<i>E. faecium</i>	<i>E. coli</i>	Log P
	NCTC 13420	NCTC 13437	NCTC 13405	ATCC 700603	JE2 (USA300)	ATCC 700221	ATCC 25922	
1	32 (32)	256 (256)	128 (128)	256 (256)	0.03 (0.06)	0.12/0.06 (0.25)	16 (16)	0.56
15^f	32/8 (>256)	128 (128)	32 (256)	>256 (>256)	0.008 (0.25)	0.25 (0.5)	64 (64)	2.20
16^g	8/16 (8)	128 (128)	8/32 (16/32)	64 (64)	0.004 (0.25)	0.01 (0.06)	16 (16)	0.61
17^d	8 (8)	64 (64)	16 (16)	16 (16)	0.01/0.03 (0.03)	0.01 (0.06)	4 (4)	1.36
18^f	16 (16)	128 (128)	64 (64)	64 (64)	0.008 (0.12)	0.01/0.06 (0.12)	16 (16)	2.01
19^e	>256	>256	>256	>256	0.0002/0.008 (0.008)	0.03/0.06 (0.25)	>256	>3.28
20^e	256 (>256)	>256	>256	>256	0.01/0.06 (0.12)	0.01 (0.12)	>256	>3.18
21^e	4 (32)	16 (16)	4/2 (32)	8 (8)	0.01 (0.01)	0.06/0.12 (0.25)	4 (4)	1.19
22^d	8 (32)	128 (128)	16 (64)	32 (32)	0.002/0.007 (0.01)	0.06/0.12 (0.25)	32 (32)	1.75
23^g	32/16 (64)	64 (64)	16 (64)	64 (64)	0.004 (0.06)	0.25 (0.5)	32 (32)	2.08

^aAssays were repeated twice. Only one value is presented unless both data are shown.

Lowest precipitation concentration at ^b4, ^c8, ^d16, ^e32 ^f64, ^g128 $\mu\text{g}/\text{mL}$.

Table S3. MIC/MBC ($\mu\text{g/mL}$)^a of Group 3 analogs having a trialkyl- or triaryl-phosphine ligand.

	<i>A. baumannii</i> NCTC 13420	<i>P. aeruginosa</i> NCTC 13437	<i>E. cloacae</i> NCTC 13405	<i>K. pneumoniae</i> ATCC 700603	<i>S. aureus</i> JE2 (USA300)	<i>E. faecium</i> ATCC 700221	<i>E. coli</i> ATCC 25922	Log P
24	4/2 (4)	16 (16)	1 (1)	4/2 (4)	0.03/0.06 (0.5)	0.12 (1)	2 (2)	0.16
25	8/4 (8)	32 (64)	8 (8)	16 (16)	0.004/0.007 (0.06)	0.03/0.06 (0.25)	8 (8)	1.74
26^c	64/32 (64)	32 (32)	>256	64 (64)	1 (2)	2 (2)	>256	>3.99
27^d	64/8 (258)	>256	>256	>256	0.5/1 (2)	2 (2)	>256	>3.94
28^c	>256	>256	>256	>256	1 (2)	2 (2)	>256	>4.09
29^d	32/16 (128)	>256	>256	>256	0.5 (2)	1/2 (2)	>256	>4.04
30^e	>256	>256	>256	>256	2/1 (8)	>256	>256	>3.94
31^g	8/4 (64)	64 (64)	2 (2)	8 (8)	0.06 (0.12)	0.12/0.25 (0.5)	4 (4)	0.35
1	32 (32)	256 (256)	128 (128)	256 (256)	0.03 (0.06)	0.12/0.06 (0.25)	16 (16)	0.56
32^e	64 (>256)	>256	>256	>256	2 (2)	4/2 (2)	>256	>4.32
33^c	>256	>256	>256	>256	1/2 (2)	4 (4)	>256	>3.87
34^c	>256	>256	>256	>256	1 (2)	4/2 (4)	>256	>3.03
35^e	>256	>256	>256	>256	1 (2)	4/2 (4)	>256	>3.04
36^e	>256	>256	>256	>256	4 (8)	32 (64)	>256	>3.55

^aAssays were repeated twice. Only one value is presented unless both data are shown.

Lowest precipitation concentration at ^b4, ^c8, ^d16, ^e32 ^f64, ^g128 $\mu\text{g/mL}$.

Table S4. MIC/MBC ($\mu\text{g/mL}$)^a of analogs having trimethylphosphine ligand.

	<i>A. baumannii</i>	<i>P. aeruginosa</i>	<i>E. cloacae</i>	<i>K. pneumoniae</i>	<i>S. aureus</i>	<i>E. faecium</i>	<i>E. coli</i>	Log P
	NCTC 13420	NCTC 13437	NCTC 13405	ATCC 700603	JE2 (USA300)	ATCC 700221	ATCC 25922	
1	32 (32)	256 (256)	128 (128)	256 (256)	0.03 (0.06)	0.12/0.06 (0.25)	16 (16)	0.56
31^g	8/4 (64)	64 (64)	2 (2)	8 (8)	0.06 (0.12)	0.12/0.25 (0.5)	4 (4)	0.35
37	8/2 (8)	>256	2/4 (8)	16 (16)	0.12/0.25 (0.5)	0.12/0.25 (0.25)	4 (4)	-1.63
38	8/4 (8)	>256 (>256)	2 (2)	16 (16)	0.12 (0.12)	0.12/0.25 (0.25)	4 (4)	-1.88
39	8/4 (8)	>256 (>256)	4/8 (8)	16 (64)	0.25 (0.25)	0.12/0.25 (0.25)	4/16 (16)	-2.03
40^f	2/1 (8)	32/8 (32)	1 (1)	4 (8)	0.12 (0.25)	0.12 (0.12)	0.5/2 (4)	-0.15

^aAssays were repeated twice. Only one value is presented unless both data are shown.

Lowest precipitation concentration at ^b4, ^c8, ^d16, ^e32 ^f64, ^g128 $\mu\text{g/mL}$.

Table S5. IC₅₀ ($\mu\text{g}/\text{mL}$)^a of auranofin and analogs against A549 cells.

1	2	3	4	5	6	7
7.7 \pm 1.3	10.5 \pm 0.2	30.9 \pm 5.8	7.9 \pm 0.1	15.2 \pm 0.3	29.8 \pm 3.2	14.4 \pm 0.0
8	9	10	11	12	13	14
15.8 \pm 0.1	30.2 \pm 0.1	16.9 \pm 0.5	17.2 \pm 1.1	29.2 \pm 4.8	32.9 \pm 4.6	31.0 \pm 2.4
15	16	17	18	19	20	21
4.3 \pm 0.2	7.5 \pm 0.1	6.5 \pm 1.1	7.2 \pm 0.7	5.5 \pm 1.3	3.1 \pm 0.2	7.3 \pm 1.7
22	23	24	25	26	27	28
6.4 \pm 2.4	4.6 \pm 0.2	15.9 \pm 0.7	9.5 \pm 0.1	4.7 \pm 0.0	12.0 \pm 1.8	9.4 \pm 1.7
29	30	31	32	33	34	35
13.2 \pm 1.0	85.6 \pm 26.1	13.1 \pm 2.3	2.5 \pm 0.3	3.1 \pm 0.1	3.8 \pm 0.1	2.9 \pm 0.1
36	37	38	39	40		
18.3 \pm 2.2	52.2 \pm 1.0	43.0 \pm 3.7	35.5 \pm 0.9	12.3 \pm 3.1		

^aData are presented as mean \pm S.E.M. from two independent experiments.