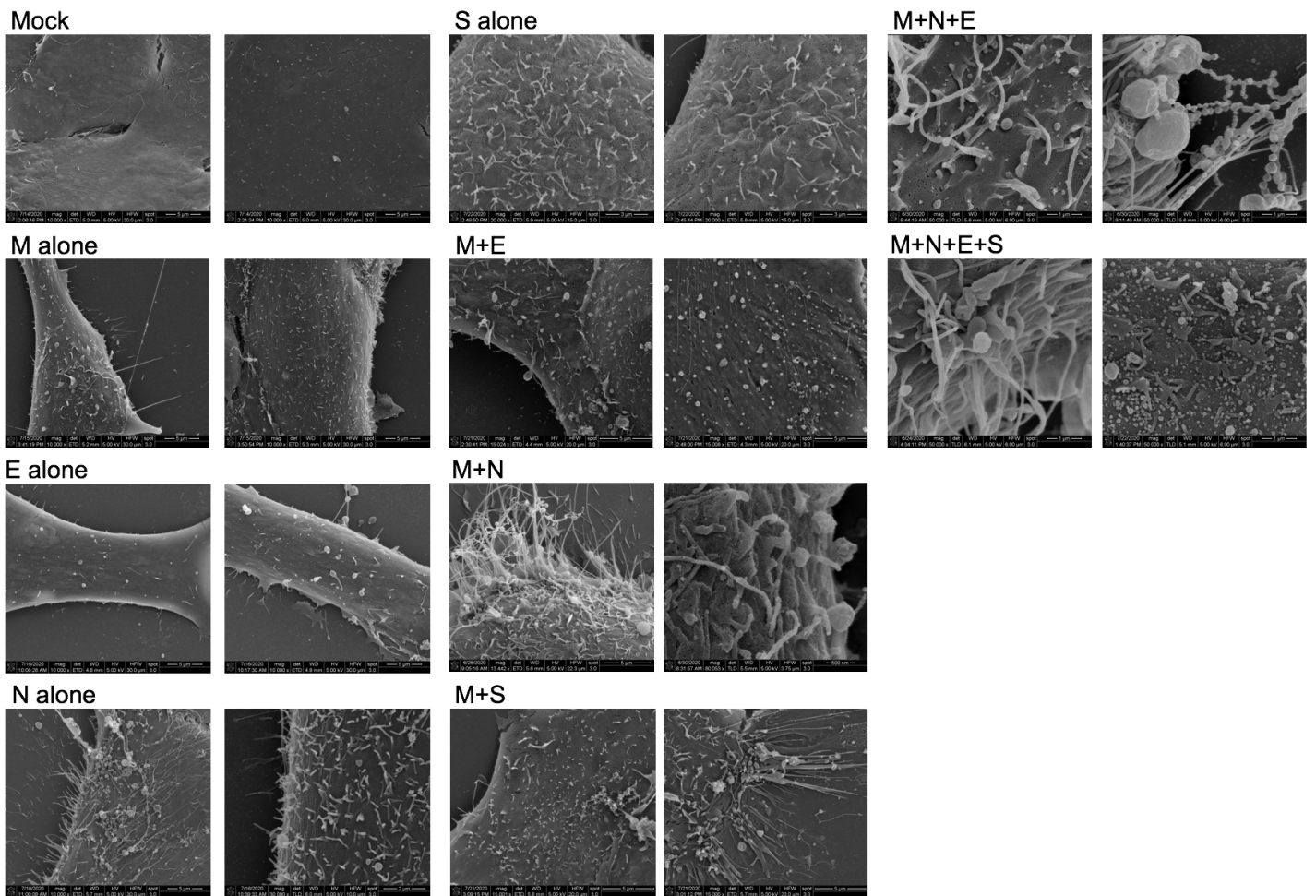


## Supporting Information for

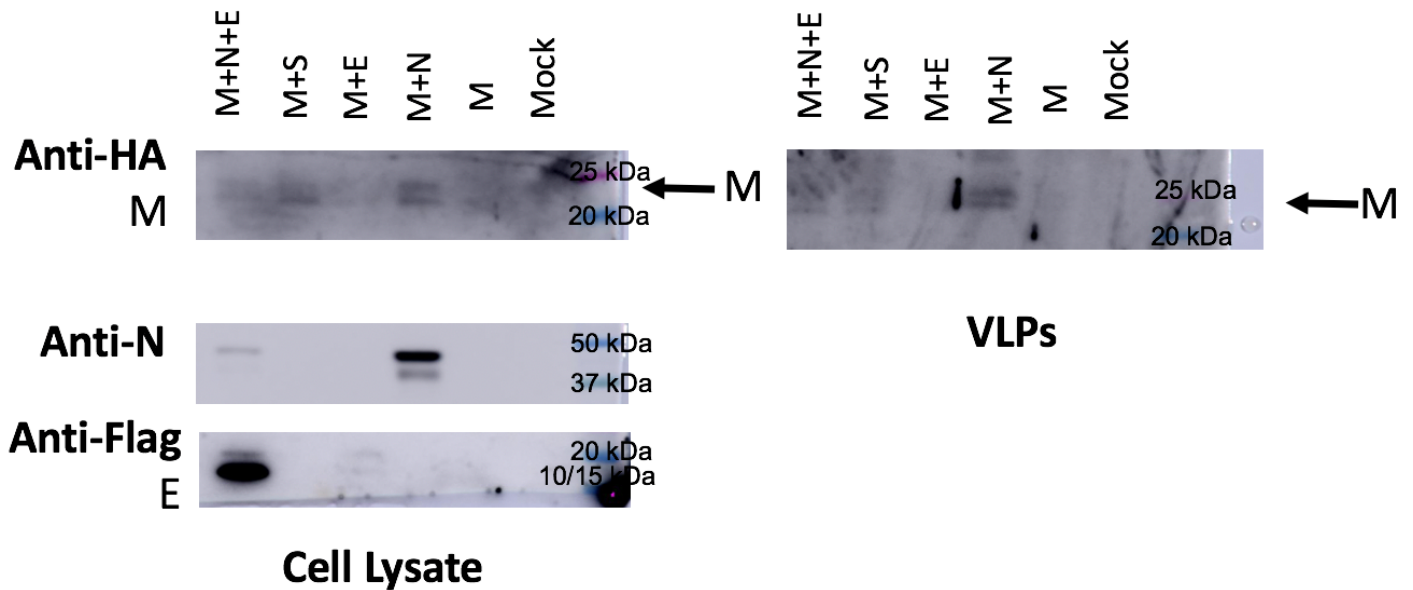
### SARS-CoV-2 viral budding and entry can be modeled using BSL-2 level virus-like particles

Caroline B. Plescia, Emily A. David, Dhableswar Patra, Ranjan Sengupta, Souad Amiar, Yuan Su, and Robert V. Stahelin\*

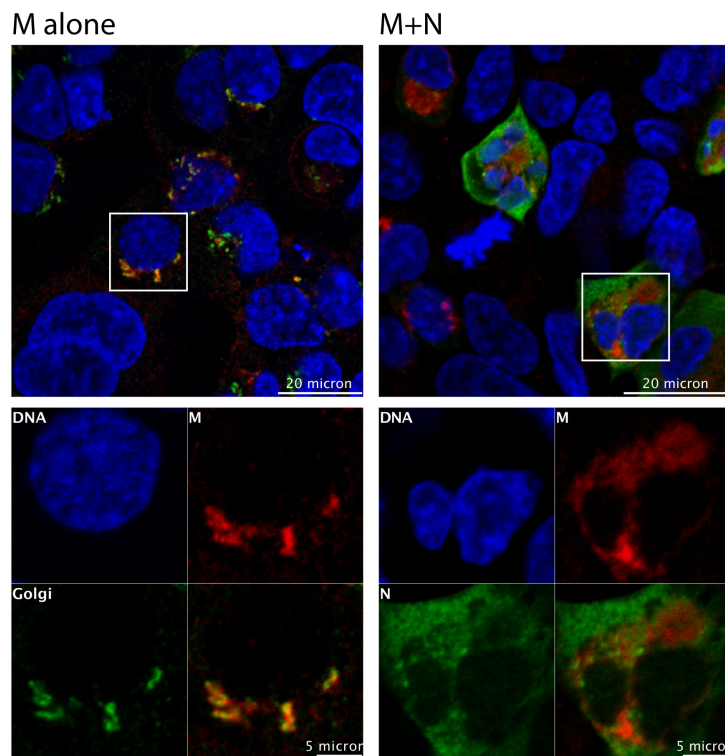
\*Corresponding Author: Robert V. Stahelin  
E-mail: rstaheli@purdue.edu



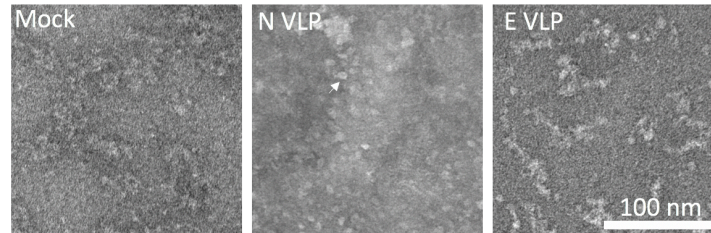
**Figure S1.** Scanning electron microscopy of SARS-CoV-2 viral structural protein expressing cells. HEK293 cells were seeded onto coverslips and transfected individually and in combination with M, N, E, and/or S. Cells were fixed with glutaraldehyde 72-hours post-transfection and kept at 4°C until fixed with osmium tetroxide. Samples were then gradually dehydrated with ethanol and completely dehydrated with a critical point dryer. Once dehydrated, samples were mounted onto aluminum pins with double-sided carbon tape, charged with silver paint, and sputter coated prior to imaging.



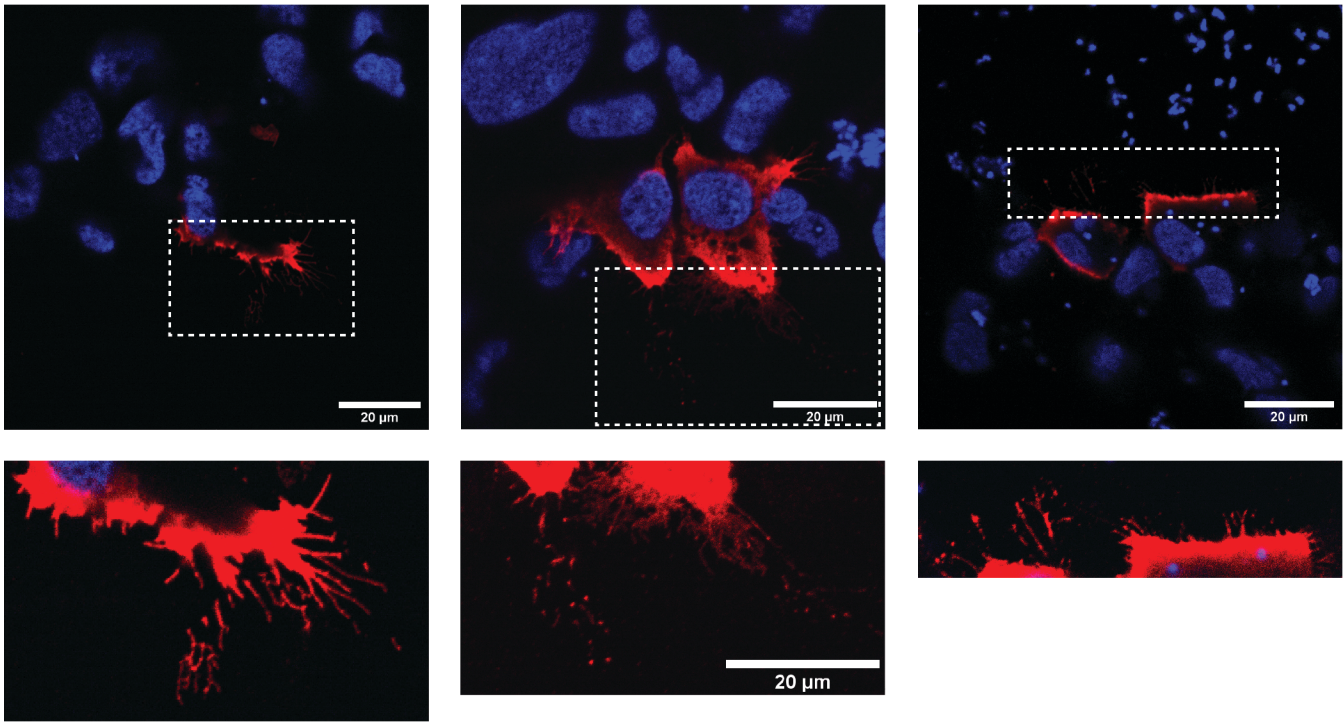
**Figure S2.** Western blot analysis of the cell lysate and VLP fractions of different combinations of M, N, E, and/or S. Total protein content of cell lysates were used to normalize loading conditions and was quantified using the Pierce bicinchoninic acid assay. VLP loading was calculated as a constant ratio to normalized cell lysates.



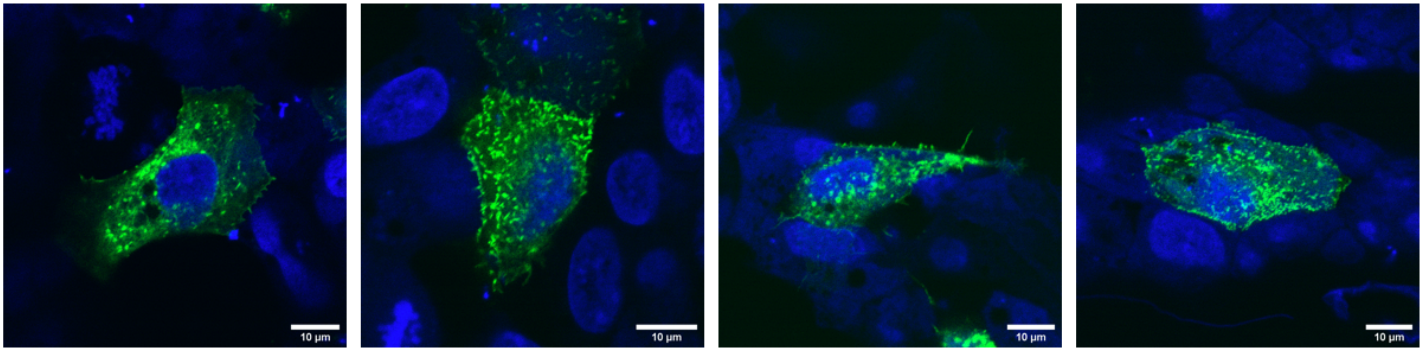
**Figure S3.** Cellular redistribution and accumulation of SARS-CoV-2 M protein in the presence of N protein. Immunofluorescence assays of HEK293 cells expressing M alone or M+N for 24 hours prior to fixation, indicated that M protein expressed alone accumulated at the Golgi apparatus (as indicated in the zoomed left inset). However, when co-expressed with N protein, M protein exhibited more diffuse and perinuclear distribution (as indicated in zoomed right inset).



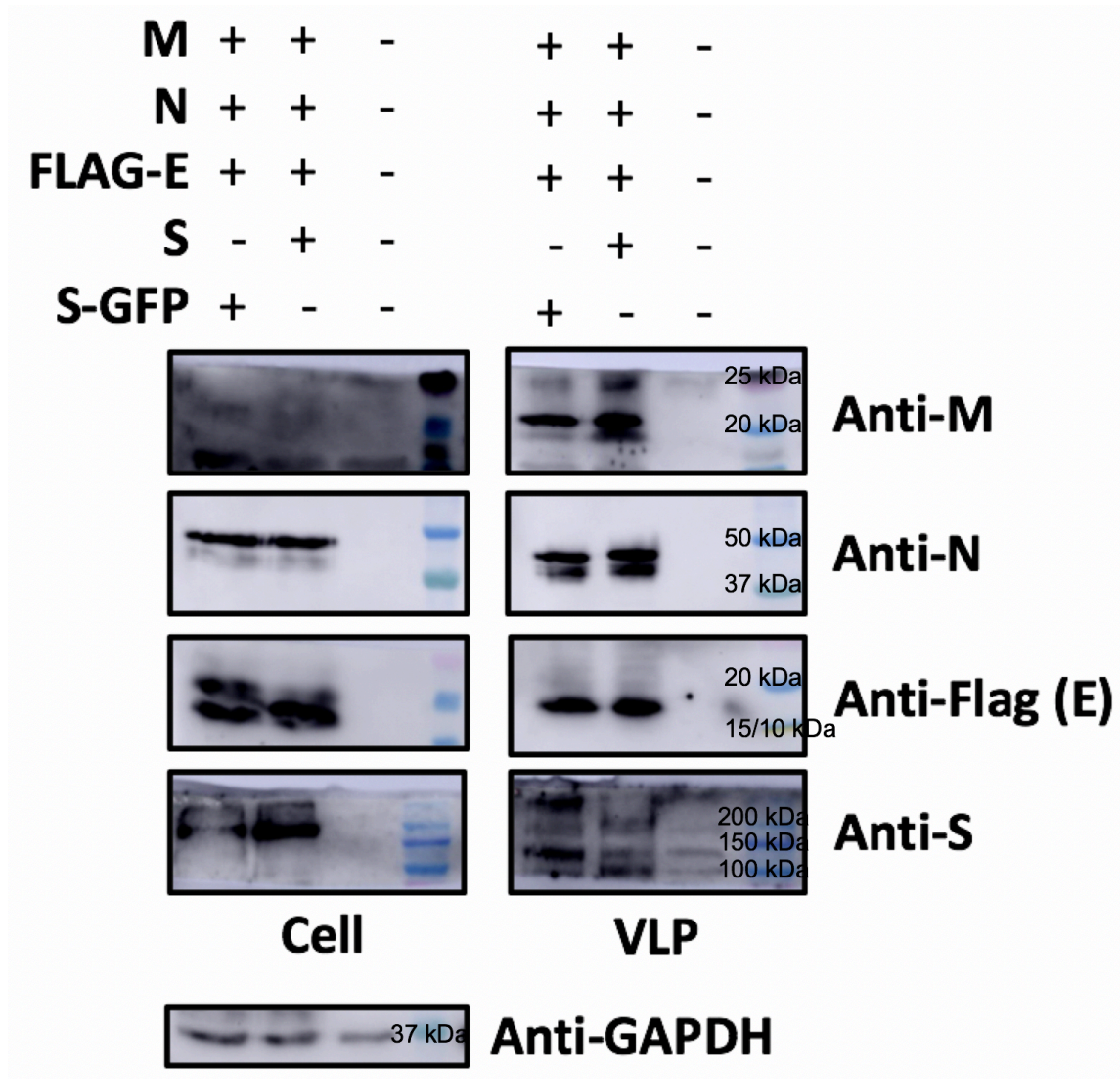
**Figure S4.** TEM analysis of VLP fractions. VLP collections were performed on mock, N, or E-transfected cells. Each VLP fraction was added to a glow discharged 400-mesh copper grid covered with carbon-coated collodion film. Grids were washed in one drop of water, stained in three drops of phosphotungstic acid (1.0% w/v), air dried, and imaged. Arrow in N VLP fraction represents large protein oligomers.



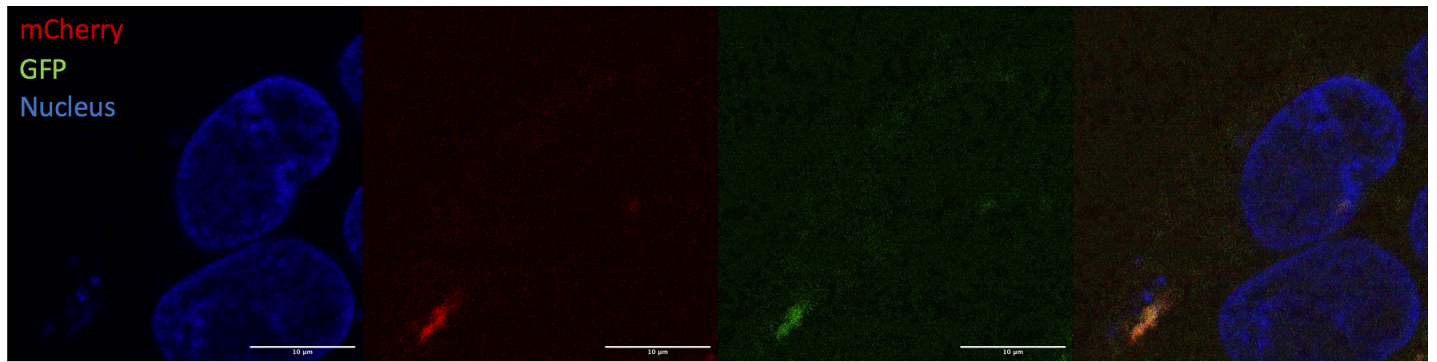
**Figure S5.** Immunofluorescence of SARS-COV-2 N in HEK293 cells. HEK293 cells expressing SARS-COV-2 N protein at 24-hours post-transfection were fixed, stained with nuclear stain (blue, Hoechst 3342) and subject to immunofluorescence assay with anti-SARS-COV-2 N antibody (1:500) followed by anti-rabbit IgG-Atto 594 (1:1000). SARS-COV-2 N protein localized in the filopodia along the cell surface.



**Figure S6:** Expression of S-GFP in HEK293 cells. Fluorescence confocal images of SARS-CoV-2 S protein in HEK293 cells. HEK293 cells transiently expressing SARS-CoV-2 S protein at 24 hours post transfection were fixed, stained with nuclear stain (blue, Hoechst 3342) and imaged with confocal microscopy.



**Figure S7.** Western blot analysis of the cell lysate and VLP fractions of VLP and GFP-producing combinations of M, N, E, and S or S-GFP. Total protein content of cell lysates were used to normalize loading conditions and was quantified using the Pierce bicinchoninic acid assay. VLP loading was calculated as a constant ratio to normalized cell lysates.



**Figure S8.** Infection of target HEK293 cells with media from S-GFP transfected cells. Media was removed from HEK293 cells expressing S-GFP 72-hours post-transfection and was used to infect target HEK293 cells. Target cells were spinoculated, incubated two hours, stained with nuclear stain (Hoechst 3342), and imaged with confocal microscopy.

<u>Plasmid</u>	<u>Construct</u>	<u>Tag</u>	<u>Construct Sequence / Reference</u>
pcDNA3-M	M	No Tag	ATGGCCGACAGCAACGGCACCATCACCGTGGAAGAGCTGAAGAACTGCTGGAACAGTGGAAACC TGGTCATCGGCTTTCTGTTCTGACCTGGATCTGCCTGCTGCAGTTCGCTTATGCCAACCGGAACC GGTTCCTGTACATCATCAAGCTGATCTTCTGTGGCTGCTGTGGCCTGTGACCTGGCTTGTTCCTGT GCTGGCCCGCTTTACCGCATCACTGGATCACAGGCGGAATCGCTATCGCAATGGCCTGCCTGG TGGGCTGTAGTGGCTGTCTACTTTCATCGCCAGCTTCCGGCTGTTTCCAGAAACCAGGAGCATGT GGTCTTCAACCCGTAGACAACATCCTGCTGAACGTGCCCTCCACGGCACAATCCTGACAAAGAC CACTGCTGGAGAGCGAGCTGGTATTGGCCCGGTATCCTTAGAGGCCACCTGAGAATCGCCGG CCACCATCTGGGCAGATGCGATATCAAGGACCTGCCTAAGGAAATACCCGTGGCCACATCCAGAA CCCTGAGCTACTACAACTGGGAGCCTCTCAAGAGTGGCCGGCGATAGCGGCTTCCGGCTTAC AGCAGATACCGGATTGGAATTAAGCTGAATACCGACCACTAGCTCTAGCGACAACATCGCC CTGCTGGTGCACTGA
pcDNA3-HA-M	HA-M	HA	TACCATACGATGTTCCAGATTACGCTGGTACCGGCGGAAGCGCCGACAGCAACGGCACCATCAC CGTGGAAGAGCTGAAGAACTGCTGGAACAGTGGAACTGGTCACTCGGCTTTCTGTTCCTGACCT GGATCTGCCTGCTGCAGTTCGCTTATGCCAACCGGAACCGGTTCTGTACATCATCAAGCTGATCT TCCTGTGGCTGCTGTGGCCTGTGACCCTGGCTTGTTCGTGCTGGCCGCGTTTACCGCATCAACT GGATCACAGGCGGAATCGCTATCGCAATGGCCTGCCTGGTGGCCTGATGTGGCTGTCTACTTC ATCGCCAGCTTCCGGCTGTTTCCAGAACCCAGGAGCATGTGGTCTTCAACCCGTAGACAACATC CTGCTGAACGTGCCCTCCAGGCACAATCTGACAAAGACCACTGCTGGAGAGCGAGCTGGTGTAT TGGCGCGTGTATCCTTAGAGGCCACCTGAGAATCGCCGGCCACCATCTGGGCAGATGCGATATCA AGGACCTGCCTAAGGAAATCACCGTGGCCACATCCAGAACCCGTGAGCTACTACAACTGGGAGCC TCTCAAAGAGTGGCCGGCGATAGCGGCTTCCGGCTTACAGCAGATACCGGATTGGAATTAACA GCTGAATACCGACCACTAGCTCTAGCGACAACATCGCCCTGCTGGTGCACTGA
pcDNA3-N	N	No Tag	ATGTCTGACAACGGCCCTCAGAACCAGCGGAATGCCCAAGAACTCACCTTCCGGCGCCCTCCGA TTCTACAGGCTCCAACCAAGAAATGGAGAGAGGTCCGGAGCACGCTCAAGCAGCGGAGACCACAG GGCCTGCCAACAAATACCGCCAGCTGGTTACCGCCCTGACACAGCACGGCAAGGAGGACCTGA AGTTTTCCAGGGGCGAGGGCGTGCCTATCAACACCAATAGCTCCCTGACGATCAGATCGGCTAC TATAGGAGGGCAACAAGGAGAATCCGGGAGGGCGACGGCAAGATGATGCTGTCCTCCCGCAGA TGGTACTTCTACTATCTGGGCACCGGACCTGAGGCAGGACTGCCATATGGCGCAATAAGGACGG AATCATCTGGTGGCAACCGAGGGCGCCCTGAACACACAAAGGATCAGATCGGCACACGCAAT CCCGCAACAATGCAGCAATCGTGTCCAGCTGCCACAGGGCACCACACTGCCAAGGGCTTTTA CGCAGAGGGCAGCAGGGGCGGCTCCAGGCCTCTAGCCGCTCCTTAGCCGGTCCAGAAAATCC TCTCGGAATTCTACCCAGGAGCTCCCGGGGCACAAGCCCTGCAAGAATGGCAGGAAACGGAG GGCAGCGCCGCTGGCCCTGCTGCTGCTGGATAGACTGAATCAGCTGGAGTCTAAGATGAGCGG CAAGGGACAGCAGCAGCGGACAGACCGGTGACAAAGAGTCTGCCCGGAGGCCAGCAAGAA GCCAAGGCAGAAAGCGCACCGCCACAAGGCCACAACTGACCCAGGCTTCCGGCAGGCGCGG ACCAGAGCAGACACAGGGCAATTTGGCGACCAGGAGCTGATCAGGCAGGGCACCAGATTATAAG CACTGGCCTCAGATCGCACAGTTCGCACCAAGCGCCTCCGCCTTCTTTGGCATGAGCAGGATCGG AATGGAGGTGACCCCATCCGGCACATGGCTGACCTACACAGGCAGCATCAAGCTGGAGCATAAG GACCCCTAATTCAAGGATCAGGTATCCTGCTGAACAAGCAGATCGATGCCTATAAGACCTTCCC CCTACAGAGCCCAAGGAGGACAAGAAAGAGGCGGATGAGACCCAGGCCCTCCCTCAGAGA CAGAAGAAGCAGCAGACCGGTGACACTGCTGCCAGCCCGCACCTGGACGATTTTCCAAGCAGCT CCAGCAGTCTATGTCTAGCGCGATAGCACCAGTGA
pCMV-3xFlag-E	3xFlag-E	3xFlag	ATGGATTACAAGGATGACGACGATAAGGACTATAAGGACGATGATGACAAGGACTACAAGATGA TGACGATAAAGCCCGGGCGGGATCCCCGGGCTGCAGGAATTCGATATCAAGCTTTACTCTTTTGT GAGCGAGGAGACCGGCACACTGATCGTGAACCTCGTGTCTGCTGTTCCGCGCTTTGGTGTTC TGCTGGTACCCTGGCAATCCTGACAGCCCTGAGGCTGTGCGCCTATTGCTGTAACATCGTGAAC GTGAGCCTGGTGAAGCCCTCTTCTACGTGTATAGCCGGGTGAAGAACCTGAATAGCTCCAGAGT GCCTGACCTGCTGGTGTGA
pCAGGS-S	S	No Tag	BEI Resources NR-52310
pcDNA3.1-S-GFP	S-GFP	GFP	Genescript MC_0101089
pcDNA3.1-S-APEX2	S-APEX2	APEX2	S-GFP construct with GFP replaced with APEX2

**Figure S9.** Table of construct sequences used for mammalian expression of SARS-CoV-2 structural proteins.