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Reporting Summary

Nature Research wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Research policies, see our Editorial Policies and the Editorial Policy Checklist.

For all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.

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n/a	Cor	nfirmed
	X	The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement
	X	A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
	×	The statistical test(s) used AND whether they are one- or two-sided Only common tests should be described solely by name; describe more complex techniques in the Methods section.
	×	A description of all covariates tested
	×	A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
	×	A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
	×	For null hypothesis testing, the test statistic (e.g. <i>F</i> , <i>t</i> , <i>r</i>) with confidence intervals, effect sizes, degrees of freedom and <i>P</i> value noted <i>Give P values as exact values whenever suitable.</i>
×		For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
x		For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
	×	Estimates of effect sizes (e.g. Cohen's <i>d</i> , Pearson's <i>r</i>), indicating how they were calculated
,		Our web collection on statistics for biologists contains articles on many of the points above

Software and code

Policy information about <u>availability of computer code</u>

Data collection

No software was used for data collection, only analysis.

Data analysis

The software for the novel analysis in this manuscript is publicly available at: https://github.com/daniel-munro/imageCCA. We used the following Python packages with corresponding version numbers: statsmodels(0.9.0). We used the following R packages with corresponding version numbers: topGO(2.30.1), Bioconductor(3.6), org.Hs.eg.db(3.5.0), GO.db(3.5.0), PMA(1.0.9), MatrixEQTL(2.1.0).

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Research guidelines for submitting code & software for further information.

Data

Policy information about availability of data

All manuscripts must include a <u>data availability statement</u>. This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A list of figures that have associated raw data
- A description of any restrictions on data availability

Tissue slide images for the BRCA and LGG samples were downloaded from the TCGA DataPortal (now available at the GDC Legacy Archive: https://portal.gdc.cancer.gov/legacy-archive/). Details of the data collection and preparation can be found in the original studies 22, 23. Tissue slide images for the GTEx samples were downloaded from the NCI Biospecimen Research Database (https://brd.nci.nih.gov/brd/image-search/searchhome). Genotype and RNA-Seq data are available upon application from dbGAP (https://www.ncbi.nlm.nih.gov/projects/gap/cgi-bin/study.cgi?study_id=phs000424.v7.p2).

Field-specific reporting						
Please select the o	ne below that is the best fit for your research. If you are not sure, read the appropriate sections before making your selection.					
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Sample size	Describe how sample size was determined, detailing any statistical methods used to predetermine sample size OR if no sample-size calculation was performed, describe how sample sizes were chosen and provide a rationale for why these sample sizes are sufficient.					
Data exclusions	Describe any data exclusions. If no data were excluded from the analyses, state so OR if data were excluded, describe the exclusions and the rationale behind them, indicating whether exclusion criteria were pre-established.					
Replication	Describe the measures taken to verify the reproducibility of the experimental findings. If all attempts at replication were successful, confirm this OR if there are any findings that were not replicated or cannot be reproduced, note this and describe why.					
Randomization	Describe how samples/organisms/participants were allocated into experimental groups. If allocation was not random, describe how covariates					

Reporting for specific materials, systems and methods

We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.

Describe whether the investigators were blinded to group allocation during data collection and/or analysis. If blinding was not possible,

Materials & experimental systems		Methods		
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X Ar	tibodies	×	ChIP-seq	
x Eu	karyotic cell lines	×	Flow cytometry	
🗶 🗌 Pa	laeontology and archaeology	x	MRI-based neuroimaging	
X Ar	nimals and other organisms			
X Hu	ıman research participants			
X Cli	nical data			
X Dr	ual use research of concern			

were controlled OR if this is not relevant to your study, explain why.

describe why OR explain why blinding was not relevant to your study.

Blinding